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Oral human papillomavirus is common in individuals with Fanconi anemia

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Abstract

Background—Fanconi Anemia (FA) is a rare genetic disorder resulting in a loss of function of the FA-related DNA repair pathway. Individuals with FA are predisposed to some cancers including oropharyngeal and gynecological cancers with known associations with human papillomavirus (HPV) in the general population. Since individuals with FA respond poorly to chemotherapy and radiation, prevention of cancer is critical.

Methods—To determine if individuals with FA are particularly susceptible to oral HPV infection, we analyzed survey-based risk factor data and tested DNA isolated from oral rinses from 126 individuals with FA and 162 unaffected first-degree family members for 37 HPV types.

Results—Fourteen individuals (11.1%) with FA tested positive, significantly more ($p=0.003$) than family members (2.5%). While HPV prevalence was even higher for sexually active individuals with FA (17.7% vs. 2.4% in family; $p=0.003$), HPV positivity also tended to be higher in the sexually inactive (8.7% in FA vs. 2.9% in siblings). Indeed, having FA increased HPV positivity 4.9 fold (95% CI: 1.6–15.4) considering age and sexual experience, but did not differ by other potential risk factors.

Conclusion—Our studies suggest that oral HPV is more common in individuals with FA. It will be essential to continue to explore associations between risk factors and immune dysfunction on HPV incidence and persistence over time.

Impact—HPV vaccination should be emphasized in those with FA as a first step to prevent oropharyngeal cancers, although additional studies are needed to determine if the level of protection it offers in this population is adequate.

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Keywords

Fanconi anemia; oral human papillomavirus; head and neck squamous cell carcinoma; oropharyngeal cancer; HPV vaccine

Introduction

Fanconi anemia (FA) is an inherited autosomal recessive or rare X-linked disorder characterized by genome instability, progressive bone marrow failure and predisposition to gynecological and head and neck squamous cell carcinomas (HNSCC). The risk of HNSCC is 1,000-fold greater compared to the general population, remains high even after bone marrow transplantation (BMT), and occurs at strikingly early ages with poor prognoses (1–4). Human papillomavirus (HPV) infection, particularly HPV16, is known to be associated with HNSCC in the general population; specifically oropharyngeal cancers (5). Interestingly, HPV positivity was significantly higher in oral samples from individuals with FA living in Brazil than in non-FA controls (6) suggesting a role for HPV in FA-associated cancer. This report was consistent with an earlier United States (US)-based study that found HPV DNA prevalence to be significantly greater in 25 FA-related HNSCC tumors compared to non-FA controls (7). However, a Dutch study failed to detect HPV DNA in any of their FA-related tumors tested (8) and HPV was not detected in 5 HNSCC or 4 anogenital squamous cell carcinoma samples from subjects with FA in a more recent study (9). Therefore, the extent to which HPV is associated with FA or HNSCC in individuals with FA remains unclear. While HPV is commonly sexually transmitted, it has been detected in healthy children and virgin adults in a number of studies (10–12), likely as a result of horizontal (non-sexual, perinatal or parenteral) transmission. Better characterization of the natural history of oral HPV in the FA population is therefore needed.

FA is a consequence of mutations in one of 16 genes whose respective protein products assemble in the nucleus to repair DNA damage (13, 14). Our recent studies indicate that under normal circumstances, activation of this important DNA repair pathway limits the HPV life cycle and prevents malignant transformation. Specifically, loss of function of the FA-related DNA repair pathway in HPV positive keratinocytes stimulates cellular and viral DNA replication (15). Conversely, rescue experiments in cells obtained from individuals with FA reduced HPV-mediated DNA damage and suppressed tumor growth (16) indicating that FA DNA repair pathway deficient keratinocytes uniquely support HPV infection and/or replication. These findings are reinforced by studies in mice demonstrating that deficiencies in the FA DNA repair pathway are associated with an increased incidence of HNSCCs, and with the ability of the HPV E7 protein to induce DNA damage (17–19). In addition, recent immunophenotyping and immune function studies in ten children with FA suggest heterogeneous immune defects (20). The extent to which FA DNA repair pathway deficiency in keratinocytes and variable or inadequate immune cell function contributes to early HPV acquisition, maintenance and/or susceptibility to cancer remains unknown. Given that HPV vaccination has been shown to offer protection against some high risk HPV types in the general population, now is a critical time to investigate HPV in these vulnerable individuals to determine the need for earlier vaccination and to develop targeted treatments.

In this study, we examined the prevalence of HPV infection in a sample of individuals with FA together with their parents who are obligate heterozygotes for the FA mutation, and siblings who are possible heterozygotes for the FA mutation. Study of family members helped to control for possibly non-sexual routes of exposure to HPV. Importantly, we observed a higher prevalence of HPV in individuals with FA as compared to their family members, particularly those that had been sexually exposed.

Materials and Methods

Informed Consent

This study was approved by Cincinnati Children's Hospital Medical Center (CCHMC) Institutional Review Board. Briefly, following a detailed verbal description of the study, written informed consent/parental permission was obtained by a trained clinical research coordinator from participants' 18 years old or participants' parent(s)/legal authorized representatives. Participants that were 8 years old were asked to sign the consent document along with their parents/guardians to indicate their assent. Younger children or children unable to provide written assent indicated their assent by willingly providing their sample(s).

