



HHS Public Access

Author manuscript

Blood Rev. Author manuscript; available in PMC 2016 July 01.

Published in final edited form as:

Blood Rev. 2015 July ; 29(4): 215–221. doi:10.1016/j.blre.2014.12.002.

Mechanisms of Thrombogenesis in Polycythemia Vera

Michael H. Kroll, MD¹, Laura C. Michaelis, MD², and Srdan Verstovsek, MD, PhD³

¹Section of Benign Hematology, Division of Internal Medicine, The University of Texas, MD Anderson Cancer Center, Houston, TX, USA

²Hematologic Malignancies Program, Division of Hematology and Oncology, Medical College of Wisconsin, Froedtert Hospital, Milwaukee, WI, USA

³Department of Leukemia, Division of Cancer Medicine, The University of Texas MD Anderson Cancer Center, Houston, TX, USA

Abstract

Thrombotic and cardiovascular events are among the leading causes of death for patients with polycythemia vera (PV), and thrombosis history is a key criterion for patient risk stratification and treatment strategy. Little is known, however, about mechanisms of thrombogenesis in patients with PV. This report provides an overview of thrombogenesis pathophysiology in patients with PV and elucidates the roles of conventional and nonconventional thrombosis risk factors. In addition to several conventional risk factors for thrombosis, clinical data have implicated increased hematocrit and red blood cell adhesiveness, activated platelets, leukocytosis, and elevated *JAK2*^{V617F} allele burden in patients with PV. Furthermore, PV-related inflammation may exacerbate thrombogenesis through varied mechanisms, including endothelial damage, inhibition of natural anticoagulant pathways, and secretion of procoagulant factors. These findings suggest a direct link between myeloproliferation and thrombogenesis in PV, which is likely to provide new opportunities for targeted antithrombotic interventions aimed at decreasing PV-related morbidity and mortality.

Keywords

Janus kinase 2; polycythemia vera; thrombosis

Address correspondence to: Srdan Verstovsek, The University of Texas MD Anderson Cancer Center, Division of Cancer Medicine, 1515 Holcombe Blvd, Unit 418, Houston, TX 77030, USA, Tel.: +1-713-745-3429, Fax: +1-713-794-4297, sverstov@mdanderson.org.

Michael H. Kroll, The University of Texas MD Anderson Cancer Center, Section of Benign Hematology, Division of Internal Medicine, 1515 Holcombe Blvd, Unit 1464, Houston, TX 77030, USA, Tel.: +1-713-794-2234, Fax: +1-713-794-4922, mkroll@mdanderson.org

Laura C. Michaelis, Medical College of Wisconsin, Froedtert Hospital, Division of Hematology and Oncology, 9200 W. Wisconsin Ave., Milwaukee, WI 53226, USA, Tel.: +1-414-805-1118, Fax: +1-414-805-4606, lmichaelis@mcw.edu

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

1 Introduction

Thrombosis is an important complication among patients with polycythemia vera (PV) and thrombotic events reflect disease burden and impact clinical outcomes. Roughly one fifth of patients with PV are diagnosed with an arterial or venous thrombotic event as a presenting feature,^{1, 2} and fatal cardiovascular events contribute to the increased mortality rate among patients with PV.¹⁻⁴ Furthermore, healthcare utilization and cost are higher for patients with PV compared with age- and gender-matched persons in a large US health insurance claims database⁵ and this is driven in part by arterial and venous thrombotic events.

Thrombotic events are so inextricably tied to the pathogenesis of PV that they are used in guiding patient risk stratification and treatment recommendations. Patients with no history of thrombotic events and who are <60 years of age are categorized as low risk,^{6, 7} whereas patients with a history of thrombotic events or are over the age of 59 years are categorized as high risk.^{6, 7} Current treatment recommendations suggest antiplatelet treatment with low-dose aspirin and phlebotomy as first-line therapy for low-risk patients.^{6, 7} For high-risk patients, recommendations also include the use of cytoreductive agents (eg, hydroxyurea or interferon- α [IFN- α]).⁶⁻⁹ However, none of the currently available treatment options for PV are designed to target the active Janus kinase (JAK)/signal transducer and activator of transcription (STAT) signaling pathway, which over the last 10 years has been identified as the molecular etiology of PV.¹⁰⁻¹³ Nearly all patients (99%) have an activating mutation in exon 12 or 14 of *JAK2*, which most commonly is *JAK2V617F*.^{6, 13, 14} Constitutive activation of *JAK2* has been associated with unregulated hematopoietic proliferation,^{11-13, 15} and *JAK2V617F* allele burden is significantly correlated with markers of inflammation.¹⁶ Inflammation is an important thrombotic risk factor.¹⁶

The risk of thrombotic events and their associated morbidity and mortality remain a primary concern in patients with PV⁶⁻⁸ despite recent advances in understanding how JAK/STAT pathway dysregulation influences disease origin and progression. The goal of this report is to provide an overview of mechanisms of thrombogenesis in patients with PV by considering both conventional and nonconventional thrombosis risk factors, including correlates of an overactive JAK/STAT pathway.

