Relationship among Glutamine, γ -Aminobutyric Acid, and Social Cognition in Autism Spectrum Disorders

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Abstract

Objective: An imbalance of excitatory and inhibitory neurotransmission in autism spectrum disorder (ASD) has been proposed. We compared glutamate (Glu), glutamine (Gln), and γ -aminobutyric acid (GABA) levels in the anterior cingulate cortex (ACC) of 13 males with ASD and 14 typically developing (TD) males (ages 13–17), and correlated these levels with intelligence quotient (IQ) and measures of social cognition.

Methods: Social cognition was evaluated by administration of the Social Responsiveness Scale (SRS) and the Reading the Mind in the Eyes Test (RMET). We acquired proton magnetic resonance spectroscopy (¹H-MRS) data from the bilateral ACC using the single voxel point resolved spectroscopy sequence (PRESS) to quantify Glu and Gln, and Mescher–Garwood point-resolved spectroscopy sequence (MEGA-PRESS) to quantify GABA levels referenced to creatine (Cr).

Results: There were higher Gln levels (p = 0.04), and lower GABA/Cre levels (p = 0.09) in the ASD group than in the TD group. There was no difference in Glu levels between groups. Gln was negatively correlated with RMET score (rho = -0.62, p = 0.001) and IQ (rho = -0.56, p = 0.003), and positively correlated with SRS scores (rho = 0.53, p = 0.007). GABA/Cre levels were positively correlated with RMET score (rho = 0.34, p = 0.09) and IQ (rho = 0.36, p = 0.07), and negatively correlated with SRS score (rho = -0.34, p = 0.09).

Conclusions: These data suggest an imbalance between glutamatergic neurotransmission and GABA-ergic neurotransmission in ASD. Higher Gln levels and lower GABA/Cre levels were associated with lower IQ and greater impairments in social cognition across groups.

Introduction

A UTISM SPECTRUM DISORDERS (ASD) ARE neurodevelopmental disorders characterized by deficits in social communication and interaction, and restricted, repetitive patterns of behavior, interests, or activities (American Psychiatric Association 2013). ASD are increasing in prevalence, with recent estimates ranging from 1.1 to 2.0% of children having a diagnosis, a 75% increase in the last 5 years (Autism and Developmental Disabilities Monitoring Network Surveillance Year 2008 Principal Investigators and Centers

for Disease Control and Prevention 2012; Blumberg et al. 2013). These disorders have significant social, public health, and economic impact, with total annual health care expenditures for ASD being \$1,700,000 per 10,000 individuals (Wang and Leslie 2010). Howlin and colleagues demonstrated that the majority of individuals diagnosed with ASD have poor outcomes in the area of social functioning. The strongest predictor of poor outcome is the deficit in social interaction at the time of diagnosis (Howlin et al. 2013).

Nevertheless, there are no medical therapies to target the core social cognition deficits in ASD. The development of such

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treatments depends upon an understanding of the pathophysiology of ASD, which has been proposed to be characterized by an imbalance in excitatory glutamatergic and inhibitory γ -aminobutyric acid (GABA)-ergic neurotransmission (Fatemi 2008). Molecular targets in the glutamate- (Glu) and GABA- signaling pathways have been the focus of recent efforts to develop such medical treatments, but development has been hampered by the lack of valid biomarkers to identify those most likely to respond to these treatments.

Glu and GABA abnormalities in ASD include postmortem differences in the expression of Glu transporters (Purcell et al. 2001), GABA-A (Fatemi et al. 2009b) and GABA-B receptors (Fatemi et al. 2009a), enzymes that convert Glu to GABA (Yip et al. 2007, 2009), metabotropic (Fatemi and Folsom 2011) and ionic (Purcell et al. 2001) Glu receptors, and regulatory proteins affecting Glu receptor activity (Fatemi and Folsom 2011). Multiple GABA receptors are underexpressed in ASD in multiple brain regions, including the anterior cingulate cortex (ACC) (Oblak et al. 2009, 2010). In clinical trials of glutamate receptor antagonists and GABA receptor agonists in individuals with Fragile X syndrome (more than half of whom had comorbid ASD), a subset of patients showed improvement in social impairments (Jacquemont et al. 2011; Berry-Kravis et al. 2012). However, there is a critical need for biomarkers that can be used a priori to predict which individuals will respond to these treatments.

After synaptic reuptake into astrocytes, Glu is stored as glutamine (Gln) until being recycled within the presynaptic terminal (Magistretti and Pellerin 1999). Because of this rapid recycling of excess Glu and the large metabolic pool of Glu that is not involved in neurotransmission, Gln concentration may be a more sensitive measure of synaptic glutamatergic neurotransmission than Glu (Ongur et al. 2008). Other studies in mood disorders have implicated the Gln/Glu ratio as a more sensitive marker of glutamatergic function (Yuksel and Ongur 2010).

Proton magnetic resonance spectroscopy (¹H-MRS) quantitatively measures regional cortical levels of Glu, Gln, and GABA, although frequently Glu and Gln are reported together as Glx because of difficulty separating out the resonances. Previous ¹H-MRS studies have demonstrated variable changes in Glx by region and age in ASD, with the majority demonstrating elevated levels. In children and adolescents with ASD, Glx has been reported to be decreased in the frontal and occipital gray matter and cerebellum (DeVito et al. 2007), and increased in the ACC (Bejjani et al. 2012). In adults with ASD, Glx has been reported to be increased in the amygdala/hippocampal region (Page et al. 2006) and auditory cortex (Brown et al. 2013), and decreased in the ACC (Bernardi et al. 2011) and basal ganglia (Horder et al. 2013). Joshi and colleagues (Joshi et al. 2013) reported increased Glu in the ACC in adolescents with ASD (similar age range to our study).

GABA levels have been studied less frequently because of the specialized editing techniques needed to segregate the GABA content; however, three studies have found decreased levels in youth with ASD (Harada et al. 2011; Gaetz et al. 2014; Rojas et al. 2014). Neither Glx nor GABA has been associated with intelligence quotient (IQ). No published studies to date have simultaneously measured Glu, Gln, and GABA in regions known to be involved in social cognition. Moreover, no studies have reported correlation of these ¹H-MRS parameters with clinical phenotype.

The ACC has been implicated in the neural basis of social impairments in ASD (Mundy 2003). Abnormalities in the ACC have been demonstrated in individuals with ASD through structural (Hyde et al. 2010), functional (Agam et al. 2010), and diffusion tensor (Thakkar et al. 2008) imaging studies, as well as neuropathologically (Oblak et al. 2009, 2010). Glucose metabolism measured by positron emission tomography in the ACC is also lower in individuals with ASD (Haznedar et al. 2000). Therefore, we have chosen the ACC as an ideal brain region to investigate glutamatergic and GABA-ergic abnormalities in ASD and their association with social cognition deficits.

