

# Toxicological and biochemical studies on *Schinus terebinthifolius* concerning its curative and hepatoprotective effects against carbon tetrachloride-induced liver injury

Rania H. Abdou, Sherif Y. Saleh<sup>1</sup>, Waleed F. Khalil<sup>2</sup>

Departments of Forensic Medicine and Toxicology, <sup>1</sup>Biochemistry and <sup>2</sup>Pharmacology, Faculty of Veterinary Medicine, University of Suez Canal, Ismailia 41522, Egypt

Submitted: 17-07-2014

Revised: 12-09-2014

Published: 27-05-2015

## ABSTRACT

**Background:** Recently, many efforts have been made to discover new products of natural origin which can limit the xenobiotic-induced hepatic injury. Carbon tetrachloride (CCl<sub>4</sub>) is a highly toxic chemical that is widely used to study hepatotoxicity in animal models. **Objective:** The present study was conducted to investigate the curative and protective effects of *Schinus terebinthifolius* ethanolic extract against CCl<sub>4</sub>-induced acute hepatotoxicity in rats. **Materials and Methods:** *S. terebinthifolius* extract was orally administered in a dose of 350 mg dried extract/kg b.wt. before and after intoxication with CCl<sub>4</sub> for curative and protective experiments, respectively. A group of hepatotoxicity indicative enzymes, oxidant-antioxidant capacity, DNA oxidation, and apoptosis markers were measured. **Results:** CCl<sub>4</sub> increased liver enzyme leakage, oxidative stress, hepatic apoptosis, DNA oxidation, and inflammatory markers. Administration of *S. terebinthifolius*, either before or after CCl<sub>4</sub> intoxication, significantly decreased elevated serum liver enzymes and reinstated the antioxidant capacity. Interestingly, *S. terebinthifolius* extract inhibited hepatocyte apoptosis as revealed by approximately 20 times down-regulation in caspase-3 expression when compared to CCl<sub>4</sub> untreated group. On the other hand, there was neither protective nor curative effect of *S. terebinthifolius* against DNA damage caused by CCl<sub>4</sub>. **Conclusion:** The present study suggests that *S. terebinthifolius* extract could be a substantially promising hepatoprotective agent against CCl<sub>4</sub> toxic effects and may be against other hepatotoxic chemical or drugs.

**Key words:** Antioxidant, apoptosis, hepatoprotective, hepatotoxicity, *Schinus terebinthifolius*

## INTRODUCTION

Liver is the most important organ responsible for xenobiotic metabolisms, acting as sensitive target site for substance modulating biotransformation.<sup>[1]</sup> During aerobic metabolic reactions, considerable amounts of reactive oxygen species (ROS) such as superoxide anion (O<sup>2-</sup>) and hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) are produced,<sup>[2]</sup> which enter a variety of chain reactions and generate free radicals such as <sup>•</sup>OH. These hydrogen species assault polyunsaturated fatty acids and thereby initiate the process of lipid peroxidation causing degradation and inactivation

of various important biomolecules.<sup>[3]</sup> This situation resulting in many complications such as; oxidative stress, antioxidant alteration, decreasing of the intracellular level of glutathione (GSH), reduction of catalase (CAT) activity which leading to tissue injury and carcinogenesis.<sup>[4,5]</sup>

Many xenobiotics (drugs and environmental chemicals) are capable of causing some degree of liver injury.<sup>[6]</sup> As lifestyle changes, hepatic injuries have become one of the most common diseases in the world attributed to great morbidity and mortality especially in developing countries.<sup>[7]</sup> Therefore, many efforts have been made recently for discovering new products of natural origin, which can limit the xenobiotic-induced hepatic injury.

Carbon tetrachloride (CCl<sub>4</sub>) is widely used to study hepatotoxicity in animal models by starting lipid peroxidation.<sup>[8]</sup> CCl<sub>4</sub> is biotransformed in the liver forming

### Address for correspondence:

Dr. Waleed Fathy Khalil, Departments of Veterinary Pharmacology, Faculty of Veterinary Medicine, Suez Canal University, Ismailia 41522, Egypt.  
E-mail: wk@vet.suez.edu.eg

### Access this article online

#### Website:

www.phcog.com

#### DOI:

10.4103/0973-1296.157705

#### Quick Response Code:



reactive metabolic trichloromethyl radicals ( $\cdot\text{CCl}_3$ ) which reacts with excess  $\text{O}_2$  producing reactive free radicals. These free radicals begin peroxidation of membrane polyunsaturated fatty acids and covalently bind microsomal lipids and proteins producing lipid peroxides leading to cellular disorders and pathological alterations.<sup>[9,10]</sup>

Herbs have been an important source of natural materials used for treatment and/or prevention of several illnesses.<sup>[11]</sup> *S. terebinthifolius*, also named Brazilian pepper, aroeira-vermelha, pink pepper, mastic tree or christmas-berry, belongs to the *Anacardiaceae* family of plants.<sup>[12]</sup> *Anacardiaceae* family comprises 800 species in 82 genera. The genus *Schinus*, which is named after the Greek name of the mastic tree (*Schinus*), is used for decoration, tanning agent and as spice. Among the 25 species, *Schinus terebinthifolius* was growing in Egypt several decades ago.<sup>[13]</sup> *S. terebinthifolius* poses antifungal, healing, analgesic, anti-inflammatory, antioxidant, anti-allergic, and insecticidal activities.<sup>[12,14-16]</sup> Marzouk *et al.*<sup>[17]</sup> reported that aqueous methanolic extract of the leaves of *Schinus molle* showed moderate to strong radical scavenging properties on lipid peroxidation, hydroxyl radical, and  $\text{O}_2^{\cdot-}$  generations. Meanwhile, the possibility of *S. terebinthifolius* to protect the liver from drugs or toxins was not studied yet. Therefore, this study was carried out to examine the possible hepatoprotective effect of *S. terebinthifolius* against  $\text{CCl}_4$  induced liver toxicity in rats.

