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Epigenetic Regulators: New Therapeutic Targets for Soft Tissue Sarcoma

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Abstract

Soft tissue sarcoma is a malignancy that develops from human soft tissues such as muscle, nerve, fat, and blood vessels. The World Health Organization classification comprises about 50 different histologic types of soft tissue sarcoma. Soft tissue sarcoma is treated most often with surgery. Chemotherapy and radiotherapy have shown only minor effects on patient survival in this disease. The overall 5-year survival rate of soft tissue sarcoma is 50%; it has not changed for the past several decades. A new class of therapeutic targets for soft tissue sarcoma was identified recently. Epigenetic regulators, such as DNA methyltransferases, histone deacetylases, and histone-modifying enzyme enhancer of zeste homolog 2, have been found to be involved in pathogenesis of various soft tissue sarcomas. Small-molecule inhibitors of these epigenetic regulators may provide a new targeted therapy approach to soft tissue sarcomas in the future.

Keywords

Soft tissue sarcoma; DNA methyltransferase; histone deacetylases; enhancer of zeste homolog 2; epigenetics

Introduction

Soft tissue sarcoma by definition is a class of tumors that develop from nonepithelial, supporting, and connective tissues of the body, including muscle, fat, fibrous tissue, nerves, lymphatic tissue, and vascular tissue. Soft tissue sarcoma can be classified on the basis of its tissue of origin into any of about 50 different histologic subtypes defined by the World Health Organization^[1]. These complex tumors are relatively rare, however. Approximately 10,000 new cases are diagnosed and 4000 deaths are attributed to soft tissue sarcoma in the USA every year^[1]. Because of its heterogeneity and rarity, soft tissue sarcoma represents a therapeutic challenge.

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For primary soft tissue sarcoma, surgical resection remains the standard of care. Surgery combined with adjuvant radiotherapy and chemotherapy has been shown to decrease the local recurrence rate and improve the disease-free and overall survival rates of patients with this tumor^[2]. For unresectable or metastastic soft tissue sarcoma, doxorubicin is the single most effective and widely used drug. A combination of doxorubicin and ifosfamide is currently the standard treatment for such cases^[3]. Although systemic treatment increases the survival rate and quality of life among patients with metastatic disease, the overall 5-year survival rate for all soft tissue sarcoma cases is still approximately 50%. In order to further improve the survival rate and quality of life for soft tissue sarcoma patients, new therapeutic options are urgently needed. This paper reviews promising areas of research on molecular regulators of sarcoma oncogenesis and progression, especially epigenetic regulators as novel therapeutic targets for this disease.

Molecular Regulation of Sarcoma Tumorigenesis

To identify novel therapeutic targets and more effective drugs for soft tissue sarcoma, better understanding of the underlying molecular and genetic mechanisms is a necessity. Activating mutations in the *c-KIT* (or *PDGFRA*) gene have been identified as key molecular switches for tumorigenesis and development of gastrointestinal stromal tumors (GIST) ^[4]. Therefore, the use of imatinib, which targets active *c-KIT*, has dramatically improved clinical outcomes of adult GIST patients bearing mutations in this gene, with minimal side effects^[5]. Although GIST tumor may develop resistance to imatinib after months or years of treatment, imatinib still results in a better survival rate than conventional chemotherapy^[6].

In addition to imatinib, a number of novel targeted therapies are currently under evaluation in various stages of clinical trials for various soft tissue sarcomas. Tyrosine kinase inhibitors, such as sorafenib, sunitinib, and panzopanib, have potent inhibitory effects on angiogenic receptor VEGFR1-3 and c-KIT. Panzopanib achieved better progression-free survival rates for non-adipocytic soft tissue sarcoma than standard therapy in Phase III clinical trials, although this drug was noted to have substantial cardiotoxicity^[7]. mTOR inhibitors have also been studied in preclinical and clinical settings. mTOR inhibitor ridaforolimus was confirmed to be have antitumor efficacy in sarcoma cell lines *in vitro* and in a xenograft animal model *in vivo*^[8]. However, it showed only modest improvements in progression-free survival and overall survival rates in clinical trials in sarcoma patients^[9]. Other new drugs, such as an MDM2 inhibitor and a CDK4 inhibitor for well-differentiated and dedifferentiated liposarcomas, are being tested in clinical trials now^[10,11].