We measured Glu, Gln, and GABA levels in the ACC of adolescents with ASD and a typically developing (TD) control group. We investigated correlations with IQ, clinical measures of social cognition, and other clinical internalizing and externalizing symptoms. We hypothesized that adolescents with ASD would have higher Glu and/or Gln levels and lower GABA levels than the TD group, and that social cognition would be negatively correlated with Glu and/or Gln and positively correlated with GABA in the ACC.

Methods

Participants

Thirteen males with ASD (mean \pm SD age: 14.9 \pm 1.5 years, range: 13.1–17.5 years) and 14 TD male controls (age: 14.7 \pm 1.8 years, range: 13.1–17.4 years) participated in the study. The participants were recruited through the Child and Adolescent Neuro-Development Initiative (CANDI) at the University of Massachusetts Medical School (UMMS) from the current pool of clinic patients, word of mouth, advertisements in local newspapers, and online advertising. A telephone screen was administered to determine participant eligibility.

Inclusion criteria for the study were: Males 13–17 years of age with Full Scale Intelligence Quotient (FSIQ) >70 as estimated by two subtests of the Weschler Abbreviated Scales of Intelligence (WASI) (Weschler 1999). Exclusionary criteria included major medical or neurological illness, primary psychiatric diagnosis other than ASD, with the exception of mild anxiety disorder (not requiring medication) and/or attention-deficit/hyperactivity disorder (ADHD), use of anticonvulsant medication or psychoactive medication with the exception of stimulants, past or current history of claustrophobia, exquisite sensitivity to noise, metal fragments or implants, or any other contraindication to magnetic resonance imaging (MRI). Parents and participants ≥ 16 years of age provided written consent, and participants < age 16 provided written assent, as approved by UMMS Institutional Review Board (Docket # H-14247).

Clinical assessment

Five individuals met *Diagnostic and Statistical Manual of Mental Disorders*, 4th ed. (DSM-IV) criteria for autistic disorder, six met criteria for Asperger's disorder, and two met criteria for pervasive developmental disorder-not otherwise specified, as determined by unstructured clinical interviews with a child psychiatrist (D.M.C.) and administration of the Autism Diagnostic Observation Schedule (ADOS) by a research- reliable trained clinician (A.F.) (American Psychiatric Association 1994). Psychiatric comorbidities were assessed by unstructured clinical interviews with a child psychiatrist (D.M.C.) and the Child Behavior Checklist (CBCL) was used to assess multiple domains of psychiatric functioning (Achenbach and Rescorla 2001).

Neurocognitive testing and measures of social cognition

All adolescents were administered two subtests of the WASI (Vocabulary and Matrix Reasoning) to obtain an estimate of FSIQ (Weschler 1999). The Social Responsiveness Scale (SRS) was administered to the primary caregiver. The SRS is a quantitative measurement of various dimensions of interpersonal behavior, communication, and repetitive/stereotypic behaviors, including subscales for Social Awareness, Social Cognition, Social Communication, Social Motivation, and Autistic Mannerisms (Constantino and Gruber 2005). Higher scores reflect greater deficits in each area. Adolescent participants were also administered the Reading the Mind in the Eyes Test (RMET) (Baron-Cohen et al. 2001). The RMET has been validated to demonstrate deficits in individuals with ASD in the ability to infer emotional state from viewing the eye region of photographed faces, which has been used in individuals as young as 12 years of age (Guastella et al. 2010). Scores are given as percentage of correct responses, so that a lower score indicates greater impairment.

NeuroImaging data collection

NeuroImaging data were acquired using a 3.0 Tesla Philips Achieva whole-body MR system (Philips Healthcare, Best, The Netherlands) with an eight-element phased-array head coil at the Advanced MRI Center, UMMS. T1-weighted anatomical MRI (MPRAGE sequence, 256×256 voxels; TR: 6.985 msec; TE: 3.15 msec; field of view [FOV]: 240 mm × 256 mm × 180 mm; 180 slices) were collected for diagnostic and localization purposes. ¹H-MRS data were acquired using the single voxel PRESS (TE: 28 msec; TR: 2000 msec; 128 averages) to quantify brain Gln, Glu, and Glx levels and MEGA-PRESS (Mescher et al. 1998; Waddell et al. 2007) (TE: 68 msec; TR: 2000 msec; 16 averages × 20 dynamic scans) to quantify brain GABA levels. Voxels (PRESS: $20 \text{ mm} \times 20 \text{ mm} \times 20 \text{ mm}; \text{ MEGA-PRESS: } 30 \text{ mm} \times 30 \text{ mm} \times 20$ mm) were placed in the bilateral pregenual ACC by a trained spectroscopist (E.M.S.) (Fig. 1). Participants taking stimulant medication for ADHD were allowed to receive their medication on the day of the scan.

Neuroimaging data analysis

A board certified UMMS neuroradiologist read the anatomic MRI scans diagnostically, and no abnormalities were identified. The spectra acquired using PRESS were fit using LCModel (Version 6.2-4E) to quantify the metabolite concentrations (Provencher

1993, 2001). LCModel analyzes *in vivo* proton spectra as a linear combination of model *in vitro* spectra from individual metabolite solutions by utilizing built-in (simulated) radial basis sets. The basis set used for this study included alanine, aspartate, creatine (Cr), phosphocreatine (PCr), GABA, glucose, Glu, Gln, glycer-ophosphocholine (GPC), phosphocholine (PCh), glutathione (GSH), inositol (Ins), lactate, *N*-acetyl aspartate (NAA), *N*-acetyl aspartyl glutamate (NAAG), scyllo-inositol, taurine, total creatine (tCr: Cr and PCr), total choline (tCh: GPC, and PCh), total NAA (tNAA: NAA and NAAG), and Glx (Glu and Gln) as well as the macromolecules and lipids. Cramer–Rao lower bounds (CRLB, estimated error of the metabolite quantification) of >25% for Gln and >10% for all other metabolites were classified as not reliably detected and were excluded from further analysis.

Quantification of GABA concentrations was done using inhouse MATLAB (The Mathworks, MA) software for obtaining the ratios with respect to Cr levels. MEGA-PRESS enables us to detect the GABA peak of the spectra by eliminating the signal from Cr. Prior to Fourier transformation of the MRS signal, 3 Hz exponential line broadening was applied. The difference-edited GABA signal at 3 ppm and the Cr peak from unsuppressed PRESS signal were fit with Gaussian and Lorentzian models respectively (r > 0.95) and the area under these model peaks was calculated to obtain GABA to tCr (GABA/Cr) ratios (Edden and Barker 2007; Dydak et al. 2011; Zhu et al. 2011; Edden et al. 2014).

