



Seroepidemiology of Toxocariasis and Its Clinical Implications in Gwangju and Jeonnam-province, Korea

Eun Jeong Won, M.D.^{1,2}, Jin Kim, M.D.³, Myung-Geun Shin, M.D.², Jong Hee Shin, M.D.¹, Soon Pal Suh, M.D.¹, and Dong Wook Ryang, M.D.¹

Department of Laboratory Medicine¹, Chonnam National University Hospital, Gwangju; Department of Laboratory Medicine², Chonnam National University Hwasun Hospital, Hwasun; Department of Parasitology³, College of Medicine, Seonam University, Namwon, Korea

We investigated the seroepidemiological, clinical, and laboratory characteristics of patients suspected to have toxocariasis in Gwangju and Jeonnam-province, Korea. In total, 228 specimens were analyzed for anti-*Toxocara canis* IgG at two university hospitals from 2010 to 2012. The overall seropositive rate was 67.1%, and the seropositive rates among the eosinophilic and non-eosinophilic groups were 76.1% (105/138) and 53.3% (48/90), respectively. Risk factors for eosinophilia and toxocariasis were male sex (odds ratios [OR]=2.632 and 3.477, respectively) and a history of ingesting raw meat (OR=2.884 and 3.274, respectively), especially raw cow liver (OR=2.089 and 10.038, respectively). *T. canis* seropositivity (OR=5.807, $P=0.004$) and a history of consuming raw cow liver (OR=2.766, $P=0.052$) were risk factors for organ involvement. The anti-*T. canis* IgG level showed weakly positive correlations with eosinophil counts ($r=0.234$, $P<0.001$) and the duration of eosinophilia ($r=0.155$, $P=0.019$). Although limited to the regions of Gwangju and Jeonnam-province, this study supports the opinion that toxocariasis is a reasonable focus as a cause of eosinophilia and that it is also associated with organ involvement.

Key Words: Toxocariasis, Raw cow liver, Organ involvement, Eosinophilia, Seroepidemiology

Received: October 16, 2014

Revision received: December 17, 2014

Accepted: April 1, 2015

Corresponding author: Dong Wook Ryang
Department of Laboratory Medicine,
Chonnam National University Medical
School, 671 Jebong-ro, Dong-gu, Gwangju
501-757, Korea
Tel: +82-62-220-5340
Fax: +82-62-224-2518
E-mail: dwryang@chonnam.ac.kr

© The Korean Society for Laboratory Medicine

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Eosinophilia, defined as an absolute eosinophil count of $>0.45 \times 10^9/L$ in peripheral blood, occurs in allergic diseases, parasitic infections, and cancer [1]. Recently, toxocariasis has emerged as a major cause of eosinophilia and induced eosinophilic infiltration in internal organs [2]. Toxocariasis caused by infection with *Toxocara canis* larvae occurs by accidental ingestion of embryonated eggs or larvae from a range of wild and domestic animals [3]. Many epidemiological studies have been performed worldwide, but most have focused on schoolchildren who are at higher risk of infection because of their play habits and typically poor hygiene [4-6]. Although less common, humans can also become infected if they eat undercooked meat from an animal infected with *T. canis* larvae [7]. Especially in Korea, it is thought that toxocariasis is likely to be more prevalent owing to the habitual intake of raw liver [2, 8, 9]. Although the epidemiology of toxocariasis is affected by regional culture, the epidemiology of

toxocariasis in the regions of Gwangju and Jeonnam-province has not been evaluated. We investigated the seroepidemiological, clinical, and laboratory characteristics of patients suspected to have toxocariasis in Gwangju and Jeonnam-province, Korea.

The medical records of 228 patients whose specimens were submitted for anti-*T. canis* IgG testing were analyzed retrospectively at Chonnam National University Hospital and Chonnam National University Hwasun Hospital from 2010 to 2012. Levels of specific IgG to *Toxocara* excretory/secretory antigen were measured by using *T. canis* ELISA kit (Bordier Affinity Products, Crissier, Switzerland) [10]. The investigated variables were age, sex, laboratory parameters such as complete blood count, liver function tests, renal function tests, total IgE, eosinophil cationic protein (ECP) levels, and history or presence of i) asthma or rhinitis, ii) cancer, iii) drug use, iv) freshwater fish, raw meat, or raw cow liver consumption, v) organ involvement (liver or lung),

and vi) other parasitic infections (clonorchiasis, paragonimiasis, cysticercosis, or sparganosis).

