REVIEW ARTICLE

Cell Therapy and Tissue Engineering Approaches for Cartilage Repair and/or Regeneration

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Articular cartilage injuries caused by traumatic, mechanical and/or by progressive degeneration result in pain, swelling, subsequent loss of joint function and finally osteoarthritis. Due to the peculiar structure of the tissue (no blood supply), chondrocytes, the unique cellular phenotype in cartilage, receive their nutrition through diffusion from the synovial fluid and this limits their intrinsic capacity for healing. The first cellular avenue explored for cartilage repair involved the in situ transplantation of isolated chondrocytes. Latterly, an improved alternative for the above reparative strategy involved the infusion of mesenchymal stem cells (MSC), which in addition to a self-renewal capacity exhibit a differentiation potential to chondrocytes, as well as a capability to produce a vast array of growth factors, cytokines and extracellular matrix compounds involved in cartilage development. In addition to the above and foremost reparative options up till now in use, other therapeutic options have been developed, comprising the design of biomaterial substrates (scaffolds) capable of sustaining MSC attachment, proliferation and differentiation. The implantation of these engineered platforms, closely to the site of cartilage damage, may well facilitate the initiation of an 'in situ' cartilage reparation process. In this mini-review, we examined the timely and conceptual development of several cell-based methods, designed to repair/regenerate a damaged cartilage. In addition to the above described cartilage reparative options, other therapeutic alternatives still in progress are portrayed.

Keywords: Cartilage damage, Repair/regeneration, Cell implantation, Biological scaffolds, Micro fracture, Novel cartilage restorative approaches

Cartilage: a peculiar type of connective tissue

Chondrocytes, the unique type of cell present in hyaline cartilage develop from the highly regulated differentiation of mesenchymal stem cells (MSC), a mesodermal-derived

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This is an open-access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/ licenses/by-nc/4.0/), which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited. stem cell present in several fetal and adult tissues. In the cartilage, chondrocytes are distributed either singularly or in clusters (recently divided) called isogenous groups. In these groups, chondrocytes are active in matrix production and display areas of cytoplasmic basophilia, which are indicative of protein synthesis and clear areas, which indicate their large Golgi apparatus (1-3).

The newly divided chondrocytes secrete the major macromolecules in cartilage matrix, including: a) collagen types II, IV, IX and XI, which are involved in the formation of a three-dimensional meshwork of the relatively thin and short matrix fibrils, and b) proteoglycans, which delineates the ground substance of hyaline cartilage, contains three types of glycosaminoglycan's (GAGs), including hyaluronans, chondroitin sulfate and keratin sulfate. The last two types are joined to a core protein to form a proteoglycan monomer, of which aggrecan is the most

significant. Other cartilage proteoglycans (decorin, biglycan, and fibromodulin), also play a role in matrix stabilization (2-5).

This unique structure, including cells and a vast network of cell regulators, gives cartilage a sort of compression rigidity and facilitates the entry of water. Thus, the tissue develops a peculiar power to absorb and dissipate tension forces constantly acting on the system bone. The high degree of hydration and the movement of water in the matrix allow the cartilage matrix to respond to varying pressure loads and contribute to cartilage's weight-bearing capacity.

Throughout life, cartilage undergoes continuous internal remodeling as the cells replace matrix molecules lost through degradation. The structure of the matrix is significant, since it acts as a signal transducer for the embedded chondrocytes. Thus, pressure loads applied to the cartilage as synovial joints create mechanical, electrical, and chemical signals that help to direct the synthetic activity of the chondrocytes (2, 3, 6).

However, as the body ages the composition of the matrix changes, and chondrocytes lose their ability to respond to these stimuli. In older chondrocytes, cytoplasmic changes are visible (shrinkage) resulting from the loss of lipid droplets and glycogen stores. In addition to ageing, there are other main ways that articular cartilage can be damaged: a sudden accidental injury, osteoarthritis, osteochondritis dissecans and infection. Conventionally, the clinical management of an osteochondral injury involves the use of mechanical symptomatic measures, in most cases associated to the use of analgesics, nonsteroidal anti-inflammatory drugs (NSAIDs) and/or chondro-protective agents (chondroitin sulfate, sulfate glucosamine, hyaluronic acid). Other protective options, involve the use of corticoids, HA, PRP, abrasion, micro fracture, radiofrequency and/or osteochondral grafts (6-8).