Participant Recruitment

Patients visiting the Cincinnati Children's FA Comprehensive Care Center (CCFACCC) were approached to participate. Further, attendees of the Fanconi Anemia Research Fund (FARF) Adult and Family Meetings were invited to participate. Individuals of all ages were eligible to participate if they reported a diagnosis of FA or were a parent or sibling of an individual with FA, and were willing to complete study-related surveys and provide oral rinse samples. A subset (N=68) also provided a venous blood sample for HPV serology and immune studies. A total of 150 subjects with FA have been enrolled in the study as well as 247 family members (biological siblings and parents). Of these, 126 individuals with FA, 41 siblings and 121 parents currently have at least one HPV test result.

Oral Sample Collection, Nucleic Acid Isolation and HPV Testing

Oral exfoliated cells or oral-rinse and gargle samples were collected in 15 mL normal saline and 2.5 mL of 100% ethanol was added immediately after collection. Samples were stored for no more than 4 days at room temperature and then spun at 3,200 RPM for 10 minutes at 4°C, the pellet washed in 10 mL of phosphate-buffered saline (PBS), spun again and resuspended in 1 mL PBS and stored in 500 µl aliquots at -80°C. In the laboratory of Darron Brown at Indiana University, DNA was isolated using the 5 Prime ArchivePure DNA kit (5 Prime Inc., Gaithersburg, MD) according to the manufacturer's protocol. DNA samples were genotyped by The Linear Array HPV Genotyping Test (Roche Molecular Diagnostics, Indianapolis, Indiana) (21). Reactions were amplified in a PerkinElmer TC9600 Thermal Cycler (PerkinElmer), and positive and negative controls (included in the assay) were performed with every polymerase chain reaction (PCR) run. The GH20/PC04 human β-globin target was co-amplified to determine sample adequacy, and detection of specific HPV types was performed as previously described (22). The 37 individual HPV types detected are comprised of high risk HPV types (16, 18, 26, 31, 33, 35, 39, 45, 51, 52, 53, 56, 58, 59, 66,

67, 68, 69, 70, 73, 82, and Subtype 82 W13B) and low-risk HPV types (6, 11, 40, 42, 54, 55, 61, 62, 64, 72, 81, 83, 84, and 89).

Blood Collection, Processing and HPV Antibody Testing

Venous blood was collected observing appropriate aseptic techniques and universal precautions by trained personnel. Fifty percent were collected at the CCFACCC and immediately placed into storage at -80 degrees Celsius. Samples collected at FARF Meetings were typically kept at room temperature and centrifuged at $2,400 \times g$ within 8 hours of collection, aliquoted and then stored on dry ice until they were shipped via Same-Day FedEx (N=20). Otherwise, they were shipped via Same-Day FedEx to CCHMC, centrifuged within 24 hours, aliquoted and stored at -80 degree Celsius (N=14). An aliquot of each serum (0.5–1mL typically) was sent via Next-Day FedEx for HPV serology testing in the HPV laboratory at the Centers for Disease Control and Prevention. The HPV4-plex virus-like particle (VLP)-based IgG enzyme-linked immunosorbent assay (ELISA) for HPV 6, 11, 16 and 18 was performed as previously described (23). Cut-off values of Median+2 standard deviations (SD) were calculated using children's sera (Gift from Joakim Diller Karolinksa Institutet, Sweden) for each type. The pseudovirion neutralization assay (PBNA) was performed as described (24).

Immune assays

Humoral immune function was analyzed to assess effective or ineffective immune responses to HPV infection and or vaccination. Patient sera were tested for lymphocyte subsets, B cell panel, immunoglobulin levels (IgA, IgG, and IgM) along with tetanus and diphtheria titers. Immunoglobulin levels were determined by standard methods in the CCHMC clinical laboratory. Tetanus and diphtheria titers were determined by Quantitative Multiplex Bead Assay in the clinical diagnostic laboratory at CCHMC. Remaining assays were performed at the Diagnostic Immunology Laboratories at CCHMC. Results were interpreted with respect to age-appropriate reference ranges established in the laboratory. Peripheral blood mononuclear cells (PBMC) were isolated from heparinized blood by centrifugation with lymphocyte separation media (MP Biomedicals, Solon, OH, USA). Evaluation of patient lymphocyte subsets was performed via routine four-color flow cytometric analysis of EDTA preserved whole blood using fluorochrome-labelled monoclonal antibodies to lineage-specific cell surface markers for T cells (CD3, CD4, CD8), B cells (CD19), and natural killer (NK) cells (CD16, CD56). All antibodies were obtained from BD Biosciences (San Jose, CA, USA). Briefly, erythrocytes were lysed by incubation in FACSLyse (BD Biosciences). Samples were then stained with antibody and analyzed on a FACSCalibur flow cytometer (BD Biosciences) using multiset software (BD Biosciences). Similar flow based approach was applied for assessment of B-Cell Panel (25).

Risk Factor Survey

Survey completion was done either by paper or by electronic survey (Research electronic data capture (REDCap) software) (26). Participants ages 15 years and older were asked to complete surveys for themselves unless there was a developmental delay, in which case a parent or guardian was asked to complete the survey. For children ages 12 to 15 years, a parent was asked either to complete the survey or assist the child in completing the survey.