Chi-square or Fisher exact test was performed to determine the distributions of categorical variables, including sex and other risk factors between groups (non-eosinophilic vs. eosinophilic group; seronegative vs. seropositive group; and organ involvement (-) vs. organ involvement (+) group). Student t-tests were used to compare continuous variables, such as age and laboratory parameters. The likelihood-ratio chi-square was used to calculate the odds ratio (OR) for eosinophilia, seropositivity, and organ involvement. Logistic regression analysis was used for multivariate analysis. Spearman correlation coefficients were used to examine relationships between the optical density levels of anti-*T. canis* IgG and the eosinophil counts or duration of eosinophilia. The correlation coefficients (*r*-values) were interpreted by Dancey and Reidy's categorization [11]. Here, *r*-value of ± 1 is interpreted as a perfect correlation; ± 0.7 to ± 0.9 as strong; ± 0.4 to ± 0.6 as moderate; ± 0.1 to ± 0.3 as weak correlation; and an *r*-value of 0 as zero correlation, implying no correlation. *P* value < 0.05 indicated significance for all analyses. All statistical analyses were performed by using PASW version 18.0 (SPSS Inc., Chicago, IL, USA).

Tables 1 and 2 describe the clinical characteristics and their

risk stratifications according to eosinophilia and toxocariasis seropositivity. Male sex was predominant in both eosinophilic and seropositive groups (72.5% and 73.2%, both $P < 0.001$). The general seroprevalence was 67.1% (153/228), and the seropositive rates in the eosinophilic and non-eosinophilic groups were 76.1% and 53.3%, respectively. Raw meat or raw cow liver ingestion occurred more frequently in the eosinophilic group (OR=2.884 and 2.089, respectively) and the seropositive group (OR=3.274 and 10.038, respectively) than in their counterparts. Other causes, including histories of other parasitic infections, asthma or allergic rhinitis, cancer, drug use, and fresh water fish ingestion, were not significantly related to eosinophilia or seropositivity. Of the 228 patients, 23 showed organ involvement findings in radiological evaluations. The presence of organ involvement was associated with seropositivity (OR=5.807) and a history of consuming raw cow liver (OR=2.766; Table 3). Multivariate analysis revealed that male and seropositivity were the independent risk factors for eosinophilia, male and the history consuming raw cow liver for seropositivity, and seropositivity for organ involvement, respectively. Patients with organ involvement had higher eosinophil levels and white blood cell counts ($P < 0.001$, both) than patients without organ involvement. Spearman correlation analysis indicated that the optical density levels

Table 1. Clinical characteristics of patients and risk stratifications according to eosinophilia

	Non-eosinophilic group (N=90)	Eosinophilic group (N=138)	<i>P</i> value	Risk stratification for eosinophilia*			
				Univariate analysis		Multivariate analysis	
				Odd ratio (95% CI)	<i>P</i> value	Odd ratio (95% CI)	<i>P</i> value
Age (yr)			0.275				
Median (range)	62 (18-84)	64 (16-92)					
Sex, N (%)			< 0.001		0.001		0.014
Female	45 (50.0)	38 (27.5)		2.632		2.093	
Male	45 (50.0)	100 (72.5)		(1.508-4.593)		(1.162-3.769)	
Presence of <i>Toxocara canis</i> IgG, N (%)			0.001		< 0.001		0.015
No	42 (46.7)	33 (23.9)		2.784		2.136	
Yes	48 (53.3)	105 (76.1)		(1.575-4.921)		(1.162-3.925)	
History of ingestion of raw meat, N (%)			0.021		0.011		
No or unknown	83 (92.2)	111 (80.4)		2.884			
Yes	7 (7.8)	27 (19.6)		(1.198-6.944)			
History of ingestion of raw cow liver, N (%)			0.09		0.064		
No or unknown	81 (90.0)	112 (81.2)		2.089			
Yes	9 (10.0)	26 (18.8)		(0.929-4.697)			

*The risk stratifications for the eosinophilia were performed by using likelihood-ratio chi-square for univariate analysis and logistic regression for multivariate analysis, respectively.

