

PEARLS

A 21st Century Perspective of Poliovirus Replication

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Why Poliovirus Replication Has Been Studied for More Than 50 Years

Poliovirus is the etiologic agent of poliomyelitis, an acute flaccid paralysis affecting 1%–2% of infected patients and, on rare occasions, causing death by paralyzing muscles that control the throat or breathing. A striking feature of infection is lifelong disabilities that may affect survivors of the acute disease. Transmitted by the fecal—oral and oral—oral route, this virus (three serotypes) was one of the most feared pathogens in industrialized countries during the 20th century affecting hundreds of thousands of children every year, via outbreaks during warm summer months. Although there are highly effective vaccines to control poliomyelitis, it remains endemic in a few countries, from which spread and outbreaks continue to occur throughout the world. Since its discovery in 1908, poliovirus has been intensively studied to better understand and control this formidable pathogen. The history of poliovirus is not, however, limited to the fight against the disease. Poliovirus replication studies also have played important roles in the development of modern virology since poliovirologists and, more generally, picornavirologists have been pioneers in many domains of molecular virology. Poliovirus was, for example, the first animal RNA virus to have its complete genome sequence determined, the first RNA animal virus for which an infectious clone was constructed, and, along with the related rhinovirus, the first human virus that had its three-dimensional structure solved by X-ray crystallography. Indeed, the history of over half a century of poliovirus replication studies is marked by major discoveries, many of which are summarized here and illustrated in [Fig 1](#).

1950–1970: The Early Years

In 1949, John Enders, Thomas Weller, and Frederick Robbins performed a landmark study showing that poliovirus could be propagated in cultured, non-neural human cells that did not correspond to the tissues infected during the disease [1]. Not only did these Nobel Prize-winning studies pave the way for the development of highly effective vaccines against poliovirus, but they also opened the door for virologists to study the molecular mechanisms of poliovirus replication in cultured cells that were much more readily manipulated than neural tissue. Isolated poliovirus genomic RNA was then shown to be infectious for susceptible HeLa cells in monolayers, demonstrating that the viral genome itself is the carrier of the biological activity responsible for infection [2]. John Holland and coworkers then reproduced this experiment with normally nonsusceptible cells, demonstrating that the block to poliovirus growth in



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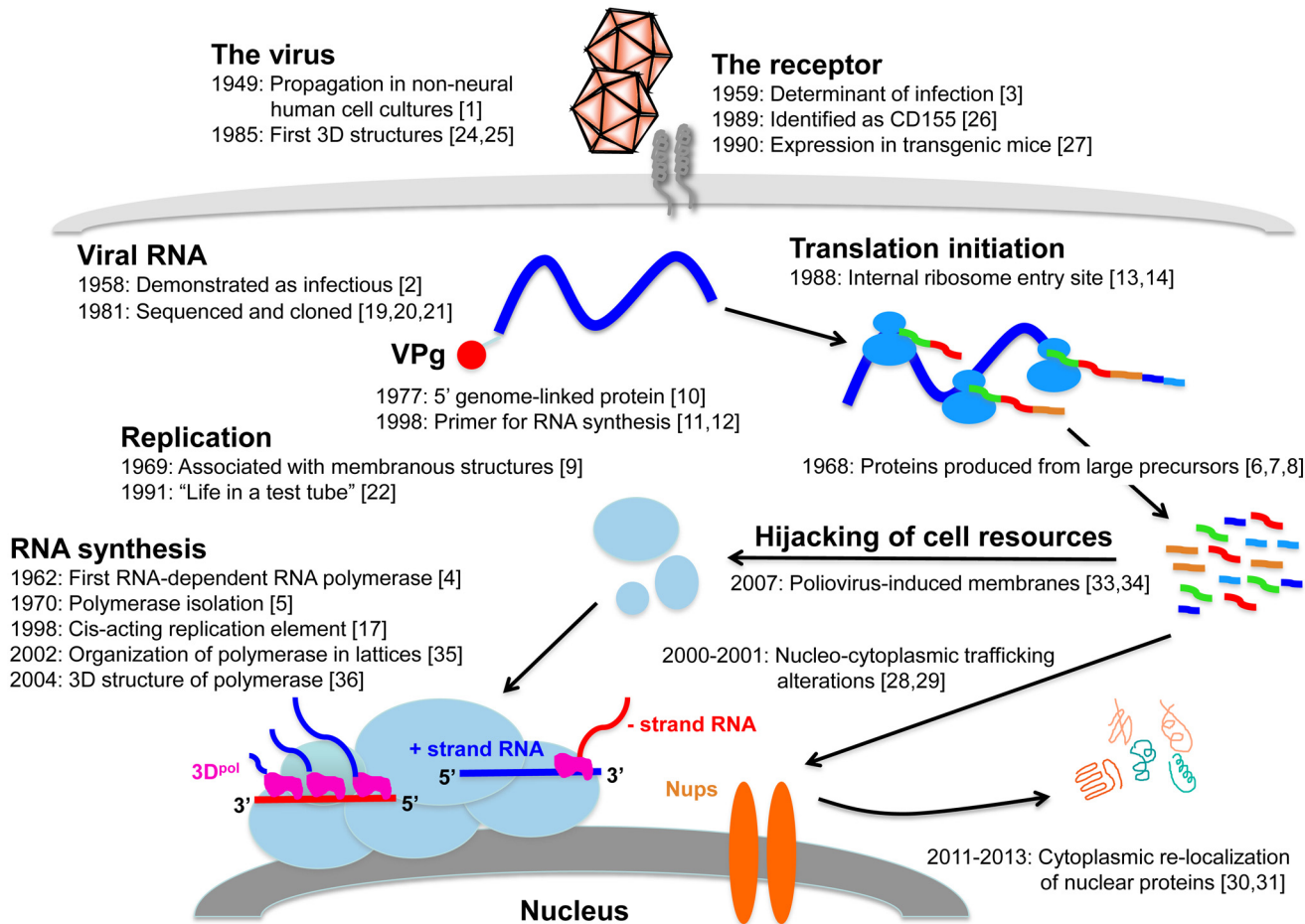