2 Thrombosis in Patients With Polycythemia Vera

In 1856, Rudolf Virchow proposed what would later be termed *Virchow's triad*, which postulates that venous thromboses occur because of abnormalities in interactions between the blood vessel wall, blood constituents, and blood flow.^{17, 18} Mechanisms underlying arterial and venous thrombogenesis have since been elucidated and include complex biochemical pathways regulating fibrin accumulation and the participation of several cell types, including platelets, endothelial cells (ECs), and leukocytes.^{17, 19-21} Perhaps of most importance is the concept that mechanisms of thrombosis differ depending on the rheology of the circuit. Arterial thrombosis, because it develops in the high-resistance, high-flow vasculature, is triggered by platelet adhesion, activation, and aggregation.²² In contrast, venous thrombosis, which develops in a high-capacitance, low-flow vascular circuit, is

triggered by the activation of the soluble coagulation system and the generation of insoluble fibrin.²²

In patients with PV, thromboses have been reported throughout the arterial (transient ischemic attack, stroke, acute myocardial infarction, and peripheral arterial thrombosis) and venous (deep vein thrombosis, superficial thrombophlebitis, and pulmonary embolism) systems (Table 1).²³ Thrombotic events also may occur in the splanchnic vasculature in patients with myeloproliferative neoplasms (MPNs) and/or *JAK2* mutation. A recent meta-analysis found that 8% to 53% of patients with hepatic vein thrombosis (Budd-Chiari syndrome) or portal vein thrombosis who did not have cirrhosis or a biliary malignancy had a MPN, which was PV in the majority of cases.²⁴

The rate of nonfatal thrombosis in patients with PV has been estimated at 3.8 per 100 patient-years.²³ Thrombotic events often occur before diagnosis and increase in frequency in the years immediately preceding diagnosis.¹ Patient sex may influence the type of thrombotic event; arterial thrombosis has been reported to be more common in men (18% vs 14%; $P=0.02$), whereas venous thrombosis has been reported to be more common in women (9.3% vs 5.4%; $P<0.01$).²

Thrombotic events, in particular arterial thrombosis, may contribute to an increased mortality risk in patients with PV. A retrospective Italian study of 1213 patients reported that 30% of 145 arterial thrombotic events and 11% of 87 venous thrombotic events were fatal, accounting for 24% and 6% of all deaths, respectively (median follow-up, 5.3 years [range, 0–32.9 years]).¹ A retrospective international study of 1545 patients reported thrombotic complications as the cause of death in 9% of patients (median follow-up, 6.9 years [range, 0–39.3 years]).² Patients with a history of thrombotic events were at further risk for mortality compared with those who had no such history (hazard ratio [HR], 1.9; 95% CI, 1.2–3.0; $P=0.007$).² These results are consistent with data from the European Collaboration on Low-Dose Aspirin in Polycythemia Vera (ECLAP) study showing that a previous thrombosis doubles PV-associated mortality.²³

3 Thrombosis Risk Factors

There are few studies elucidating molecular mechanisms of thrombogenesis in PV. Several clinical studies, however, have identified risk factors associated with thrombosis in patients with PV. There are multiple risk factors, including both conventional risks and PV-specific risks such as *JAK2*^{V617F} allele burden. These latter elements may someday inform the design of future studies investigating better treatments of thrombogenesis in this setting.

3.1 Conventional Risk Factors

3.1.1 Thrombotic Event History—Patients with PV share several thrombosis risk factors with otherwise healthy individuals (Table 2).²⁵ Perhaps of greatest importance is a history of thrombotic events. The prospective ECLAP study reported increased risk of cardiovascular events among patients with a history of thrombosis (HR, 2.09; 95% CI, 1.55–2.81)²³ and that risk for major thrombosis was associated with varied prior thrombotic events (major thrombosis [HR, 1.69; 95% CI, 1.21–2.36], arterial thrombosis [HR, 1.66;

