

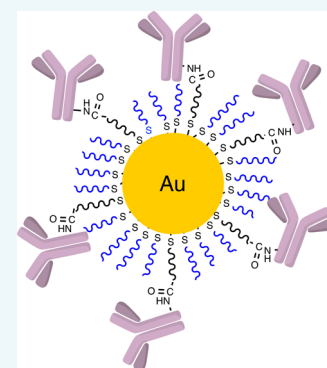
Lack of *In Vivo* Antibody Dependent Cellular Cytotoxicity with Antibody Containing Gold Nanoparticles

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S Supporting Information

ABSTRACT: Antibody-dependent cellular cytotoxicity (ADCC) is a cytolytic mechanism that can elicit *in vivo* antitumor effects and can play a significant role in the efficacy of antibody treatments for cancer. Here, we prepared cetuximab, panitumumab, and rituximab containing gold nanoparticles and investigated their ability to produce an ADCC effect *in vivo*. Cetuximab treatment of EGFR-expressing H1975 tumor xenografts showed significant tumor regression due to the ADCC activity of the antibody *in vivo*, while the control antibody, panitumumab, did not. However, all three antibody containing nanoparticles are not able to suppress tumor growth in the same *in vivo* mouse model. The antibody containing nanoparticles localized in the tumors and did not suppress the immune function of the animals, so the lack of tumor growth suppression of the cetuximab containing nanoparticle suggests that immobilizing antibodies onto a nanoparticle significantly decreases the ability of the antibody to promote an ADCC response.



INTRODUCTION

A number of targeted nanoparticles have now been investigated in human clinical trials.^{1,2} At this time, there is no clinical example of a full antibody targeted nanoparticle. Since immunotherapies are finding increased importance in cancer, the use of a full antibody targeted nanoparticle could be interesting. This type of therapeutic could potentially elicit immunotherapeutic functions such as antibody-dependent cellular cytotoxicity (ADCC) and complement dependent cytotoxicity (CDC) in addition to targeting the nanoparticles to cancer cell surface receptors and blocking cell signaling from those receptors. While antibody fragments can elicit the latter two functions, they do not stimulate immunotherapeutic pathways.

Numerous preclinical studies utilize full antibody targeted nanoparticles. However, only one investigation has specifically explored the possibility of stimulating an ADCC response.³ Rituximab is an IgG1 antibody that binds to the CD20 receptor, and rituximab containing lipid nanoparticles were investigated both *in vitro* and *in vivo* for their ability to elicit ADCC. Rituximab nanoparticles exhibited ADCC cell lysis *in vitro*, but the observed *in vivo* therapeutic efficacy of the antibody-lipid conjugates could not be ascribed to ADCC function.³

Natural killer (NK) cell based immunotherapies have shown considerable potential for cancer therapy in the clinic.^{4,5} ADCC is an immune mechanism dependent on the activity of CD56^{dim} CD16⁺ NK cells. Transgenic mouse models deficient in the CD16 receptor, also known as the activating Fcγ (FcγRIIIa/CD16) receptor, are unable to inhibit tumor growth in the presence of IgG1 antibodies that primarily work by inducing an ADCC response.⁶ Various types of peripheral blood mononuclear cells (PBMCs) have been studied for their antitumor

ADCC activities *in vitro*, and NK cells have been found to induce the most potent ADCC response.⁷

Cetuximab and panitumumab are two antibodies that specifically target the epidermal growth factor receptor I (EGFR) and possess similar EGFR binding affinities.^{8,9} In contrast to cetuximab, panitumumab is not able to elicit an ADCC response.¹⁰ Here, we address the question as to whether full antibodies that are displayed on the surface of nanoparticles can elicit an ADCC response *in vivo*. In order to observe antitumor effects that would be specific to an ADCC response, we selected a lung cancer cell line (H1975) that does not show any *in vitro* antiproliferative effects upon treatment with either cetuximab or panitumumab. Thus, any antitumor behavior observed *in vivo* can be ascribed to an ADCC function (positive for cetuximab and negative for panitumumab). Since gold nanoparticles will not have antitumor effects, antibody containing gold nanoparticles were prepared using cetuximab, panitumumab, and rituximab (negative control) and investigated *in vivo* with xenografts of the EGFR-expressing H1975 lung cancer cell line in athymic nude mice. While cetuximab alone reveals significant ADCC dependent antitumor behavior, the lack of antitumor function with the cetuximab containing gold nanoparticles shows that the ADCC function from antibody containing nanoparticles maybe be difficult to achieve *in vivo*.