However, answers to the sexual questions were ascertained by study staff interview whenever possible. Surveys collected demographic and clinical data including FA complementation group, disease severity, and date/age at bone marrow transplant (if any). In addition, other potential risk factors for HPV-related disease such as vaginal versus Cesarean section mode of delivery at birth, gestational age at birth, tobacco smoke exposure, alcohol consumption and sexual history (including questions about oral sex and kissing), family history of FA and cancer, history of personal immunization and/or HPV testing, and presence and frequency of oral lesions were ascertained by survey and/or staff interview. When available, biological mothers of participants were asked whether they have had genital or anal warts, a dysplastic Pap smear, or had a positive HPV test within a 3-year period before the birth of the individual(s) with FA and siblings.

Statistical Analysis

Statistical analyses were performed using Statistical Analysis Software (SAS), version 9.3 (SAS Institute Inc., Cary, NC). Prior to analyses, data qualities were examined. Demographic variables and potential risk factors of HPV were first summarized as median and interquartile range (IQR) or frequencies (proportions) and compared in FA, siblings and parents. These factors were then compared between HPV negative and positive individuals with FA. Using Fisher's exact tests, the impact of FA status on HPV prevalence was stratified by sexual experience or HPV vaccination status. Using a multi-variable logistic regression, we further tested the effects of HPV risk factors on the prevalence of HPV, adjusting for sexual experience, HPV vaccination status, and FA status in the model. Finally, the association between BMT and oral HPV was assessed considering HPV infection and sexual experience. Effects were considered significant if p-value ≤ 0.05 .

Results

Population characteristics

A total of 288 individuals with oral rinse samples and risk factor questionnaires were included in this study. Of these, 126 individuals had FA, 121 were parents of individuals with FA (74 mothers and 47 fathers), and 41 were siblings of the individuals with FA (Table 1). Participants with FA ranged in age from 6 months to 55 years. Family members' ages ranged from 4 months to 79 years. As expected, ages were significantly different between subjects with FA and family members ($p < 0.0001$). There were more females in each group than males, and more Caucasians than any other race. Individuals with FA were living in families across all income levels (26% $< 40K$, 36% $40-90K$, 39% $> 90K$) and most had health insurance (32% public, 54% private and 11% had both public and private insurance) (Table 1).

HPV prevalence in FA

Among those individuals with FA, 14 (11.1%) tested positive for one or more HPV types. Indeed, oral HPV positivity in FA was significantly greater than in non-FA family members (parents and siblings; 2.5%; p -value=0.003; Fisher's exact test; Figure 1A), whose prevalence's were consistent with published levels in the general population (range from 1.9% to 7% (10–12)). Also consistent with published studies, the most common types of

HPV detected among those with FA were HPV 6 (50%) and HPV 16 (64%), although HPV 18, 51, 66, 84 and CP6 were also detected. Only three of the HPV positive individuals with FA had previously been vaccinated for HPV (all receiving the GARDASIL Quadrivalent HPV Vaccine, Merck & Co., Inc., West Point, PA, USA) and of these three vaccinated HPV positive individuals, only a single person with FA tested positive for an HPV type included in the vaccine (HPV16). We also examined HPV positivity in people based on HPV vaccine history. As shown in Figure 1B, in both vaccinated and unvaccinated groups, individuals with FA tested positive for HPV more often. Importantly, in our study population, we observed significantly higher rates of vaccination in individuals with FA compared to family members (38% vs. 21% in siblings and 1% of parents, $p < 0.0001$, Fisher's exact test, Table 2). It is therefore interesting that HPV prevalence is higher in FA despite greater rates of vaccination.

Typically transmission of high risk carcinogenic HPV types in the general population occurs predominantly via sexual contact including oral sex and open-mouthed kissing outside of vaginal sex (27). When we examined only those individuals who were sexually experienced, we observed significantly more oral HPV infections in those with FA (17.7%; p -value = 0.003) compared to family members (2.4%; Figure 1C). In participants that were not yet sexually experienced, individuals with FA also tested positive for oral HPV (8.7%) more often compared to siblings (2.9%), though the difference did not reach statistical significance. We next assessed the effect of sex on the association between oral HPV positivity and FA. While having sex did not seem to increase HPV positivity in family members, individuals with FA exhibited a higher prevalence of HPV when they reported having had sexual experience (17.7% vs 8.7%), but this association again was not statistically significant at the 5% level. In multivariable logistic regression analysis including sexual experience, HPV vaccination and FA status, only having FA was significantly associated with oral HPV positivity ($p = 0.002$). Indeed, having FA increased the odds of HPV positivity by 4.9 fold (95% CI: 1.6–15.4). Greater HPV prevalence was observed in those with FA compared to family members regardless of age. Importantly, among those individuals with FA that were not sexually active, 7 children less than 13 years of age (10.9%) also tested HPV positive (Figure 1D).

Other potential between-group confounding or risk factors

We next evaluated several other factors associated with HPV positivity or HNSCC in the general population and compared those with FA to their family members (Table 2). We observed that 71% of individuals with FA compared to 54% of siblings and 90% of parents were delivered vaginally ($p = 0.0001$; Table 2). The biological mothers of those with FA compared to sibling group as a whole were not more likely to have reported having an abnormal Pap smear, positive HPV test or other sexually transmitted disease within 3 years of their children's birth ($p = 0.75$). Not surprising given the differences in age as well as strength of the anti-smoking and anti-alcohol guidelines and education provided to those with FA by the Fanconi Anemia Research Fund (28), primary smoking and alcohol use were lower for those with FA and their siblings compared to parents ($p = 0.0003$ and 0.0001 , respectively; Table 2). While the frequency of genital warts and common warts did not significantly differ between groups ($p = 0.87$ and $p = 0.81$, respectively; Table 2), significantly

more individuals with FA reported oral sores (18%) compared to 8% of siblings and 4% of parents ($p=0.006$).

