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Plasmodium knowlesi – an emerging pathogen

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Ten years have passed since the publication of a large focus of Plasmodium knowlesi infections in the human population. The discovery was made during a molecular investigation of atypical P. malariae cases in the Kapit Health Division, Sarawak, Malaysian Borneo. Patients were more symptomatic with higher parasite counts than expected in P. malariae infections. The investigation found only P. knowlesi DNA present in patient blood samples. Morphological similarity had allowed P. knowlesi to masquerade as P. malariae during routine diagnostic microscopy for malaria. P. knowlesi, a malaria parasite of macaque monkeys, had entered the human population. The subsequent development of P. knowlesi species-specific PCR assays soon demonstrated that the entry was not confined to the Kapit Division but extended across island and mainland Southeast Asia. Relevant clinical descriptions and guidelines for the treatment and management of patents with P. knowlesi malaria were not available. Nor was it clear whether P. knowlesi had undergone a host switch event into the human population or if infections were zoonotic. The outputs of studies on P. knowlesi malaria during the past 10 years will be summarized, highlighting major findings within the context of pathophysiology, virulence, host switch events, treatment, control and importantly malaria elimination.

Key words: Plasmodium, knowlesi, malaria, pathophysiology, virulence, zoonosis

Introduction

Malaria parasites, member species of the genus *Plasmodium*, are blood-borne and pose a substantial risk in blood transfusion related infection [1–5]. Donor blood infected with any of the human-host-adapted *Plasmodium* species, *P. falciparum*, *P. vivax*, *P. ovale* and *P. malariae* or newly emergent species such as *P. knowlesi* would cause malaria in the untreated non-immune recipient. Most medical texts and guidelines for malaria provide detailed information on the human–host-adapted malaria parasites but not on emergent species. *P. knowlesi*, a parasite of the long and pig-tailed macaques of Southeast Asia, is not vertebrate host restricted, and humans, under

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experimental conditions, were permissive to infection [6]. Clinical information collected from therapeutically induced P. knowlesi infections in patients with tertiary syphilis provides information on human-parasite interaction using blood passage of laboratory lines of P. knowlesi [6]. However, naturally acquired P. knowlesi malaria was not recognized and therefore clinically overlooked, until a large focus of P. knowlesi was described in the human population in Sarawak, Malaysian Borneo, in 2004 [7]. P. knowlesi is morphologically indistinguishable from P. malariae, one of the more benign human-host-adapted Plasmodium species, and infections were misdiagnosed until the recent development of P. knowlesi-specific PCR-based assays [7-9]. The use of these assays indicate that zoonotic malaria, caused by P. knowlesi, is widespread in Southeast Asia and, in stark contrast to P. malariae, is not benign [10-13].

Plasmodium knowlesi pathophysiology

Serial blood passage of a limited number of experimental lines of *P. knowlesi* in neurosyphilis patients, pre-antibiotic pyretic treatment in the 1950s, was associated with

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increased parasite virulence [6]. The next account of P. knowlesi pathophysiology came with the first documented case of naturally acquired P. knowlesi published in 1965 [14]. The case was diagnosed retrospectively and, while the patient was symptomatic, he recovered fully following treatment and no other cases were confirmed. In fact, when we published the large focus of P. knowlesi in patients with single-species P. knowlesi infections in Kapit, Sarawak, we stated that P. knowlesi caused the usual symptoms of malaria; all patients responded to treatment and no deaths were reported [7]. This changed during a subsequent study when four P. knowlesi deaths were confirmed retrospectively. Only one of these was reported as a malaria death, P. falciparum, the others were attributed to other causes [11]. There was no information on severe knowlesi malaria in the literature or medical texts to guide healthcare professionals towards correct diagnosis and treatment of these patients. P. knowlesi malaria was new to the medical field, clinicians and other healthcare professionals were unaware