In most cases, the above therapeutic options are reparative but not renewable and associated to the creation of a more fibrous than hyaline tissue. However, the better understanding of the molecular, biological and structural components comprised in cartilage structure and function, has permitted the development of new prospects aimed to design biological therapies aimed to repair and/or regenerate a damaged cartilage (9, 10).

Cell-based therapies for the treatment of chondral lesions

Based on the better understanding of the cellular, molecular and micro environmental features of cartilage, it became evident that a damaged cartilage certainly could be a select target to develop tissue engineering procedures utilizing cell-based strategies. Most of these procedures have been designed to generate a neo-cartilage in an attempt to offer patients with chondral injuries, either an improvement in quality of life or a definitive cure.

Autologous chondrocytes implantation (ACI)

This early restorative procedures involves the arthroscopic procurement of a biopsy (8 mm) from the femoral groove of a healthy cartilage, area that normally is not subjected to load. Retrieved tissue is enzymatically treated to obtain a population of healthy isolated chondrocytes, which are then ex vivo expanded under conditions that preserve cell viability and function. The resulting population of chondrocytes is then injected under the periosteum, where they should grow and mature over time (11, 12).

Results from several clinical studies, including small cohorts of patients suffering diverse types of cartilage damage (early osteoarthritis, femoral condyle defects, knee joints defects, others), have shown that ACI treatment prompts pain reduction, improves quality of life and in many cases delay the need of joint replacement (13, 14). Despite these encouraging conclusions, the results of a similar but more comprehensive study revealed that effect (s) elicited by ACI are quite similar to those attained after osteochondral grafts treatment (mosaicplasty) (15).

Mesenchymal stem cells (MSC) implantation

Due to several cellular and molecular traits, mesenchymal stem cells (MSC) have been proposed to be an attractive candidate for cartilage repair. Among many others, MSC attributes comprise its abundance in various tissue sources (bone marrow, adipose tissue, umbilical cord blood, cord blood, others), self-renewal and a vast differentiation potential towards a chondrogenic. In addition, MSC produces a variety of extracellular matrix macromolecules involved in cartilage function, including collagen (s), fibronectin, glycosylaminoglycans (GAGs) and proteoglycans, as well as a vast repertoire of cytokines, growth factors, colony stimulating factors and chemokines (16, 17).

Based on the cellular and molecular features of MSC, biomedical actors developed the notion that the implantation of MSC may represent an appealing clinical alternative for regeneration of articular cartilage defects. Results of several clinical studies utilizing MSC for cartilage repair have evidenced that the procedure is feasible and safe. In addition, the intra-articular injection of MSC

proved to be effective in terms of reducing pain, improving tissue function and a robust capability to regenerate hyaline-like cartilage (18, 19).

Table 1 summarizes the outcomes of a group of comprehensive but not all-inclusive clinical studies, utilizing cell-based therapeutic approaches for the treatment of cartilage lesions. As compared to conventional procedures utilized to regenerate a damaged articular cartilage (micro-fracture, perforations, abrasion and/or mosaicplasty), it is without doubt that cell-based therapies, using either chondrocytes or MSC, represent an appealing curative alternative for cartilage regeneration.

However, there are several issues dealing with the isolation and manipulation of the 'curative' cell that require additional attentiveness. Specifically, in the case of MSC: a) selection of the most appropriate tissue source (s) (bone marrow, fat, umbilical cord blood, placenta, others), b) validation of a proper delivery route to the damaged tissue and c) a clear understanding that the curative effect of a MSC-based protocol, resides on the quality (biological attributes) and not necessarily on the quantity of the 'curative' cell (22). Accordingly, provisions should be taken during the ex-vivo processing of MSC (expansion), to protect stemness and avoid the expression of senescence-associated features (23, 24). As indicated in a recent publication, there are several challenges that must be overcome before MSC-based tissue engineering can become an effective cartilage regeneration therapy (17).