In addition, structural MRI scans were analyzed using Statistical Parameter Mapping (SPM8- http://www.fil.ion.ucl.ac.uk/spm/ software/spm8/); and white or gray matter or cerebrospinal fluid (CSF) in each MRS voxel were estimated using Matlab code (provided by Drs. Nia Goulden and Paul Mullins at Bangor University [Gasparovic et al. 2006], http://biu.bangor.ac.uk/projects .php.en).

Statistical analysis

Demographic and clinical characteristics of the participants were compared using parametric *t* tests. One participant with ASD had a CRLB for Gln > 25% and was excluded from further analysis. Data for each metabolite concentration were screened for values that were beyond two standard deviations of the respective group mean. No more than one value was excluded for any

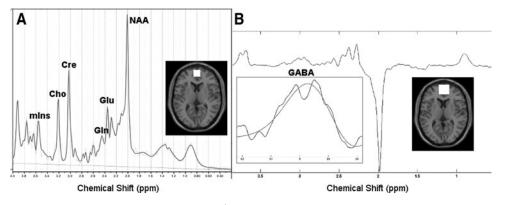


FIG. 1. Sample proton magnetic resonance spectroscopy (¹H-MRS) spectrum acquired using (**A**) Point resolved spectroscopy sequence (PRESS). (**B**) Mescher–Garwood point-resolved spectroscopy sequence (MEGA-PRESS) with sample voxel placements on the anterior cingulate cortex (ACC). (**A**) We investigated brain *N*-acetyl aspartate (NAA), glutamate (Glu), glutamine (Gln), creatine (Cr), choline (Cho), and myo-inositol (mIns) concentrations. Thick line represents the spectral fit. (**B**) Spectrum represents the case where editing pulse was on. The signal around the γ -aminobutyric acid (GABA) resonance was fit using a Gaussian function (see inside the square for a blown-up view of the signal with the fit).

metabolite. Analyses were performed using the R statistical programming language (R Core Team 2014) and the contributed packages "coin" (Hothorn et al. 2006, 2008) and "ggplot2" (Wickham 2009).

Comparisons of MRS values across groups and subsequent correlations of those values with clinical measures (without regard to group membership) were made with rank-based statistics: the Wilcoxon rank sum test with p value based on the exact distribution of the test statistic and Spearman rank-order correlations. Tabular and graphic displays of the data show parametric means and standard deviations. Nonparametric methods were used for betweengroup comparisons and correlations given the small sample size and non-normal distribution of data. Mean MRS values and associated standard deviations were similar to median values and median absolute deviation (MAD). The former are shown in the tables because of their familiarity. For all analyses, the threshold for statistical significance (α) was 0.05. A Bonferroni correction was applied for multiple comparisons of a priori hypothesized correlations of a subset of metabolites (Glu, Gln, GABA/Cr) with primary clinical measures of FSIQ, RMET score, and SRS Total Score. Exploratory correlations of these metabolites with subscales of the clinical measures were uncorrected.

Results

Demographics and clinical characteristics of the participants are listed in Table 1. The two groups did not differ significantly in age. Two participants were diagnosed with an anxiety disorder (one ASD, one TD), and seven were diagnosed with ADHD (six ASD, one TD). Three participants in the ASD group were currently taking stimulant medication (two methylphenidate extended release [ER], one lisdexamfetamine dimesylate), and no other participants were taking psychoactive medication. Mean estimated FSIQ was 103.1 ± 17.0 in the ASD group, and 121.6 ± 13.4 in the TD group

(p=0.006, Wilcoxon rank sum test), a clinically significant between-group difference. The ASD group demonstrated clinically significant lower accuracy on the RMET test and higher scores on the SRS compared to the TD group. All clinical scales except Somatic Complaints and Aggressive Behaviors on the CBCL were significantly higher in the ASD group, although the mean scores for all subscales remained below the threshold for clinical significance ($t \ge 70$). Seven individuals in the ASD group had clinically significant *t* scores in at least one subscale of the CBCL (two Attention Problems, two Withdrawn-Depressed, one Social Problems, one Thought Problems, one Anxious-Depressed/Thought Problems/ Attention Problems).

Table 2 lists the ¹H-MRS Glu, Gln, and GABA levels in the ACC in the two groups. There was no between-group difference in Glu levels. There were significantly higher Gln levels (p = 0.04), and a trend toward lower GABA/Cr levels (p = 0.09) in the ASD group (Table 2) in comparison with the TD group. There were no significant differences in levels of other metabolites (NAA, Cr, myoinositol [mIns], choline [Cho]) between groups (data not shown; p > 0.05). In addition, there was no significant difference between the two groups for voxel tissue content (i.e., gray matter, white matter, and CSF; p > 0.05).

The association between Glu, Gln, and GABA levels and IQ and social cognition measures was examined (see Table 3). The main analysis focused on the FSIQ, RMET score, and total SRS score. Correcting for multiple comparisons (p=0.0055 corresponds to α =0.05), there were significant negative correlations between Gln level and IQ (rho = -0.56, p=0.003, Fig. 2A) and between Gln level and RMET score (rho = -0.62, p=0.001, Fig. 2B). There was a trend toward significance (rho=0.53, p=0.007, Fig. 2C) for a positive correlation between Gln levels and SRS total score. There was a nonsignificant positive correlation between GABA/Cr levels and IQ (rho=0.36, p=0.07, Fig. 2D) and between GABA/Cr levels and RMET score (rho=0.42, p=0.034, Fig. 2E)

