

Review Article

Neutral Sphingomyelinase and Breast Cancer Research

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Our understanding of the functions of neutral sphingomyelinase (N-SMase) signaling has advanced over the past decade. In this review, we focus on the roles and regulation of N-SMase 1, N-SMase 2, N-SMase 3, an enzyme that generates the bioactive lipid ceramide through the hydrolysis of the membrane lipid sphingomyelin. A large body of work has now implicated N-SMase 2 in a diverse set of cellular functions, physiological processes, and disease pathologies. We focus on different aspects of this enzyme's regulation from transcriptional, post-translational, and biochemical. Furthermore, we expected N-SMase involvement in cellular processes including inflammatory signaling, cell growth, apoptosis, and tumor necrosis factor which in turn play important roles in pathologies such as cancer metastasis, variable disease, and other organ system disorders. Lastly, we examine avenues where targeted N-SMase inhibition may be clinically beneficial in disease scenarios. (J Menopausal Med 2015;21:24-27)

Key Words: Breast neoplasms, Sphingomyelin phosphodiesterase, Tumor necrosis factor-alpha

Introduction

Sphingomyelin hydrolysis is catalyzed by a class of enzymes referred to as sphingomyelinases (SMases) to generate ceramide. Ceramide and sphingolipid metabolites are well-established regulators of many important cellular signaling pathways and are implicated in human health and disease. SMases are classified based on their pH optima of activity into acid, neutral, and alkaline subtypes. Of the four different mammalian neutral SMases that have been identified; neutral SMase 1 (N-SMase 1), N-SMase 2. N-SMase 3. Mitochondrial-Associated Neutral SMase (MA-N-SMase) appears to be the predominant N-SMase in cellular systems, physiologies, and pathologies. 2,3 Breast cancer is still the most common type of cancer in women and Gail model (risk measurements) thought prediction increase role. 4 Surgical resection method is the best way for a correct diagnosis but recently, in surgery incisions

and shorter time of more precise lesions, scars and could result from reduced recommend the future of mammotome.⁵ According to a recent study, breast cancer characteristics of Korean postmenopausal women have a high incidence is also higher hormone receptor—positive rate of breast cancer.⁶ Breast cancer patients are not recommended hormone therapy. Because it is hormone therapy increases of the risk and affect the existing not expected to not diagnosis another breast cancer.⁷

This review will focus on the roles and regulation of this enzyme emphasizing recent findings implicating N-SMase in disease processes.

Classification of Neutral SMase

1. N-SMase 1

 $N-SMase\ 1$ was the first discover mammalian N-SMase

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based on sequence homology to bacterial SMases. It is not activated by phospholipids. N-SMase 1 activity on sphingomyelin *in vitro*, overexpression in cells does not affect sphingomyelin metabolism. N-SMase 1 in sphingolipid metabolism is unclear. Although a function for N-SMase 1 is not apparent, With regards to N-SMase 1, this may be particularly relevant given the different subcellular localizations of endogenous and overexpressed proteins.

2. N-SMase 2

N-SMase 2 is to be known to study mammalian N-SMase and has emerged as a key mediator of cellular stress-induced generation of ceramide. Several characterizations have identified a number of mechanisms for activation and regulation of N-SMase 2. These studies provide a base to investigate other N-SMase isoforms, as well as potential areas and modes for therapeutic intervention,

3. N-SMase 3

Human N-SMase 3 was be come to in 2006, ¹⁰ relatively little work has been reported since regarding further biochemical and functional characterization. The peptide sequence only matches 7 of the 11 residues in the identified human protein and bovine homolog. ¹¹ The question if the original purified bovine and identified human proteins correspond to the same protein. N-SMase 3 shares no sequence homology with any N-SMases or any other characterized type of enzyme catalytic domain. The region comprising the catalytic domain is yet to be identified. A C-terminal transmembrane helix is predicted to embed N-SMase 3 in the membrane. ¹⁰

Two conflicting reports have characterized N–SMase 3 activity is reported to occur at neutral pH and require ${\rm Mg}^{2+}$ or ${\rm Mn}^{2+}$. The observed activity was slightly enhanced, approximately two–fold, by the phospholipid. In a later study, MCF–7 cells transiently and stably overexpressing N–SMase 3 did not have significant N–SMase activity over vector controls. We suppose to future studies will determine the underlying reasons behind this major discrepancy. N–SMase 3 suggested that may activity a role in tumor necrosis factor– α (TNF– α) mediated signaling. 10,12 However, another report

found that N–SMase 2 was the primary N–SMase activated by TNF– α in MCF–7 cells. ^{11,13,14}

4. MA-N-SMase

MA-N-SMase is the most recently identified mammalian N-SMase being discovered in 2010¹⁵ by sequence homology with N-SMase 2 and a zebrafish mitochondrial N-SMase. 16 The subcellular localization of overexpressed MA-N-SMase protein varied with cell type, showing strong or partial co-localization with mitochondrial markers, in addition to co-localization with endoplasmic reticulum markers. 15 Present in mitochondria providing a putative mechanism for activation and/or regulation of MA-N-SMase activity in vivo. 15 MA-N-SMase is little known about beyond basic properties. However, the identification of a mammalian mitochondrial N-SMase presents another potential endogenous mechanism, in addition to action of ceramide synthesis, for mitochondrial ceramide generation. This is exciting considering the numerous studies linking ceramide to mitochondrial activation of apoptosis. 17 Another potential role for MA-N-SMase is in fertilization. Activation of an unidentified SMase during fertilization was inferred by a corresponding decrease in sphingomyelin and increase in ceramide levels. 18