To identify more drug targets for sarcoma therapies, genomic, genetic and molecular approaches have been used. Recent reports have shown that mutations in retinoblastoma protein (Rb1) and isocitrate dehydrogenases 1 and 2 (IDH1/2) are involved in sarcoma initiation^[12,13]. Mitotic spindle checkpoint kinase aurora A/B, cytoskeleton protein vimentin, growth factor midkine, hepatoma-derived growth factor, Wnt/β-catenin, and the AKT pathway have also been identified as potential drug targets for sarcoma^[14-20]. Hypoxia-induced enzyme procollagen-lysine, 2-oxoglutarate 5-dioxygenase 2 has been shown to promote sarcoma metastasis but not primary tumor growth^[21]. MicroRNAs such

as oncogenic miR-155 and tumor suppressive miR-143, miR-340, and miR-29 also were identified as potential therapeutic targets for sarcoma^[22-25].

Epigenetic Regulators as Targets in Sarcoma

Among the novel and promising therapeutic targets for sarcoma are epigenetic regulators, which have been discovered recently as a new class of drug targets for human malignancies^[26]. Epigenetic regulation of gene expression refers to the mechanisms that activate or suppress gene expression without changing underlying DNA sequences. In contrast to permanent and heritable genomic DNA changes, such as mutation, insertion or deletion; epigenetic changes of genomic DNA are temporal, spatial, reversible and not heritable. The molecular mechanisms of epigenetically gene regulation in cells include DNA methylation, histone modification, and RNA-associated silencing. Epigenetic regulation plays a vital role in stem cell maintenance, cell differentiation, and cell senescence^[27].

DNA methylation of the CpG islands in the promoter region by DNA methyltransferases is an important regulator of gene transcription. Experimental evidence has revealed that aberrant DNA methylation of the promoter region of a gene is associated with abnormal gene expression, leading to various diseases and developmental defects^[28]. Aberrant DNA methylation patterns on genomic DNA have been shown to cause a variety of human cancers and are shown in two distinct forms: hypermethylation and hypomethylation compared to non-tumor tissue^[28]. Therfore, DNA-demethylating agents were developed and have been shown to be effective therapeutics for some types of hematological malignancies. For example, 5-azacytidine and 5-aza-2'-deoxycytidine, the most successful DNAdemethylating drugs, are currently used as the first-line treatment for high-risk myelodysplastic syndromes^[29]. However, DNA methylation inhibitors have shown only minor effects on osteosarcoma cell proliferation and tumor growth^[30]. Combinations of these agents with histone deacetylase (HDAC) inhibitors have exhibited synergistic effects in osteosarcoma cells in preclinical studies^[30]. So far, no DNA methylation inhibitors have been tested in clinical trials for soft tissue sarcoma.

Histone modifications are also critically important for regulation of oncogenic and tumor suppressive genes. As the nucleosome core of the DNA assembly in chromatin, histones can be acetylated, methylated, ubiquitylated, phosphorylated, sumoylated, citrullinated, and ADP-ribosylated at multiple sites by different histone-modifying enzymes^[27]. The histone acetylation balance, mediated by histone acetyltransferase and deacetylase, is crucial for maintenance of normal cell growth^[31]. Deregulation induced by HDAC overexpression or malfunction occurs in many malignancies, resulting in abnormal modulation of target gene transcriptional activity. Therefore, HDAC inhibitors have been evaluated as anticancer therapeutics.