Examining only those individuals with FA, no other factor related to FA (complementation group, having had a transfusion or having a family history of cancer) or those known to contribute to HPV or HNSCCs (i.e. tobacco and alcohol use, oral, genital and common warts, and dental hygiene) were found to be significantly different between oral HPV positive or oral HPV negative individuals with FA in our study (Table 3). While some have hypothesized that BMT might be associated with the increased risk of HNSCC in individuals with FA, we found that HPV positivity was only slightly higher (12.7%) post-BMT compared to pre-BMT (9.5%) and this association was not statistically significant ($p=0.78$; Figure 2). When we considered BMT and sexual experience simultaneously, we again observed higher levels of oral HPV positivity in those post-BMT particularly if they had had sexual experience, but again these differences were not statistically significant ($p=1.0$; Figure 2). Secondhand smoke exposure, however, was marginally statistically significantly higher in those that were oral HPV positive (29%) compared to oral HPV negative (9%; $p=0.052$).

Characterization of individuals with FA who are HPV positive

There was a broad range of ages (from 6 to 51 years of age) among the 14 individuals with FA who tested oral HPV positive (Table 3). Importantly, 7 (50%) were under the age of 13 years and 8 individuals with FA indicated that they were not sexually active. The level of oral HPV positivity was similar for males and females and there were no differences by whether or not they had a BMT (57%). While 9 (64%) had the FANCA complementation group, C, F, I, and D2 complementation groups were also represented in those that tested oral HPV positive (Table 4). Only a single oral HPV positive participant (HPV 84) reported having had a diagnosis of oropharyngeal cancer prior to sampling.

Further, among all 8 oral HPV positive individuals with FA who had corresponding immune data, we observed 6 study participants to have normal IgG levels and 5 participants with normal absolute and/or memory B cells (Table 5). Most tetanus and diphtheria titers were also normal. Only 3 of the 8 subjects had been vaccinated for HPV prior to blood sampling. All vaccinated individuals had positive titers to all four HPV types included in the vaccine (all Gardasil) as well as normal tetanus and diphtheria levels (Table 5). Two of the oral HPV positive individuals who indicated that they had not yet been vaccinated for HPV were also serologically positive to types other than identified in their oral sample suggesting prior natural infections with these types. Three of the oral HPV positive participants had no detectable HPV titers. Two of these individuals tested positive for both oral HPV6 and HPV16. Interestingly, these individuals were either deficient in their absolute B cell count or memory B cells. Overall, there were no unifying characteristics that were shared by those that were oral HPV or serologically positive with FA. In fact, these data support our previous report that individuals with FA have heterogeneous immune responses (20).

Discussion

Our cross sectional study demonstrates a higher frequency of oral HPV positivity in individuals with FA compared to family members. Further, while HPV prevalence was significantly higher for those with FA who reported sexual exposure compared to family members, HPV prevalence tended to also be higher in those who were not yet exposed to sex.

A possible role for HPV in FA (29) has been previously suggested following the report of higher prevalence of HPV in FA-related head and neck tumors (7). In contrast, another equally small study reported the absence of HPV in FA-related HNSCC tumors and cell lines derived from them (8). These opposing results may be due to experimental variability such as types of primers used, quality and content of DNA tested, analysis of tumors or tumor-derived cell lines, or geographic differences. More recently, Alter et al were unable to detect HPV in HNSCCs from individuals with FA (9). However, de Araujo et al. have shown an increased prevalence of oral HPV isolated from 42 oral cytobrush samples from individuals with FA without oral malignant lesions (6). One possibility is that the Brazilian population spanned a younger age range comparable to our study population. A second explanation for the discrepancy could be that some/many individuals with FA successfully clear the virus or suppress its replication before tumors develop, thus limiting detection in the tumors themselves. These studies, together with our data presented here, suggest that studies of the association between HPV and FA might have to be conducted on younger individuals, and call for continued longitudinal studies across the lifespan to determine if early life exposure to HPV impacts the development of HNSCCs in individuals with FA later in life. Further, longitudinal studies would better address potential routes of transmission outside of sexual exposure. We have currently not detected significant differences in breastfeeding, birthing method (c-section vs. vaginal), BMT/blood transfusions, and alcohol and tobacco exposure, although in some cases, the data collected was incomplete and likely underpowered to ascertain a true association.

While studies in human subjects remain controversial, there is growing evidence from cell lines, animal studies, expression analysis and computational biology approaches that support a strong association between the consequences of HPV infection in the context of deficiency in the FA proteins and development of cancer. Organotypic raft cultures of HPV-oncogene immortalized, FANCA deficient patient-derived keratinocytes demonstrate increased DNA damage, and excessive hyperplasia compared to FANCA complemented cell populations (16). In addition, the FA pathway seems to limit the ability of HPV to replicate in cultured keratinocytes (15). Indeed, FANCD2 deficient mice overexpressing the HPV E7 oncoprotein have increased DNA damage and are more likely to develop HNSCCs (17) and cervical and vaginal cancer (18). These effects of E7 are due to inactivation of the Rb family of tumor suppressors that limit DNA damage (19). Microarray analysis of vulvar tissue infected with both low-risk and/or high-risk HPV types indicated that FANCA, FANCD2, and other DNA damage markers were significantly induced following high-risk HPV infection, along with increased DNA damage in the tissues (30). And finally, using a computational biology approach, a link between HPV16 and the FA DNA repair pathway was identified and then validated experimentally by evaluating the impact of overexpressing

E6 or E7 proteins in primary fibroblasts and keratinocytes using global gene expression analysis (31).