that they were dealing with a newly described virulent form of malaria that, in practice, was most often misdiagnosed as the more benign P. malariae. A prospective clinical study on PCR-confirmed single-species P. knowlesi infected patients confirmed the need for knowlesi-specific treatment and management guidelines [12]. In that study 107 patients had P. knowlesi malaria, ten had complications and two died [12]. Complications included >100 000 parasites/ μ l blood, acute renal impairment, jaundice, hypotension, lactate acidosis, acute and late onset respiratory distress syndrome. All of these complications are included in the WHO guidelines for severe falciparum malaria in the non-immune [15]. Notably in this and subsequent studies, severe malaria with coma was not described in severe P. knowlesi infections [11, 12, 16, 17]. Severe malarial anaemia was reported in children but not adults and was not as severe as in the P. falciparum comparator group [18]. In a recent study, 232 adult patients with PCRconfirmed single-species P. knowlesi infections were recruited in Sarawak Malaysian Borneo [19]. Twenty-eight

Table 1 Summary of patients with complicated *Plasmodium knowlesi* malaria (n = 28)

Parasites >100,000 /uL blood	T. bilirubin 43-100 umol/ul	T. bilirubin >100 umol/L	Serum creatinine >265 umol/L	Plasma lactate >6.0 mmol/L	SOB/ ARDs	SystolicB P <80 mmHg	Metab. acidosis	Other	n =
•••		•••							3
•••	•••		•••						3
••		••	••						2
••			••						1;1
•			•			•			1
•	•								1
•			•	•					1
•									1
				•			•		1
•	•				•				1
	•		•						1
		•	•						1
•			•	•	•				1
			•						1
	•	••••							5
								•••	3
								Total	28

Red = fatal cases. Others = Low Sodium, co-morbidity with parasitaemia >100,000 parasites/uL, patient ill and referred.

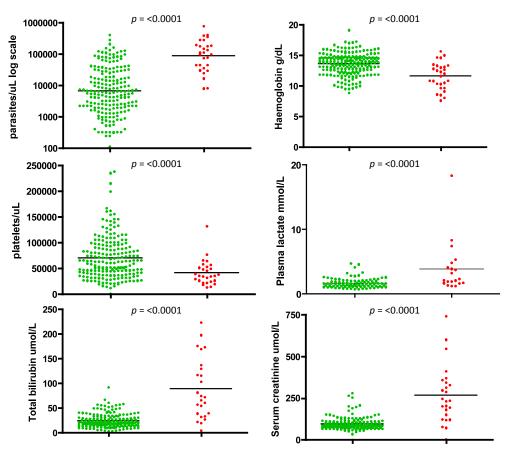


Fig. 1 Complicated vs. uncomplicated *Plasmodium knowlesi* disease groups. Patients with *P. knowlesi* malaria grouped by uncomplicated disease [U = Green] and complicated disease [C = Red]. Plasma lactate results from a subset of patients (U = 91, C = 20, n = 111). P values were calculated using the Mann–Whitney *U*-test for nonparametric data and the unpaired *t*-test for normally distributed data (Prism 4 for Macintosh; GraphPad Software, Inc. San Diego).

(12%) of these had complications and four (1.7%) patients died, Table 1. Parasitaemia, total bilirubin, serum creatinine and plasma lactate were significantly higher in the complicated group, Fig. 1. Parasitaemia was not used as a criterion to score severe disease. The study of P. knowlesi malaria is relatively new, and markers of severe disease are preliminary but suggest that patients with parasitaemia $>35~000~parasites/\mu l$, bilirubin $>43~\mu mol/l$, serum creatinine >256 μ mol/l or platelets <45 000/ μ l are at risk [12, 19]. P. knowlesi is unusual among the parasites that infect human and non-human primates because the asexual part of the life cycle, the erythrocytic cycle associated with clinical signs and symptoms, takes 24 h to complete rather than 48 and 72 h in other types of malaria. Therefore, parasitaemia increases daily in P. knowlesi infections, increased parasitaemia is associated with disease severity, and while multiplication rates are probably variant specific and the number of merozoites produced will need to be factored in, patients with suspected P. knowlesi infection require urgent diagnosis and rapid access to optimal treatment.