## MATERIALS AND METHODS

### Animals

A total of 42 adult male Wistar rats with initial body weights of  $206 \pm 12$  g and 8 weeks age were obtained from the Animal House Colony of the National Research Center, Dokki, Cairo, Egypt. Rats were allowed to acclimatize to their new environment for 2 weeks prior to the start of the experiment. The rats were housed in stainless steel cages under proper environmental conditions on a 12 h light-dark cycle. Rats were received a commercial pellet diet and water *ad libitum*.

### Ethics statement

The laboratory animal ethical committee of Veterinary Medicine Faculty has approved this study; rats were maintained and received human care in accordance with the guiding principles for the Care and Use of Animals. The experimental design was approved by the Scientific Ethical Committee, Faculty of Veterinary Medicine, Suez Canal University (Protocol 014/2013). All the efforts were taken to minimize the animal suffering and to decrease the number of animals used.

### Chemicals and reagents

Kits of alanine aminotransferase (ALT), aspartate aminotransferase (AST), total bilirubin (TB),  $\gamma$  glutamyl transferase (GGT), lactate dehydrogenase (LDH), GSH S-transferase reduced GSH, malondialdehyde (MDA), superoxide dismutase (SOD), CAT, glutathione peroxidase (GPx), and glutathione S-transferase (GST) were purchased from Biodiagnostics Co. (Cairo, Egypt). C-reactive protein (CRP) and  $\alpha_1$ -fetoprotein (AFP) levels were analyzed using diagnostic kits (Roche Diagnostics, Cairo, Egypt). Immunohistochemical kits and antibodies were obtained from abcam<sup>®</sup> (Cambridge, MA, USA). 8-hydroxydeoxyguanosine (8-OHdG) was measured by the Oxi Select<sup>™</sup> oxidative DNA damage ELISA kit (Cell Biolabs, Inc., USA).  $\text{CCl}_4$  and all other chemicals of analytical grade were purchased from Sigma Chemical Co. (St Louis, Mo, USA).

### Plant collection and extract preparation

The aerial parts of *S. terebinthifolius* were collected from Ismailia, Egypt. The leaves were washed by fresh water to remove the contaminants, and then dried in hot air oven at  $40^\circ\text{C}$ . The dried leaves were grounded into powder with a laboratory electrical mill, de-fattened by *n*-hexane and then extracted in 70% ethanol at  $4^\circ\text{C}$  for 24 h. The extract was filtered and evaporated till dryness under reduced pressure using a rotatory evaporator (rotavapor RE111, BÜCHI). The obtained dried extract (each 1 kg dry leaves yield 126 g dry ethanolic extract) was stored in dry cold place until used in the experiment.<sup>[18]</sup> Ethanol: Deionized water (70:30) was selected as an organic polar solvent to be able to extract the vacuolar components (such as total phenolic compounds including flavonoids exist in glycosides form) of *S. terebinthifolius* leaves.<sup>[19]</sup>

### Phytochemical constituents detection and quantification

Total flavonoid contents (TFC) and total phenolic contents (TPC) were measured in *Schinus terebinthifolius* extract following the method described by Park *et al.*<sup>[20]</sup> and Kim *et al.*<sup>[21]</sup> respectively. The TFC and the TPC were expressed as mg of rutin equivalents (RE) and mg of gallic acid equivalents per gram dried extract, respectively. Thin layer chromatography (TLC) was performed for detection and semi-quantification of other phytochemicals such as alkaloids, cardiac glycosides, saponins, tannins, terpenoids, quinones, and essential oils. Silica gel 60 F254 (0.25 mm thickness) plates (Merck Co, Darmstadt, Germany) were used as a stationary phase. Four mobile phases were used for separations; *n*-hexane: Ethyl acetate (8:2), chloroform: Methanol: Water (8:2:0.2), ethyl acetate: Chloroform: Methanol: Water (12:8:8:2) and chloroform: Methanol: Water (65:35:10). The spots have been detected under UV at 254 and 366 nm and sprayed with specific reagents to visualize some major phytochemicals.

These reagents were Dragendorff reagent for alkaloids, Kedde reagent for cardiac glycosides, potassium hydroxide reagent for anthraquinones (red), anthonies (yellow), and coumarins group (blue) at UV-366 nm. Also, anisaldehyde reagent and vanillin-sulfuric reagent were used for detection of terpenoids, steroids, and essential oils. Saponins and tannins were detected using forth and gelatin-salt block tests respectively.<sup>[22]</sup>

### Acute oral toxicity

Median lethal dose (LD<sub>50</sub>) of *S. terebinthifolius* ethanolic extract was determined using 12 male Wistar rats. Graduated doses between 0.5 and 5.0 g dried extract/kg b.wt. were reconstituted in 1 mL saline and administered orally using the stomach tube. Rats were observed for clinical signs of toxicity and mortality for 24 h. LD<sub>50</sub> for the extract was determined according to the modified up-and-down technique<sup>[23]</sup> to reduce the number of animals used. LD<sub>50</sub> value was calculated using AOT425 statistical program (Version 1.0, Westat, NW, Washington DC, USA) developed by Westat for US EPA.

### Experimental design

#### Preventive study

Fifteen rats were divided into three groups (five animals of each). Control group was injected interapretonially (i.p.) with 1 ml physiological saline instead of CCl<sub>4</sub>. CCl<sub>4</sub> intoxicated group was given a single i.p. dose of 1 ml CCl<sub>4</sub>/kg b.wt. according to Manoj and Aqued.<sup>[24]</sup> Preventive group was orally administered 350 mg *S. terebinthifolius* dried extract/kg b.wt dissolved in 1 ml saline for 7 consecutive days prior to CCl<sub>4</sub> injection.

#### Treatment study

Control and CCl<sub>4</sub> groups were treated same as in preventive study. Treatment group was orally administered 350 mg *S. terebinthifolius* dried extract/kg b.wt. reconstituted in 1 ml saline for 14 consecutive days after CCl<sub>4</sub> injection. Number of animals used for treatment study was 15 rats.

Dose of *S. terebinthifolius* was decided according to preliminary pilot study aimed to determine the least effective dose. This preliminary study revealed a good protective effect at a dose of 300 mg dried extract/kg b.wt and a moderate curative effect at a dose of 400 mg extract/Kg b.w.