Together, these diverse approaches point to a common theme. It appears that HPV infection results in elevated DNA damage that then triggers the FA pathway to repair this DNA damage. In individuals where this pathway is defective, it is likely that the DNA damage will not be repaired, and as a consequence increases the likelihood of tumor development in the long term. It becomes important to consider these studies again in the context of the conflicting human data. Even if HPV is suppressed or cleared, and is undetectable by PCR assays, one might speculate that the resulting DNA damage is the trigger for increased HNSCCs and anogenital carcinomas many years later. Additional studies will be required to support this hypothesis.

Similar to the Brazilian study (6), we found that oral HPV16 was the most common type detected. We also detected less common HPV types in those with FA (36% of oral samples). It is plausible, although speculative, that less common high-risk types, or even low-risk HPV types, might unusually initiate a cancer cascade in those with FA who have both a DNA repair defect and heterogeneous immune deficiency, suggesting a need to evaluate the broad spectrum of HPV types rather than HPV16 and 18 alone in oral rinse and possibly other sample types. Indeed, among 67 throat swabs from individuals with FA collected at the same time as our oral rinse samples in this report, we identified 5 individuals positive for HPV 33, HPV 84, HPV 66, 51, 52, 62 82 and HPV 51; a 60% agreement between sample types (32).

An important variable to consider is the immune response to HPV infection in individuals with FA. We have previously demonstrated that individuals with FA have fewer B cells and NK cells, as well as decreased NK cell function and cytotoxic T cell activity (20). It is conceivable that in the absence of a fully functional FA pathway, the ability to suppress or clear HPV is also delayed. Based on the data from Table 5, there seems to be variability in the immune responses in HPV positive individuals with FA. While some subjects with FA had normal immune responses, others seem to be unable to mount a strong response (based on B cell and memory B cell numbers, IgG levels, and response to tetanus and diphtheria vaccines). These data indicate that while not all individuals with FA have measurably impaired immune responses, those that do could conceivably be at higher risk. It is reasonable to speculate that viral persistence due to the inability of the immune system to clear the infection or due to fluctuations in host immunity as a result of progressive bone marrow failure or FA-related illnesses, further increases the duration of HPV infection and extent of DNA damage, thereby resulting in an increased risk for carcinomas. Our data showed no differences in the type and distribution of factors known to cause cancers in individuals with Fanconi anemia regardless of their HPV infection status. This would further suggest that it might be the ability of the individual to suppress or clear the infection that most likely determines HPV prevalence, incidence, and susceptibility to HNSCCs.

In conclusion, continued longitudinal collection of oral samples and broad testing for HPV types along with immunological studies to better characterize response remains critical to address these questions. In addition, studies are needed to determine prospectively if HPV

infection is associated with increased HNSCCs in individuals with FA, and if vaccination at a younger age could be effective in preventing early infection.

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The findings and conclusions in this report are those of the authors and do not necessarily represent the official position of the CDC.

Abbreviations

HPV	human papillomavirus
HNSCC	head and neck squamous cell carcinoma
FA	Fanconi anemia
BMT	bone marrow transplant
CCHMC	Cincinnati Children's Hospital Medical Center
CCFACCC	Cincinnati Children's FA Comprehensive Care Center
REDCap	Research Electronic Data Capture
FARF	Fanconi Anemia Research Fund
IQR	interquartile range
PBS	phosphate-buffered saline
PBNA	pseudovirion neutralization assay
VLP	virus-like particle
ELISA	enzyme-linked immunosorbent assay
DNA	deoxyribonucleic acid
PCR	polymerase chain reaction
SD	standard deviation
STD	sexually transmitted disease
IgG	immunoglobulin G
Tet	tetanus
Dip	diphtheria

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Key Points

- Increased oral HPV prevalence, particularly of HPV16, in individuals with Fanconi Anemia
- No obvious risk factors for oral HPV positivity among individuals with FA
- Those with FA have heterogeneous general immune responses and positive HPV serology