Virulence

Plasmodium knowlesi isolates collected from human infections are genetically diverse [7, 9]. Parasitaemia is associated with disease severity and we asked the question are some P. knowlesi variants more virulent than others? We sequenced two P. knowlesi genes responsible for parasite invasion of erythrocytes and found two genetically distinct clusters of one of the P. knowlesi genes encoding erythrocyte invasion proteins, P. knowlesi normocyte binding protein xa (Pknbpxa) [19, 21]. We found that 44% and 56% of patients were infected with Pknbpxa cluster 1 and Pknbpxa cluster 2, respectively. The clusters contained single nucleotide polymorphisms that allowed for differentiation of haplotype groups with 2 or three alleles among the P. knowlesi isolates. Analysis of the alleles with the clinical and laboratory variables in the patient cohort found that patients infected with particular allelic forms of cluster 2 had increased markers of disease severity including parasitaemia [19]. Not all patients with complications were

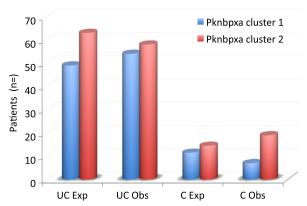


Fig. 2 The number of expected and observed cases of Plasmodium knowlesi malaria infected with parasites with Plasmodium knowlesi normocyte binding protein (pknbp) xa cluster 1 or cluster 2 parasites. Pknbpxa haplotypes were generated for 138 patient isolates in a subset of 147 patients. In the study, 44% of patients were infected with Pknbpxa cluster 1 type parasites and 56% with Pknbpxa cluster 2 type parasites. The patients were then grouped into uncomplicated (UC) n = 112 and complicated (C) n = 28 groups. The expected (Exp) number of patients in the UC and C groups was calculated based on the 44:56 ration of cluster 1 to cluster infections and compared to the number observed (Obs). More patients with complicated disease (C Obs) were infected with Pknbpxa cluster 2 parasites. Chi-square P = 0.048; with Yates correction P = 0.08 (http://vassarstats.net/tab 2x2.html).

infected with cluster 2 parasites but there were more patients than expected with complications in this cluster (Fig. 2). This suggests that, in nature, some P. knowlesi variants may be more virulent in the human-host population than others. This study was conducted in Sarawak, Malaysian Borneo, and may explain some of the apparent differential susceptibility of humans to P. knowlesi infection in Malaysia and other areas within the region. More work is required to determine genetic diversity of P. knowlesi across the Southeast Asian region and associated parasite virulence.

Host switch events

Plasmodium knowlesi infections occur throughout the Southeast Asian region. The region is separated by high mountain ranges and large expanses of water including the South China Sea, the Sulu Sea and Andaman Sea [21]. P. knowlesi is a parasite of non-human primates that do not cross oceans and rural human populations at risk tend to be local. It seems likely that, following the primary emergence of P. knowlesi and ancient dispersal events, P. knowlesi, populations have evolved in local geographical isolation. Each location reporting human infections is most likely dealing with different genetic variants of P. knowlesl [22]. The recent discovery of P. knowlesi malaria in humans was in Kapit, Malaysian Borneo, and further study found that naturally acquired P. knowlesi infections occurred throughout Malaysian Borneo, Peninsular Malaysia, all geographically distinct areas, and subsequently in most parts of Southeast Asia [7, 11, 21, 23]. Therefore, it is important not to extrapolate studies on genetic diversity and virulence between geographically distinct transmission sites.