RESULTS AND DISCUSSION

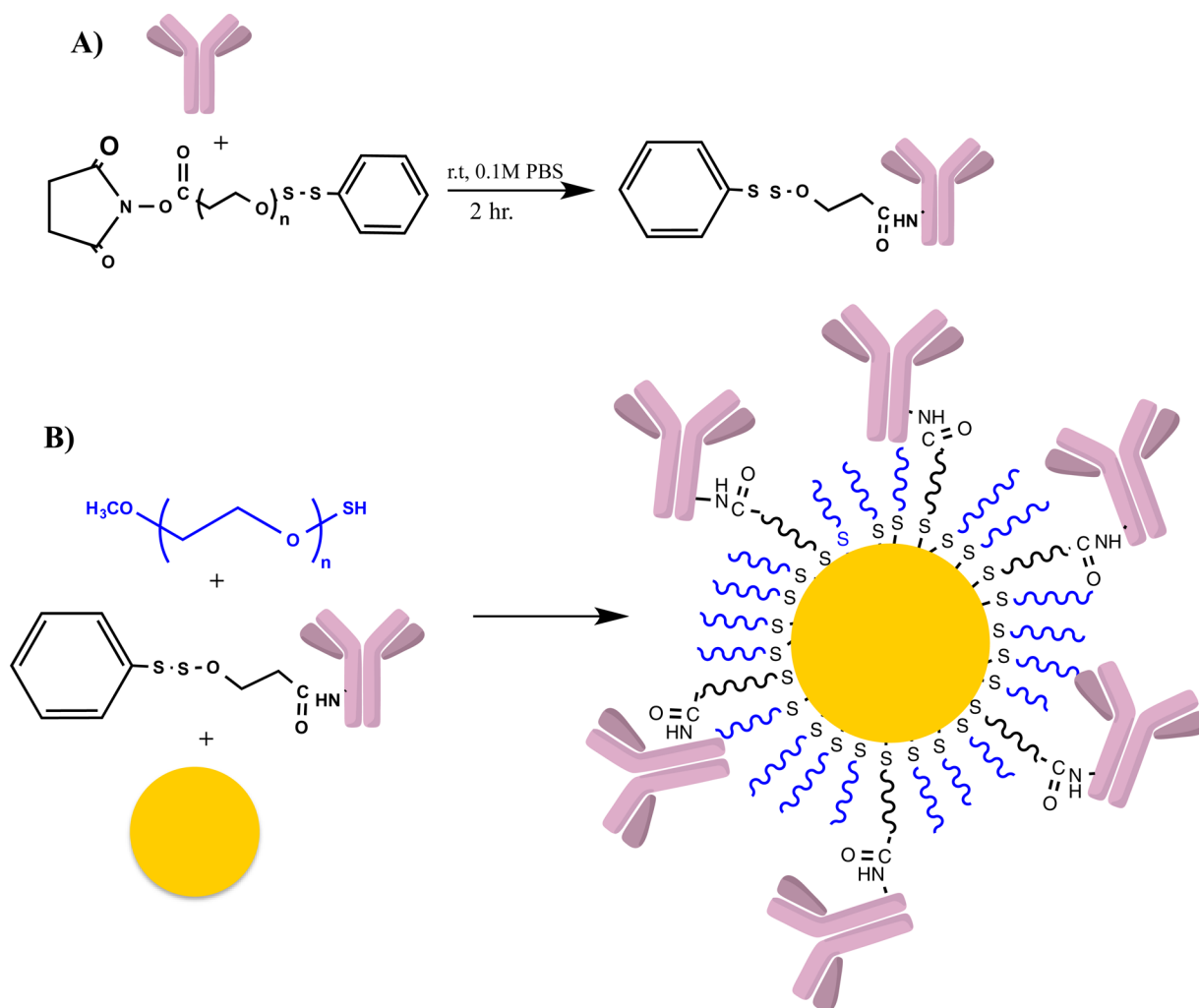
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The assembly of the antibody containing gold nanoparticles

Received: March 17, 2015

Revised: April 9, 2015

Published: April 16, 2015

Scheme 1. Assembly of Antibody Containing Gold Nanoparticles^a

^a(A) Antibodies first reacted with the NHS-PEG-OPSS and then purified. (B) In a second step, the antibody conjugates were then combined with mPEG-SH and assembled onto the surface of the gold nanoparticles.

