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Further evidence of association of *ABCA4* gene with cleft lip/palate

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Abstract

Nonsyndromic cleft lip with or without cleft palate (CL/P) is a common birth defect with complex etiology. Numerous genes and environmental factors and their interactions are thought to play a role in the susceptibility to CL/P. A recent genome-wide association study with several populations revealed markers in/near *MAFB* and *ABCA4* genes as new susceptibility loci for CL/P. We hypothesized that these genes could also contribute to CL/P in a Brazilian population, hence we evaluated if the associated SNPs in *MAFB* [rs13041247 and rs11696257] and *ABCA4* [rs560426 and rs481931] were associated with CL/P in our case-control dataset. We genotyped 812 Caucasian individuals (400 cases and 412 controls) from Brazil. Allele frequencies were compared for cases and controls as well as for cleft subgroups and controls. *ABCA4* rs540426 showed strong association with CL/P, unilateral and right CL/P, and bilateral CL/P, whereas SNP rs481931 showed borderline association with CL/P, and bilateral CL/P. No association was found for *MAFB*. Our results support a potential role for *ABCA4* in the etiology of CL/P in individuals from Brazil.

Keywords

cleft lip/cleft palate; association; *ABCA4* gene; SNP

Craniofacial anomalies, and in particular oral-facial clefts, are major human birth defects with a worldwide frequency of 1 in 700 live births and substantial clinical impact. The possible etiologies are many, including single-gene disorders, chromosome aberrations, exposure to teratogens, and sporadic conditions of unknown cause (1). Oral-facial clefts can be further classified as nonsyndromic (isolated) or syndromic based on the presence of other structural anomalies. Approximately 30% of all clefts are associated with one of more than 400 described syndromes (2) while the remaining 70% are isolated defects. It is generally accepted that cleft lip with or without cleft palate (CL/P) and cleft palate only (CPO) are

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CONFLICTS OF INTEREST

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genetically distinct phenotypes. CL/P is more common, affecting 1–2/1000 births and presenting considerable differences in prevalence, with Native Americans and Asians showing the highest rate and Africans the lowest. On the other hand, CPO is less common, with a prevalence of approximately 1/1500–2000 births in Caucasians, less variable among different ethnic backgrounds (3). These observations suggest that the relative contribution of individual susceptibility genes may vary across different populations, thus reinforcing the need of replication of association studies in different populations. Further, numerous lines of evidence now suggest that the phenotypic spectrum of nonsyndromic CL/P is more complex than previously realized and therefore genetic studies should include a more accurate description of the cleft phenotype, such as cleft type and laterality, as well as presence of subclinical phenotypes, such as defects in the orbicularis oris muscle and dental anomalies (4,5).

Several loci and genes - including, but not limited to, *MSX1*, *IRF6*, *CRISPLD2*, *FOXE1*, *AXIN2*, and members of the WNT, FGF, and MMP gene families - have been associated with oral clefts (6–17). Additionally, recent advances in research methodologies have accelerated the discovery of loci conferring susceptibility to isolated CL/P through the use of genome-wide association studies (GWAS). The first three GWAS found strong evidence for association of an intergenic marker (rs987525) in the 8q24 chromosomal region with CL/P (18–20), and this association has been independently validated in additional populations, including a population from Brazil (21). Recently, a third GWAS identified associations with markers in/nearby *ABCA4* and *MAFB* genes, located on chromosomes 1p22.1 and 20q11.1-q13.1, respectively, with CL/P in multiple populations (22). In two subsequent studies, the originally associated SNP in *ABCA4* (rs540026) was associated with increased risk of CL/P in US and South American populations (23, 24) whereas a SNP in *MAFB* (rs13041247) was associated with increased risk of CL/P in Chinese (25). Intriguingly, a study with a Nigerian population did not find evidence of association for either *ABCA4* or *MAFB* genes with CL/P (26).

Due to allelic heterogeneity among populations, in order to validate the findings of genetic association studies, it is necessary to independently attempt to replicate these findings in multiple populations. Hence, to further investigate a possible role for *ABCA4* and *MAFB* in the susceptibility to CL/P in a population from Brazil, we tested the previously associated SNPs in *ABCA4* (rs560426 and rs481931), and *MAFB* (rs13041247 and rs11696257) genes (Table 1) for association with CL/P in our case-control dataset.