	$\begin{array}{l} ASD \ (n = 13) \\ Mean \ (SD) \end{array}$	TD (n = 14) Mean (SD)	p value
Age	14.9 (1.5)	14.7 (1.8)	0.51
FSIQ	103.1 (17.0)	121.6 (13.4)	0.006
ADHD (n)	6/13	1/14	
Anxiety disorder (<i>n</i>)	1/13	1/14	
Stimulant medication (<i>n</i>)	3/13	0/14	
RMET Score (% Correct)	0.556 (0.203)	0.736 (0.079)	0.025
SRS - Total Score	83.0 (29.8)	15.9 (9.6)	< 0.0001
SRS - Soc Awr	11.2 (3.6)	4.4 (2.0)	< 0.0001
SRS - Soc Cog	16.1 (7.1)	1.6 (1.5)	< 0.0001
SRS - Soc Comm	29.9 (12.3)	4.9 (3.9)	< 0.0001
SRS - Soc Mot	12.0 (5.3)	3.6 (3.1)	0.0001
SRS - Aut Man	13.8 (5.8)	1.4 (1.5)	< 0.0001
CBCL - Anxious/Depressed	57.8 (7.7)	51.3 (2.6)	0.0006
CBCL - Withdrawn/Depressed	59.3 (8.5)	52.4 (3.1)	0.006
CBCL - Somatic Complaints	55.2 (6.5)	52.4 (4.8)	0.17
CBCL - Social Problems	58.9 (6.4)	50.6 (2.1)	< 0.0001
CBCL - Thought Problems	59.8 (8.4)	50.7 (1.3)	0.0001
CBCL - Attention Problems	60.7 (7.0)	51.4 (2.1)	0.0001
CBCL - Rule-Breaking Behavior	55.1 (5.5)	50.9 (2.1)	0.0038
CBCL - Aggressive Behaviors	54.1 (6.3)	50.3 (0.5)	0.06

TABLE 1. CLINICAL CHARACTERISTICS OF STUDY PARTICIPANTS

ADHD, attention-deficit hyperactivity disorder; ASD, autism spectrum disorder; CBCL, Child Behavior Checklist; FSIQ, Full-Scale Intelligence Quotient; RMET, Reading the Mind in the Eyes Test; Soc Awr, Social Awareness; Soc Cog, Social Cognition; Soc Comm, Social Communication; Soc Mot, Social Motivation; Aut Man, Autistic Mannerisms; SRS, Social Responsiveness Scale; TD, typically developing.

TABLE 2. PROTON MAGNETIC RESONANCE SPECTROSCOPY				
METABOLITE LEVELS IN PREGENUAL MIDLINE ANTERIOR				
CINGULATE CORTEX VOXEL				

	$\begin{array}{l} ASD \ (n = 13) \\ Mean \ (SD) \end{array}$	TD (n=14) Mean (SD)	p value
Glu	10.6 (0.7)	10.6 (0.7)	0.76
Gln	4.1 (1.1)	3.4 (0.4)	0.04
GABA/Cr	0.075 (0.013)	0.083 (0.011)	0.09

ASD, autism spectrum disorder; GABA/Cr, γ -aminobutyric acid/creatine; Gln, glutamine; Glu, glutamate; TD, typically developing.

and a nonsignificant negative correlation between GABA/Cr levels and SRS total score (rho = -0.34, p = 0.09, Fig. 2F). There were no significant correlations or trends for Glu levels and clinical characteristics. Follow-up analyses were performed with Gln/Glu ratios, with similar but less significant group differences and correlations compared with analysis of Gln alone (data not shown).

In exploratory subanalyses (Table 3), the most significant correlations for the various SRS subscales were for the Social Communication scale with Gln and the Social Cognition scale with Gln and GABA/Cre (rho=0.58, p=0.002; rho=0.53, p=0.006; and rho = -0.42, p=0.03, respectively). For the CBCL subscale scores, there were positive correlations between Gln levels and scores in Social Problems (rho=0.55, p=0.005), Attention Problems (rho=0.55, p=0.005), Rule-Breaking Behavior (rho=0.50, p=0.01), Anxiety/Depression (rho=0.41, p=0.04), and Thought Problems (rho=0.42, p=0.04). There were trends toward negative correlations between GABA/Cr levels and Rule-Breaking Behavior (rho=-0.37, p=0.06), Thought Problems (rho=-0.34, p=0.09), and Attention Problems (rho=-0.34, p=0.09). Given the explor-

atory nature of these subanalyses, attempts were not made to correct *p* values for multiple comparisons.

Discussion

Adolescents with ASD had higher Gln levels in the pregenual ACC and a trend toward lower GABA/Cr levels than the TD control group. However, there was no difference in Glu levels between the two groups. Lower IQ and greater impairment in social cognition (lower RMET score, higher SRS score) were associated with higher Gln levels and lower GABA/Cre levels across groups (see Fig. 2).

Our findings support the growing literature indicating an imbalance in excitatory (glutamatergic) and inhibitory (GABA-ergic) neurotransmission in ASD. This is the first report to our knowledge suggesting that the increased glutamatergic activity may be best characterized spectroscopically by an increase in Gln levels rather than by Glu. Gln is involved in the Glu-Gln cycle, in which Glu released by neurons is taken up by astrocytes and then converted into Gln and stored. Gln is eventually returned to neurons where it is hydrolyzed to Glu (Bak et al. 2006; Schousboe and Waagepetersen 2006). Thus, Gln exists as an intracellular buffer to prevent the buildup of excessive glutamate. This, in addition to the large metabolic pool contributing to Glu levels, may contribute to Gln being a more sensitive marker of glutamatergic activity (Mlynarik et al. 2008; Pereira et al. 2008; Iltis et al. 2009). Elevated Gln may suggest an abnormality in glial cell number or function in ASD. Increased glial cell numbers and upregulation of glial cell gene expression have been demonstrated in postmortem brain tissue from individuals with ASD (Edmonson et al. 2014). ¹H-MRS studies in mood disorders have implicated the Gln/Glu ratio as a more accurate and sensitive marker of glutamatergic neurotransmission (Yuksel and Ongur 2010). However, in the current study, an analysis of Gln/ Glu ratios yielded similar but less significant results than the analysis

Spectroscopy Metabolites with Clinical Characteristics				
	Glu	Gln	GABA/Cr	
A priori predicted correlations				
FSIQ	0.02 (0.9)	-0.56 (0.003)**	0.36 (0.07)^	
RMET Score	-0.17(0.4)	-0.62 (0.0009)*	0.42 (0.03)*	
SRS - Total Score	0.13 (0.5)	0.53 (0.007)*	-0.34 (0.09)^	
Exploratory subanalyses				
SRS - Soc Awr	0.12 (0.6)	0.51 (0.01)*	-0.35 (0.08)^	
SRS - Soc Cog	0.11 (0.6)	0.53 (0.006)*	-0.42 (0.03)*	
SRS - Soc Comm	0.24 (0.2)	0.58 (0.002)*	-0.34 (0.09)^	
SRS - Soc Mot	0.14 (0.5)	0.45 (0.02)*	-0.24(0.2)	
SRS - Aut Man	-0.01(0.9)	0.38 (0.06)	-0.21(0.3)	
CBCL - Anxious/Depressed	0.07 (0.7)	0.41 (0.04)*	-0.23(0.3)	
CBCL - Withdrawn/Depressed	0.04 (0.8)	0.17 (0.4)	0.06 (0.8)	
CBCL - Somatic Complaints	0.26 (0.2)	0.10 (0.6)	0.10 (0.6)	
CBCL - Social Problems	0.15 (0.5)	0.55 (0.005)*	-0.32(0.1)	
CBCL - Thought Problems	0.10(0.6)	0.42 (0.04)*	-0.34 (0.09)^	
CBCL - Attention Problems	0.13 (0.5)	0.55 (0.005)*	-0.34 (0.09)^	
CBCL - Rule-breaking Behavior	-0.03 (0.9)	0.50 (0.01)*	-0.37 (0.06)^	
CBCL - Aggressive Behaviors	0.27 (0.2)	0.16 (0.4)	-0.13 (0.5)	