Fig 1. Replication cycle of poliovirus, annotated with references to key findings during the past 65 years. Abbreviations and symbols: 3D, three-dimensional; receptor proteins (grey cylinders spanning plasma membrane); RNA, ribonucleic acid; VPg, viral protein, genome-linked (red oval); ribosomes (medium blue oval shapes bound to viral RNA); + strand RNA, positive-sense genomic RNA (blue horizontal or wavy line); - strand RNA, negative-sense antigenomic RNA (red horizontal or wavy line); 3D^{pol}, viral RNA-dependent RNA polymerase (pink shapes); membranous vesicles (light blue oval shapes); Nups, nucleoporins (orange, oblong oval shapes).

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nonpermissive, nonprimate cells was due to the absence of specific receptors, defining cell determinants of poliovirus infection [3].

The first evidence for an RNA-dependent RNA polymerase used by the virus for RNA genome replication was reported in 1962 by Baltimore and Franklin [4]. Until then, it was believed that the genome of RNA viruses was replicated in the cell nucleus by a cellular DNA-dependent RNA polymerase. Studies on another picornavirus, mengovirus, revealed the existence of an actinomycin D-resistant replication activity in the cytoplasm of infected cells that was later also identified in poliovirus-infected cells [4]. This virus-specific enzyme was isolated eight years later [5].

In the late 1960s, Summers and Maizel [6], and others [7,8], showed that the genomic RNA of poliovirus is translated to produce very large polypeptides that are then specifically cleaved into smaller functional proteins. This discovery was the first demonstration that picornavirus proteins were produced from large precursors proteolytically processed in infected cells [6]. At the same time, cellular fractionation studies revealed that poliovirus RNAs are synthesized in

replication complexes bound to distinct membranous structures in the cytoplasm of infected cells [9].