MATERIALS AND METHODS

Subjects

A convenience sample consisting of 812 unrelated Caucasian individuals was included in this study. Of these, cases comprised 400 individuals (average age 17.3 yr, 252 males, 148 females) with isolated CL/P: 246 with unilateral CL/P and 154 cases with bilateral CL/P whereas 412 individuals without CL/P or family history of CL/P (average age 24.8 years, 165 males, 247 females) served as controls. Individuals were recruited at the Hospital of Rehabilitation and Craniofacial Anomalies and Bauru Dental School at the University of São Paulo, Brazil, after signing an informed consent. All individuals were from the

Southeast region of Brazil, who are mostly European descendants from Portugal and Spain and have been suggested as bearing ~99% Caucasian ethnicity (27). Only cases with CL/P were recruited for the study, regardless of cleft side. Cases with cleft palate alone were not included. This study was approved by the University of Sao Paulo and University of Texas Health Science Center Institutional Review Boards.

Sample collection and genotyping

Saliva samples were collected from each individual as source of genomic DNA using Oragene kits (DNA Genotek, Ontario, CA). Genotyping was performed using Taqman chemistry (28) in 5uL reactions and detected on a 7900HT Sequence Detection Instrument (Applied Biosystems, Foster City, CA, USA). Assays and reagents were supplied by Applied Biosystems (Applied Biosystems). For quality control purposes, negative control reactions were performed using no nucleic acid template; positive control reactions included samples of known genotypes. Genotyping was performed blind to sample status.

Statistical Analyses

Power calculations were performed using the Genetic Power Calculator (29) and indicate that the sample size would provide approximately 80% statistical power to detect an association with an alpha of 0.05, if the markers selected are in linkage disequilibrium with the causal factor ($D'=0.8$) and their frequencies are around 20%.

Statistical analyses were performed using PLINK software (v.1.06) (30). For each SNP, We tested for deviation from Hardy-Weinberg equilibrium in cases and controls using a Pearson's chi-square test. Association analyses were performed comparing differences in genotype and allele frequencies for each SNP between CL/P cases and controls, and between unilateral and bilateral CL/P cases and controls. We applied Bonferroni correction for multiple testing considering the number of tests and variables (0.05/7) and $P < 0.007$ was considered statistically significant.

RESULTS and DISCUSSION

There was no evidence of deviation from Hardy-Weinberg equilibrium for any of the SNPs investigated (data not shown). The results of the association analyses are summarized in Tables 2 and 3. We found evidence of genotypic and allelic association for the SNPs in *ABCA4* and CL/P. SNP rs560426 showed association with CL/P ($P=0.0002$ for genotype, $P=0.00007$ for allele), particularly bilateral ($P=0.0006$ for genotype, $P=0.001$ for allele) and also unilateral ($P=0.004$ for genotype, $P=0.001$ for allele) CL/P. Additional associations were also found for *ABCA4* SNP rs481931 alleles and bilateral ($P=0.006$) and unilateral ($P=0.009$) CL/P. We did not find association of *MAFB* SNPs and CL/P in our population (Tables 2 and 3).

Much progress has been made in the identification of putative candidate genes for CL/P. Recent genome-wide association studies (GWAS) have identified several novel loci associated with CL/P (18–20, 22). Nonetheless, to validate the findings of these studies and understand the overall impact of these results in the general population, it is necessary to independently replicate these findings in multiple populations. Our replication study

suggests a role for *ABCA4* and not *MAFB* in this Brazilian population of Caucasian ethnicity. Similarly, YUAN et al. (23) found evidence of association of *ABCA4* with CL/P, whereas *MAFB* presented only nominal association values in their study with non-Hispanic white and Hispanic CL/P families. Moreover, both *ABCA4* SNPs rs560426 and rs481931 showed association in the Hispanics, although the association in the non-hispanic whites was stronger for SNP rs481931.

The *ABCA4* gene is located on chromosome 1p22.1 and belongs to a superfamily of transmembrane proteins expressed exclusively in retinal photoreceptors (31, 32). So far, there is little biological evidence to support a role for *ABCA4* in craniofacial morphogenesis, particularly since *Abca4* null mice do not exhibit cleft palate (22, 33). It is possible that the associated variants in *ABCA4* are in linkage disequilibrium with a causal variant located in another gene, and act as indirect surrogates for a true etiologic variant in CL/P cases. Nevertheless, the previous associations of *ABCA4* with CL/P in both GWAS (22, 33) and association studies in different populations (23, 24), including the population of the present study, warrants additional investigations on the possible role for this gene in the etiology of CL/P.