TABLE 3. SPEARMAN CORRELATION COEFFICIENTS FOR MAGNETIC RESONANCE SPECTROSCOPY METABOLITES WITH CLINICAL CHARACTERISTICS

Uncorrected p values in parentheses.

^Trend to significance at p < 0.1 level, uncorrected for multiple comparisons.

*Significant at p < 0.05 level, uncorrected for multiple comparisons.

**Significant at α <0.05 level, after Bonferroni correction for nine comparisons.

CBCL, Child Behavior Checklist; FSIQ, Full-Scale Intelligence Quotient; GABA/Cr, γ-aminobutyric acid/creatine; Gln, glutamine; Glu, glutamate; RMET, Reading the Mind in the Eyes Test; RMET, Reading the Mind in the Eyes Test; Soc Awr, Social Awareness; Soc Cog, Social Cognition; Soc Comm, Social Communication; Soc Mot, Social Motivation; Aut Man, Autistic Mannerisms;SRS, Social Responsiveness Scale.

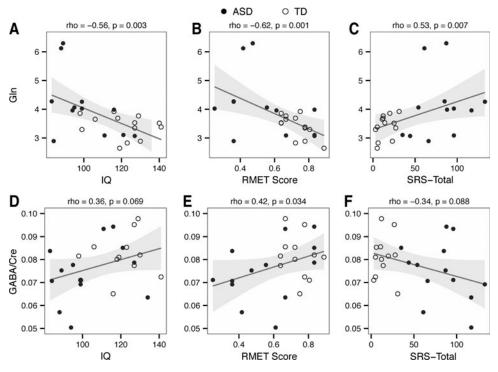


FIG. 2. (A–F) Bivariate scatterplots of metabolite values by clinical measures and associated Spearman rank order correlations and p values. A p value of 0.0055 corresponds to a Bonferroni corrected significance level (α) of 0.05. The line of linear best fit is shown, along with a shaded region representing the 95% confidence region for the slope of the line. For visualization purposes, data points are coded by group designation. (A) Glutamine (Gln) and intelligence quotient (IQ); (B) Gln and Reading the Mind in the Eyes Test (RMET) score (% Correct); (C) Gln and Social Responsiveness Scale (SRS)-Total score; (D) γ -aminobutyric acid/creatine (GABA/Cr) and IQ; (E) GABA/Cr and RMET score (% Correct); (F) GABA/Cr and SRS-Total score. ASD, autism spectrum disorder; TD, typically developing.

of Gln alone, likely because of a very high correlation between Gln/Glu and Gln (r=0.96, p<0.0001; data not shown).

Previous ¹H-MRS studies in ASD are equivocal in the direction of abnormality of glutamatergic compounds, particularly when measuring combined Glx compounds (Page et al. 2006; DeVito et al. 2007; Bernardi et al. 2011; Bejjani et al. 2012; Brown et al. 2013; Horder et al. 2013; Joshi et al. 2013). Differences may be associated with regional differences or age-related changes in Glx; however, the current study suggests that differences may also reflect a lack of sensitivity for combined Glx to adequately reflect glutamatergic neurotransmission, which may be better characterized by Gln levels. Joshi and colleagues (Joshi et al. 2013) reported increased Glu in the ACC in adolescents with ASD (similar age range to that in our study). However, limitations of the previous study include a smaller sample size (approximately half the size of our sample) and greater psychiatric comorbidity, including anxiety disorder, ADHD, mood disorder, or a combination. Also, 57% of the ASD group were taking antidepressant and/or antipsychotic medications, which are known to affect glutamatergic activity (Golembiowska and Dziubina 2000; Tarazi et al. 2003; Musazzi et al. 2010; Goto et al. 2012).

Only three previous studies have examined GABA levels in ASD using ¹H-MRS, and our findings support the previous results of decreased GABA levels in the frontal lobe (Harada et al. 2011), auditory (Gaetz et al. 2014; Rojas et al. 2014), and motor (Gaetz et al. 2014) cortices. Decreased cortical GABA levels appear to be the most consistent and reproducible ¹H-MRS finding in ASD.

Our study is the first to investigate concurrent Glu, Gln, and GABA levels in a cortical region known to be associated with social

cognitive deficits in ASD, and to correlate these levels with social cognition and IQ. Those individuals with lower IQ and greater social cognitive deficits tended to have higher Gln levels and lower GABA/Cr levels in the ACC. Exploratory analyses indicated that the correlation was most significant for SRS subscales measuring social cognition (Gln and GABA/Cr) and social communication (Gln). Therefore, these ¹H-MRS measures may provide a neuro-imaging biomarker for social cognition deficits in ASD.

There are several limitations to the current study. First, there were clinical differences between the two groups aside from the ASD characteristics (Table 1). Most problematic was the difference in IQ between the groups, a confounding limitation of this study. The significant group difference in IQ and the relatively small sample size precludes the ability to separate out the effect size of IQ and social cognition on the ¹H-MRS measures, as it would be inappropriate to include IQ as a covariate (see Miller and Chapman [2001] for a review of this misapplication of analysis of covariance). However, there is no prior literature suggesting a relationship between Gln and/or GABA levels and intelligence. Future studies are needed with a larger sample size in which groups are matched for IQ to determine whether the association between social cognition and ¹H-MRS measures remains significant.

Another difference between the groups was the comorbidity of ADHD. Almost half the ASD group had diagnoses of ADHD, but only three were taking stimulant medication on the day of the imaging acquisition. By contrast, in the TD group, one had a diagnosis of ADHD and none were taking medication. Prior studies have demonstrated decreased Glx (Perlov et al. 2007) and increased Glu (Hammerness et al. 2012) in the ACC of individuals with

ADHD, with evidence that treatment with stimulants normalizes the difference in Glu (Hammerness et al. 2012). Therefore, our ability to discern differences in Glu may have been hampered by the impact of the stimulant medications in the small subgroup. No ADHD studies have examined Gln or GABA in the ACC, although decreased GABA levels have been reported in somatosensory and motor cortices (Edden et al. 2012). The current study did not have sufficient sample size to statistically control for differences in medication or diagnosis between groups.