1970–2000: The Late 20th Century

This period corresponds to several major advances in understanding the mechanisms of translation and replication of poliovirus genomic RNA. In 1977, a small protein of 22 amino acids, called VPg, was discovered to be covalently linked to the 5' end of poliovirus RNA [10]. Its absence when the virus genome is translated on polysomes indicated that VPg is removed prior to or during translation, whereas its presence at the 5' end of the newly synthesized positive- and negative-strand RNAs suggested that VPg might be involved in the initiation of RNA synthesis [11]. This hypothesis was confirmed later by Paul et al., who demonstrated that VPg linked by the poliovirus RNA polymerase 3D^{pol} to the 5'-terminal uridylic acid of the virus genome is the key player in protein-primed initiation of poliovirus RNA synthesis [12]. Experiments with dicistronic vectors subsequently led to the discovery of a novel mechanism of initiation of translation through ribosome binding to RNA secondary structures within the long 5' nontranslated region of the virus genome. This internal ribosome entry site (or IRES) allows recognition by the host translation apparatus in the absence of a 5' cap structure on viral mRNAs [13,14]. Since infection by poliovirus and many other picornaviruses leads to the shut off of cap-dependent translation [15,16], the IRES allows the virus to effectively compete for the cellular translation machinery via cap-independent mechanisms. Finally, a cis-acting replication element (or cre) was identified in picornavirus genomic RNAs. Cre sequences are RNA stem-loop structures almost exclusively located within the coding region, and they are required for viral RNA replication. These elements bind viral proteins involved in RNA replication complex formation, allowing specific recognition of viral RNAs in the cytoplasm of infected cells among a myriad of poly(A)-containing host cellular mRNAs. Cre sequences promote uridylation of VPg, the primer for initiation of viral RNA synthesis, by 3D^{pol}. Their functions appear to be strand-specific, since the cre is required for positive-strand RNA synthesis but may not be essential for negative-strand RNA synthesis. A cre was first found in the capsid coding region of human rhinovirus (HRV) RNA and later in the 2C coding region of poliovirus RNA [17,18]. Since then, similar types of internal recognition elements have been detected for other positive-strand RNA viruses.

The complete sequence of poliovirus genomic RNA was reported in 1981 by two different groups [19,20]. Poliovirus RNA was shown to be more than 7,400 nucleotides long, polyadenylated at the 3' terminus, and composed of an open reading frame (ORF) of 2,207 consecutive triplets, spanning over 89% of the total nucleotide sequence. This single ORF encodes the viral polyprotein that is ultimately cleaved into more than 15 intermediate and mature viral polypeptides. Poliovirus was also the first RNA animal virus for which an infectious, cloned complementary DNA copy of the RNA genome was constructed. Transfection of this clone into mammalian cells produced infectious poliovirus and, via genetic manipulation, led to new insights about the functions of viral proteins and RNA sequences and their roles in the picornavirus intracellular replication cycle [21].

Transcribed viral RNA from a plasmid-derived cDNA was also used *in vitro* for the *de novo* synthesis of infectious poliovirus, a first for an animal RNA virus. Published in 1991 by Eckard Wimmer's group, "Life in a Test Tube" resulted from the addition of viral RNA to a crude cytoplasmic extract of uninfected HeLa cells. These extracts contain all essential elements for poliovirus replication, including cytoplasmic membranes and components required for virion assembly [22]. Implementation of this system allowed the use of "in vitro genetics" with defined templates and fractionated extracts to understand many aspects of the virus life

cycle, especially the mechanism of initiation of RNA synthesis to yield VPg-linked progeny RNAs [23].

Based upon X-ray crystallography studies, poliovirus and its relative, HRV, were the first animal viruses for which the three-dimensional virus structure was solved in 1985 by the groups of Hogle and Rossmann [24,25]. And in 1989, Racaniello and coworkers identified CD155, a member of the immunoglobulin superfamily, as the poliovirus receptor [26]. This finding was followed by the generation of mice carrying CD155 as a transgene, allowing studies of poliovirus infection and pathogenesis in vivo in a nonprimate model [27].

2000–2015: The 21st Century

Numerous studies carried out during this period uncovered the ability of poliovirus to usurp cellular components and structures for its own benefit, reflecting the necessity for this virus with a very limited coding capacity to hijack cell resources during infection. Relocalization of nuclear proteins in the cytoplasm during poliovirus infection was first reported in 2000–2001. Cleavage of nuclear pore complex components such as Nup153 and p62 by the viral 2A proteinase led to the accumulation in the cytoplasm of virus-infected cells of a number of proteins normally present in the nucleus [28,29]. These relocalized proteins often manifest RNA binding capabilities and function in host cell RNA metabolism steps. For example, the nuclear protein SRp20, a cellular splicing factor, is relocalized to the cytoplasm of poliovirus-infected human cells. It was identified as an important IRES trans-acting factor (ITAF) for poliovirus translation [30]. Another cellular protein that is redistributed in the cytoplasm during poliovirus infection is 5'-tyrosyl-DNA phosphodiesterase-2 (TDP2), a DNA repair enzyme identified as the source of VPg unlinkase activity that cleaves the protein-RNA covalent linkage of VPg at the 5' end of virion RNA [31,32]. VPg unlinkase/TDP2 activity may be essential to provide a balance between the translation, replication, and packaging functions of viral RNA.