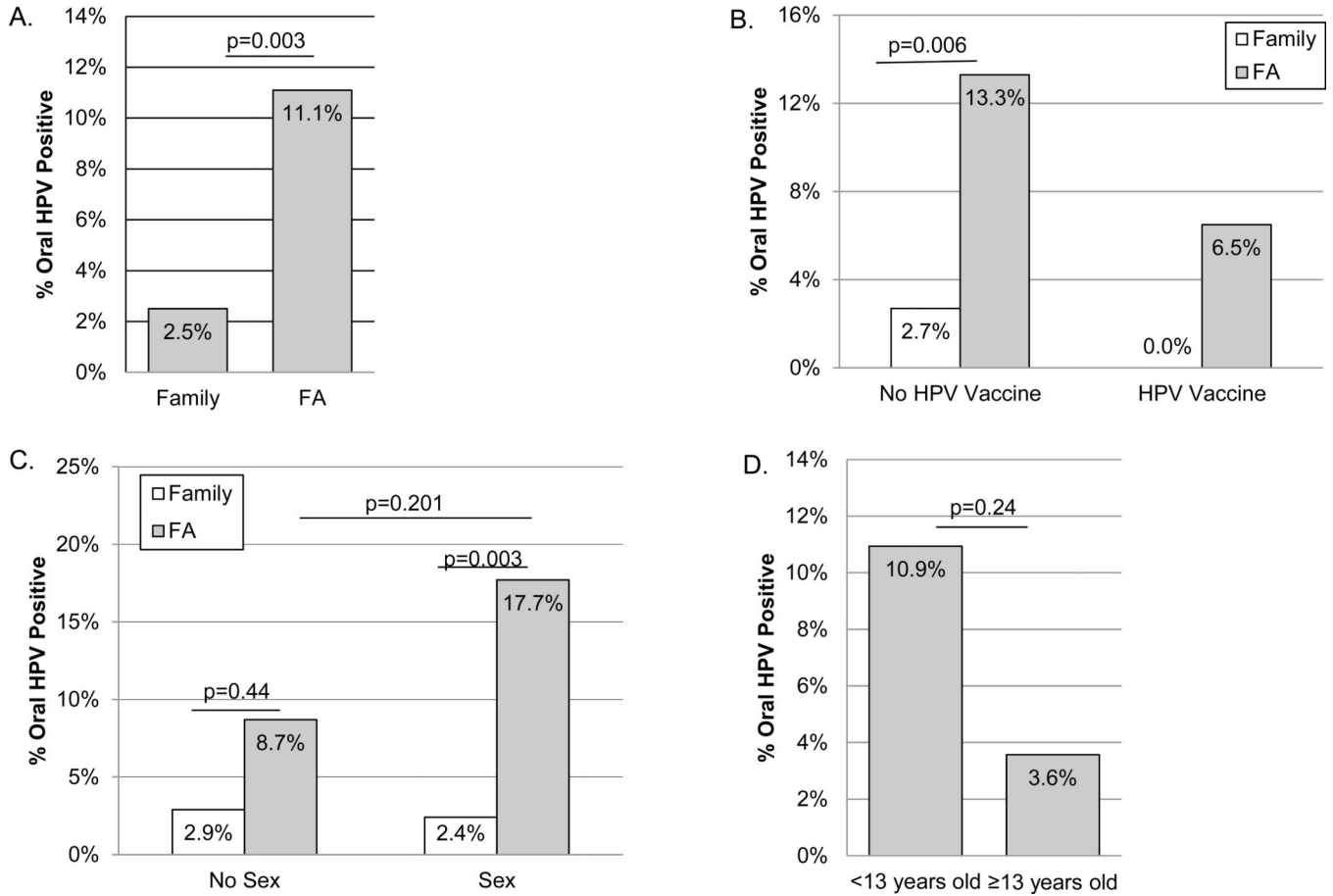


Figure 1.

Associations between oral HPV positivity in individuals with FA vs. their family members. A) Markedly greater oral HPV positivity considering all HPV-tested individuals with FA vs. their family members. B) Oral HPV positivity stratified by HPV vaccination (Gardasil). C) Oral HPV positivity stratified by sexual experience. Sex indicates that the participant reported having had sexual contact (oral, genital, and/or anal) via paper or online survey at the time of oral sampling. Those not having reported sexual contact are in the no sex group. D) Oral HPV positivity among those in the no sex group stratified by age. For all comparisons, Oral HPV positivity was determined by Roche Linear Array and the percentage of positive individuals are shown. A fisher's exact test was used to assess statistical differences (2-sided p-values).