An HDAC inhibitor was recently studied for its effects on soft tissue sarcoma cell lines *in vitro* and animal xenograft models *in vivo*. Experimental data showed that HDAC inhibitor PCI-24781 inhibited cell proliferation of various soft tissue sarcoma cell lines *in vitro*^[32]. The single-agent effects on tumor growth and metastasis *in vivo* were modest. However, combination of an HDAC inhibitor with chemotherapy produced substantial antitumor effects, and the HDAC inhibitor enhanced the effects of the chemotherapy agent on drug-

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resistant sarcoma cells^[32]. In malignant peripheral nerve sheath tumor (MPNST), neurofibromatosis 1–related (NF1-related) MPNST cell lines were sensitive to HDAC inhibitor treatment, which inhibited cell proliferation and induced cell apoptosis *in vitro* and blocked tumor growth in a xenograft mice model. However, sporadic (non-NF1-related) MPNST cell lines are resistant to HDAC inhibition *in vitro* and *in vivo* because HDAC inhibitor induced autophagy in these cells. When combined with an autophagy inhibitor, however, the HDAC inhibitor resumed strong inhibition of proliferation of sporadic MPNST cells *in vitro* and tumor growth *in vivo*^[33]. HDAC inhibitors were also found to block growth of clear cell sarcoma, synovial sarcoma, and uterine sarcoma and to induce apoptosis and differentiation *in vitro* and *in vivo*^[34-36]. These studies suggested that HDAC inhibition is a promising therapeutic strategy for sarcomas.

Another important histone modification enzyme is enhancer of zeste homolog 2 (EZH2), a histone methyltransferase that trimethylates histone H3 lysine 27^[37]. EZH2 forms polycomb-repressor complex 2 (PRC2) with two other core proteins, SUZ12 and EED. PRC2 functions as a transcription repressor to critically coordinate gene expression and repression during many physiological and pathological processes^[38]. Specifically in cancer, EZH2 has been identified as an oncogene in breast, lung, liver, prostate, blood and pancreatic cancers^[38,39]. In mesenchyme-originating sarcomas, evidences show that EZH2 is involved in tumorigenesis and progression. High EZH2 expression is correlated with greater tumor size, distant metastasis, and poor prognosis in synovial sarcoma, and EZH2 mediates repression by SYT-SSX of the tumor suppressor ERG1^[40]. In rhabdomyosarcoma, EZH2 is overexpressed and suppresses skeletal muscle differentiation through myofibrillary genes and miR-29 inhibition^[41]. High EZH2 expression has also been found in Ewing sarcoma, in which EWS/FLI1 directly activates EZH2 expression and inhibits tumor cell differentiation^[42]. Recently, the function of EZH2 and the molecular mechanisms that are regulated by EZH2 in MPNST pathogenesis have been investigated^[43]. EZH2 expression is significantly higher in MPNST tumor samples than in neurofibromas and normal nerve tissues. In multiple MPNST cell lines, EZH2 protein expression is also higher than that in Schwann cells which are the potential origin of MPNST. Genetic knockdown of EZH2 in NF1-related and non-NF1-related MPNST cell lines induces cell death in vitro and inhibits tumor growth in vivo^[43]. Evidences demonstrate that upregulated EZH2 in MPNST cells inhibits miR-30d expression via binding to miR-30d promoter. Decreased miR-30d expression leads to enhanced expression of KPNB1, because KPNB1 is inhibited by miR-30d targeting of the KPNB1 3'-untranslated region^[43]. KPNB1 expression in MPNST cell lines and normal Schwann cells are positively correlated to EZH2 expression, and negative associated with mIR-30d expression^[43]. These data suggest that EZH2 may play a critical role in the initiation and progression of MPNST. Compelling data from all these studies show the oncogenic function of EZH2 in different sarcomas and suggested that EZH2 inhibition may be a novel therapeutic approach for soft tissue sarcoma.

Conclusion

Taken together, epigenetic regulation mechanisms, specifically DNA methylation and histone modification, have been implicated as having important roles in sarcoma pathology. Preclinical studies have demonstrated that DNA methylation and histone modification

inhibitors have promise as new targeted therapies for sarcoma, either as single agents or combined with chemotherapy agents.