In summary, our results provide additional evidence for the association of *ABCA4* with CL/P in a population from Brazil. Given the genetic heterogeneity across populations, it is important to investigate the association of previously reported genes with CL/P in multiple populations.

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REFERENCES

1. Murray JC. Gene/environment causes of cleft lip and/or palate. *Clin Genet.* 2002; 61:248–256. [PubMed: 12030886]
2. Gorlin, RJ.; Cohen, MM.; Hennekam, RCM. *Syndromes of the head and neck.* New York: Oxford University Press; 2001.
3. Mossey, PA.; Little, J. Epidemiology of oral clefts: an international perspective. In: Wyszynski, DF., editor. *Cleft lip and palate: from origin to treatment.* New York, NY: Oxford University Press; 2002. p. 127-158.
4. Weinberg SM, Neiswanger K, Martin RA, Mooney MP, Kane AA, Wenger SL, Losee J, Deleyiannis F, Ma L, De Salamanca JE, Czeizel AE, Marazita ML. The Pittsburgh Oral-Facial Cleft study: expanding the cleft phenotype. Background and justification. *Cleft Palate Craniofac J.* 2006; 43:7–20. [PubMed: 16405378]
5. Letra A, Menezes R, Granjeiro JM, Vieira AR. Defining subphenotypes for oral clefts based on dental development. *J Dent Res.* 2007; 86:986–991. [PubMed: 17890676]
6. Jezewski PA, Vieira AR, Nishimura C, Ludwig B, Johnson M, O'Brien SE, Daack-Hirsch S, Schultz RE, Weber A, Nepomucena B, Romitti PA, Christensen K, Orioli IM, Castilla EE, Machida J, Natsume N, Murray JC. Complete sequencing shows a role for *MSX1* in nonsyndromic cleft lip and palate. *J Med Genet.* 2003; 40:399–407. [PubMed: 12807959]
7. Zucchero TM, Cooper ME, Maher BS, Daack-Hirsch S, Nepomuceno B, Ribeiro L, Caprau D, Christensen K, Suzuki Y, Machida J, Natsume N, Yoshiura K, Vieira AR, Orioli IM, Castilla EE, Moreno L, Arcos-Burgos M, Lidral AC, Field LL, Liu YE, Ray A, Goldstein TH, Schultz RE, Shi