Despite excluding anyone with a psychiatric diagnosis other than ADHD or mild anxiety disorder, all of the clinical subscales on the CBCL were significantly higher in the ASD group. Seven individuals in the ASD group had at least one CBCL subscale score in the clinically significant range, although these score elevations could be attributed to ASD (Social Problems, Thought Problems, Withdrawn-Depressed), ADHD (Attention Problems), or Anxiety Disorder (Anxious-Depressed). For example, two individuals were clinically elevated on the Withdrawn-Depressed subscale, but did not meet diagnostic criteria for a depressive disorder. Correlations of ¹H-MRS parameters with CBCL subscale scores were much less significant than the correlations with social cognition measures, with the exception of a strong positive correlation between Gln and CBCL-Attention Problems (rho = 0.55, p = 0.005).

Another limitation in this study is the inherent variance associated with Gln measurements. For all reported peaks other than Gln, we included only the findings where CRLB was < 10%. For Gln, we included data when CRLB was < 25%; two thirds of the CRLB values were between 15% and 25%. Therefore, the values for Gln are not as precise as those of other ¹H-MRS parameters. However, the novel findings of differences in Gln associated with ASD and its correlation with social cognition deficits provide further impetus for the refinement of ¹H-MRS analysis techniques to better characterize Gln as a separate metabolite in future studies.

Conclusions and Clinical Significance

This study further highlights the potential importance of an imbalance in glutamatergic and GABA-ergic neurotransmission driving the social cognition deficits in ASD. The novel finding of a correlation of ¹H-MRS parameters with social cognition suggests that ¹H-MRS can be used to develop neuroimaging biomarkers of ASD as a measure of severity of the social deficits, although larger studies are needed that control for IQ, psychiatric diagnosis (particularly ADHD), and medication effects.

Our findings have interesting treatment implications for individuals with ASD. If further research supports the connection between excitatory-inhibitory imbalance and social cognitive deficits in ASD, then further development of medications that target glutamatergic and GABA-ergic function may provide treatments that impact the core social deficits of ASD. Likewise, repetitive transcranial magnetic stimulation (rTMS) has been postulated to address excitatory-inhibitory imbalance. A promising randomized controlled trial of rTMS in the bilateral dorsomedial prefrontal cortex in adults with ASD demonstrated a reduction in socialrelating impairment and socially related anxiety (Enticott et al. 2014). Importantly, treatment studies thus far have demonstrated response in only a subset of patients, and ¹H-MRS may provide a noninvasive method of investigating differences in treatment responders and nonresponders, thus providing information that may allow for the prediction of individualized treatment response.

The integration of ¹H-MRS methods into future clinical studies should be an ongoing area of research development, as the potential

for medical interventions targeting the core social communication deficits in ASD could revolutionize management of these chronic, debilitating disorders.

Disclosures

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References

- Achenbach TM, Rescorla L: The Manual for the ASEBA School-Age Forms and Profiles. Burlington (VT), University of Vermont, Research Center for Children, Youth & Families, 2001.
- Agam Y, Joseph RM, Barton JJ, Manoach DS: Reduced cognitive control of response inhibition by the anterior cingulate cortex in autism spectrum disorders. Neuroimage 52:336–347, 2010.
- American Psychiatric Association: Diagnostic and Statistical Manual of Mental Disorders, 4th ed. Washington, DC: American Psychiatric Association; 1994.
- American Psychiatric Association: Statistical Manual of Mental Disorders, 5th ed. Washington, DC: American Psychiatric Association; 2013.
- Autism and Developmental Disabilities Monitoring Network Surveillance Year 2008 Principal Investigators and Centers for Disease Control and Prevention: Prevalence of autism spectrum disorders autism and developmental disabilities monitoring network, 14 sites, United States, 2008. MMWR Surveill Summ 61:1–19, 2012.
- Bak LK, Schousboe A, Waagepetersen HS: The glutamate/GABA– glutamine cycle: Aspects of transport, neurotransmitter homeostasis and ammonia transfer. J Neurochem 98:641–653, 2006.
- Baron–Cohen S, Wheelwright S, Hill J, Raste Y, Plumb I: The "reading the mind in the eyes" test revised version: A study with normal adults, and adults with asperger syndrome or highfunctioning autism. J Child Psychol Psychiatry 42:241–251, 2001.
- Bejjani A, O'Neill J, Kim JA, Frew AJ, Yee VW, Ly R, Kitchen C, Salamon N, McCracken JT, Toga AW, Alger JR, Levitt JG: Elevated glutamatergic compounds in pregenual anterior cingulate in pediatric autism spectrum disorder demonstrated by 1H MRS and 1H MRSI. PLoS One 7: e38786, 2012.
- Bernardi S, Anagnostou E, Shen J, Kolevzon A, Buxbaum JD, Hollander E, Hof PR, Fan J: In vivo 1H-magnetic resonance spectroscopy study of the attentional networks in autism. Brain Res 1380:198–205, 2011.
- Berry–Kravis EM, Hessl D, Rathmell B, Zarevics P, Cherubini M, Walton-Bowen K, Mu Y, Nguyen DV, Gonzalez–Heydrich J, Wang PP, Carpenter RL, Bear MF, Hagerman RJ: Effects of STX209 (arbaclofen) on neurobehavioral function in children and adults with fragile X syndrome: A randomized, controlled, phase 2 trial. Sci Transl Med 4:152ra127, 2012.
- Blumberg SJ, Bramlett MD, Kogan MD, Schieve LA, Jones JR, Lu MC: Changes in prevalence of parent-reported autism spectrum disorders in school-aged U.S. children: 2007 to 2011–2012. National Health Statistics Reports No. 65; Hyattsville (MD), National Center for Health Statistics, 2013.
- Brown MS, Singel D, Hepburn S, Rojas DC: Increased glutamate concentration in the auditory cortex of persons with autism and first–degree relatives: A (1)H-MRS study. Autism Res 6:1–10, 2013.
- Constantino J, Gruber C: Social responsiveness scale (SRS). Los Angeles, Western Psychological Services, 2005.
- DeVito TJ, Drost DJ, Neufeld RW, Rajakumar N, Pavlosky W, Williamson P, Nicolson R: Evidence for cortical dysfunction in

autism: A proton magnetic resonance spectroscopic imaging study. Biol Psychiatry 61:465–473, 2007.