#### Blood and liver sampling

After 72 h (in preventive study) and after 2 weeks (in treatment study) of CCl<sub>4</sub> administration, all animals were lightly anesthetized using diethyl ether. Blood samples were collected from postorbital plexus and divided into two parts, first part was collected in heparinized tubes and used for the determination of selected hematological

parameters, while the second part was allowed to clot at room temperature then centrifuged at 1200 × g gravitational force (G) for 10 min to separate the serum which was used for biochemical estimation of liver integrity enzymes, antioxidants and oxidative stress biomarkers, and inflammatory markers. Rats then were sacrificed by diethyl ether anesthesia and the liver was removed from each rat and washed several times with normal saline. Part of the liver was taken for biochemical estimations of antioxidant and oxidative stress biomarkers and the remaining tissue was kept in 10% formalin buffer and used for immunohistochemical examination.

### Processing of the liver

Liver sample was washed twice with saline solution, crushed and homogenized in ice-cold 0.01 M sodium-potassium phosphate buffer (pH 7.4). The homogenate was centrifuged at 860 × g for 20 min at 4°C, and the resultant supernatant was frozen at -20°C and used for hepatic oxidative stress markers.

### Analysis of 8-Hydroxydeoxyguanosine in DNA

A commercially available ELISA kit (Cell Biolabs, inc) was used to evaluate the levels of 8-OH-dG in DNA. The assays were performed following the instructions of the manufacturer.<sup>[25]</sup> Briefly, DNA was extracted then incubated at 95°C for 5 min and rapidly chilling on ice. Digest DNA sample to nucleosides by incubating the denatured DNA with 5–20 units of nuclease P1 for 2 h at 37°C in 20 mM sodium acetate (pH 5.2), then treated with 5–10 units of alkaline phosphatase for 1 h at 37°C in 100 mM Tris buffer (pH 7.5). The reaction mixture was centrifuged for 5 min at 6000 × g, and the supernatant was used for the 8-OHdG ELISA assay. Briefly, 50 µL of unknown sample or 8-OHdG standard was incubated at room temperature for 10 min. 50 µL of the diluted anti-8-OHdG antibody was added and incubated at room temperature for 1 h. Washed 3 times, then 100 µL of the diluted secondary antibody-enzyme conjugate was added and incubate at room temperature for 1 h before re-washing. 100 µL of substrate solution was added to each well and incubate at room temperature for 20 min. Enzyme reaction was stopped by adding 100 µL of stop solution and absorbance was measured using a spectrophotometer at 450 nm (Shimadzu, model UV-3000, Kyoto-Japan).

### Activated caspase-3 immunohistochemistry

Formalin liver samples were embedded in paraffin. The paraffin blocks were cut into 5–7 µm thickness. Deparaffinization and antigen recovery were carried out. Endogenous peroxidase activity was blocked by incubation for 5 min with 3% H<sub>2</sub>O<sub>2</sub>. Sections were incubated with the primary antibody for caspase-3 overnight at 4°C. Detection kit was used for visualization of the antibody reaction to

detect and counting the number of apoptotic cells. The visualization performed according to the manufacturer instructions (Sigma, St. Louis, MO). Revelation done using 3',3'-diaminobenzidine tetrahydrochloride (Sigma, St. Louis, MO). Sections washed in distilled water, and counter stained with Mayer's Hematoxylin for 5 min then examined using Olympus CX 21 microscope under 400 × powers.

### Statistical analysis

The results were expressed as mean ± standard deviation for five animals in each group. One-way ANOVA was used followed by Holm-Sidak Multiple comparison test. Pearson's Correlation test was done for correlating parametric variables. Statistical significance was accepted at  $P \leq 0.05$ .

## RESULTS

### Phytochemical constituents detection and quantification

The quantitative and semi-quantitative phytochemical compositions of *S. terebinthifolius* ethanolic extract are shown in Table 1. TPC was found to be the major constitute, it represented about half of the total ethanolic extract dry weight. From the TPC, the TFC constituted about 8% with a concentration of  $38.33 \pm 1.45$  mg REs/g dried extract. Tannins, saponins, and essential oils were also detected in medium to high concentrations in the extract while no alkaloids, cardiac glycosides, terpenoids, quinones, anthraquinones or coumarins could be detected in the extract.

### Acute oral toxicity

Oral administration of *S. terebinthifolius* ethanolic extract up to 5 g/kg b.wt. did not show any toxic signs or deaths within 24 h from extract administration. Moreover, no signs of delayed toxicity were perceived when rats were observed for further 14 days.

### Serum hepatic parameters

The effects of *S. terebinthifolius* on serum transaminases, bilirubin, and LDH, are summarized in Table 2. There

were significant increases ( $P \leq 0.05$ ) in liver enzymes leakage (AST, ALT, GGT and LDH) as well as blood bilirubin concentrations in  $\text{CCl}_4$  intoxicated rats when compared to control groups in both preventive and treatment experiments. Administration of *S. terebinthifolius*, either for 1 week before or for 2 weeks after  $\text{CCl}_4$  injection, significantly ( $P \leq 0.05$ ) decreased liver enzymes leakage and restored the altered bilirubin to approximately the control levels.

### Inflammatory markers

While there were significant increases in CRP levels in all  $\text{CCl}_4$  intoxicated groups, AFP was almost unchanged in comparison with the control group [Table 2]. Using of *S. terebinthifolius* either as preventive or as a treatment in rats intoxicated with  $\text{CCl}_4$  significantly decreased the CRP levels when compared to  $\text{CCl}_4$  groups.