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References

- Fletcher, CDM.; Bridge, JA.; Hogendoorn, P.; Mertens, F. WHO Classification of Tumours. 4th Edition. Vol. 5. IARC Press; Lyon, France: 2013. WHO Classification of Tumours of Soft Tissue and Bone..
- Yang JC, Chang AE, Baker AR, Sindelar WF, Danforth DN, Topalian SL, et al. Randomized prospective study of the benefit of adjuvant radiation therapy in the treatment of soft tissue sarcomas of the extremity. J. Clin. Oncol. 1998; 16:197–203. PMid:9440743. [PubMed: 9440743]
- Casali PG, Blay J-Y, experts ObotECECPo. Soft tissue sarcomas: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. Ann. Oncol. 2010; 21:v198–v203. http:// dx.doi.org/10.1093/annonc/mdq209 PMid:20555081. [PubMed: 20555081]
- Hirota S, Isozaki K, Moriyama Y, Hashimoto K, Nishida T, Ishiguro S, et al. Gain-of-Function Mutations of c-kit in Human Gastrointestinal Stromal Tumors. Science. 1998; 279:577–80. http:// dx.doi.org/10.1126/science.279.5350.577 PMid:9438854. [PubMed: 9438854]
- Joensuu H, Eriksson M, Sundby Hall K, et al. One vs three years of adjuvant imatinib for operable gastrointestinal stromal tumor: A randomized trial. JAMA. 2012; 307:1265–72. http://dx.doi.org/ 10.1001/jama.2012.347 PMid:22453568. [PubMed: 22453568]
- Antonescu CR, Besmer P, Guo T, Arkun K, Hom G, Koryotowski B, et al. Acquired Resistance to Imatinib in Gastrointestinal Stromal Tumor Occurs Through Secondary Gene Mutation. Clin. Cancer Res. 2005; 11:4182–90. http://dx.doi.org/10.1158/1078-0432.CCR-04-2245 PMid: 15930355. [PubMed: 15930355]
- van der Graaf WTA, Blay J-Y, Chawla SP, Kim D-W, Bui-Nguyen B, Casali PG, et al. Pazopanib for metastatic soft-tissue sarcoma (PALETTE): a randomised, double-blind, placebo-controlled phase 3 trial. Lancet. 2012; 379:1879–86. http://dx.doi.org/10.1016/S0140-6736(12)60651-5. [PubMed: 22595799]
- Squillace RM, Miller D, Cookson M, Wardwell SD, Moran L, Clapham D, et al. Antitumor Activity of Ridaforolimus and Potential Cell-Cycle Determinants of Sensitivity in Sarcoma and Endometrial Cancer Models. Mol. Cancer Ther. 2011; 10:1959–68. http://dx.doi.org/ 10.1158/1535-7163.MCT-11-0273 PMid:21825008. [PubMed: 21825008]
- Demetri GD, Chawla SP, Ray-Coquard I, Le Cesne A, Staddon AP, Milhem MM, et al. Results of an International Randomized Phase III Trial of the Mammalian Target of Rapamycin Inhibitor Ridaforolimus Versus Placebo to Control Metastatic Sarcomas in Patients After Benefit From Prior Chemotherapy. J. Clin. Oncol. 2013; 31:2485–92. http://dx.doi.org/10.1200/JCO.2012.45.5766 PMid:23715582. [PubMed: 23715582]
- Dickson MA, Tap WD, Keohan ML, D'Angelo SP, Gounder MM, Antonescu CR, et al. Phase II Trial of the CDK4 Inhibitor PD0332991 in Patients With Advanced CDK4-Amplified Well-Differentiated or Dedifferentiated Liposarcoma. J. Clin. Oncol. 2013; 31:2024–8. http://dx.doi.org/ 10.1200/JCO.2012.46.5476 PMid:23569312 PMCid:PMC3661937. [PubMed: 23569312]
- Ray-Coquard I, Blay J-Y, Italiano A, Le Cesne A, Penel N, Zhi J, et al. Effect of the MDM2 antagonist RG7112 on the P53 pathway in patients with MDM2-amplified, well-differentiated or dedifferentiated liposarcoma: an exploratory proof-of-mechanism study. Lancet Oncol. 2012; 13:1133–40. http://dx.doi.org/10.1016/S1470-2045(12)70474-6. [PubMed: 23084521]
- Liu Y, Sánchez-Tilló E, Lu X, Clem B, Telang S, Jenson AB, et al. Rb1 family mutation is sufficient for sarcoma initiation. Nat Commun. 2013; 4:2650. PMid:24150016. [PubMed: 24150016]