Even so the multiple entry events recorded across Southeast Asia may have been the result of parasite adaptation to the human host and clonal expansion but the infections are not clonal or related to any one index case or location [7]. A study on P. knowlesi isolated from the natural macaque hosts and human infections in Sarawak did not identify genetic clustering of parasites to either host, strongly suggesting a zoonotic origin for the infections in Sarawak [9]. However, children in Sabah were infected without any obvious risk behaviour for acquiring a zoonotic infection, such as spending time in the forest [23]. Also, and perhaps most worrying, Anopheles balabacensis is a vector of P. knowlesi in Sabah [16]. An balabacensis is better known as a vector of P. falciparum in Sabah and therefore well placed to channel a host switch event for *P. knowlesi* from macaque-to-human to human-to-human malaria transmission at this location [24, 25]. In Vietnam, another geographically distant and distinct location, children have asymptomatic low-level P. knowlesi infections mixed with P. falciparum and P. vivax [26]. Anopheles dirus is the main malaria vector in this region of Vietnam and transmits all three parasite species, P. knowlesi, P. falciparum and P. vivax, concurrently [26, 27]. P. knowlesi warrants careful observation to pre-empt or at least contain a host switch event in any number of Southeast Asian locations quite a monumental task.

Treatment

Earlier studies reported that P. knowlesi responds well to currently available antimalarial compounds including chloroquine [7, 12, 28]. The level of sensitivity supports an absence of drug selection pressure implying a recent emergence or zoonotic origin for P. knowlesi. Studies designed to properly assess clinical and parasitological responses of P. knowlesi to treatment highlight some important differences in response. In 2005, inherent mefloquine resistance of P. knowlesi in experimentally infected Rhesus macaques was reported [29]. Also, P. knowlesi isolates from human infections responded poorly to mefloquine ex vivo [30]. Furthermore, the parasitaemia in a patient, who presented with severe P. knowlesi malaria during a second infection, increased following mefloquine treatment [31]. Nonetheless, other patients with uncomplicated P. knowlesi infections responded to mefloquine treatment [32, 33].

Plasmodium knowlesi responds to chloroquine albeit with a higher IC_{50} ex vivo [30]. Chloroquine should be used with caution because in the absence of species-specific diagnostic tests in most regions, the risk of misdiagnosing *P. falciparum*, mostly chloroquine resistant, as *P. knowlesi* would put patients at risk.

The most compelling evidence supports the use of artemisinin derivatives either as intravenous artesunate or as oral combination therapies to treat complicated and uncomplicated P. knowlesi infections. Patients in Sabah with severe knowlesi malaria were more likely to survive when given intravenous artesunate than quinine [28]. In another study in Sabah patients with all cause, malaria were given oral artemisinin combination therapy or artesunate IV immediately on diagnosis. Most of the patients had knowlesi malaria, 39% had severe symptoms and all survived [16]. Both studies report shorter parasite clearance times for the artemisinin based therapies than chloroquine or quinine. Also P. knowlesi responded better to artemisinin derivatives than all other compounds ex vivo [30]. In other studies, IV artesunate performed better than quinine in treating severe falciparum malaria [34]. It should be noted that patients with P. knowlesi infections can be symptomatic at low parasitaemia, lower than the sensitivity of thick film microscopy [12]. Currently, there is no specific or sensitive diagnostic test for P. knowlesi malaria [35].

Control of zoonotic malaria

Human cases of P. knowlesi malaria occur across Southeast Asia [10]. This region is known for human, animal, plant, insect, geographical and geological diversity [36]. Apart from parts of Malaysia and Vietnam, there is little information on P. knowlesi transmission, reservoirs of infection, vectors and risk of human infection [27, 37, 38]. Without this information, it will be difficult to design control measures for zoonotic P. knowlesi acquired in the jungle setting, over and beyond use of prophylaxis when spending time in areas where other patients with P. knowlesi have been infected. The pragmatic solution would be to encourage location-specific vector, parasite and natural host monitoring and surveillance. These measures are currently costly, require manpower and are logistically difficult. Perhaps a meeting of minds is required to develop alternative sampling and monitoring methods for forest settings that would capture the required data on parasite prevalence, vector capacity, natural hosts but using minimal manpower. Not an easy problem to solve on the backdrop of competing public health problems at each location. In areas where P. knowlesi is transmitted in human settlements or even human-to-human, existing malaria control measures would be expected to be effective.