Poliovirus replication sites on cellular membranes (first described in 1969) were also shown to be the result of viral hijacking of components of cellular membrane metabolic pathways, leading to intracellular membrane remodeling and generation of specialized sites distinct in protein and lipid composition from that of the host cell. Belov, Altan-Bonnet, and colleagues demonstrated that viral proteins promoted membrane-binding of members of the Arf family of small guanosine triphosphatases (GTPases), leading to the formation of phosphatidylinositol-4-phosphate lipid-enriched organelles that are essential for poliovirus RNA replication [33, 34]. Indeed, these poliovirus-induced membranes contain viral RNA-dependent RNA polymerases organized in lattices of hundreds of molecules observed for the first time in 2002 by electron microscopy [35]. The two-dimensional planar and tubular oligomeric arrays of poliovirus polymerase display cooperative RNA binding and elongation. Understanding the nature of polymerase interface domains, polymerase activation, and template RNA/rNTP interactions has been greatly facilitated by the complete structure of the poliovirus RNA polymerase, which was reported in 2004 [36]. Combining our knowledge of polymerase structure, function, and genetics has also revealed novel insights into how poliovirus polymerase fidelity contributes to the quasi-species nature of RNA virus populations [37,38].

Why Poliovirus Replication Must Still Be Studied

Despite a dramatic reduction in the number of cases of poliomyelitis (350,000 cases in 1988 versus 416 cases in 2013), as well as of polio endemic countries (125 countries in 1988 versus 3 countries in 2014), poliovirus is not yet eradicated from our planet and is still the target of a massive worldwide vaccination campaign (<http://www.polioeradication.org/>). Moreover, vaccine-associated paralytic poliomyelitis resulting from the use of a live, attenuated vaccine

complicates any poliovirus endgame strategy [39]. Finally, enteroviruses other than poliovirus have emerged as serious threats to public health. These include enterovirus 71, responsible in infants and young children for hand, foot, and mouth disease with the potential for severe central nervous system complications, and enterovirus D68, detected in children hospitalized with severe lower respiratory symptoms and asthma. In this context, there is an absolute necessity to identify potential new targets for novel antienteroviral drugs and develop new, safe, and effective vaccines. Given the plethora of cellular, molecular, and genetic tools produced during the study of poliovirus replication for more than half a century, it is paramount to continue to use these invaluable research tools to expand our knowledge of the replication and virulence determinants of emerging enteroviruses. In addition, there is still much to be learned about the pathogenic mechanisms of enteroviruses as well as the mechanistic details of poliovirus replication, particularly how the virus weaves the functions of host cell activities into a complex web of intracellular events in its reproductive cycle. Such information may be required to implement the final push that could ultimately result in the eradication of this legendary human pathogen.