- Dydak U, Jiang YM, Long LL, Zhu H, Chen J, Li WM, Edden RA, Hu S, Fu X, Long Z, Mo XA, Meier D, Harezlak J, Aschner M, Murdoch JB, Zheng W: In vivo measurement of brain GABA concentrations by magnetic resonance spectroscopy in smelters occupationally exposed to manganese. Environ Health Perspect 119:219–224, 2011.
- Edden RA, Barker PB: Spatial effects in the detection of gammaaminobutyric acid: Improved sensitivity at high fields using inner volume saturation. Magn Reson Med 58:1276–1282, 2007.
- Edden RA, Crocetti D, Zhu H, Gilbert DL, Mostofsky SH: Reduced GABA concentration in attention-deficit/hyperactivity disorder. Arch Gen Psychiatry 69:750–753, 2012.
- Edden RAE, Puts NAJ, Harris AD, Barker PB, Evans CJ: Gannet: A batch-processing tool for the quantitative analysis of gammaaminobutyric acid?edited MR spectroscopy spectra. J Magn Reson Imaging 40:1445–1452, 2014.
- Edmonson C, Ziats MN, Rennert OM: Altered glial marker expression in autistic post-mortem prefrontal cortex and cerebellum. Mol Autism 5:3, 2014.
- Enticott PG, Fitzgibbon BM, Kennedy HA, Arnold SL, Elliot D, Peachey A, Zangen A, Fitzgerald PB: A double-blind, randomized trial of deep repetitive transcranial magnetic stimulation (rTMS) for autism spectrum disorder. Brain Stimul 7:206–211, 2014.
- Fatemi SH: The hyperglutamatergic hypothesis of autism. Prog Neuropsychopharmacol Biol Psychiatry 32:911–912, 2008.
- Fatemi SH, Folsom TD: Dysregulation of fragile x mental retardation protein and metabotropic glutamate receptor 5 in superior frontal cortex of individuals with autism: A postmortem brain study. Mol Autism 2:6, 2011.
- Fatemi SH, Folsom TD, Reutiman TJ, Thuras PD: Expression of GABA(B) receptors is altered in brains of subjects with autism. Cerebellum 8:64–69, 2009a.
- Fatemi SH, Reutiman TJ, Folsom TD, Thuras PD: GABA(A) receptor downregulation in brains of subjects with autism. J Autism Dev Disord 39:223–230, 2009b.
- Gaetz W, Bloy L, Wang DJ, Port RG, Blaskey L, Levy SE, Roberts TP: GABA estimation in the brains of children on the autism spectrum: Measurement precision and regional cortical variation. Neuroimage 86:1–9, 2014.
- Gasparovic C, Song T, Devier D, Bockholt HJ, Caprihan A, Mullins PG, Posse S, Jung RE, Morrison LA: Use of tissue water as a concentration reference for proton spectroscopic imaging. Magn Reson Med 55:1219–1226, 2006.
- Golembiowska K, Dziubina A: Effect of acute and chronic administration of citalopram on glutamate and aspartate release in the rat prefrontal cortex. Pol J Pharmacol 52:441–448, 2000.
- Goto N, Yoshimura R, Kakeda S, Nishimura J, Moriya J, Hayashi K, Katsuki A, Hori H, Umene–Nakano W, Ikenouchi–Sugita A, Korogi Y, Nakamura J: Six-month treatment with atypical antipsychotic drugs decreased frontal-lobe levels of glutamate plus glutamine in early-stage first-episode schizophrenia. Neuropsychiatr Dis Treat 8:119–122, 2012.
- Guastella AJ, Einfeld SL, Gray KM, Rinehart NJ, Tonge BJ, Lambert TJ, Hickie IB: Intranasal oxytocin improves emotion recognition for youth with autism spectrum disorders. Biol Psychiatry 67:692– 694, 2010.
- Hammerness P, Biederman J, Petty C, Henin A, Moore CM: Brain biochemical effects of methylphenidate treatment using proton magnetic spectroscopy in youth with attention-deficit hyperactivity disorder: A controlled pilot study. CNS Neurosci Ther 18:34–40, 2012.
- Harada M, Taki MM, Nose A, Kubo H, Mori K, Nishitani H, Matsuda T: Non-invasive evaluation of the GABAergic/glutamatergic sys-

tem in autistic patients observed by MEGA-editing proton MR spectroscopy using a clinical 3 tesla instrument. J Autism Dev Disord 41:447–454, 2011.

- Haznedar MM, Buchsbaum MS, Wei TC, Hof PR, Cartwright C, Bienstock CA, Hollander E: Limbic circuitry in patients with autism spectrum disorders studied with positron emission tomography and magnetic resonance imaging. Am J Psychiatry 157:1994–2001, 2000.
- Horder J, Lavender T, Mendez MA, O'Gorman R, Daly E, Craig MC, Lythgoe DJ, Barker GJ, Murphy DG: Reduced subcortical glutamate/glutamine in adults with autism spectrum disorders: A [(1)H]MRS study. Transl Psychiatry 3:e279, 2013.
- Hothorn K, Hornik K, van de Wiel MA, Zeileis A: Implementing a class of permutation tests: The coin package. J Stat Softw 28:1–23, 2008.
- Hothorn T, Hornik K, van de Wiel MA, Zeileis A: A lego system for conditional inference. Am Stat 60:257–263, 2006.
- Howlin P, Moss P, Savage S, Rutter M: Social outcomes in mid- to later adulthood among individuals diagnosed with autism and average nonverbal IQ as children. J Am Acad Child Adolesc Psychiatry 52:572–81.e1, 2013.
- Hyde KL, Samson F, Evans AC, Mottron L: Neuroanatomical differences in brain areas implicated in perceptual and other core features of autism revealed by cortical thickness analysis and voxelbased morphometry. Hum Brain Mapp 31:556–566, 2010.
- Iltis I, Koski DM, Eberly LE, Nelson CD, Deelchand DK, Valette J, Ugurbil K, Lim KO, Henry PG: Neurochemical changes in the rat prefrontal cortex following acute phencyclidine treatment: An in vivo localized (1)H MRS study. NMR Biomed 22:737–744, 2009.
- Jacquemont S, Curie A, des Portes V, Torrioli MG, Berry–Kravis E, Hagerman RJ, Ramos FJ, Cornish K, He Y, Paulding C, Neri G, Chen F, Hadjikhani N, Martinet D, Meyer J, Beckmann JS, Delange K, Brun A, Bussy G, Gasparini F, Hilse T, Floesser A, Branson J, Bilbe G, Johns D, Gomez–Mancilla B: Epigenetic modification of the FMR1 gene in fragile X syndrome is associated with differential response to the mGluR5 antagonist AFQ056. Sci Transl Med 3:64ra1, 2011.
- Joshi G, Biederman J, Wozniak J, Goldin RL, Crowley D, Furtak S, Lukas SE, Gonenc A: Magnetic resonance spectroscopy study of the glutamatergic system in adolescent males with high-functioning autistic disorder: A pilot study at 4T. Eur Arch Psychiatry Clin Neurosci 263:379–384, 2013.
- Magistretti PJ, Pellerin L: Cellular mechanisms of brain energy metabolism and their relevance to functional brain imaging. Philos Trans R Soc Lond B Biol Sci 354:1155–1163, 1999.
- Mescher M, Merkle H, Kirsch J, Garwood M, Gruetter R: Simultaneous in vivo spectral editing and water suppression. NMR Biomed 11:266–272, 1998.
- Miller GA, Chapman JP: Misunderstanding analysis of covariance. J Abnorm Psychol 110:40–48, 2001.
- Mlynarik V, Kohler I, Gambarota G, Vaslin A, Clarke PG, Gruetter R: Quantitative proton spectroscopic imaging of the neurochemical profile in rat brain with microliter resolution at ultra-short echo times. Magn Reson Med 59:52–58, 2008.
- Mundy P: Annotation: The neural basis of social impairments in autism: The role of the dorsal medial-frontal cortex and anterior cingulate system. J Child Psychol Psychiatry 44:793–809, 2003.
- Musazzi L, Milanese M, Farisello P, Zappettini S, Tardito D, Barbiero VS, Bonifacino T, Mallei A, Baldelli P, Racagni G, Raiteri M, Benfenati F, Bonanno G, Popoli M: Acute stress increases depolarizationevoked glutamate release in the rat prefrontal/frontal cortex: The dampening action of antidepressants. PLoS One 5:e8566, 2010.
- Oblak A, Gibbs TT, Blatt GJ: Decreased GABAA receptors and benzodiazepine binding sites in the anterior cingulate cortex in autism. Autism Res 2:205–219, 2009.