Abbreviation: CI, confidence interval.

Table 2. Clinical characteristics of patients and risk stratifications according to seropositivity for toxocariasis

	Sero-negative group (N = 75)	Sero-positive group (N = 153)	P value	Risk stratification for seropositivity*			
				Univariate analysis		Multivariate analysis	
				Odd ratio (95% CI)	P value	Odd ratio (95% CI)	P value
Age (yr)			0.089				
Median (range)	62 (18-84)	62 (16-92)					
Sex, N (%)			<0.001		<0.001		<0.001
Female	42 (56.0)	41 (26.8)		3.477		3.021	
Male	33 (44.0)	112 (73.2)		(1.948-6.207)		(1.665-5.483)	
History of ingestion of raw meat, N (%)			0.017		0.009		
No or unknown	70 (93.3)	124 (81.0)		3.274			
Yes	5 (6.7)	29 (19.0)		(1.213-8.841)			
History of ingestion of raw cow liver, N (%)			<0.001		<0.001		0.014
No or unknown	73 (97.3)	120 (78.4)		10.038		6.829	
Yes	2 (2.7)	33 (21.6)		(2.339-43.076)		(1.479-31.54)	

*The risk stratifications for the seropositivity for toxocariasis were performed using likelihood-ratio chi-square for univariate analysis and logistic regression for multivariate analysis, respectively.
Abbreviation: CI, confidence interval.

Table 3. Analysis of clinical and laboratory characteristics for the risk of organ involvement

	Organ involvement (-) (N = 205)	Organ involvement (+) (N = 23)	P value	Risk stratification for organ involvement*			
				Univariate analysis		Multivariate analysis	
				Odd ratio (95% CI)	P value	Odd ratio (95% CI)	P value
Age (yr)							
Median (range)	62 (16-92)	67 (40-83)	0.329				
Sex, N (%)			0.17				
Female	78 (38.0)	5 (21.7)					
Male	127 (62.0)	18 (78.3)					
anti- <i>Toxocara canis</i> IgG, Mean ± SD	18.7 ± 22.0	28.0 ± 30.2	0.069				
WBC count, 10 ⁹ /L, Mean ± SD	7.9 ± 4.2	11.6 ± 7.0	<0.001				
Eosinophils, 10 ³ /L, Mean ± SD	1,080.7 ± 2,434.1	4,030 ± 5,946.8	<0.001				
Presence of <i>Toxocara canis</i> IgG, N (%)			0.009		0.004		0.038
No	73 (35.6)	2 (8.7)		5.807		4.906	
Yes	132 (64.4)	21 (91.3)		(1.324-25.466)		(1.091-22.07)	
History of ingestion of raw meat, N (%)			0.125		0.139		
No or unknown	177 (86.3)	17 (73.9)		2.231			
Yes	28 (13.7)	6 (26.1)		(0.811-6.141)			
History of ingestion of raw cow liver, N (%)			0.06		0.052		
No or unknown	177 (86.3)	16 (69.6)		2.766			
Yes	28 (13.7)	7 (30.4)		(1.045-7.321)			

*The risk stratifications for the organ involvement were performed using likelihood-ratio chi-square for univariate analysis and logistic regression for multivariate analysis, respectively.
Abbreviation: CI, confidence interval.

of IgG showed a weak correlation with eosinophil counts ($r=0.234$, $P<0.001$) and the duration of eosinophilia ($r=0.155$, $P=0.019$; Supplemental Data Figure S1). The eosinophil counts alone also showed a moderate correlation with the duration of

eosinophilia ($r=0.585$, $P<0.001$; data not shown).