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References

1. Enders J. F., Weller T. H., and Robbins F. C.. 1949. Cultivation of the Lansing strain of poliomyelitis virus in cultures of various human embryonic tissues. *Science* 109:85–87. PMID: [17794160](#)
2. Alexander H. E., Koch G., Mountain I. M., and Van Damme O.. 1958. Infectivity of ribonucleic acid from poliovirus in human cell monolayers. *J. Exp. Med.* 108:493–506. PMID: [13575680](#)
3. Holland J. J., M. L. C., and Syverton J. T.. 1959. Mammalian cell-virus relationship. III. Poliovirus production by non-primate cells exposed to poliovirus ribonucleic acid. *Proc. Soc. Exp. Biol. Med.* 100:843–845. PMID: [13645739](#)
4. Baltimore D., and Franklin R. M.. 1962. Preliminary data on a virus-specific enzyme system responsible for the synthesis of viral RNA. *Biochem. Biophys. Res. Commun.* 9:388–392. PMID: [13966258](#)
5. Ehrenfeld E., Maizel J. V., and Summers D. F.. 1970. Soluble RNA polymerase complex from poliovirus-infected HeLa cells. *Virology* 40:840–846. PMID: [4317360](#)
6. Summers D. F., and Maizel J. V.. 1968. Evidence for large precursor proteins in poliovirus synthesis. *Proc. Natl. Acad. Sci. U.S.A.* 59:966–971. PMID: [4296044](#)
7. Holland J. J., and Kiehn E. D.. 1968. Specific cleavage of viral proteins as steps in the synthesis and maturation of enteroviruses. *Proc. Natl. Acad. Sci. U.S.A.* 60:1015–1022. PMID: [4299264](#)
8. Jacobson M. F., and Baltimore D.. 1968. Polypeptide cleavages in the formation of poliovirus proteins. *Proc. Natl. Acad. Sci. U.S.A.* 61:77–84. PMID: [4301595](#)
9. Caligiuri L. A., and Tamm I.. 1969. Membranous structures associated with translation and transcription of poliovirus RNA. *Science* 166:885–886. PMID: [4310189](#)
10. Lee Y. F., Nomoto A., Detjen B. M., and Wimmer E.. 1977. A protein covalently linked to poliovirus genome RNA. *Proc. Natl. Acad. Sci. U.S.A.* 74:59–63. PMID: [189316](#)
11. Nomoto A., Detjen B., Pozzatti R., and Wimmer E.. 1977. The location of the polio genome protein in viral RNAs and its implication for RNA synthesis. *Nature* 268:208–213. PMID: [196204](#)
12. Paul A. V., van Boom J. H., Filippov D., and Wimmer E.. 1998. Protein-primed RNA synthesis by purified poliovirus RNA polymerase. *Nature* 393:280–284. PMID: [9607767](#)
13. Pelletier J., and Sonenberg N.. 1988. Internal initiation of translation of eukaryotic mRNA directed by a sequence derived from poliovirus RNA. *Nature* 334:320–325. PMID: [2839775](#)
14. Jang S. K., Krausslich H. G., Nicklin M. J., Duke G. M., Palmenberg A. C., and Wimmer E.. 1988. A segment of the 5' nontranslated region of encephalomyocarditis virus RNA directs internal entry of ribosomes during *in vitro* translation. *J. Virol.* 62:2636–2643. PMID: [2839690](#)