### Antioxidant activities and oxidative DNA damage

Significant decreases in the erythrocyte and hepatic GSH in addition to hepatic SOD, CAT, GPx and GST were observed in  $\text{CCl}_4$  intoxicated rats when compared with normal control rats. Oral administration of *S. terebinthifolius* extract in both preventive and treatment groups reinstated the GSH levels in comparison with  $\text{CCl}_4$  group [Table 3]. The rats received  $\text{CCl}_4$  alone showed elevated lipid peroxidation demonstrated by high serum and liver MDA levels as compared to the control rats. Once more, administration of *S. terebinthifolius* extract before or after  $\text{CCl}_4$  showed significant decreases in serum MDA levels as compared to those in  $\text{CCl}_4$  intoxicated groups. Unlike in the preventive group, liver MDA level in the treatment group was still higher than that in the control group. The oxidative DNA damage marker (8-OHdG) was significantly elevated in the rats intoxicated with  $\text{CCl}_4$  in comparison with that in control rats. There was neither protective nor curative effect of *S. terebinthifolius* against DNA damage caused by  $\text{CCl}_4$  [Table 3]. When data were analyzed for correlations using Pearson's correlation analysis, our results showed negative correlations between liver enzymes (AST, ALT, and GGT) and the hepatic antioxidant parameters GSH, SOD, CAT, GPx, and GST.

**Table 1: Quantitative and semi-quantitative phytochemical compositions of *S. terebinthifolius* ethanolic extract**

Phytochemical	Phytochemical compositions										
	TFC (mg RE/ g DE)	TPC (mg GAE/ g DE)	Tannins	Saponins	Essential oils	Alkaloids	Cardiac glycosides	Terpenoids	Quinones	Anthraquinones	Coumarins
Amount	38.33± 1.45	488.67± 13.02	+++	++	++	--	--	--	--	--	--

For quantitative evaluation of phytochemical contents, either mean±SE (n=3) or descriptive signs were used, when (--) means absent, (+) means low, (++) means medium and (+++) means high content. TFC: Total flavonoids content, TPC: Total phenolic content, RE: Rutin equivalents, GAE: Gallic acid equivalents, *S. terebinthifolius*: *Schinus terebinthifolius*



Meanwhile, only MDA showed positive correlations with these liver enzymes [Table 4].

**Blood parameters**

There was a significant decrease in blood platelets while there was a significant increase in WBCs count in rats

treated with CCl<sub>4</sub> compared to control rats. Although pre-treatment with *S. terebinthifolius* restored these values to approximately normal values, administration of *S. terebinthifolius* after CCl<sub>4</sub> significantly decreased the WBCs but not platelets counts when compared to CCl<sub>4</sub> group [Table 5]. When the correlation coefficient (*r*) was

**Table 2: Liver serum parameters and inflammatory markers in control rats, rats intoxicated with CCl<sub>4</sub> and rats received *S. terebinthifolius* either before CCl<sub>4</sub> (prevention group) or after CCl<sub>4</sub> (treatment group)**

Parameters	Experimental groups					
	Preventive study			Treatment study		
	Control group	CCl <sub>4</sub> group	Prevention group	Control group	CCl <sub>4</sub> group	Treatment group
AST (u/L)	21.8±2.77	55.2±12.8*	24.4±3.85 <sup>#</sup>	20.8±2.17	58.4±12.5*	38.6±6.58 <sup>#</sup>
ALT (u/L)	27.6±5.13	128±10.6*	32.0±3.16 <sup>#</sup>	27.2±3.96	134±13.6*	55.2±5.97 <sup>#</sup>
GGT (u/L)	4.20±2.05	9.50±1.35*	4.80±1.48 <sup>#</sup>	3.80±1.48	9.82±0.89*	7.20±1.48 <sup>#</sup>
TB	0.24±0.11	1.22±0.41*	0.26±0.15 <sup>#</sup>	0.22±0.13	1.14±0.23*	0.38±0.08 <sup>#</sup>
Serum LDH (u/L)	322±47	830±77*	416±51 <sup>#</sup>	341±42	815±86*	667±65 <sup>#</sup>
Liver LDH (u/g)	453±59	718±68*	506±67 <sup>#</sup>	441±62	737±76*	622±68 <sup>#</sup>
Serum/liver LDH ratio	0.71±0.08	1.16±0.11*	0.82±0.09 <sup>#</sup>	0.77±0.09	1.15±0.10*	1.07±0.10
CRP (mg/L)	1.60±0.43	5.76±1.28*	1.82±0.36 <sup>#</sup>	1.66±0.37	5.88±0.83*	3.60±0.88 <sup>#</sup>
AFP (IU/ml)	3.70±0.80	3.60±0.92	3.68±0.91	3.56±0.86	3.80±0.75	3.72±1.13

Data are expressed as means±SD (n=5). One-way ANOVA followed by Holm-Sidak multiple comparison test was used: \*P≤0.05 versus control group, <sup>#</sup>P≤0.05 versus CCl<sub>4</sub> group. CCl<sub>4</sub>: Carbon tetrachloride; AST: Aspartate aminotransferase; ALT: Alanine aminotransferase; GGT: Gama glutamyl transferase; TB: Total bilirubin; LDH: Lactate dehydrogenase; CRP: C-reactive protein; AFP: α<sub>1</sub>-fetoprotein; *S. terebinthifolius*: *Schinus terebinthifolius*

**Table 3: Oxidant and antioxidant parameters in control rats, rats intoxicated with CCl<sub>4</sub> and rats received *S. terebinthifolius* either before CCl<sub>4</sub> (prevention group) or after CCl<sub>4</sub> (treatment group)**

Parameters	Experimental groups					
	Preventive study			Treatment study		
	Control group	CCl <sub>4</sub> group	Prevention group	Control group	CCl <sub>4</sub> group	Treatment group
Serum MDA (µmol/L)	3.44±0.73	7.82±1.22*	3.42±0.37 <sup>#</sup>	3.74±0.29	7.90±0.71*	6.54±0.62 <sup>#</sup>
Erythrocyte GHS (mg/dl)	20.6±2.07	12.4±0.86*	18.6±3.05 <sup>#</sup>	22.4±2.88	12.2±0.96*	16.0±2.00 <sup>#</sup>
Liver MDA (µmol/g)	2.48±0.36	5.58±1.16*	2.34±0.30 <sup>#</sup>	2.58±0.30	5.44±1.19*	4.16±0.61
Liver GSH (mg/g)	29.0±7.69	18.74±1.25*	27.92±4.59 <sup>#</sup>	29.28±6.75	17.1±2.02*	24.1±1.78 <sup>#</sup>
Liver SOD (u/g)	448±30.5	214±17.6*	399±27.7 <sup>#</sup>	431±28.9	239±20.7*	310±34.5 <sup>#</sup>
Liver CAT (u/g)	1.87±0.07	0.79±0.009*	1.61±0.09 <sup>#</sup>	1.79±0.08	0.84±0.01*	1.23±0.06 <sup>#</sup>
Liver GPx (u/g)	1.38±0.05	0.81±0.03*	1.22±0.05 <sup>#</sup>	1.34±0.06	0.84±0.04*	1.12±0.05 <sup>#</sup>
Liver GST (µmol/g)	1529±123	735±53*	1168±97 <sup>#</sup>	1486±112	760±64*	981±73 <sup>#</sup>
Hepatic 8-OHdG (pg/ml)	348±18.1	471±25.9*	492±87.7	350±23.5	635±34.4*	617±58.1