Table 1. Properties of Antibody Containing AuNPs

sample	size (nm) by DLS ^a	size (nm) by NTA ^b	ζ potential (mV) ^c	no. of antibodies on surface
cetuximab-AuNPs	79.5 \pm 4	72.6 \pm 0.4	-18.7 \pm 1	19 \pm 6
panitumumab-AuNPs	72.8 \pm 1.9	74.8 \pm 1.6	-19.2 \pm 2.1	20 \pm 5
rituximab-AuNPs	82.5 \pm 5.5	73.2 \pm 1.3	-20.5 \pm 1.7	17 \pm 3
mPEG-AuNPs	64.1 \pm 9.5	72.3 \pm 0.9	-19.4 \pm 4.3	0

^aDLS (Dynamic Light Scattering), measured in phosphate buffered saline (PBS). ^bNTA (Nanoparticle Tracking Analysis), measured in deionized water. ^cMeasured in 1 mM KCl solution in deionized water.

was accomplished as follows. Conjugates of polyethylene glycol (PEG) and cetuximab, panitumumab, and rituximab were prepared by antibody reaction with NHS-PEG-OPSS (reacts with amine groups of antibodies to yield antibody-PEG conjugates through amide bond formation (Scheme 1)). High-pressure liquid-phase chromatography (HPLC) purified antibody-PEG conjugates were analyzed by MALDI-TOF-MS and confirmed to be mono-PEGylated. 50 nm gold nanoparticles (AuNPs) were then functionalized with the mono-PEGylated antibody-PEG conjugates and mPEG-SH (Scheme 1) and were analyzed for their average hydrodynamic diameter and surface charge (Table 1). The quantitative number of antibodies per nanoparticle was obtained using two different

methods. The results from the two were consistent with each other (Supporting Information, Table S1), and the mean values obtained from the two methods are presented in Table 1. PEGylated AuNPs containing approximately 15–20 antibodies per particle have negative ζ potential values and are stable in deionized water and saline solutions.

In Vitro Cytotoxicity of Antibodies and Antibody-Functionalized Gold Nanoparticles. The H1975 cell line harbors a double mutation in the kinase domain of EGFR that makes these cells insensitive to treatments with tyrosine kinase inhibitors (TKIs). Also, EGFR specific monoclonal antibodies do not produce antiproliferative effects with the H1975 cells in vitro.¹¹ The in vivo antitumor effects observed with cetuximab