Biological scaffolds and its use in the treatment of chondral lesions

As indicated above, the limited cell proliferation and

differentiation capacity of chondrocytes in conjunction with a low production of cartilage-specific extracellular matrix have seriously limited their use in regenerative strategies. Accordingly, attempts have been oriented to develop other reparative options aimed to achieve an effective regeneration of hyaline cartilage.

The enhanced understanding of the molecular structure and functional role of extracellular matrix components in cartilage dynamics (25), encouraged the construction of sophisticated platforms (scaffolds) mimicking cartilage microenvironment. As a result, these prototypes seem to represent an appealing clinical device to be used in the treatment of chondral defects. To facilitate the binding of a cartilage-repair cell prototype (chondrocytes, MSC, others) these biodegradable scaffolds have been designed to include both a proximal cell-binding surface and a distal one to facilitate their loading in the proximity of a cartilage damage site (26).

The commercial availability of a vast array of these scaffolds, have prompted the initiation of a number of clinical studies (Table 2) to explore their use in the treatment of diverse cartilage lesions. The result of several preclinical and clinical studies put forward the notion that these elaborated structures embody a safe and promising clinical option for cartilage repair.

Nonetheless, in the case that these scaffolds turn out to be loaded with MSC entails the validation of several issues including the quality of the ex vivo expanded MSC (23), the assurance that mature and not a hypertrophic chondrocytes will be generated (17), and last but not least, the absence of ancillary factors that may alter MSC's cartilage repair potential (9).

Table 1. Clinical studies assessing the capability of cell-based therapies to the repair cartilage defects: an assortment of illustrative studies

| Cartilage lesion | Cell type ¹ | Number of patients ² | Most significant findings | Reference |
|---------------------|------------------------|---------------------------------|--|-----------|
| Knee articular | ACI | 431 | Mild or no effects | 15 |
| Knee osteoarhritis | MSC | 41 | Significant improvement in knee evaluation tests and MRI scores | 18 |
| Knee articular | ACI or MSC | 72 | Both cell types produce no significant differences in knee evaluation tests (IKDC, Lysholm, Tegner). However, patients receiving MSC, but not ACI, improve evaluation tests and require less surgery | |
| Knee osteoarthritis | MSC | 18 | No adverse events, improvement in knee evaluation tests, size of defect decreased, hyaline-like cartilage regeneration | 20 |
| Knee osteoarthritis | MSC | 6 | In 3/6 patients, cartilage thickness and knee evaluation tests improved (6 months); increase in extension of repair tissue; decrease in edematous subchondral patches | |

^{1:} ACI: chondrocyte; MSC: mesenchymal stem cells, 2: Include both treated and control patients.

| Scaffold and cell type used | Chondral lesion, number of patients and clinical outcome | Reference |
|--|--|-----------|
| Collagen I/III-based/bone marrow cells | Knee large lesions, 52/54 patients, after 1-5 year significant improvement in all knee functional scores | 27 |
| Hyaluronic acid/ chondrocytes | Knee, 141 patients, after an average follow up time (8 months) more than 70% of patients had no pain and mobility problems, histological analysis revealed hyaline-like cartilage, no side effects | 28 |
| Hyaluronic or Collagen-based/chondrocytes | In both groups (10 patients each), clinical outcome (24 months) was similar in MRI of cartilage repaired tissue, relaxation times for healthy surrounding cartilage and Zonal evaluation. However, global T2 was significantly higher in the hyaluronic group. Thus, functional outcome seems to be related to the type of scaffold used | 29 |

Table 2. Treatment of cartilage defects by using diverse types of biological scaffolds seeded with cartilage-repair cells

The use of cell-based therapies in conjunction with biological scaffolds to repair a damaged cartilage: new challenges to overcome

It is without doubt that the development of cell-based therapies aimed to repair a damaged cartilage, using either isolated chondrocytes or MSC as such or in conjunction with biological scaffolds has been an area of intensive clinical research in the last years. The clinical results of these studies, some of them depicted in Tables 1 and 2, have revealed that these procedures are feasible, safe and in most cases beneficial in the management of patients suffering chondral defects (13-15).