95% CI, 1.20–2.32], venous thrombosis [HR, 1.74; 95% CI, 1.06–2.87]), prior stroke or transient ischemic attack (HR, 1.81; 95% CI, 1.27–2.57), and prior deep vein thrombosis (HR, 2.04; 95% CI, 1.22–3.39).²⁶ Large retrospective studies have also reported significantly greater risk for thrombotic events in patients with a history of thrombosis.^{1, 27}

3.1.2 Erythrocytosis—A recent prospective randomized clinical trial analyzed 365 patients with PV, and identified a lower incidence of cardiovascular events among patients who maintained hematocrit <45% compared with 45% to 50% (4.4% vs 10.9%; $P=0.02$) (Table 2).²⁸ Furthermore, the lower hematocrit group was associated with a lower death rate from cardiovascular causes or major thrombosis (1.1 vs 4.4 per 100 person-years). This important prospective clinical study clarified the target hematocrit level for clinicians to pursue in everyday practice. The issue was previously debated because two retrospective analyses reported no correlation between patient hematocrit and subsequent arterial or venous thrombotic events.^{27, 29} Further research is necessary to untangle the complex mechanisms by which small differences in hematocrit, which have little effect on viscosity and blood fluidity,³⁰ could influence thrombosis in patients with PV.

There may also be qualitative changes in the red blood cells of patients with PV that promote thrombosis. Recent data suggest that erythrocytes from patients with PV are more adhesive because *JAK2^{V617F}* drives phosphorylation of the red blood cell adhesion receptor Lutheran/basal cell-adhesion molecule (Lu/BCAM), which enhances red blood cell binding to subendothelial laminin.³¹

3.1.3 Leukocytosis—In recent years, leukocytosis has emerged as a potential risk factor for thrombosis in patients with PV. An unadjusted analysis of ECLAP study data identified an increased risk for myocardial infarction during follow-up in patients with elevated white blood cell (WBC) counts ($>15 \times 10^9/L$ vs 10.1 to $15 \times 10^9/L$; $P=0.015$) that was confirmed in multivariate analyses adjusted for age, sex, disease duration, prior thrombosis, prior hemorrhage, conventional risk factors, number of comorbidities, or cytoreductive and antithrombotic treatment.²⁶ In agreement, a prospective study of 187 patients with PV or essential thrombocythemia (ET) reported increased risk of thrombosis among patients with a WBC count $>9.5 \times 10^9/L$ (median) at baseline ($P=0.03$)³² and a nonsignificant trend toward increased thrombotic risk among patients with leukocytosis in the PV subgroup ($P=0.07$). Finally, a retrospective study of 459 patients with PV reported a significant association between baseline leukocyte count $15 \times 10^9/L$ and venous thrombotic events during a median follow-up of 64 months (range, 0–562 months; $P=0.005$) (Table 2).²⁷

The association between leukocyte count and recurrent thrombotic risk may be limited to patients aged <60 years. A retrospective study of 494 patients with PV ($n=235$) or ET ($n=259$) reported increased risk for recurrent thrombosis in patients with leukocyte count in the highest versus lowest quartile (HR, 3.55; 95% CI, 1.02–12.25) only for patients aged <60 years.³³

3.1.4 Other Conventional Risk Factors—Several other conventional risk factors have also been reported in patients with PV. Analysis of the ECLAP study data identified older age (>65 years; HR, 2.89; 95% CI, 1.98–4.22) and smoking (HR, 1.55; 95% CI, 0.99–2.43)

as risk factors for major thrombosis.²⁶ A retrospective study of 459 patients with PV reported that hypertension and tobacco use were associated with arterial thrombosis ($P = 0.02$), and diabetes mellitus was associated with venous thrombosis ($P=0.002$).²⁷

3.2 Nonconventional Risk Factors

3.2.1 *JAK2*^{V617F} Allele Burden—The genetic ratio of the *JAK2*^{V617F} mutation to *JAK2* wild-type in granulocytes, termed the *JAK2*^{V617F} allele burden,^{34, 35} may also be prognostic for thrombotic risk (Table 2). Patients with a *JAK2*^{V617F} allele burden >75% compared with 25% have a seven-fold greater risk for thrombosis according to a prospective study that followed 173 patients with PV for a median of 24 months. Although a more recent prospective study (n=338 patients with PV followed for a median of 3.2 years) was unable to confirm an association between *JAK2*^{V617F} allele burden and thrombotic risk, this analysis compared patients with an allele burden >50% versus 50%.¹⁴ Taken together, these results suggest that a *JAK2*^{V617F} allele burden threshold between 50% and 75% may identify patients at high risk for thrombotic events.