Data are expressed as means±SD (n=5). One-way ANOVA followed by Holm-Sidak multiple comparison test was used: \*P≤0.05 versus control group, <sup>#</sup>P≤0.05 versus CCl<sub>4</sub> group. CCl<sub>4</sub>: Carbon tetrachloride; GSH: Glutathione; MDA: Malondialdehyde; SOD: Superoxide dismutase; CAT: Catalase; GPx: Glutathione peroxidase; GST: Glutathione S-transferase; 8-OHdG: 8-hydroxydeoxyguanosine; *S. terebinthifolius*: *Schinus terebinthifolius*

**Table 4: Pearson’s correlation coefficients among the serum liver enzymes and the oxidant-antioxidant markers in the prevention and treatment studies**

Parameters	Experimental groups					
	Preventive study			Treatment study		
	AST	ALT	GGT	AST	ALT	GGT
MDA	r=0.834*	r=0.945*	r=0.832*	r=0.697*	r=0.800*	r=0.614
GSH	r=-0.830*	r=-0.832*	r=-0.872*	r=-0.525	r=-0.800*	r=-0.815*
SOD	r=-0.716*	r=-0.947*	r=-0.799*	r=-0.639*	r=-0.847*	r=-0.672*
CAT	r=-0.727*	r=-0.971*	r=-0.770*	r=-0.721*	r=-0.946*	r=-0.760*
GPx	r=-0.741*	r=-0.969*	r=-0.773*	r=-0.871*	r=-0.993*	r=-0.881*
GST	r=-0.789*	r=-0.938*	r=-0.654*	r=-0.775*	r=-0.903*	r=-0.724*

\*Significant at P≤0.05. AST: Aspartate aminotransferase; ALT: Alanine aminotransferase; GGT: Gama glutamyl transferase; MDA: Malondialdehyde; GSH: Glutathione; SOD: Superoxide dismutase; CAT: Catalase; GPx: Glutathione peroxidase; GST: Glutathione S-transferase; r: Correlation coefficient

**Table 5: Blood parameters in control rats, rats intoxicated with CCl<sub>4</sub> and rats received *S. terebinthifolius* either before (prevention group) or after (treatment group) CCl<sub>4</sub> intoxication**

Parameters	Experimental groups					
	Preventive study			Treatment study		
	Control group	CCl <sub>4</sub> group	Prevention group	Control group	CCl <sub>4</sub> group	Treatment group
Hb (g/dL)	12.28±0.72	12.0±1.18	13.0±0.80	12.5±0.57	12.2±0.84	12.7±0.85
BP (10 <sup>3</sup> /μL)	1128±40.5	891±53.6*	1121±58.6 <sup>#</sup>	1117±53.2	899±65.6*	947±55.0
WBCs (10 <sup>3</sup> /μL)	11.3±0.68	16.8±1.51*	10.1±0.74 <sup>#</sup>	11.5±0.75	16.6±1.30*	13.5±0.60 <sup>#</sup>

Data are expressed as means±SD (n=5). One-way ANOVA followed by Holm-Sidak Multiple comparison test was used: \*P≤0.05 versus control group, <sup>#</sup>P≤0.05 versus CCl<sub>4</sub> group. SD: Standard deviation; CCl<sub>4</sub>: Carbon tetrachloride; Hb: Hemoglobin; BP: Blood platelets; WBCs: White blood cells; *S. terebinthifolius*: *Schinus terebinthifolius*

calculated using Person's correlation analysis, our data showed significant negative correlations between blood platelets count and TB ( $r = -0.824$  and  $-0.442$  in prevention and treatment studies, respectively), whereas there were significant positive correlations between WBCs count and CRP levels in prevention and treatment studies ( $r = 0.889$  and  $0.778$ , respectively) using Pearson's correlation.

### Immunohistochemical findings

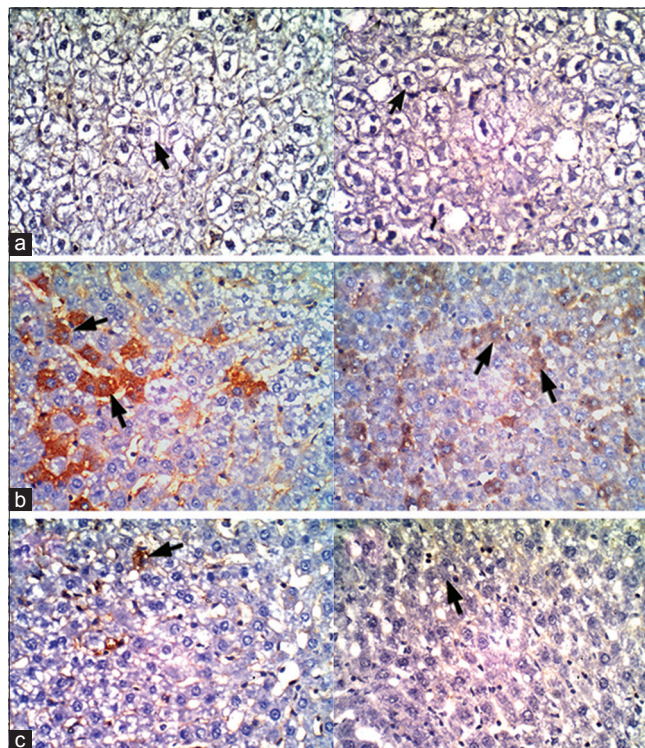
The expression levels of cleaved caspase-3 in the livers of CCl<sub>4</sub> intoxicated rats were relatively high ( $16.8 \pm 1.48$  cells/HPF in prevention study and  $22.6 \pm 3.36$  cells/HPF in treatment study). Interestingly, these levels were significantly decreased when CCl<sub>4</sub> intoxicated rats were administered *S. terebinthifolius* extract ( $1.2 \pm 0.45$  cells/HPF in prevention study and  $0.4 \pm 0.55$  cell/HPF in treatment study) as detected by immunohistochemistry [Figure 1].