- M, Johnson MK, Kondo S, Schutte BC, Marazita ML, Murray JC. Interferon regulatory factor 6 (IRF6) gene variants and the risk of isolated cleft lip or palate. *N Engl J Med*. 2004; 351:769–780. [PubMed: 15317890]
8. Chiquet BT, Lidral AC, Stal S, Mulliken JB, Moreno LM, Arcos-Burgos M, Valencia-Ramirez C, Blanton SH, Hecht JT. CRISPLD2: a novel NSCLP candidate gene. *Hum Mol Genet*. 2007; 16:2241–2248. [PubMed: 17616516]
 9. Chiquet BT, Blanton SH, Burt A, Ma D, Stal S, Mulliken JB, Hecht JT. Variation in WNT genes is associated with non-syndromic cleft lip with or without cleft palate. *Hum Mol Genet*. 2008; 17:2212–2218. [PubMed: 18413325]
 10. Riley BM, Mansilla MA, Ma J, Daack-Hirsch S, Maher BS, Raffensperger LM, Russo ET, Vieira AR, DodÉ C, Mohammadi M, Marazita ML, Murray JC. Impaired FGF signaling contributes to cleft lip and palate. *Proc Natl Acad Sci USA*. 2007; 104:4512–4517. [PubMed: 17360555]
 11. Menezes R, Marazita ML, Goldstein Mchenry T, Cooper ME, Bardi K, Brandon C, Letra A, Martin RA, Vieira AR. AXIS inhibition protein 2 orofacial clefts and a family history of cancer. *J Am Dent Assoc*. 2009; 140:80–84. [PubMed: 19119171]
 12. Menezes R, Letra A, Kim AH, KÜCHLER EC, Day A, Tannure PN, Gomes DA, Motta L, Paiva KB, Granjeiro JM, Vieira AR. Studies with Wnt genes and nonsyndromic cleft lip and palate. *Birth Defects Res A Clin Mol Teratol*. 2010; 88:995–1000. [PubMed: 20890934]
 13. Moreno LM, Mansilla MA, Bullard SA, Cooper ME, Busch TD, Machida J, Johnson MK, Brauer D, Krahn K, Daack-Hirsch S, L'Heureux J, Valencia-Ramirez C, Rivera D, LÓpez AM, Moreno MA, Hing A, Lammer EJ, Jones M, Christensen K, Lie RT, Jugessur A, Wilcox AJ, Chines P, Pugh E, Doheny K, Arcos-Burgos M, Marazita ML, Murray JC, Lidral AC. *FOXE1* association with both isolated cleft lip with or without cleft palate and isolated cleft palate. *Hum Mol Genet*. 2009; 18:4879–4896. [PubMed: 19779022]
 14. Letra A, Menezes R, Govil M, Fonseca RF, Mchenry T, Granjeiro JM, Castilla EE, Orioli IM, Marazita ML, Vieira AR. Follow-up association studies of chromosome region 9q and nonsyndromic cleft lip/palate. *Am J Med Genet A*. 2010; 152A:1701–1710. [PubMed: 20583170]
 15. Letra A, Menezes R, Fonseca RF, Govil M, Mchenry T, Murphy MJ, Hennebold JD, Granjeiro JM, Castilla EE, Orioli IM, Martin R, Marazita ML, Bjork BC, Vieira AR. Novel cleft susceptibility genes in chromosome 6q. *J Dent Res*. 2010; 89:927–932. [PubMed: 20511563]
 16. Letra A, Bjork B, Cooper ME, Szabo-Rogers H, Deleyiannis FW, Field LL, Czeizel AE, Ma L, Garlet GP, Poletta FA, Mereb JC, Lopez-Camelo JS, Castilla EE, Orioli IM, Wendell S, Blanton SH, Liu K, Hecht JT, Marazita ML, Vieira AR, Silva RM. Association of *AXIN2* with non-syndromic oral clefts in multiple populations. *J Dent Res*. 2012; 91:473–478. [PubMed: 22370446]
 17. Letra A, Silva RM, Motta LG, Blanton SH, Hecht JT, Granjeiro JM, Vieira AR. Association of MMP3 and TIMP2 promoter polymorphisms with nonsyndromic oral clefts. *Birth Defects Res A Clin Mol Teratol*. 2012; 94:540–548. [PubMed: 22730240]
 18. Birnbaum S, Ludwig KU, Reutter H, Herms S, Steffens M, Rubini M, Baluardo C, Ferrian M, Almeida DE, Assis N, Alblas MA, Barth S, Freudenberg J, Lauster C, Schmidt G, Scheer M, Braumann B, BergÉ SJ, Reich RH, Schiefke F, Hemprich A, PÖTzsch S, Steegers-Theunissen RP, PÖTzsch B, Moebus S, Horsthemke B, Kramer FJ, Wienker TF, Mossey PA, Propping P, Cichon S, Hoffmann P, Knapp M, NÖThen MM, Mangold E. Key susceptibility locus for nonsyndromic cleft lip with or without cleft palate on chromosome 8q24. *Nat Genet*. 2009; 41:473–477. [PubMed: 19270707]
 19. Grant SF, Wang K, Zhang H, Glaberson W, Annaiah K, Kim CE, Bradfield JP, Glessner JT, Thomas KA, Garris M, Frackelton EC, Otieno FG, Chiavacci RM, Nah HD, Kirschner RE, Hakonarson H. A genome-wide association study identifies a locus for nonsyndromic cleft lip with or without cleft palate on 8q24. *J Pediatr*. 2009; 155:909–913. [PubMed: 19656524]
 20. Mangold E, Ludwig KU, Birnbaum S, Baluardo C, Ferrian M, Herms S, Reutter H, De Assis NA, Chawa TA, Mattheisen M, Steffens M, Barth S, Kluck N, Paul A, Becker J, Lauster C, Schmidt G, Braumann B, Scheer M, Reich RH, Hemprich A, PÖTzsch S, Blaumeiser B, Moebus S, Krawczak M, Schreiber S, Meitinger T, Wichmann HE, Steegers-Theunissen RP, Kramer FJ, Cichon S, Propping P, Wienker TF, Knapp M, Rubini M, Mossey PA, Hoffmann P, NÖThen MM. Genome-

wide association study identifies two susceptibility loci for nonsyndromic cleft lip with or without cleft palate. *Nat Genet.* 2010; 42:24–26. [PubMed: 20023658]