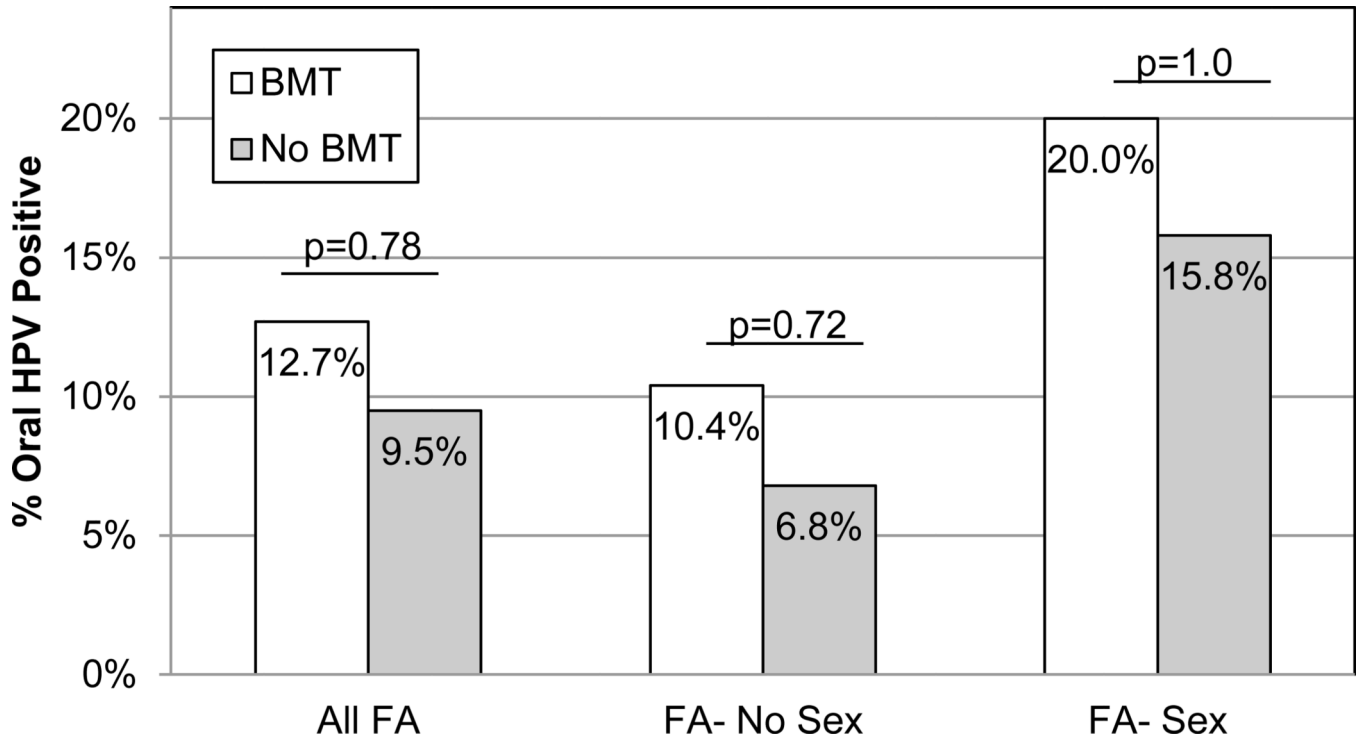


Figure 2. Oral HPV positivity stratified by both history of bone marrow transplant (BMT) history and sexual experience. Oral HPV positivity was determined by the Roche Linear Array and the percentage of positive individuals are shown. BMT history was reported via paper or online survey at the time of oral sampling. Sex indicates that the participant reported having had sexual contact (oral, genital, and/or anal). Those not having reported sexual contact are in the no sex group. A fisher’s exact test was used to assess statistical differences (2-sided p-values).

Table 1

Demographic and Socioeconomic Characteristics.

	Fanconi Anemia		Siblings		Parents		P
	All N=126		All N=41		N=121 All		
Age (years)	12	(9, 22)	9	(5, 16)	43	(37, 50)	<0.0001
Age Range (years)							<0.0001
0-4	13	(10%)	6	(15%)	0	(0%)	
5-12	51	(40%)	19	(46%)	0	(0%)	
13-18	25	(20%)	10	(24%)	0	(0%)	
19+	37	(29%)	6	(15%)	121	(100%)	
Sex							0.034
Male	56	(44%)	9	(22%)	47	(39%)	
Female	70	(56%)	32	(78%)	74	(61%)	
Race							0.020
White/Caucasian	107	(86%)	34	(94%)	113	(95%)	
Black/African American	6	(5%)	2	(6%)	5	(4%)	
Other/Mixed	11	(9%)	0	(0%)	1	(1%)	
Income							0.14
Under \$39,999	28	(26%)	3	(8%)	21	(18%)	
\$40,000-\$89,999	39	(36%)	16	(40%)	42	(37%)	
Over \$90,000	42	(39%)	21	(53%)	52	(45%)	
Insurance/Medical							0.001
Self-Pay/Other	4	(3%)	0	(0%)	10	(9%)	
Public/Government	37	(32%)	7	(21%)	19	(16%)	
Private	63	(54%)	26	(79%)	83	(72%)	
Private and Public	13	(11%)	0	(0%)	4	(3%)	

Age was tested using Kruskal-Wallis test; other variables were tested by Fisher's exact tests. All analyses were performed on non-missing data. Information on race, income and health insurance was missing in 9, 24 and 22 subjects, respectively. In bold are those P values that are less than 0.05.

Differences in HPV Risk Factors Between Individuals with Fanconi Anemia and Their Family Members

Table 2

Risk Factors	FA N=126	Siblings N=41	Parents N=121	P-value
HPV Vaccination (N=279)	46 (38%)	8 (21%)	1 (1%)	< 0.0001
Sexual Experience (N=288)	34 (27%)	6 (15%)	121 (100%)	< 0.0001
Vaginal Birth/Delivery (N=276)	87 (71%)	22 (54%)	101 (90%)	< 0.0001
Primary Tobacco User (N=285)	5 (4%)	1 (2%)	21 (18%)	0.0003
Secondhand Smoke Exposure (N=283)	14 (11%)	3 (7%)	14 (12%)	0.8120
Alcohol Use (N=285)	14 (11%)	2 (5%)	43 (36%)	< 0.0001
Genital Warts (N=282)	4 (3%)	1 (2%)	2 (2%)	0.8733
Common Warts (N=64)	9 (30%)	2 (22%)	9 (36%)	0.8123
Oral Sores (N=224)	14 (18%)	3 (8%)	5 (4%)	0.006
Breastfed as an infant (N=217)	60 (71%)	30 (81%)	46 (48%)	0.0003
Mother Abnormal Pap/ HPV/STD at Birth (N=109)	8 (11%)	5 (14%)	-	0.7554

All variables were binary. The fisher's exact test was used to assess differences (2-sided p-values). Pap- Papanicolaou, STD – other sexually transmitted disease (trichomoniasis, chlamydia, gonorrhea, and genital herpes). As some variables have missing data, frequencies are provided for each. In bold are those p-values that are less than 0.05.