## DISCUSSION

In this study, the TPC was found to be the major constitute of *S. terebinthifolius* ethanolic extract. Besides, the TFC was found to be  $38.33 \pm 1.45$  mg (REs) per gram of dried extract. These two constituents are good indicators for the antioxidant activity of plant extracts. Many researchers reported a significantly high correlation between the high antioxidant activity of plant extracts and its high TPC and TFC.<sup>[26]</sup>

Median lethal dose was estimated to evaluate the acute toxicity of *S. terebinthifolius* ethanolic extract aiming to use one-twentieth of LD<sub>50</sub> in our experiment. Our result revealed neither toxic signs nor deaths up to a dose of 5 g dried extract/kg b.wt. Substances of oral LD<sub>50</sub> < 5000 mg/kg are considered practically nontoxic.<sup>[27]</sup> Accordingly, the dose of *S. terebinthifolius* in this study was decided based on its effective not on its toxicity.

Carbon tetrachloride is one of the most frequently used hepatotoxins to study liver diseases in experimental animal models; it has been reported to show many hepatic disorders similar to that perceived in viral hepatitis



**Figure 1:** Immunohistochemical staining of the liver sections (×400) stained with caspase 3 antibodies for apoptotic cells and hematoxylin counterstaining of the nuclei and non-apoptotic cells. (a) control groups, (b) CCl<sub>4</sub> groups and (c) rats received ethanolic extract of *Schinus terebinthifolius* either before [left side pictures] or after [right side pictures] CCl<sub>4</sub>-intoxications. Arrowheads in figures (b) point out caspase-3 positive apoptotic cells

or chemically-induced liver injury.<sup>[28-31]</sup> In liver, CCl<sub>4</sub> transformed into trichloromethyl radicals those react with excess O<sub>2</sub> resulting in the production of reactive free radicals and stimulation of lipid peroxidation followed by cellular damage, inflammation, and pathological changes.<sup>[10,32-33]</sup> Administration of CCl<sub>4</sub> significantly increased the leakage of liver ALT, AST, GGT, and LDH enzymes into the serum; it also increased blood TB. ALT, AST, GGT, and LDH are sensitive indicators for the hepatic function and their excessive leakage into the blood is usually associated with impaired hepatocellular function and designates the disruption of integrity of hepatic cell

membranes.<sup>[34-37]</sup> In the present study, CCl<sub>4</sub> administration increased lipid peroxidation via elevation of serum and hepatic MDA levels and decreasing GSH antioxidant capacity in erythrocytes and liver as well as decreasing the activities of hepatic SOD, CAT, GPx, and GST [Table 3]. These effects are implicated in hepatic oxidative stress and toxicity. This pointed out that CCl<sub>4</sub>-mediated hepatic injury occurred as a result of excessive production of ROS, which attack various biological molecules including lipids and causing lipid peroxidation. These results are parallel with those obtained by many other researchers.<sup>[8,38,39]</sup> CRP is an acute-phase protein that produced by hepatocytes and it is a sensitive marker for inflammation, infection, and tissue damage.<sup>[40]</sup> The results in this study revealed that CCl<sub>4</sub> administration induced liver inflammation as there were significant increases in CRP levels coupled with an increase in WBCs count. Administration of *S. terebinthifolius* pre and post CCl<sub>4</sub> intoxication significantly reduced the elevated liver enzymes, CRP, and WBCs count; these findings designate the protection of the hepatocytes integrity or renewal of injured hepatocytes.<sup>[34]</sup> In addition, there were elevations in antioxidant enzymes and reduction of the lipid peroxidation in blood and livers of groups those received *S. terebinthifolius* compared to those of CCl<sub>4</sub>-intoxicated groups. These findings verify the antioxidant effects of *S. terebinthifolius* and explain its restorative effect that could be attributed to its ability to minimize the production of free radicals and/or to enhance the scavenger activities. The antioxidant activity of *S. terebinthifolius* was confirmed by other researchers.<sup>[14,16,17]</sup>

8-OHdG is a specific marker for DNA oxidation.<sup>[41]</sup> The current study showed significant elevation in 8-OHdG in CCl<sub>4</sub>-intoxicated groups when compared to the control groups. Such elevation in 8-OHdG levels can be explained by the oxidative alteration of macromolecules and consequently genomic unsteadiness caused by the formation of excessive ROS.<sup>[42]</sup> In our study, *S. terebinthifolius* failed to protect hepatocyte DNA from oxidative damage or reverse the damage. This could be attributed to the pharmacokinetic properties of the antioxidant substances (mainly the flavonoids) inside the liver cells. When the CCl<sub>4</sub> is a non-polar compound and can easily pass both cellular and nuclear membranes causing damage to DNA, it seemed that the flavonoids in *S. terebinthifolius* extract become more polar and less fat soluble inside the liver cells through binding with one or more intracellular compounds such as proteins (which its concentrations are 8 times higher than extracellular concentrations). Specially that the polarity of flavonoids depends to a large degree on whether they occur bound to one or more polar moiety (such as sugar), which renders them highly polar, or whether they occur in the free form in which case they are much less polar.<sup>[19]</sup>

As a general, the present data showed a significant positive correlation between liver function enzymes and the oxidant-antioxidant markers, which confirms the antioxidant mechanism of *S. terebinthifolius* as a hepatoprotective agent.