Simultaneously, several attempts have been initiated to further improve chondrogenic recovery, after the utilization of several types of cell-loaded biomaterial scaffolds. Among them, there are several efforts to improve ex vivo expansion of autologous chondrocytes (15) or intended to find alternative sources of MSC aimed to obtain cell products exhibiting a solid chondrogenic differentiation program with less hypertrophic differentiation (17, 22, 23).

The above, may well permit the development of protocols easily scalable, translatable to the clinic and capable of resisting variable biomechanical loading.

Despite the remarkable development of techniques intended to be utilized in chondrogenic cellular therapies, robust data still is lacking in terms of assuring the patient a proficient therapy capable of recapitulate hyaline cartilage tissue (9, 17, 25).

Recent biomedical advances in articular cartilage repair

Cartilage structure and function can be fairly harmed by a variety of causes resulting in injury, inflammation, pain, limited movement and significant joint damage and deformity. The therapeutic modalities previously examined sustain the regeneration of a damaged cartilage only in those conditions where the extent of chondral damage is limited and/or adjacent to a tissue region that preserves its full functional structure.

Most recently, a generation of innovative biomedical procedures has been explored in an attempt to treat not only focal defects, but even large-scale osteoarthritic degenerative changes. Among them, the following can be mentioned.

The 'one step' cell free cartilage reparative method

This novel and promising therapeutic option has matured from the expertise gained after the implementation of the so called 'two-steps' procedures, as shown in Tables 1 and 2. The 'one step' cell free reparative method starts with the direct implantation of a cell-free scaffold neighboring the site of the chondral lesion. The above procedure is followed by a mechanical arthroscopic maneuver intended to create a micro fracture within the bone underlying the damaged cartilage zone. The above procedure facilitates the in situ release, migration and attachment of endogenous MSC (the 'repair' cell) to the nearby implanted cell-free scaffold. This sort of combined reparative process represents an appealing therapeutic option for the treatment of small to medium-sized cartilage defects (30, 31).

Attempts to further improve the therapeutic capacity of MSC to play a part in cartilage repair

It is well known that the binding capacity of MSC to extracellular-like molecules (collagen I/II, hyaluronic acid, fibronectin, others) is dependent, among other factors, on the cellular expression of β 1-integrins, a type of transmembrane receptors (16). In addition, recent studies have shown that escalation of integrin expression facilitates the attachment of MSC to precise regions of a damaged cartilage (32). In the same vein, cellular studies have shown that the presence of L-Ascorbic acid 2-phosphate (a cul-

ture media components often utilized to grow MSC) modulates not only the differentiation of adipose-derived MSC to chondrocytes, but stimulate the production of chondrogenic growth factors (33).

Without a doubt, the translation to the clinic of specific cellular and molecular information may pave the way to the development of novel and possible more effective cell-based therapies aimed to repair osteochondral defects.

Multidisciplinary strategies associated to the development of innovative articular cartilage repair procedures

As discussed in previous sections, numerous attempts have been developed to repair focal chondral and/or osteochondral defects. However, major approaches aimed to repair large-scale osteoarthritic degenerative changes are still under development. In this respect, a number of preclinical studies have been initiated to investigate whether migratory progenitor cells and/or gene-based approaches may be valuable to repair major articular cartilage (34, 35).

Conclusions

Taken as a whole, both the current cartilage reparative options as well as several pre-clinical studies, still under development (36, 37), epitomize an evolving approach for the generation of therapies targeted to bring welfare to a large population of patients who suffer from articular cartilage damage. However, the biomedical efforts associated with the implementation of all these new medications, will bring together (as usual) new challenges linked to the safe and highly regulated translation of such procedures into the clinic (38).

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Potential conflict of interest

The authors report no financial or other conflict of interest relevant to the subject of this article.