- Lu C, Venneti S, Akalin A, Fang F, Ward PS, DeMatteo RG, et al. Induction of sarcomas by mutant IDH2. Gene Dev. 2013; 27:1986–98. http://dx.doi.org/10.1101/gad.226753.113 PMid: 24065766 PMCid:PMC3792475. [PubMed: 24065766]
- Brewer Savannah KJ, Demicco EG, Lusby K, Ghadimi MP, Belousov R, Young E, et al. Dual Targeting of mTOR and Aurora-A Kinase for the Treatment of Uterine Leiomyosarcoma. Clin. Cancer Res. 2012; 18:4633–45. http://dx.doi.org/10.1158/1078-0432.CCR-12-0436 PMid: 22821997 PMCid:PMC3432751. [PubMed: 22821997]
- Lahat G, Zhu Q-S, Huang K-L, Wang S, Bolshakov S, Liu J, et al. Vimentin Is a Novel Anti-Cancer Therapeutic Target; Insights from In Vitro and In Vivo Mice Xenograft Studies. PLoS ONE. 2010; 5:e10105. http://dx.doi.org/10.1371/journal.pone.0010105 PMid:20419128 PMCid:PMC2855704. [PubMed: 20419128]
- 16. Jin Z, Lahat G, Korchin B, Nguyen T, Zhu Q-S, Wang X, et al. Midkine Enhances Soft-Tissue Sarcoma Growth: A Possible Novel Therapeutic Target. Clin. Cancer Res. 2008; 14:5033–42. http://dx.doi.org/10.1158/1078-0432.CCR-08-0092 PMid:18698021. [PubMed: 18698021]
- Yang Y, Li H, Zhang F, Shi H, Zhen T, Dai S, et al. Clinical and biological significance of hepatoma-derived growth factor in Ewing's sarcoma. J Pathol. 2013; 231(3):323–34. PMid: 23878072. [PubMed: 23878072]
- Shan W, Akinfenwa PY, Savannah KB, Kolomeyevskaya N, Laucirica R, Thomas DG, et al. A Small-Molecule Inhibitor Targeting the Mitotic Spindle Checkpoint Impairs the Growth of Uterine Leiomyosarcoma. Clin. Cancer Res. 2012; 18:3352–65. http://dx.doi.org/ 10.1158/1078-0432.CCR-11-3058 PMid:22535157. [PubMed: 22535157]
- Trautmann, M.; Sievers, E.; Aretz, S.; Kindler, D.; Michels, S.; Friedrichs, N., et al. SS18-SSX fusion protein-induced Wnt/[beta]-catenin signaling is a therapeutic target in synovial sarcoma.. Oncogene. 2013. doi: 10.1038/onc.2013.443. http://dx.doi.org/10.1038/onc.2013.443
- Zhu Q-S, Ren W, Korchin B, Lahat G, Dicker A, Lu Y, et al. Soft Tissue Sarcoma Cells Are Highly Sensitive to AKT Blockade: A Role for p53-Independent Up-regulation of GADD45α. Cancer Res. 2008; 68:2895–903. http://dx.doi.org/10.1158/0008-5472.CAN-07-6268 PMid: 18413758. [PubMed: 18413758]