- Oblak AL, Gibbs TT, Blatt GJ: Decreased GABA(B) receptors in the cingulate cortex and fusiform gyrus in autism. J Neurochem 114:1414–1423, 2010.
- Ongur D, Jensen JE, Prescot AP, Stork C, Lundy M, Cohen BM, Renshaw PF: Abnormal glutamatergic neurotransmission and neuronal-glial interactions in acute mania. Biol Psychiatry 64:718– 726, 2008.
- Page LA, Daly E, Schmitz N, Simmons A, Toal F, Deeley Q, Ambery F, McAlonan GM, Murphy KC, Murphy DG: In vivo 1H-magnetic resonance spectroscopy study of amygdala-hippocampal and parietal regions in autism. Am J Psychiatry 163:2189–2192, 2006.
- Pereira FC, Rolo MR, Marques E, Mendes VM, Ribeiro CF, Ali SF, Morgadinho T, Macedo TR: Acute increase of the glutamate-glutamine cycling in discrete brain areas after administration of a single dose of amphetamine. Ann N Y Acad Sci 1139:212–221, 2008.
- Perlov E, Philipsen A, Hesslinger B, Buechert M, Ahrendts J, Feige B, Bubl E, Hennig J, Ebert D, Tebartz van Elst L: Reduced cingulate glutamate/glutamine-to-creatine ratios in adult patients with attention deficit/hyperactivity disorder — a magnet resonance spectroscopy study. J Psychiatr Res 41:934–941, 2007.
- Provencher SW: Automatic quantitation of localized in vivo 1H spectra with LCModel. NMR Biomed 14: 260–264, 2001.
- Provencher SW: Estimation of metabolite concentrations from localized in vivo proton NMR spectra. Magn Reson Med 30:672–679, 1993.
- Purcell AE, Jeon OH, Zimmerman AW, Blue ME, Pevsner J: Postmortem brain abnormalities of the glutamate neurotransmitter system in autism. Neurology 57:1618–1628, 2001.
- R Core Team: R: A Language and Environment for Statistical Computing. http://www.R-project.org/. Vienna, R Foundation for Statistical Computing, 2014.
- Rojas DC, Singel D, Steinmetz S, Hepburn S, Brown MS: Decreased left perisylvian GABA concentration in children with autism and unaffected siblings. Neuroimage 86:28–34, 2014.
- Schousboe A, Waagepetersen HS: Glial modulation of GABAergic and glutamatergic neurotransmission. Curr Top Med Chem 6:929– 934, 2006.
- Tarazi FI, Baldessarini RJ, Kula NS, Zhang K: Long-term effects of olanzapine, risperidone, and quetiapine on ionotropic glutamate

receptor types: Implications for antipsychotic drug treatment. J Pharmacol Exp Ther 306:1145–1151, 2003.

- Thakkar KN, Polli FE, Joseph RM, Tuch DS, Hadjikhani N, Barton JJ, Manoach DS: Response monitoring, repetitive behaviour and anterior cingulate abnormalities in autism spectrum disorders (ASD). Brain 131:2464–2478, 2008.
- Waddell KW, Avison MJ, Joers JM, Gore JC: A practical guide to robust detection of GABA in human brain by J-difference spectroscopy at 3 T using a standard volume coil. Magn Reson Imaging 25:1032–1038, 2007.
- Wang L, Leslie DL: Health care expenditures for children with autism spectrum disorders in Medicaid. J Am Acad Child Adolesc Psychiatry 49:1165–1171, 2010.
- Weschler D: Weschler Abbreviated Scale of Intelligence. San Antonio, NCS Pearson, 1999.
- Wickham H: Ggplot2: Elegant graphics for data analysis. New York, Springer, 2009.
- Yip J, Soghomonian JJ, Blatt GJ: Decreased GAD65 mRNA levels in select subpopulations of neurons in the cerebellar dentate nuclei in autism: An in situ hybridization study. Autism Res 2:50–59, 2009.
- Yip J, Soghomonian JJ, Blatt GJ: Decreased GAD67 mRNA levels in cerebellar purkinje cells in autism: Pathophysiological implications. Acta Neuropathol 113:559–568, 2007.
- Yuksel C, Ongur D: Magnetic resonance spectroscopy studies of glutamate-related abnormalities in mood disorders. Biol Psychiatry 68:785–794, 2010.
- Zhu H, Edden RA, Ouwerkerk R, Barker PB: High resolution spectroscopic imaging of GABA at 3 tesla. Magn Reson Med 65:603– 609, 2011.

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