Malaria elimination

According to the World Health Organization 'Malaria elimination is the interruption of local mosquito-borne malaria transmission, i.e. the reduction to zero of the incidence of malaria infection in a defined geographical area.' [39]. Malaysia has an excellent record in malaria control; however, Malaysia and other countries in Southeast Asia, pursuing malaria elimination, need to carefully consider P. knowlesi. There is some evidence that P. knowlesi enters human populations where national human-host-adapted, malaria control initiatives are successful, a good example of this is Malaysia [13, 40]. This poses two questions: (i) Will removal of the human-hostadapted species open a new host niche for P. knowlesi? or (ii) By removing human host challenge, particularly with P. vivax, are humans more susceptible to P. knowlesi through loss of cross protection?

Both scenarios are worth taking seriously. The relatively large number of *P. knowlesi* cases was discovered in Malaysia [7]. The study was prompted by the report of atypical cases of *P. malariae* in the area. It is possible that these cases were more visible as cases of *P. falciparum* and *P. vivax* were controlled. Or were there more of these cases as immunity to malaria declined along with *P. vivax* and *P. falciparum*? Returning to *P. knowlesi* induced pyretic therapy in tertiary syphilis. Patients who were previously treated with *P. vivax* were partially resistant to subsequent *P. knowlesi* infection suggesting a degree cross immunity [6].

There is some debate about the prevalence of zoonotic malaria – Is *P. knowlesi* being better recognized because of heightened awareness and PCR identification? If so, then the prevalence of *P. knowlesi* may not have increased, we are simply looking harder, difficult to argue against. However, the number of cases of *P. knowlesi* in Sabah do seem to be increasing [13]. Regardless, awareness brings the opportunity to deal with a large entry and host switch events that might threaten malaria elimination in the area.

If the ultimate goal is malaria eradication, the WHO definition is much more specific 'Malaria *eradication* is defined as the permanent reduction to zero of the world-wide incidence of malaria infection caused by a particular malaria parasite species.' [39]. If eradication is the goal, then it would be possible to eradicate *P. falciparum*, *P. vivax*, *P. malariae* and *P. ovale* while still tolerating *P. knowlesi* in Southeast Asia.

Summary

Plasmodium knowlesi has entered the human population in Southeast Asia. People who spend time the sylvan

transmission sites are at risk. P. knowlesi remains largely zoonotic, but transmission near human settlements is suspected in Sabah. P. knowlesi is diverse and causes severe disease in at least 10% of those infected in Sabah and Sarawak, Patients with severe and fatal P. knowlesi infections fulfil the WHO criteria for severe falciparum malaria but without coma or severe adult anaemia. Healthcare professionals should suspect P. knowlesi in patients and blood donors with a history of time spent in the jungles of Southeast Asia. Blood donor screening protocols, to detect malaria exposure, use recombinant P. falciparum and P. vivax antigen. While there may be some cross reactivity between P. knowlesi and the phylogenetically related P. vivax, these reagents could not be used reliably to detect antibodies to P. knowlesi. At the moment, the risk of donor blood infection with P. knowlesi would be expected to be low; however, a history of time spent in the jungles of Southeast Asia may warrant donor referral for a 1-year period.

Importantly, evidence suggests that rapid diagnosis and immediate treatment with artemisinin derivatives is appropriate for this potentially virulent parasite with a short erythrocytic life cycle.

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Conflicting interests

The authors have no conflicts to report.

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References

- 1 Freimanis G, Sedegah M, Owusu-Ofori S, et al.: Investigating the prevalence of transfusion transmission of Plasmodium within a hyperendemic blood donation system. Transfusion 2013; 53:1429-1441
- 2 Owusu-Ofori AK, Bates I: Impact of inconsistent policies for transfusion-transmitted malaria on clinical practice in Ghana. PLoS ONE 2012; 7:e34201
- 3 Noubouossie D, Tagny CT, Same-Ekobo A, et al.: Asymptomatic carriage of malaria parasites in blood donors in Yaounde. Transfus Med 2012; 22:63-67
- 4 Tagny CT, Owusu-Ofori S, Mbanya D, et al.: The blood donor in sub-Saharan Africa: a review. *Transfus Med* 2010; **20**:1–10
- 5 Owusu-Ofori AK, Parry C, Bates I: Transfusion-transmitted malaria in countries where malaria is endemic: a review of