- Eisinger-Mathason TSK, Zhang M, Qiu Q, Skuli N, Nakazawa MS, Karakasheva T, et al. Hypoxia-Dependent Modification of Collagen Networks Promotes Sarcoma Metastasis. Cancer Discov. 2013; 3:1190–205. http://dx.doi.org/10.1158/2159-8290.CD-13-0118 PMid:23906982. [PubMed: 23906982]
- 22. Balkhi MY, Iwenofu OH, Bakkar N, Ladner KJ, Chandler DS, Houghton PJ, et al. miR-29 Acts as a Decoy in Sarcomas to Protect the Tumor Suppressor A20 mRNA from Degradation by HuR. Sci Signal. 2013; 6:ra63. http://dx.doi.org/10.1126/scisignal.2004177 PMid:23901138 PMCid:PMC3885907. [PubMed: 23901138]
- Zhou X, Wei M, Wang W. MicroRNA-340 suppresses osteosarcoma tumor growth and metastasis by directly targeting ROCK1. Biochem. Biophys. Res. Commun. 2013; 437:653–8. http:// dx.doi.org/10.1016/j.bbrc.2013.07.033 PMid:23872151. [PubMed: 23872151]
- 24. Ugras S, Brill E, Jacobsen A, Hafner M, Socci ND, DeCarolis PL, et al. Small RNA Sequencing and Functional Characterization Reveals MicroRNA-143 Tumor Suppressor Activity in Liposarcoma. Cancer Res. 2011; 71:5659–69. http://dx.doi.org/10.1158/0008-5472.CAN-11-0890 PMid:21693658 PMCid:PMC3165140. [PubMed: 21693658]
- Zhang P, Bill K, Liu J, Young E, Peng T, Bolshakov S, et al. MiR-155 Is a Liposarcoma Oncogene That Targets Casein Kinase-1α and Enhances β-Catenin Signaling. Cancer Res. 2012; 72:1751– 62. http://dx.doi.org/10.1158/0008-5472.CAN-11-3027 PMid:22350414 PMCid:PMC3319789. [PubMed: 22350414]
- Helin K, Dhanak D. Chromatin proteins and modifications as drug targets. Nature. 2013; 502:480– 8. http://dx.doi.org/10.1038/nature12751 PMid:24153301. [PubMed: 24153301]
- Kouzarides T. Chromatin Modifications and Their Function. Cell. 2007; 128:693–705. http:// dx.doi.org/10.1016/j.cell.2007.02.005 PMid:17320507. [PubMed: 17320507]
- Baylin SB, Jones PA. A decade of exploring the cancer epigenome biological and translational implications. Nat Rev Cancer. 2011; 11:726–34. http://dx.doi.org/10.1038/nrc3130 PMid: 21941284 PMCid:PMC3307543. [PubMed: 21941284]