Toxocariasis has been recognized as the most commonly neglected parasitic infection in the United States, and its global importance might be greatly underestimated [12]. Although a nationwide survey has not been performed in Korea, regional studies enable estimates of the prevalence of toxocariasis as follows (Supplemental Data Table S1): 86.7% (Seoul), 68.0% (Seoul), 62.1% (Pohang), and 45.5% (Chungnam-province) [2, 8, 9, 13]. A previous study reported that the seroprevalence in healthy rural Korean adults might be around 5% [14], but this study showed 53.3% seroprevalence in the non-eosinophilic group. This might be because many healthy subjects display residual antibodies and the IgG assay cannot rule out past infection within the non-eosinophilic group. Thus, specific IgE for *T. canis* would be useful for differentiating acute and past infections, although it is not widely used in clinical laboratories because a commercial kit is not yet available [15]. The cross-reactivity to other helminthic infections should be considered a factor for high seroprevalence in this study [10]. We could not find any evidence of filariasis in this study but this is limited owing to the retrospective nature of our study. Although filariasis is well controlled in Korea [16], the potential cross-reactivity should be considered in the clinical use of this ELISA kit.

It is noteworthy that seropositivity may be an independent risk factor for eosinophilia. Considering the high prevalence of toxocariasis in eosinophilic patients, a diagnostic approach to toxocariasis should be performed with a view to diagnose idiopathic hypereosinophilic syndrome [17]. Male sex is considered a risk factor for both eosinophilia and toxocariasis; this is likely because of the habit of ingesting raw meat [2]. This study also showed that the ingestion of raw cow liver substantially increased the risk of toxocariasis, more than 10 times, which is in line with other studies [2, 8, 9]. A recent meta-analysis found that a higher prevalence of *T. canis* infection was associated with asthma [18], but we could not find significant association between seropositivity and asthma. In addition to asthma, other causes of eosinophilia such as other allergic diseases, underlying cancer, and other parasitic infections should also be ruled out for the evaluation of eosinophilia. We found that 57% (25/44) of eosinophilic patients, seronegative for toxocariasis, had evidence of allergic diseases, underlying cancer, or other parasitic infections.

Almost all patients with organ involvement were seropositive for toxocariasis, except for two patients diagnosed as having paragonimiasis and the Drug Reaction with Eosinophilia and Systemic Symptom syndrome. This was suggestive of the phe-

nomenon of visceral larva migrans, because the lesions resolved or moved on follow-up imaging. These findings highlight that clinicians should be aware of toxocariasis serology in eosinophilic patients with organ involvement [19]. High anti-*T. canis* IgG levels also correlated with high eosinophil counts and persistent eosinophilia. This supports the need for proper treatment of toxocariasis to control the levels of anti-*T. canis* IgG associated with eosinophilia-related symptoms in these patients. In addition, anti-*T. canis* IgG levels seemed to diminish after six months when we followed up serially. Especially in two patients, seroconversion from positive to gray zone occurred after 21 months (data not shown). Although the duration of seropositivity for *T. canis* is not well known, serial follow-up would be recommended with an interval of more than six months.

This study had some limitations. The clinical, laboratory parameters might not have been fully investigated in all patients, because this study was performed retrospectively. ECP and total IgE levels have been thought to be markers for toxocariasis [20], but we did not demonstrate correlations with eosinophilia or toxocariasis. In particular, 120 of 153 seropositive patients had no evidence of raw cow liver consumption. It seems likely that history taking for risk factors may have been performed incompletely. Thus, thorough evaluation with laboratory parameters and history taking should be done when toxocariasis is suspected. Regrettably, the current data may not reflect the seroprevalence of toxocariasis exactly, because the sample population included patients with suspected toxocariasis at two university hospitals. Although regional data are used in this study, these data are consistent with other Korean data for the epidemiology of toxocariasis in eosinophilic patients. The current study highlights that toxocariasis is a reasonable focus as a cause of eosinophilia and is associated with organ involvement. A serological evaluation for toxocariasis is essential for patients with eosinophilia, considering the cultural habit of ingesting raw cow liver in Korea.

Authors' Disclosures of Potential Conflicts of Interest

No potential conflicts of interest relevant to this article were reported.

Acknowledgments

This work was partially supported by the Fund of Annals of Clinical Microbiology.