- the literature from sub-Saharan Africa. Clin Infect Dis 2010; 51:1192-1198
- 6 Coatney GR, Collins WE, Warren M, et al.: Plasmodium knowlesi; in Coatney GR (ed.): The Primate Malarias. Washington, DC, USA, Government Printing Office, 1971:317-
- 7 Singh B, Kim Sung L, Matusop A, et al.: A large focus of naturally acquired Plasmodium knowlesi infections in human beings. Lancet 2004; 363:1017-1024
- 8 Divis PC, Shokoples SE, Singh B, et al.: A TaqMan real-time PCR assay for the detection and quantitation of Plasmodium knowlesi. Malar J 2010; 9:344
- 9 Lee KS, Divis PC, Zakaria SK, et al.: Plasmodium knowlesi: reservoir hosts and tracking the emergence in humans and macaques. PLoS Pathog 2011; 7:e1002015
- 10 Cox-Singh J: Zoonotic malaria: Plasmodium knowlesi, an emerging pathogen. Curr Opin Infect Dis 2012; 25:530-536
- 11 Cox-Singh J, Davis TM, Lee KS, et al.: Plasmodium knowlesi malaria in humans is widely distributed and potentially life threatening. Clin Infect Dis 2008; 46:165-171
- 12 Daneshvar C, Davis TM, Cox-Singh J, et al.: Clinical and laboratory features of human Plasmodium knowlesi infection. Clin Infect Dis 2009; 49:852-860
- 13 William T, Rahman HA, Jelip J, et al.: Increasing incidence of Plasmodium knowlesi malaria following control of P. falciparum and P. vivax Malaria in Sabah, Malaysia. PLoS Negl Trop Dis 2013: 7:e2026
- 14 Chin W, Contacos PG, Coatney GR, et al.: A naturally acquited quotidian-type malaria in man transferable to monkeys. Science 1965; 149:865
- 15 2013: World Health Organization: Management of Severe malaria - A Practical Handbook, 3rd edn. http://www.who.int/ malaria/publications/atoz/9789241548526/en/ [last accessed 15th September 2014].
- 16 Barber BE, William T, Grigg MJ, et al.: A prospective comparative study of knowlesi, falciparum, and vivax malaria in Sabah, Malaysia: high proportion with severe disease from Plasmodium knowlesi and Plasmodium vivax but no mortality with early referral and artesunate therapy. Clin Infect Dis 2013; 56:383-397
- 17 Cox-Singh J, Hiu J, Lucas SB, et al.: Severe malaria a case of fatal Plasmodium knowlesi infection with post-mortem findings: a case report. Malar J 2010; 9:10
- 18 Barber BE, William T, Jikal M, et al.: Plasmodium knowlesi malaria in children. Emerg Infect Dis 2011; 17:814-820
- 19 Ahmed MA, Pinheiro MM, Divis PC, Siner A, Zainudin R, et al.: Disease progression in Plasmodium knowlesi malaria is linked to variation in invasion gene family members. PLoS neglected tropical diseases 2014; 8: e3086.
- 20 Willmann M, Ahmed A, Siner A, et al.: Laboratory markers of disease severity in Plasmodium knowlesi infection: a casecontrol study. Malar J 2012; 11:363
- 21 Meyer EV, Semenya AA, Okenu DM, et al.: The reticulocyte binding-like proteins of P. knowlesi locate to the micronemes of merozoites and define two new members of this invasion ligand family. Mol Biochem Parasitol 2009; 165: 111 - 121