Apoptosis of hepatocytes is mainly initiated by activation of the caspase family of cysteine proteases, which is a major form of cell death.<sup>[43]</sup> Our findings indicated that a substantial number of hepatocytes undergo apoptosis after CCl<sub>4</sub> administration; this finding was in consistent with those obtained by other researchers.<sup>[44-46]</sup> Our results also revealed that *S. terebinthifolius* intensely inhibited hepatocyte apoptosis by down-regulating the expression of caspase-3 compared with CCl<sub>4</sub> group. Besides the decreased LDH leakage, which could be a beneficial marker for the integrity of the outer cell membrane, that result confirmed the protective and therapeutic effects of *S. terebinthifolius* against CCl<sub>4</sub>-induced hepatotoxicity.

In summary, the present study affirmed a beneficial ability of *S. terebinthifolius* ethanolic extract to reduce CCl<sub>4</sub>-induced liver damage. The markers of protection include significantly decreased serum hepatic enzyme, increased antioxidant capacity, and decreased hepatocyte apoptosis when compared with the CCl<sub>4</sub> treatment group. This hepatoprotective effect might be attributed to its antioxidant and free radical scavenging effects. Such findings put up *S. terebinthifolius* as a promising hepatoprotective agent against CCl<sub>4</sub> toxic effects and may be against other hepatotoxic chemicals or drugs, both in animal and human.

## ACKNOWLEDGMENTS

The authors would like to thank Dr. Ahmed M. Abdel-Azeem (Botany and Microbiology Department, Faculty of Science, Suez Canal University, Egypt) for identification and authentication of *S. terebinthifolius* and supervising its collection and preparation.

## REFERENCES

1. Madkour FF, Khalil WF, Dessouki AA. Protective effect of ethanol extract of *Sargassum Dentifolium* (*Phaeophyceae*) in carbon tetrachloride induced hepatitis in rats. *Int J Pharm Pharm Sci* 2012;4:637-41.
2. Gupta M, Mazumder UK, Thamilselven V, Manikandan L, Senthilkumar GP. Potential hepatoprotective effect and antioxidant role of ethanol extract of *Oldenlandia umbellata* in carbon tetrachloride induced hepatotoxicity in Wistar rats. *Iran J Pharmacol Ther* 2007;6:5-9.
3. Pari L, Murugan P. Protective role of tetrahydrocurcumin against erythromycin estolate-induced hepatotoxicity. *Pharmacol Res* 2004;49:481-6.
4. Halliwell B, Gutteridge JM. *Free Radicals in Biology and Medicine*. 3<sup>rd</sup> ed. New York: Oxford University Press; 1999. p. 617-783.