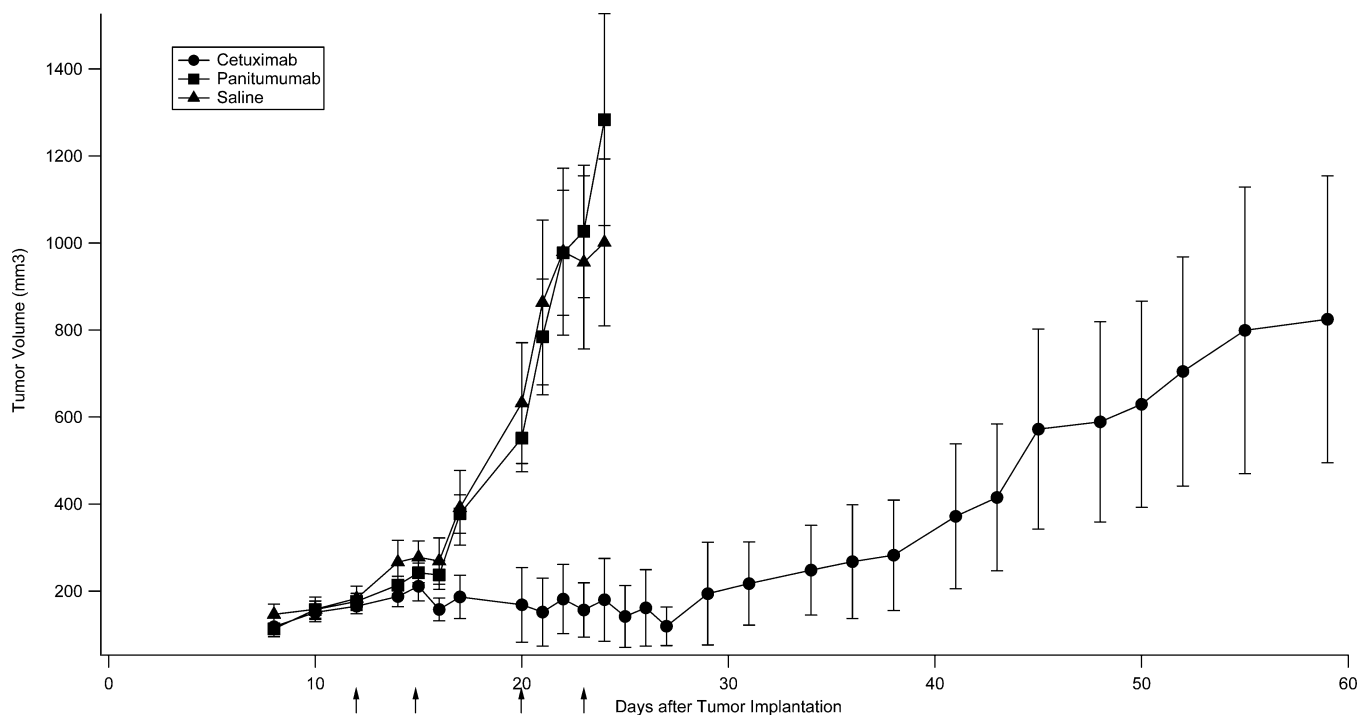


Figure 1. Effects of antibody treatment on H1975 xenograft tumors in nude mice. Mice were treated with cetuximab (circles), panitumumab (squares), or saline (triangles) as a negative control. Arrows indicate the days treatment occurred.