- 22 Moyes CL, Henry AJ, Golding N, et al.: Defining the geographical range of the Plasmodium knowlesi reservoir. PLoS Negl Trop Dis 2014; 8:e2780
- 23 Silva JC, Egan A, Friedman R, et al.: Genome sequences reveal divergence times of malaria parasite lineages. Parasitology 2011; 138:1737–1749
- 24 Barber BE, William T, Dhararaj P, et al.: Epidemiology of Plasmodium knowlesi malaria in north-east Sabah, Malaysia: family clusters and wide age distribution. Malar J 2012; 11:401
- 25 Hii JL, Chew M, Sang VY, et al.: Population genetic analysis of host seeking and resting behaviors in the malaria vector, Anopheles balabacensis (Diptera: Culicidae). J Med Entomol 1991; 28:675–684
- 26 Vythilingam I, Chan ST, Shanmugratnam C, et al.: The impact of development and malaria control activities on its vectors in the Kinabatangan area of Sabah, East Malaysia. Acta Trop 2005; 96:24–30
- 27 Marchand RP, Culleton R, Maeno Y, et al.: Co-infections of Plasmodium knowlesi, P. falciparum, and P. vivax among Humans and Anopheles dirus Mosquitoes, Southern Vietnam. Emerg Infect Dis 2011; 17:1232–1239
- 28 Nakazawa S, Marchand RP, Quang NT, et al.: Anopheles dirus co-infection with human and monkey malaria parasites in Vietnam. Int J Parasitol 2009; 39:1533–1537
- 29 William T, Menon J, Rajahram G, et al.: Severe Plasmodium knowlesi malaria in a tertiary care hospital, Sabah, Malaysia. Emerg Infect Dis 2011; 17:1248–1255
- 30 Tripathi R, Awasthi A, Dutta GP: Mefloquine resistance reversal action of ketoconazole a cytochrome P450 inhibitor, against mefloquine-resistant malaria. *Parasitology* 2005; 130:475–479
- 31 Fatih FA, Staines HM, Siner A, et al.: Susceptibility of human Plasmodium knowlesi infections to anti-malarials. Malar J 2013; 12:425

- 32 Lau YL, Tan LH, Chin LC, et al.: Plasmodium knowlesi reinfection in human. Emerg Infect Dis 2011; 17:1314–1315
- 33 Bronner U, Divis PC, Farnert A, et al.: Swedish traveller with Plasmodium knowlesi malaria after visiting Malaysian Borneo. Malar J 2009; 8:15
- 34 Tanizaki R, Ujiie M, Kato Y, et al.: First case of Plasmodium knowlesi infection in a Japanese traveller returning from Malaysia. Malar J 2013; 12:128
- 35 Dondorp A, Nosten F, Stepniewska K, *et al.*: Artesunate versus quinine for treatment of severe falciparum malaria: a randomised trial. *Lancet* 2005; 366:717–725
- 36 Foster D, Cox-Singh J, Mohamad DS, *et al.*: Evaluation of three rapid diagnostic tests for the detection of human infections with *Plasmodium knowlesi*. *Malar J* 2014; 13:60
- 37 Woodruff DS: Biogeography and conservation in Southeast Asia: how 2.7 million years of repeated environmental fluctuations affect today's patterns and the future of the remaining refugial-phase biodiversity. *Biodivers Conserv* 2010; 19:919– 941
- 38 Tan CH, Vythilingam I, Matusop A, et al.: Bionomics of Anopheles latens in Kapit, Sarawak, Malaysian Borneo in relation to the transmission of zoonotic simian malaria parasite *Plasmodium knowlesi*. Malar J 2008; 7:52
- 39 Vythilingam I: *Plasmodium knowlesi* in humans: a review on the role of its vectors in Malaysia. *Trop Biomed* 2010; 27:1–12
- 40 WHO: Malaria. World Health Organization. Overview of malaria elimination, 2014. http://www.who.int/malaria/areas/ elimination/overview/en/ [last accessed 15th September 2014].
- 41 Braima KA, Sum JS, Ghazali AR, et al.: Is there a risk of suburban transmission of malaria in Selangor, Malaysia? PLoS ONE 2013; 8:e77924