5. Stal P, Olson J. Ubiquinone: Oxidative stress, and liver carcinogenesis. In: Kagan VE, Quinn DJ, editors. Coenzyme Q: Molecular Mechanisms in Health and Disease. Boca Raton: CRC Press; 2000. p. 317-29.
6. Sturgill MG, Lambert GH. Xenobiotic-induced hepatotoxicity: Mechanisms of liver injury and methods of monitoring hepatic function. Clin Chem 1997;43:1512-26.
7. Lau DT, Membreno FE. Antiviral therapy for treatment-naïve hepatitis B virus patients. Gastroenterol Clin North Am 2004;33:581-99, ix.
8. Tirkey N, Pilkhwai S, Kuhad A, Chopra K. Hesperidin, a citrus bioflavonoid, decreases the oxidative stress produced by carbon tetrachloride in rat liver and kidney. BMC Pharmacol 2005;5:2.
9. Hiraganahalli BD, Chinampudur VC, Detha S, Mundkinajeddu D, Pandre MK, Balachandran J, et al. Hepatoprotective and antioxidant activity of standardized herbal extracts. Pharmacogn Mag 2012;8:116-23.
10. Nazima B, Manoharan V, Prabu SM. Cadmium toxicity: Oxidative stress and organ dysfunction. Res Rev J Toxicol 2014;4:1-18.
11. Feroz Z, Khan RA, Amber F, Mahayrookh A. Hepatoprotective effect of herbal drug on CCl<sub>4</sub> (4) induced liver damage. Pak J Pharm Sci 2013;26:99-103.
12. Affonso CR, Fernandes RM, Oliveira JM, Martins MC, Lima SG, Sousa Júnior GR, et al. Effects of the essential oil from fruits of *Schinus terebinthifolius* Raddi (*Anacardiaceae*) on reproductive functions in male rats. J Braz Chem Soc 2012;23:180-5.
13. Glen HF. Cultivated Plants of Southern Africa. Botanical Names, Common Names, Origins, Literature. Johannesburg, South Africa: Jacana; 2002. p. 404.
14. Chowdhury AR, Tripathi S. Essential oil from leaves of *S. terebinthifolius* Raddi. Indian Perfumer 2001;4:257-9.
15. Di Stasi LC, Oliveira GP, Carvalhaes MA, Queiroz M Jr, Tien OS, Kakinami SH, et al. Medicinal plants popularly used in the Brazilian Tropical Atlantic Forest. Fitoterapia 2002;73:69-91.
16. Cavalher-Machado SC, Rosas EC, Brito Fde A, Heringe AP, de Oliveira RR, Kaplan MA, et al. The anti-allergic activity of the acetate fraction of *Schinus terebinthifolius* leaves in IgE induced mice paw edema and pleurisy. Int Immunopharmacol 2008;8:1552-60.
17. Marzouk MS, Moharram FA, Haggag EG, Ibrahim MT, Badary OA. Antioxidant flavonol glycosides from *Schinus molle*. Phytother Res 2006;20:200-5.
18. Jirayus W, Kanok-Orn I, Korakod I. Antioxidant activity and cytotoxicity of six selected, regional, Thai vegetables. Am Eurasian J Toxicol Sci 2012;4:108-17.
19. Bohm BA. Introduction to flavonoids. Chemistry and Biochemistry of Organic Natural Products. Vol 2. Amsterdam: Harwood Academic Publishers; 1998. p. 176-7.
20. Park YS, Jung ST, Kang SG, Heo BK, Arancibia-Avila P, Toledo F, et al. Antioxidants and proteins in ethylene-treated kiwifruits. Food Chem 2008;107:640-8.
21. Kim DO, Jeong SW, Lee CY. Antioxidant capacity of phenolic phytochemicals from various cultivars of plums. Food Chem 2003;81:321-6.
22. Harborne JB. Phytochemical Methods. A Guide to Modern Techniques of Plant Analysis. London, UK: Chapman and Hall; 1998. p. 302.
23. Bruce RD. A confirmatory study of the up-and-down method for acute oral toxicity testing. Fundam Appl Toxicol 1987;8:97-100.
24. Manoj B, Aqueed K. Protective effect of *Lawsonia inermis* against CCl<sub>4</sub> induced hepatic damage in albino rats. Indian J Exp Biol 2003;41:85-7.
25. Shen J, Deiningner P, Hunt JD, Zhao H. 8-Hydroxy-2'-deoxyguanosine (8-OH-dG) as a potential survival biomarker in patients with nonsmall-cell lung cancer. Cancer 2007;109:574-80.
26. Sarepoua E, Tangwongchai R, Suriharn B, Lertrat K. Relationships between phytochemicals and antioxidant activity in corn silk. Int Food Res J 2013;20:2073-9.
27. Hodge HC, Sterner JH. Combined tabulation of toxicity classes. In: Spector WS, editor. Handbook of Toxicology. Vol. 1. Philadelphia: W. B. Saunders Company; 1956. p. 37.
28. Sannigrahi S, Mazumder UK, Pal D, Mishra SL. Hepatoprotective potential of methanol extract of *Clerodendrum infortunatum* Linn. against CCl<sub>4</sub> induced hepatotoxicity in rats. Pharmacogn Mag 2009;5 Suppl S1:394-9.
29. Williams AT, Burk RF. Carbon tetrachloride hepatotoxicity: An example of free radical-mediated injury. Semin Liver Dis 1990;10:279-84.
30. Johnston DE, Kroening C. Mechanism of early carbon tetrachloride toxicity in cultured rat hepatocytes. Pharmacol Toxicol 1998;83:231-9.
31. Shenoy KA, Somayaji SN, Bairy KL. Hepatoprotective effects of *Ginkgo Biloba* against tetrachloride induced hepatic injury in rats. Indian J Pharmacol 2001;33:260-6.
32. Basu S. Carbon tetrachloride-induced lipid peroxidation: Eicosanoid formation and their regulation by antioxidant nutrients. Toxicology 2003;189:113-27.
33. Fu Y, Zheng S, Lin J, Ryerse J, Chen A. Curcumin protects the rat liver from CCl<sub>4</sub>-caused injury and fibrogenesis by attenuating oxidative stress and suppressing inflammation. Mol Pharmacol 2008;73:399-409.
34. Patrick-Iwuanyanwu KC, Wegwu MO, Ayalogu EO. Prevention of CCl<sub>4</sub>-induced liver damage by ginger, garlic and vitamin E. Pak J Biol Sci 2007;10:617-21.
35. Firdous SM, Sravanthi K, Debnath R, Neeraja K. Protective effect of ethanolic extract and its ethylacetate and n-butanol fractions of fruits against carbon tetrachloride induced hepatic injury rats. Int J Pharm Pharm Sci 2012;4:354-9.
36. Bhakta T, Mukherjee PK, Mukherjee K, Banerjee S, Mandal SC, Maity TK, et al. Evaluation of hepatoprotective activity of *Cassia fistula* leaf extract. J Ethnopharmacol 1999;66:277-82.
37. Wafay H, El-Saeed G, El-Toukhy S, Youness E, Ellaithy N, Agaibi M, et al. Potential effect of garlic oil and Silymarin on carbon tetrachloride-induced liver injury. Aust J Basic Appl Sci 2012;6:409-14.
38. Khan RA, Khan MR, Sahreen S. CCl<sub>4</sub>-induced hepatotoxicity: Protective effect of rutin on p53, CYP2E1 and the antioxidative status in rat. BMC Complement Altern Med 2012;12:178.
39. Al-Fartosi KG, Majid A, Auda AM, Hussein MH. The Role of Camel's Milk against some oxidant-antioxidant markers of male rats treated with CCl<sub>4</sub>. Int J Res Pharm Biomed Sci 2012;3:385-9.
40. Husain MT, Kim HD. C-Reactive protein and erythrocyte sedimentation rate in orthopaedics. Univ Pa Orthop J 2002;15:13-6.
41. Subash P, Gurumurthy P, Sarasabharathi A, Cherian KM. Urinary 8-OHdG: A marker of oxidative stress to DNA and total antioxidant status in essential hypertension with South Indian population. Indian J Clin Biochem 2010;25:127-32.
42. Azad N, Rojanasakul Y, Vallyathan V. Inflammation and lung cancer: Roles of reactive oxygen/nitrogen species. J Toxicol Environ Health B Crit Rev 2008;11:1-15.
43. Inoue S, Browne G, Melino G, Cohen GM. Ordering of caspases in cells undergoing apoptosis by the intrinsic pathway. Cell Death Differ 2009;16:1053-61.



44. Shi J, Aisaki K, Ikawa Y, Wake K. Evidence of hepatocyte apoptosis in rat liver after the administration of carbon tetrachloride. *Am J Pathol* 1998;153:515-25.
45. Lee TY, Chang HH, Wang GJ, Chiu JH, Yang YY, Lin HC. Water-soluble extract of *Salvia miltiorrhiza* ameliorates carbon tetrachloride-mediated hepatic apoptosis in rats. *J Pharm Pharmacol* 2006;58:659-65.
46. Kuwahata M, Kubota H, Kanouchi H, Ito S, Ogawa A, Kobayashi Y, *et al.* Supplementation with branched-chain amino acids attenuates hepatic apoptosis in rats with chronic liver disease. *Nutr Res* 2012;32:522-9.

**Cite this article as:** Abdou RH, Saleh SY, Khalil WF. Toxicological and biochemical studies on *Schinus terebinthifolius* concerning its curative and hepatoprotective effects against carbon tetrachloride-induced liver injury. *Phcog Mag* 2015;11:93-101.

**Source of Support:** Nil, **Conflict of Interest:** None declared.