Neutral SMase Inhibitor

Many subtypes N-SMase about N-SMase 2 have only inhibitor, GW4869 and variety inhibitor. GW4869 has been widely used as a tool to identify and confirm N-SMase 2-specific functions. GW4869 is inference thought to inhibit N-SMase 2 by interfering with APL activation, but this pathway still not become known to inhibitor.

Mechanism of Pathway Action

1. Response to TNF- α

TNF- α triggers the activation of the sphingomyelin–ceramide pathway through stimulation of SMases. ²¹ TNF- α pathway is induced by the hydrolysis of sphingomyelin to ceramide, which is considered as a putative bioactive



sphingolipid in cell signaling. 22 TNF– $\!\alpha\!$ has been reported to activate acidic and N-SMases 2 and 3. 10,21,23 Activation of N-SMases in response to the cytokine TNF- α was initially observed in the HL-60 cell line (leukemia cell line). The TNF- α receptor-1 (TNFR-1), ordinarily known to the p55 receptor. 24-27 It is pathway a distinct region of the p55 receptor was found to mediate the increase in N-SMase activity and was the neutral N-SMase activation domain (NSAD). 24 Factor associated with N-SMase (FAN-SMase) that influences N-SMase activity in response to TNF- α . ^{24,25} Indeed, overexpression of a dominant-negative form of FAN-SMase alters TNF- α induced N-SMase 3 activation in MCF-7 (human breast cancer cell line) cells. 10 In addition to the elucidation of the molecular partners, other investigations have focused on cellular factors that influence TNF- α activation of N-SMase 2. Reported that the levels of glutathione, which can inhibit neutral SMase activity, are critical for TNF- α activation. Functionally, the activation of N-SMase 2 has implications in the inflammatory response of TNF- α . HeLa cells were known that N-SMase 2 acts on the PI3K/Akt pathway to activate endothelial nitric oxide synthase (eNOS). This suggests that N-SMase 2-derived ceramide can have bioactive action by itself in response to TNF- α or can be metabolized into other lipids to induce differential responses.

2. N-SMase: apoptosis

Recent findings have demonstrated a functional regulation of N–SMase 2 in response to lung insults and in certain lung diseases. Initially, reported that $\rm H_2O_2$ stimulation and reactive oxygen species (ROS) formation activates N–SMase 2 to mediate apoptosis in human airway epithelium, while glutathione pretreatment prevents apoptosis.

N-SMase and Breast Cancer

Recent studies are interested in N-SMase about related with of cancer. However, gynecologic diseases related research has been studied a little, among the rest active in the research of breast cancer. Hormone-dependent growth has been observed in some human cancers including breast cancer.

N-SMase enzyme activity and ceramide level regulate of breast cancer cell line (MCF-7, MDA-MB-231 cells). In agreement with the study N-SMase 2 is known that functions as an N-SMase that N-SMase 2 of over expression caused decrease in cell growth in MCF-7 breast cancer cells.²³

Conclusion

In summary, recently, reported N-SMase related with cell signal and immune response about many studies and research proceed. TNF- α and apoptosis pathway, cell responsive involvement and associated with various diseases. Nowadays, cancer research or research processing of breast cancer has been known slightly studying furthermore in front of various gynecological disease relative thinking.

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Conflict of Interest

No potential conflict of interest relevant to this article was reported.

References

- 1. Wu BX, Clarke CJ, Hannun YA. Mammalian neutral sphingomyelinases: regulation and roles in cell signaling responses. Neuromolecular Med 2010; 12: 320–30.
- 2. Hannun YA, Obeid LM. Many ceramides. J Biol Chem 2011; 286: 27855–62.
- Ogretmen B, Hannun YA. Biologically active sphingolipids in cancer pathogenesis and treatment. Nat Rev Cancer 2004; 4: 604–16.
- Lee ES, Seo JS, Hong YP, Park HM. Estimation of the relative risk for breast cancer in Korean women using gail model. J Korean Soc Menopause 2012; 18: 180-6.