REFERENCES

1. Rothenberg ME. Eosinophilia. *N Engl J Med* 1998;338:1592-600.
2. Seo M and Yoon SC. A seroepidemiological survey of toxocariasis among eosinophilia patients in Chungcheongnam-do. *Korean J Parasitol* 2012;50:249-51.
3. Macpherson CN. The epidemiology and public health importance of toxocariasis: a zoonosis of global importance. *Int J Parasitol* 2013; 43:999-1008.
4. Mendonça LR, Figueiredo CA, Esquivel R, Fiaccone RL, Pontes-de-Carvalho L, Cooper P, et al. Seroprevalence and risk factors for *Toxocara* infection in children from an urban large setting in Northeast Brazil. *Acta Trop* 2013;128:90-5.
5. Romero Núñez C, Mendoza Martínez GD, Yañez Arteaga S, Ponce Macotela M, Bustamante Montes P, Ramírez Durán N. Prevalence and risk factors associated with *Toxocara canis* infection in children. *Scientific-WorldJournal* 2013;2013:572089.
6. Maraghi S, Rafiei A, Hajihosseini R, Sadjjadi SM. Seroprevalence of toxocariasis in hypereosinophilic individuals in Ahwaz, south-western Iran. *J Helminthol* 2012;86:241-4.
7. Woodhall DM, Eberhard ML, Parise ME. Neglected parasitic infections in the United States: toxocariasis. *Am J Trop Med Hyg* 2014;90:810-3.
8. Ryu BH, Park JS, Jung YJ, Kang SK, Lee SH, Choi SJ. Clinical and serological findings in patients with toxocariasis in the Pohang region: The features of toxocariasis in Pohang. *Korean J Med* 2013;84:203-10.
9. Choi D, Lim JH, Choi DC, Paik SW, Kim SH, Huh S. Toxocariasis and ingestion of raw cow liver in patients with eosinophilia. *Korean J Parasitol* 2008;46:139-43.
10. Jacquier P, Gottstein B, Stingelin Y, Eckert J. Immunodiagnosis of toxocariasis in humans: evaluation of a new enzyme-linked immunosorbent assay kit. *J Clin Microbiol* 1991;29:1831-5.
11. Dancey C and Reidy J. *Statistics without maths for psychology: using SPSS for Windows*. London: Prentice Hall, 2004.
12. Hotez PJ and Wilkins PP. Toxocariasis: America's most common neglected infection of poverty and a helminthiasis of global importance? *PLoS Negl Trop Dis* 2009;3:e400.
13. Kwon NH, Oh MJ, Lee SP, Lee BJ, Choi DC. The prevalence and diagnostic value of toxocariasis in unknown eosinophilia. *Ann Hematol* 2006; 85:233-8.
14. Park HY, Lee SU, Huh S, Kong Y, Magnaval JF. A seroepidemiological survey for toxocariasis in apparently healthy residents in Gangwon-do, Korea. *Korean J Parasitol* 2002;40:113-7.
15. Park HJ, Choi SJ, Kim HM, Choi GS, Sung JM, Lee JW, et al. Diagnostic value of serum specific IgE to *Toxocara canis* in patients with eosinophilia. *Korean J Asthma Allergy Clin Immunol* 2009;29:105-11.
16. Cheun HI, Kong Y, Cho SH, Lee JS, Chai JY, Lee JS, et al. Successful control of lymphatic filariasis in the Republic of Korea. *Korean J Parasitol* 2009;47:323-35.
17. Lim JH. Foodborne eosinophilia due to visceral larva migrans: a disease abandoned. *J Korean Med Sci* 2012;27:1-2.
18. Li L, Gao W, Yang X, Wu D, Bi H, Zhang S, et al. Asthma and toxocariasis. *Ann Allergy Asthma Immunol* 2014;113:187-92.
19. Kang YR, Kim SA, Jeon K, Koh WJ, Suh GY, Chung MP, et al. Toxocariasis as a cause of new pulmonary infiltrates. *Int J Tuberc Lung Dis* 2013; 17:412-7.
20. Magnaval JF, Berry A, Fabre R, Morassin B. Eosinophil cationic protein as a possible marker of active human *Toxocara* infection. *Allergy* 2001; 56:1096-9.