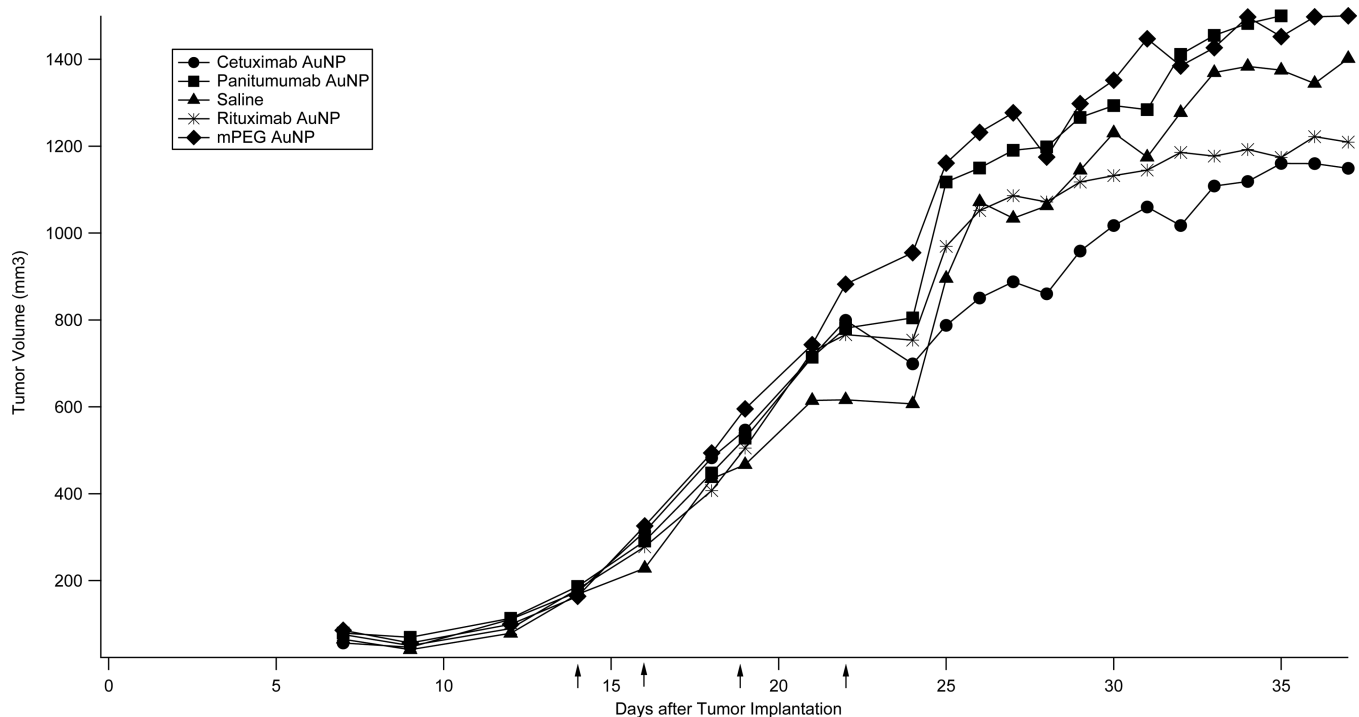


Figure 2. H1975 xenograft tumor growth in mice treated with antibody containing AuNPs. Nonantibody containing AuNPs, rituximab containing AuNPs, and saline were used as negative controls. Arrows indicate the days treatment occurred.

are thought to be associated with the ADCC activity of the antibody. Here, the antiproliferative effects of the antibodies cetuximab, panitumumab, and rituximab and the corresponding antibody containing AuNPs were investigated with the EGFR-TKI resistant H1975 lung cancer cell line via a cell viability assay. The H1975 cells remained viable in the presence of all antibodies and antibody containing AuNPs (Supporting

Information, Figure S1) 72 h post-treatment. The results are consistent with previous reports that show no *in vitro* cell death upon exposure of H1975 lung cancer cells to cetuximab or panitumumab¹⁰ and reveal that multiple antibodies on the surface of the AuNPs also do not produce any antiproliferative effects *in vitro*.

Antitumor Effects of Antibodies and Antibody Containing AuNPs. To investigate the role of ADCC *in vivo*, mice bearing established H1975 xenograft tumors were dosed twice a week for 2 weeks, and tumor growth was measured over a period of 2 months. Cetuximab treated mice showed significant tumor regression, while panitumumab treated mice did not (Figure 1). Thus, cetuximab treatments inhibit *in vivo* H1975 tumor growth, and these results are consistent with literature reports.

Antibody containing AuNPs were investigated for their antitumor activities in nude mice bearing H1975 tumor xenografts in order to compare to the results of treatments with the antibodies alone. The data illustrated in Figure 2 show that neither the cetuximab nor panitumumab containing AuNPs produced an antitumor effect *in vivo* (compared to saline, rituximab containing AuNPs, and AuNPs with no antibody (mPEG AuNP)). These results show that attaching the antibody to the surface of the AuNPs abolished cetuximab's ADCC activity *in vivo*.

Lack of ADCC with Cetuximab Containing AuNPs Is Not Due to Loss of Tumor Accumulation or Presence of Immune Cells. The strong tumor regression observed in cetuximab treated mice (Figure 1) is attributed to the ADCC activity of cetuximab. As expected, no antitumor effects are observed with panitumumab, which does not elicit an ADCC effect. This difference in the antitumor effect between the two antibodies was completely abolished when they were attached to the AuNPs (Figure 2). Unfortunately, we were not able to construct an *in vitro* model system for testing the ADCC activity of cetuximab or panitumumab. However, we were able to do so for trastuzumab using the BT474M1 cell line and immortalized NK cells (Supporting Information, Materials and Methods). These model data show that trastuzumab and trastuzumab AuNPs give an *in vitro* ADCC response (Supporting Information, Figure S2). Since we used the same conjugation chemistry with cetuximab and panitumumab as with trastuzumab, we assume that at least some fraction of the antibodies displayed on the surfaces of the nanoparticles are able to bind to their cancer cell surface receptors and stimulate an ADCC response *in vitro*. Thus, in order to better understand the factors that cause the *in vivo* loss in antitumor effects for the cetuximab that is contained on the AuNPs, we must first prove that the AuNPs are in fact reaching the tumors and that these tumors possess immune cells.