References

- 1. Zhang Z. Chondrons and the pericellular matrix of chondrocytes. Tissue Eng Part B Rev 2014 [Epub ahead of print]
- 2. Dvir-Ginzberg M, Reich E. Chopping off the chondrocyte proteome. Biomarkers 2014:1-7 [Epub ahead of print]

- 3. Responte DJ, Natoli RM, Athanasiou KA. Collagens of articular cartilage: structure, function, and importance in tissue engineering. Crit Rev Biomed Eng 2007;35:363-411
- 4. Roughley PJ. The structure and function of cartilage proteoglycans. Eur Cell Mater 2006;12:92-101
- 5. Hollander AP, Dickinson SC, Kafienah W. Stem cells and cartilage development: complexities of a simple tissue. Stem Cells 2010;28:1992-1996
- 6. Cole BJ, Pascual-Garrido C, Grumet RC. Surgical management of articular cartilage defects in the knee. Instr Course Lect 2010;59:181-204
- 7. Kon E, Mandelbaum B, Buda R, Filardo G, Delcogliano M, Timoncini A, Fornasari PM, Giannini S, Marcacci M. Platelet-rich plasma intra-articular injection versus hyaluronic acid viscosupplementation as treatments for cartilage pathology: from early degeneration to osteoarthritis. Arthroscopy 2011;27:1490-1501
- 8. Rodríguez-Merchán EC. The treatment of cartilage defects in the knee joint: microfracture, mosaicplasty, and autologous chondrocyte implantation. Am J Orthop (Belle Mead NJ) 2012;41:236-239
- 9. Bhardwaj N, Devi D, Mandal BB. Tissue-engineered cartilage: the crossroads of biomaterials, cells and stimulating factors. Macromol Biosci 2015;15:153-182
- 10. Athanasiou KA, Responte DJ, Brown WE, Hu JC. Harnessing biomechanics to develop cartilage regeneration strategies. J Biomech Eng 2015;137:020901
- 11. Richardson JB, Caterson B, Evans EH, Ashton BA, Roberts S. Repair of human articular cartilage after implantation of autologous chondrocytes. J Bone Joint Surg Br 1999;81: 1064-1068
- 12. NICE updates guidance on the use of autologous chondrocyte implantation in treating cartilage defects in knee joints. 2005. Available from: www.nice.org.uk/guidance/ta89.
- 13. Minas T, Gomoll AH, Solhpour S, Rosenberger R, Probst C, Bryant T. Autologous chondrocyte implantation for joint preservation in patients with early osteoarthritis. Clin Orthop Relat Res 2010;468:147-157
- 14. Peterson L, Vasiliadis HS, Brittberg M, Lindahl A. Autologous chondrocyte implantation: a long-term follow-up. Am J Sports Med 2010;38:1117-1124
- 15. Vasiliadis HS, Wasiak J. Autologous chondrocyte implantation for full thickness articular cartilage defects of the knee. Cochrane Database Syst Rev 2010;(10):CD003323
- 16. Minguell JJ, Erices A, Conget P. Mesenchymal stem cells. Exp Biol Med (Maywood) 2001;226:507-520
- 17. Somoza RA, Welter JF, Correa D, Caplan AI. Chondrogenic differentiation of mesenchymal stem cells: challenges and unfulfilled expectations. Tissue Eng Part B Rev 2014;20: 596-608
- 18. Wakitani S, Okabe T, Horibe S, Mitsuoka T, Saito M, Koyama T, Nawata M, Tensho K, Kato H, Uematsu K, Kuroda R, Kurosaka M, Yoshiya S, Hattori K, Ohgushi H. Safety of autologous bone marrow-derived mesenchymal stem cell transplantation for cartilage repair in 41 patients with 45 joints followed for up to 11 years and 5 months.