21. Brito LA, Paranaiba LM, Bassi CF, Masotti C, Malcher C, Schlesinger D, Rocha KM, Cruz LA, BARbara LK, Alonso N, Franco D, Bagordakis E, Martelli H Jr, Meyer D, Coletta RD, Passos-Bueno MR. Region 8q24 is a susceptibility locus for nonsyndromic oral clefting in Brazil. *Birth Defects Res A Clin Mol Teratol.* 2012; 94:464–468. [PubMed: 22511506]
22. Beaty TH, Murray JC, Marazita ML, Munger RG, Ruczinski I, Hetmanski JB, Liang KY, Wu T, Murray T, Fallin MD, Redett RA, Raymond G, Schwender H, Jin SC, Cooper ME, Dunnwald M, Mansilla MA, Leslie E, Bullard S, Lidral AC, Moreno LM, Menezes R, Vieira AR, Petrin A, Wilcox AJ, Lie RT, Jabs EW, Wu-Chou YH, Chen PK, Wang H, Ye X, Huang S, Yeow V, Chong SS, Jee SH, Shi B, Christensen K, Melbye M, Doheny KF, Pugh EW, Ling H, Castilla EE, Czeizel AE, Ma L, Field LL, Brody L, Pangilinan F, Mills JL, Molloy AM, Kirke PN, Scott JM, Arcos-Burgos M, Scott AF. A genome-wide association study of cleft lip with and without cleft palate identifies risk variants near MAFB and ABCA4. *Nat Genet.* 2010; 42:525–529. [PubMed: 20436469]
23. Yuan Q, Blanton SH, Hecht JT. Association of ABCA4 and MAFB with non-syndromic cleft lip with or without cleft palate. *Am J Med Genet A.* 2011; 155A:1469–1471. [PubMed: 21567910]
24. Lennon CJ, Birkeland AC, NuEz JA, Su GH, Lanzano P, Guzman E, Celis K, Eisig SB, Hoffman D, Rendon MT, Ostos H, Chung WK, Haddad J Jr. Association of candidate genes with nonsyndromic clefts in Honduran and Colombian populations. *Laryngoscope.* 2012 Jul 2. [Epub ahead of print].
25. Pan Y, Zhang W, Du Y, Tong N, Han Y, Zhang H, Wang M, Ma J, Wan L, Wang L. Different roles of two novel susceptibility loci for nonsyndromic orofacial clefts in a Chinese Han population. *Am J Med Genet A.* 2011; 155A:2180–2185. [PubMed: 21834038]
26. Butali A, Mossey PA, Adeyemo WL, Jezewski PA, Onwuamah CK, Ogunlewe MO, Ugboko VI, Adejuyigbe O, Adigun AI, Abdur-Rahman LO, Onah II, Audu RA, Idigbe EO, Mansilla MA, Dragan EA, Petrin AL, Bullard SA, Uduezue AO, Akpata O, Osaguona AO, Olosoji HO, Ligali TO, Kejeh BM, Iseh KR, Olaitan PB, Adebola AR, Efunkeye E, Adesina OA, Oluwatosin OM, Murray JC. Genetic studies in the nigerian population implicate an MSX1 mutation in complex oral facial clefting disorders. *Cleft Palate Craniofac J.* 2011; 48:646–653. [PubMed: 21740177]
27. Pena SD, Di Pietro G, Fuchshuber-Moraes M, Genro JP, Hutz MH, Kehdy FDES, Kohlrausch F, Magno LA, Montenegro RC, Moraes MO, De Moraes ME, De Moraes MR, Ojopi EB, Perini JA, Racciopi C, Ribeiro-Dos-Santos AK, Rios-Santos F, Romano-Silva MA, Sortica VA, Suarez-Kurtz G. The genomic ancestry of individuals from different geographical regions of Brazil is more uniform than expected. *PLoS One.* 2011; 6:e17063. [PubMed: 21359226]
28. Ranade K, Chang MS, Ting CT, Pei D, Hsiao CF, Olivier M, Pesich R, Hebert J, Chen YD, Dzau VJ, Curb D, Olshen R, Risch N, Cox DR, Botstein D. High-throughput genotyping with single nucleotide polymorphisms. *Genome Res.* 2001; 11:1262–1268. [PubMed: 11435409]
29. Purcell S, Cherny SS, Sham PC. Genetic power calculator: design of linkage and association genetic mapping of complex traits. *Bioinformatics.* 19:149–150. [PubMed: 12499305]
30. Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MA, Bender D, Bender D, Maller J, Sklar P, De Bakker PI, Daly MJ, Sham PC. PLINK: a toolset for whole-genome association and population-based linkage analysis. *Am J Hum Genet.* 2007; 81:559–575. [PubMed: 17701901]
31. Sun H, Molday RS, Nathans J. Retinal stimulates ATP hydrolysis by purified and reconstituted ABCR, the photoreceptor-specific ATP-binding cassette transporter responsible for Stargardt disease. *J Biol Chem.* 1999; 274:8269–8281. [PubMed: 10075733]
32. Cideciyan AV, Swider M, Aleman TS, Tsybovsky Y, Schwartz SB, Windsor EAM, Roman AJ, Sumaroka A, Steinberg JD, Jacobson SG, Stone EM, Palczewski K. ABCA4 disease progression and a proposed strategy for gene therapy. *Hum Molec Genet.* 2009; 18:931–941. [PubMed: 19074458]
33. Weng J, Mata NL, Azarian SM, Tzekov RT, Birch DG, Travis GH. Insights into the function of Rim protein in photoreceptors and etiology of Stargardt's disease from the phenotype in *Abcr* knockout mice. *Cell.* 1999; 98:13–23. [PubMed: 10412977]