- 29. Yang X, Lay F, Han H, Jones PA. Targeting DNA methylation for epigenetic therapy. Trends Pharmacol Sci. 2010; 31:536–46. http://dx.doi.org/10.1016/j.tips.2010.08.001 PMid:20846732 PMCid:PMC2967479. [PubMed: 20846732]
- Thayanithy V, Park C, Sarver AL, Kartha RV, Korpela DM, Graef AJ, et al. Combinatorial Treatment of DNA and Chromatin-Modifying Drugs Cause Cell Death in Human and Canine Osteosarcoma Cell Lines. PLoS ONE. 2012; 7:e43720. http://dx.doi.org/10.1371/journal.pone. 0043720 PMid:22957032 PMCid:PMC3434163. [PubMed: 22957032]
- Khan O, La Thangue NB. HDAC inhibitors in cancer biology: emerging mechanisms and clinical applications. Immunol Cell Biol. 2012; 90:85–94. http://dx.doi.org/10.1038/icb.2011.100 PMid: 22124371. [PubMed: 22124371]
- Lopez G, Liu J, Ren W, Wei W, Wang S, Lahat G, et al. Combining PCI-24781, a Novel Histone Deacetylase Inhibitor, with Chemotherapy for the Treatment of Soft Tissue Sarcoma. Clin. Cancer Res. 2009; 15:3472–83. http://dx.doi.org/10.1158/1078-0432.CCR-08-2714 PMid:19417021. [PubMed: 19417021]
- 33. Lopez G, Torres K, Liu J, Hernandez B, Young E, Belousov R, et al. Autophagic Survival in Resistance to Histone Deacetylase Inhibitors: Novel Strategies to Treat Malignant Peripheral Nerve Sheath Tumors. Cancer Res. 2011; 7:185–96. http://dx.doi.org/ 10.1158/0008-5472.CAN-10-2799 PMid:21084276 PMCid:PMC3064267. [PubMed: 21084276]
- Liu S, Cheng H, Kwan W, Lubieniecka JM, Nielsen TO. Histone deacetylase inhibitors induce growth arrest, apoptosis, and differentiation in clear cell sarcoma models. Mol Cancer Ther. 2008; 7:1751–61. http://dx.doi.org/10.1158/1535-7163.MCT-07-0560 PMid:18566246. [PubMed: 18566246]
- Su L, Cheng H, Sampaio AV, Nielsen TO, Underhill TM. EGR1 reactivation by histone deacetylase inhibitors promotes synovial sarcoma cell death through the PTEN tumor suppressor. Oncogene. 2010; 29:4352–61. http://dx.doi.org/10.1038/onc.2010.204 PMid:20514024. [PubMed: 20514024]
- 36. Hrzenjak A, Moinfar F, Kremser M-L, Strohmeier B, Petru E, Zatloukal K, et al. Histone deacetylase inhibitor vorinostat suppresses the growth of uterine sarcomas in vitro and in vivo. Mol Cancer. 2010; 9:49. http://dx.doi.org/10.1186/1476-4598-9-49 PMid:20202195 PMCid:PMC2843655. [PubMed: 20202195]
- Cao R, Zhang Y. The functions of E(Z)/EZH2-mediated methylation of lysine 27 in histone H3. Curr Opin Genet Dev. 2004; 14:155–64. http://dx.doi.org/10.1016/j.gde.2004.02.001 PMid: 15196462. [PubMed: 15196462]
- Simon JA, Lange CA. Roles of the EZH2 histone methyltransferase in cancer epigenetics. Mutat Res. 2008; 647:21–9. http://dx.doi.org/10.1016/j.mrfmmm.2008.07.010 PMid:18723033. [PubMed: 18723033]
- 39. Crea F, Fornaro L, Bocci G, Sun L, Farrar W, Falcone A, et al. EZH2 inhibition: targeting the crossroad of tumor invasion and angiogenesis. Cancer Metastasis Rev. 2012; 31:753–61. 2012. http://dx.doi.org/10.1007/s10555-012-9387-3 PMid:22711031. [PubMed: 22711031]
- 40. Lubieniecka JM, de Bruijn DRH, Su L, van Dijk AHA, Subramanian S, van de Rijn M, et al. Histone Deacetylase Inhibitors Reverse SS18-SSX–Mediated Polycomb Silencing of the Tumor Suppressor Early Growth Response 1 in Synovial Sarcoma. Cancer Res. 2008; 68:4303–10. http:// dx.doi.org/10.1158/0008-5472.CAN-08-0092 PMid:18519690. [PubMed: 18519690]
- Wang H, Garzon R, Sun H, Ladner KJ, Singh R, Dahlman J, et al. NF-κB–YY1–miR-29 Regulatory Circuitry in Skeletal Myogenesis and Rhabdomyosarcoma. Cancer Cell. 2008; 14:369– 81. http://dx.doi.org/10.1016/j.ccr.2008.10.006 PMid:18977326 PMCid:PMC3829205. [PubMed: 18977326]
- 42. Richter GHS, Plehm S, Fasan A, Rössler S, Unland R, Bennani-Baiti IM, et al. EZH2 is a mediator of EWS/FL11 driven tumor growth and metastasis blocking endothelial and neuro-ectodermal differentiation. Proc Natl Acad Sci U S A. 2009; 106:5324–9. http://dx.doi.org/10.1073/pnas. 0810759106 PMid:19289832 PMCid:PMC2656557. [PubMed: 19289832]
- 43. Zhang P, Garnett J, Creighton CJ, Al Sannaa GA, Igram DR, Lazar A, et al. EZH2–miR-30d– KPNB1 pathway regulates malignant peripheral nerve sheath tumour cell survival and tumourigenesis. J Pathol. 2014; 232:308–18. http://dx.doi.org/10.1002/path.4294 PMid:24132643. [PubMed: 24132643]