- J Tissue Eng Regen Med 2011;5:146-150
- Nejadnik H, Hui JH, Feng Choong EP, Tai BC, Lee EH. Autologous bone marrow-derived mesenchymal stem cells versus autologous chondrocyte implantation: an observational cohort study. Am J Sports Med 2010;38:1110-1116
- 20. Jo CH, Lee YG, Shin WH, Kim H, Chai JW, Jeong EC, Kim JE, Shim H, Shin JS, Shin IS, Ra JC, Oh S, Yoon KS. Intra-articular injection of mesenchymal stem cells for the treatment of osteoarthritis of the knee: a proof-of-concept clinical trial. Stem Cells 2014;32:1254-1266
- Emadedin M, Aghdami N, Taghiyar L, Fazeli R, Moghadasali R, Jahangir S, Farjad R, Baghaban Eslaminejad M. Intra-articular injection of autologous mesenchymal stem cells in six patients with knee osteoarthritis. Arch Iran Med 2012;15:422-428
- 22. Minguell JJ, Allers C, Lasala GP. Mesenchymal stem cells and the treatment of conditions and diseases: the less glittering side of a conspicuous stem cell for basic research. Stem Cells Dev 2013;22:193-203
- Hoch AI, Leach JK. Concise review: optimizing expansion of bone marrow mesenchymal stem/stromal cells for clinical applications. Stem Cells Transl Med 2014;3:643-652
- Allers C, Lasala GP, Minguell JJ. Presence of osteoclast precursor cells during ex vivo expansion of bone marrow-derived mesenchymal stem cells for autologous use in cell therapy. Cytotherapy 2014;16:454-459
- Harvey A, Yen TY, Aizman I, Tate C, Case C. Proteomic analysis of the extracellular matrix produced by mesenchymal stromal cells: implications for cell therapy mechanism. PLoS One 2013;8:e79283
- Brittberg M. Cell carriers as the next generation of cell therapy for cartilage repair: a review of the matrix-induced autologous chondrocyte implantation procedure. Am J Sports Med 2010;38:1259-1271
- Skowroński J, Skowroński R, Rutka M. Large cartilage lesions of the knee treated with bone marrow concentrate and collagen membrane--results. Ortop Traumatol Rehabil 2013;15:69-76
- 28. Marcacci M, Berruto M, Brocchetta D, Delcogliano A, Ghinelli D, Gobbi A, Kon E, Pederzini L, Rosa D, Sacchetti GL, Stefani G, Zanasi S. Articular cartilage engineering with Hyalograft C: 3-year clinical results. Clin

- Orthop Relat Res 2005;(435):96-105
- 29. Welsch GH, Mamisch TC, Zak L, Blanke M, Olk A, Marlovits S, Trattnig S. Evaluation of cartilage repair tissue after matrix-associated autologous chondrocyte transplantation using a hyaluronic-based or a collagen-based scaffold with morphological MOCART scoring and biochemical T2 mapping: preliminary results. Am J Sports Med 2010;38:934-942
- 30. Freymann U, Petersen W, Kaps C. Cartilage regeneration revisited: entering of new one-step procedures for chondral cartilage repair. OA Orthopaedics 2013;1:6
- 31. Giannini S, Buda R, Battaglia M, Cavallo M, Ruffilli A, Ramponi L, Pagliazzi G, Vannini F. One-step repair in talar osteochondral lesions: 4-year clinical results and t2-mapping capability in outcome prediction. Am J Sports Med 2013;41:511-518
- Zwolanek D, Flicker M, Kirsta ter E, Zaucke F, van Osch GJVM, Erben RG. β1 integrins mediate attachment of mesenchymal stem cells to cartilage lesions. BioResearch 2015 [Epub ahead of print]
- Lee CS, Watkins E, Burnsed OA, Schwartz Z, Boyan BD. Tailoring adipose stem cell trophic factor production with differentiation medium components to regenerate chondral defects. Tissue Eng Part A 2013;19:1451-1464
- 34. Johnstone B, Alini M, Cucchiarini M, Dodge GR, Eglin D, Guilak F, Madry H, Mata A, Mauck RL, Semino CE, Stoddart MJ. Tissue engineering for articular cartilage repair--the state of the art. Eur Cell Mater 2013;25:248-267
- Schminke B, Miosge N. Cartilage repair in vivo: the role of migratory progenitor cells. Curr Rheumatol Rep 2014;16:461
- 36. Steinert AF, Nöth U, Tuan RS. Concepts in gene therapy for cartilage repair. Injury 2008;39 Suppl 1:S97-S113
- 37. Frisch J, Venkatesan JK, Rey-Rico A, Schmitt G, Madry H, Cucchiarini M. Determination of the chondrogenic differentiation processes in human bone marrow-derived mesenchymal stem cells genetically modified to overexpress transforming growth factor-β via recombinant adeno-associated viral vectors. Hum Gene Ther 2014;25:1050-1060
- Muthuswamy V. Ethical issues in clinical research. Perspect Clin Res 2013;4:9-13.