15. Fernandez-Munoz R., and Darnell J. E.. 1976. Structural difference between the 5' termini of viral and cellular mRNA in poliovirus-infected cells: possible basis for the inhibition of host protein synthesis. *J. Virol.* 18:719–726. PMID: [178904](#)
16. Ehrenfeld E., and Lund H.. 1977. Untranslated vesicular stomatitis virus messenger RNA after poliovirus infection. *Virology* 80:297–308. PMID: [196392](#)
17. McKnight K. L., and Lemon S. M.. 1998. The rhinovirus type 14 genome contains an internally located RNA structure that is required for viral replication. *RNA.* 4:1569–1584. PMID: [9848654](#)
18. Goodfellow I., Chaudhry Y., Richardson A., Meredith J., Almond J. W., Barclay W., and Evans D. J.. 2000. Identification of a cis-acting replication element within the poliovirus coding region. *J. Virol.* 74:4590–4600. PMID: [10775595](#)
19. Kitamura N., Semler B. L., Rothberg P. G., Larsen G. R., Adler C. J., Dorner A. J., Emimi E. A., Hanecak R., Lee J. J., van der Werf S., Anderson C. W., and Wimmer E.. 1981. Primary structure, gene organization and polypeptide expression of poliovirus RNA. *Nature* 291:547–553. PMID: [6264310](#)
20. Racaniello V. R., and Baltimore D.. 1981. Molecular cloning of poliovirus cDNA and determination of the complete nucleotide sequence of the viral genome. *Proc. Natl. Acad. Sci. U.S.A.* 78:4887–4891. PMID: [6272282](#)
21. Racaniello V. R., and Baltimore D.. 1981. Cloned poliovirus complementary DNA is infectious in mammalian cells. *Science* 214:916–919. PMID: [6272391](#)
22. Molla A., Paul A. V., and Wimmer E.. 1991. Cell-free, de novo synthesis of poliovirus. *Science* 254:1647–1651. PMID: [1661029](#)
23. Barton D. J., and Flanagan J. B.. 1993. Coupled translation and replication of poliovirus RNA in vitro: synthesis of functional 3D polymerase and infectious virus. *J. Virol.* 67:822–831. PMID: [8380467](#)
24. Hogle J. M., Chow M., and Filman D. J.. 1985. Three-dimensional structure of poliovirus at 2.9 Å resolution. *Science* 229:1358–1365. PMID: [2994218](#)
25. Rossmann M. G., Arnold E., Erickson J. W., Frankenberger E. A., Griffith J. P., Hecht H. J., Johnson J. E., Kamer G., Luo M., Mosser A. G., and et al. 1985. Structure of a human common cold virus and functional relationship to other picornaviruses. *Nature* 317:145–153. PMID: [2993920](#)
26. Mendelsohn C. L., Wimmer E., and Racaniello V. R.. 1989. Cellular receptor for poliovirus: molecular cloning, nucleotide sequence, and expression of a new member of the immunoglobulin superfamily. *Cell* 56:855–865. PMID: [2538245](#)
27. Ren R. B., Costantini F., Gorgacz E. J., Lee J. J., and Racaniello V. R.. 1990. Transgenic mice expressing a human poliovirus receptor: a new model for poliomyelitis. *Cell* 63:353–362. PMID: [2170026](#)
28. Belov G. A., Evstafieva A. G., Rubtsov Y. P., Mikitas O. V., Vartapetian A. B., and Agol V. I.. 2000. Early alteration of nucleocytoplasmic traffic induced by some RNA viruses. *Virology* 275:244–248. PMID: [10998323](#)
29. Gustin K. E., and Sarnow P.. 2001. Effects of poliovirus infection on nucleocytoplasmic trafficking and nuclear pore complex composition. *EMBO J.* 20:240–249. PMID: [11226174](#)
30. Fitzgerald K. D., and Semler B. L.. 2011. Re-localization of cellular protein SRp20 during poliovirus infection: bridging a viral IRES to the host cell translation apparatus. *PLoS Pathog.* 7:e1002127. doi: [10.1371/journal.ppat.1002127](#) PMID: [21779168](#)
31. Virgen-Slane R., Rozovics J. M., Fitzgerald K. D., Ngo T., Chou W., van der Heden van Noort G. J., Filippov D. V., Gershon P. D., and Semler B. L.. 2012. An RNA virus hijacks an incognito function of a DNA repair enzyme. *Proc. Natl. Acad. Sci. U.S.A.* 109:14634–14639. doi: [10.1073/pnas.1208096109](#) PMID: [22908287](#)
32. Ambros V., Pettersson R. F., and Baltimore D.. 1978. An enzymatic activity in uninfected cells that cleaves the linkage between poliovirion RNA and the 5' terminal protein. *Cell* 15:1439–1446. PMID: [215328](#)
33. Belov G. A., Altan-Bonnet N., Kovtunovych G., Jackson C. L., Lippincott-Schwartz J., and Ehrenfeld E.. 2007. Hijacking components of the cellular secretory pathway for replication of poliovirus RNA. *J. Virol.* 81:558–567. PMID: [17079330](#)
34. Hsu N. Y., Ilnytska O., Belov G., Santiana M., Chen Y. H., Takvorian P. M., Pau C., van der Schaar H., Kaushik-Basu N., Balla T., Cameron C. E., Ehrenfeld E., van Kuppeveld F. J., and Altan-Bonnet N.. 2010. Viral reorganization of the secretory pathway generates distinct organelles for RNA replication. *Cell* 141:799–811. doi: [10.1016/j.cell.2010.03.050](#) PMID: [20510927](#)
35. Lyle J. M., Bullitt E., Bienz K., and Kirkegaard K.. 2002. Visualization and functional analysis of RNA-dependent RNA polymerase lattices. *Science* 296:2218–2222. PMID: [12077417](#)
36. Thompson A. A., and Peersen O. B.. 2004. Structural basis for proteolysis-dependent activation of the poliovirus RNA-dependent RNA polymerase. *EMBO J.* 23:3462–3471. PMID: [15306852](#)

37. Pfeiffer J. K., and Kirkegaard K.. 2003. A single mutation in poliovirus RNA-dependent RNA polymerase confers resistance to mutagenic nucleotide analogs via increased fidelity. *Proc. Natl. Acad. Sci. U.S.A.* 100:7289–7294. PMID: [12754380](#)
38. Vignuzzi M., Stone J. K., Arnold J. J., Cameron C. E., and Andino R.. 2006. Quasispecies diversity determines pathogenesis through cooperative interactions in a viral population. *Nature* 439:344–348. PMID: [16327776](#)
39. Orenstein W. A. 2015. Eradicating polio: how the world's pediatricians can help stop this crippling illness forever. *Pediatrics* 135:196–202. doi: [10.1542/peds.2014-3163](#) PMID: [25548328](#)