- 5. Jeong JH, Kim HG, Kim KH, Choi OH. The clinical experience of an ultrasound-guided vacuum-assisted resection (mammotome) for benign breast lesions through a core needle biopsy. J Korean Soc Menopause 2013; 19: 9-17.
- Son JB, Jeong JE, Joo JK, Kim KH, Lee KS. Clinical characteristics of breast cancer detected during hormone therapy in Korean women. J Korean Soc Menopause 2012; 18: 52-9
- The Korean Society of Menopause Scientific Board. Hormone replacement therapy and breast cancer in postmenopausal women. J Korean Soc Menopause 2011; 17: 125-6.
- Tomiuk S, Hofmann K, Nix M, Zumbansen M, Stoffel W. Cloned mammalian neutral sphingomyelinase: functions in sphingolipid signaling? Proc Natl Acad Sci U S A 1998; 95: 3638–43.
- Sawai H, Domae N, Nagan N, Hannun YA. Function of the cloned putative neutral sphingomyelinase as lyso-platelet activating factor-phospholipase C. J Biol Chem 1999; 274: 38131-9.
- Krut O, Wiegmann K, Kashkar H, Yazdanpanah B, Krönke M. Novel tumor necrosis factor—responsive mammalian neutral sphingomyelinase—3 is a C-tail—anchored protein. J Biol Chem 2006; 281: 13784—93.
- Clarke CJ, Cloessner EA, Roddy PL, Hannun YA. Neutral sphingomyelinase 2 (nSMase2) is the primary neutral sphingomyelinase isoform activated by tumour necrosis factor—alpha in MCF-7 cells. Biochem J 2011; 435: 381-90.
- 12. Corcoran CA, He Q, Ponnusamy S, Ogretmen B, Huang Y, Sheikh MS. Neutral sphingomyelinase—3 is a DNA damage and nongenotoxic stress—regulated gene that is deregulated in human malignancies. Mol Cancer Res 2008; 6: 795–807.
- 13. Ito H, Tanaka K, Hagiwara K, Kobayashi M, Hoshikawa A, Mizutani N, et al. Transcriptional regulation of neutral sphingomyelinase 2 in all-trans retinoic acid-treated human breast cancer cell line, MCF-7. J Biochem 2012; 151: 599-610.
- 14. Ito H, Murakami M, Furuhata A, Gao S, Yoshida K, Sobue S, et al. Transcriptional regulation of neutral sphingomyelinase 2 gene expression of a human breast cancer cell line, MCF-7, induced by the anti-cancer drug, daunorubicin. Biochim Biophys Acta 2009; 1789: 681–90.
- 15. Wu BX, Rajagopalan V, Roddy PL, Clarke CJ, Hannun YA. Identification and characterization of murine mitochondria—associated neutral sphingomyelinase (MA-nSMase), the mammalian sphingomyelin phosphodiesterase 5. J Biol

- Chem 2010; 285: 17993-8002.
- Yabu T, Shimuzu A, Yamashita M. A novel mitochondrial sphingomyelinase in zebrafish cells. J Biol Chem 2009; 284: 20349–63.
- Mullen TD, Obeid LM. Ceramide and apoptosis: exploring the enigmatic connections between sphingolipid metabolism and programmed cell death. Anticancer Agents Med Chem 2012; 12: 340-63.
- Petcoff DW, Holland WL, Stith BJ. Lipid levels in sperm, eggs, and during fertilization in Xenopus laevis. J Lipid Res 2008; 49: 2365–78
- 19. Luberto C, Hassler DF, Signorelli P, Okamoto Y, Sawai H, Boros E, et al. Inhibition of tumor necrosis factor—induced cell death in MCF7 by a novel inhibitor of neutral sphingomyelinase. J Biol Chem 2002; 277: 41128–39.
- 20. Canals D, Perry DM, Jenkins RW, Hannun YA. Drug targeting of sphingolipid metabolism: sphingomyelinases and ceramidases. Br J Pharmacol 2011; 163: 694-712.
- Wiegmann K, Schütze S, Machleidt T, Witte D, Krönke M. Functional dichotomy of neutral and acidic sphingomyelinases in tumor necrosis factor signaling. Cell 1994; 78: 1005–15.
- 22. Ségui B, Andrieu-Abadie N, Jaffrézou JP, Benoist H, Levade T. Sphingolipids as modulators of cancer cell death: potential therapeutic targets. Biochim Biophys Acta 2006; 1758: 2104-20.
- 23. Marchesini N, Luberto C, Hannun YA. Biochemical properties of mammalian neutral sphingomyelinase 2 and its role in sphingolipid metabolism. J Biol Chem 2003; 278: 13775–83.
- 24. Adam-Klages S, Adam D, Wiegmann K, Struve S, Kolanus W, Schneider-Mergener J, et al. FAN, a novel WD-repeat protein, couples the p55 TNF-receptor to neutral sphingomyelinase. Cell 1996; 86: 937-47.
- 25. Adam-Klages S, Schwandner R, Adam D, Kreder D, Bernardo K, Krönke M. Distinct adapter proteins mediate acid versus neutral sphingomyelinase activation through the p55 receptor for tumor necrosis factor. J Leukoc Biol 1998; 63: 678-82.
- 26. Belka C, Wiegmann K, Adam D, Holland R, Neuloh M, Herrmann F, et al. Tumor necrosis factor (TNF)—alpha activates c—raf—1 kinase via the p55 TNF receptor engaging neutral sphingomyelinase. EMBO J 1995; 14: 1156—65.
- 27. Yang Z, Costanzo M, Golde DW, Kolesnick RN. Tumor necrosis factor activation of the sphingomyelin pathway signals nuclear factor kappa B translocation in intact HL– 60 cells. J Biol Chem 1993; 268: 20520–3.