Table 3

Associations with HPV In Individuals with Fanconi Anemia

Potential Risk Factors	FA HPV- N=112	FA HPV+ N=14	P-value
Age 13 years (N=126)	55 (49%)	7 (50%)	1.000
Male Sex (N=126)	48 (43%)	8 (57%)	0.396
White/Caucasian (N=124)	95 (86%)	12 (86%)	1.000
Income < \$39,999/year (N=109)	25 (26%)	3 (23%)	1.000
Private Medical Insurance (N=117)	51 (50%)	12 (86%)	0.0195
HPV Vaccination (N=121)	43 (40%)	3 (23%)	0.366
Sex Experience (N=126)	28 (25%)	6 (43%)	0.201
Delivered Vaginally (N=123)	77 (71%)	10 (71%)	1.000
Primary Tobacco User (N=126)	4 (4%)	1 (7%)	0.451
Secondhand Smoke Exposure (N=125)	10 (9%)	4 (29%)	0.052
Alcohol User (N=126)	12 (11%)	2 (14%)	0.655
Genital Warts (N=124)	3 (3%)	1 (8%)	0.362
Common Warts (N=30)	7 (26%)	2 (67%)	0.207
Oral Sores (N=76)	13 (19%)	1 (17%)	1.000
Breastfed (N=85)	53 (68%)	7 (100%)	0.100
Family History of Cancer (N=119)	48 (46%)	7 (50%)	0.783
Mother Abnormal Pap/HPV/STD at Birth (n=73)	7 (11%)	1 (13%)	1.000
Bone Marrow Transplant (n=126)	55 (49%)	8 (57%)	0.778
Blood Transfusions (n=121)	67 (63%)	9 (64%)	1.000

All variables were dichotomized. The Fisher's exact test was used to assess group differences (2-sided p-values). Among the 14 HPV+ individuals, 1 person was missing data on HPV vaccination and genital warts, 11 were missing common warts, 8 missing oral sores, 7 subjects were missing breastfeeding data and 6 subjects did not have biological mother Pap/sexually transmitted disease (STD) data at the participant's birth available. Those with private medical insurance were compared to those with no-pay/self-pay/public policies. In bold are those p-values that are less than 0.05.

Table 4

Individual Characteristics of HPV+ Individuals with Fanconi Anemia

#	Age (years)	Sex	Sexual Exposure	BMT	FANC Group	HPV Vaccine	HPV Types+
1	32	F	+	-	A	-	6, 16
2	7	F	-	+	A	-	66
3	23	F	+	+	Unknown	-	84
4	20	F	+	-	A	+	84
5	14	F	-	+	A	+	16
6	12	F	-	+	A	+	CP6
7	7	M	-	-	Other	-	6, 16
8	27	M	+	+	Other	-	6, 16
9	10	M	-	-	A	-	6, 16
10	6	M	-	+	Other	-	6, 16
11	51	M	+	-	A	-	16
12	6	M	-	-	Other	-	6, 16
13	12	M	-	+	A	-	6, 16, 18
14	35	M	+	+	A	-	51
Sum	7 <13y	8 M's	6+'s	8+'s	8 A's	3+'s	9 HPV16's

M – Male sex, F-female sex, BMT – history of Bone Marrow Transplant, FANC- FA complementation group, + indicates presence of exposure/history of BMT or HPV vaccine. Other complementation groups represented include C, F, I, and D2. BMT, bone marrow transplant

Table 5

Immune and HPV-serological Responses

#	Age (years) at:			General Immune Response					HPV-Specific Immune Responses							
	BMT	Oral rinse	Blood	Absolute B-cell Count	Memory B-cells	IgG	Tet	Dip	Oral HPV Types +	GARDASIL HPV Vaccine (#shots)	HPV6 VLP-ELISA	HPV11 VLP-ELISA	HPV16 VLP-ELISA	HPV18 VLP-ELISA	HPV16 PBNA	HPV18 PBNA
1	7	10	10	Normal	Normal	Down	++	+	6,16	-/+ (2)*	+	+	+	+	+	+
2	18	27	30	Normal	Normal	Normal	+	-	6, 16	-/+ (3)*	+	+	+	+	+	-
3	4	10	13	Normal	Normal	Normal	+	++	6,16	-	+	+	+	+	+	+
4	4	6	9	Down	Up	Normal	++	++	6, 16	-	-	-	-	-	-	-
5	5	32	35	Normal	Down	Normal	+	+	6, 16	-	-	-	-	-	-	-
6	5	7	7	Up	Normal	Normal	+	+	66	-	-	-	-	-	-	-
7	11	23	23	Normal	Normal	Up	+	-	84	-	-	-	+	+	+	+
8	20	23	23	Down	Up	Normal	++	+	84	3	+	+	+	+	+	+

* Did not have the human papillomavirus (HPV) vaccine at time of oral rinse collection, but had received the vaccine at time of blood collection; tetanus (Tet)/diphtheria (Dip) responses ≥ 1.0 IU/mL is considered protective (++); 0.1 to 0.99 IU/mL (+); <0.1 IU/mL (-) is considered a non-responder; absolute CD3 and absolute CD16/56 were normal for all 8 participants. BMT-bone marrow transplant; VLP- Virus-Like Particles; ELISA- enzyme-linked immunosorbent assay; PBNA-pseudovirion-based neutralization assay.