AuNPs in tissue can be visualized by silver staining and imaging by light microscopy. Images of silver stained tissues harvested from mice treated with antibody containing AuNPs (Supporting Information, Figure S3) show the presence of AuNPs in all treated tumor tissues, as well as in liver, spleen, and small amounts in kidney (nontumor organs were imaged to confirm that the AuNPs were reaching all organs expected with NPs). No staining was evident for tissues from mice treated with saline (negative control). Thus, the AuNPs are localizing to the tumors in these mice. The presence of immune cells within the tumor tissue was investigated by staining for CD45 (pan-leukocyte) and CD11b (NK) immune cell markers. In comparison to saline treated tumors, no significant difference in the presence of immune cells in tumor sections was found for antibody or antibody containing AuNP treated mice (Supporting Information, Figures S4 and S5).

Lack of ADCC with Cetuximab Containing AuNPs Is Due to Loss of ADCC Function. The *in vivo* antitumor data shown here reveal that an antibody that can elicit an ADCC

effect when used alone loses this function when it is conjugated onto a nanoparticle. Cetuximab injected at 9.3 mg/kg showed significant tumor regression due to an ADCC effect, while cetuximab containing AuNPs dosed at 0.44 mg of antibody/kg did not show any antitumor effects. It should be noted that cetuximab maintains antitumor activities in nude mice over the range of 0.4–40 mg/kg.¹²

The mice dosed with 0.44 mg/kg of cetuximab containing AuNPs (0.299 nmoles of gold/gram of body weight) showed significant accumulation of AuNPs in dermis, spleen, liver, and kidney, as revealed by sharp changes in skin color and silver staining of tissues from treated mice. Similar deposition of targeted and nontargeted PEGylated AuNPs in mouse dermis and lymph nodes was previously observed when studied at a much lower dose of AuNPs per mouse.¹³ Most importantly, the AuNPs also trafficked to the tumors (Supporting Information, Figure S3). Thus, the lack of antitumor effects with the cetuximab containing AuNPs is not because the NPs did not reach the tumor.

It has been argued that introducing NPs into animals can modify their immune response via secretion of anti- or proinflammatory cytokines, which in turn has an impact on immune cell surveillance in tumor tissues.^{14–16} Here, the presence of immune cells in tumor tissues was studied using antibodies against CD45 (for all leukocytes) and CD11b (for mouse NK cells). Immune cell surveillance in the tumors used here was not significantly different regardless of treatment (Supporting Information, Figures S4 and S5). CD45 and CD11b labeled immune cells were abundant in H1975 tumor xenografts, indicating that the inability of the cetuximab containing AuNPs to show an antitumor effect *in vivo* was not due to a suppression of the animal's immune response.

The results presented here lead to the conclusion that covalent immobilization of an antibody on a nanoparticle surface can eliminate the ADCC function of the antibody. Here, we conjugated the antibody to PEG via reactions with amines on the antibody. Because of the presence of several amines on the antibody, the conjugation of the PEG component will occur at various sites on the antibody. The antibody–PEG conjugates were purified by HPLC to obtain a mono-PEGylated fraction for attachment to the nanoparticle surface. However, the orientation and distribution of antibodies on the nanoparticle surface, such as whether the Fc region is accessible to immune cell binding may play an important role in eliminating the ADCC effects *in vivo*. Results from a model *in vitro* system suggest that at least a fraction of the antibodies that are contained on the surfaces of the gold nanoparticles can bind to the appropriate cancer cell surface receptor and stimulate an *in vitro* ADCC response. Thus, at this time, the mechanistic origin of the *in vivo* loss in ADCC with nanoparticle containing antibodies remains unknown. The ability to perform site-specific conjugation to the antibody like what is presently done with antibody–drug conjugates¹⁷ may help resolve some of these issues.

■ ASSOCIATED CONTENT

📄 Supporting Information

Materials and instrumentation, experimental procedures, and analytical data. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

We thank Kristin Anderson for performing MTS cell viability assays, Mona Shahgholi for MALDI-TOF-MS training, and Andres Collazo (Beckman Imaging Center) for confocal microscopy training. This work was supported by the National Cancer Institute Grant CA 151819 and the National Science and Engineering Research Council of Canada (NSERC) (fellowship for M.A.).

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