Table 1

Details of the SNPs investigated in this study.

Gene	Chromosome	SNP	SNP Function	Alleles ^a
<i>ABC4</i>	1p22	rs560426	intron	c/T
		rs481931	intron	G/t
<i>MAFB</i>	20q11.2-q13.1	rs13041247	intergenic	c/T
		rs11696257	intergenic	C/t

^aMinor allele indicated as lower case, according to NCBI dbSNP.

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Table 2

Results of genotypic association analysis between single nucleotide polymorphisms (SNPs) in *ABCA4* and *MAFB* genes and nonsyndromic cleft lip/palate (CL/P).

Gene	SNP	CL/P (n=400)			P- value ^b	CL/P Unilateral (n=246)			P- value ^b	CL/P Bilateral (n=154)			P- value ^b
		Genotype ^a				Genotype ^a				Genotype ^a			
		AA	Aa	aa		AA	Aa	aa		AA	Aa	aa	
<i>ABCA4</i>	rs560426	116/74	118/203	86/123	0.0002	70/74	118/203	55/123	0.004	45/74	54/203	31/123	0.0006
	rs481931	184/154	155/192	44/57	0.02	113/154	104/192	29/57	0.15	71/154	49/192	14/57	0.05
<i>MAFB</i>	rs13041247	182/166	165/180	38/42	0.45	110/166	108/180	25/42	0.83	13/42	55/180	71/166	0.24
	rs11696257	185/181	163/185	32/41	0.44	115/181	107/185	22/41	0.78	69/181	54/185	10/41	0.29

^a Order of genotypes, AA/Aa/aa where a is the minor allele

^b Fisher exact test, significant if P < 0.05. Undetermined genotypes not included in analysis.

Table 3
Results of allelic association analysis between single nucleotide polymorphisms (SNPs) in *ABCA4* and *MAFB* genes and nonsyndromic cleft lip/palate (CL/P).

Gene	SNP		CL/P		P-value ^b	CL/P Unilateral		P-value ^b	CL/P Bilateral		P-value ^b
			Allele ^a			Allele ^a			Allele ^a		
			A	a		A	a		A	a	
<i>ABCA4</i>	rs560426	Case/control	406/351	346/449	0.00007	258/351	228/449	0.001	144/351	116/449	0.001
	rs481931	Case/control	523/500	243/306	0.06	330/500	162/306	0.009	191/500	77/306	0.006
<i>MAFB</i>	rs13041247	Case/control	529/512	241/264	0.25	328/512	158/264	0.58	197/512	81/264	0.14
	rs11696257	Case/control	533/547	227/267	0.21	337/547	151/267	0.48	192/547	74/267	0.13

^a Order of alleles, a where a is the minor allele

^b Fisher exact test, significant if P < 0.05.