JAK2^{V617F} may also identify patients specifically at risk for splanchnic vein thromboses. A meta-analysis of patients with splanchnic vein thromboses reported the presence of the *JAK2*^{V617F} mutation in 26% to 70% of patients with Budd-Chiari syndrome and 10% to 46% of patients with portal vein thrombosis.²⁴

3.2.2 Inflammation—Inflammation may predict thrombosis in patients with PV. A retrospective study identified patients with elevated inflammation using the serum biomarker C-reactive protein (CRP). Patients with high serum CRP concentration had significantly greater risk for thrombosis ($P=0.01$),¹⁶ which was confirmed in a multivariable analysis that adjusted for age, sex, ET or PV diagnosis, cardiovascular risk factors, *JAK2*^{V617F} mutation status, and hydroxyurea treatment (all $P = 0.045$).

3.2.3 Blood Cell Activation—Laboratory data suggest that activation of leukocytes, platelets, and ECs may promote a prothrombotic state;^{19, 36–41} markers for activation of these cell types have been observed in patients with PV. A prospective study of 37 patients with ET and 34 with PV reported significant elevations in markers for leukocyte activation (CD11b, leukocyte alkaline phosphatase, elastase activity, plasma elastase, and plasma myeloperoxidase; $P<0.01$) and EC activation (von Willebrand factor [vWF] and thrombomodulin; $P<0.01$) compared with healthy adults with no history of thrombohemorrhagic events.⁴² The aggregation of leukocytes with platelets is a marker for platelet activation⁴³ and is thought to be a key component of leukocyte-driven thrombosis.⁴⁴ A follow-up study that included 34 patients with PV reported increased levels of markers for leukocyte-platelet aggregates (CD11b/CD42b, CD11b/CD62P; $P = 0.001$) and a higher percentage of circulating activated platelets than controls (CD62P; $P = 0.001$).⁴³ In patients with PV, treatment with hydroxyurea did not influence marker levels for leukocyte activation, endothelial activation, or hypercoagulability.⁴⁴ More studies are needed to understand whether these markers can be used to guide treatment decision making, either with cytoreduction, *JAK2* inhibition, or with anticoagulation.

A connection between *JAK2*^{V617F} and platelet activation in human disease is suggested by a mouse model in which a megakaryocyte-specific knock-in of *JAK2*^{V617F} resulted in the production of platelets that were hyperreactive to thrombin and collagen.⁴⁵ Paradoxically, these platelets were hyporesponsive to adenosine diphosphate, suggesting that purinergic blockade may be less effective than aspirin for preventing acute arterial thrombosis in PV.⁴⁵

3.2.4 Microparticles—Serum microparticles are small membrane vesicles that are derived from activated or apoptotic cells, including platelets, monocytes, erythrocytes, and ECs.⁴⁶ Among patients with PV, serum microparticles originating from platelets, erythrocytes, granulocytes, and ECs are elevated compared with healthy controls.^{47, 48} In particular, patients with MPNs that had the *JAK2*^{V617F} mutation had significantly higher plasma concentrations of tissue factor–positive microparticles and erythrocyte microparticles ($P<0.05$).⁴⁸ Among patients with MPNs, serum concentrations of platelet, erythrocyte, and EC microparticles were higher in patients with thrombotic complications ($P<0.05$).⁴⁸

3.2.5 Coagulation System Activation—Patients with PV have demonstrated significantly higher levels of plasma coagulation markers (prothrombin fragment F1+2, thrombin-antithrombin complexes, and D-dimers) than controls ($P<0.05$).⁴² Furthermore, the combined coagulant activity of microparticles, erythrocytes, and platelets is increased in patients with PV, as evidenced by a significantly reduced clotting time ($P<0.01$).⁴⁷

3.3 Proposed Mechanism for Thrombogenesis in Polycythemia Vera

Mechanisms of thrombogenesis in patients with PV remain unclear but appear to be complex, involving multiple components (Figure 1). Statistically significant correlations of *JAK2*^{V617F} allele burden with leukocytosis and elevated hematocrit suggest that several drivers of thrombogenesis may be unique to patients with PV.³⁴ Elevation in hematocrit pushes platelets closer to the vessel wall and has been shown to increase the probability of adhesive wall collisions mediated by platelets binding to vWF and collagen.^{49, 50} Furthermore, under higher shear conditions, as exist with elevated hematocrit and in mid-sized arteries affected by atherothrombotic narrowing, platelets are activated directly⁵¹ and indirectly through leukocyte-platelet interactions mediated by cathepsin G⁴¹ and CD62P (P-selectin).⁵² In addition, constitutively active *JAK2* signaling may directly drive activation of platelets and granulocytes,^{53, 54} and indirectly lead to endothelial activation via platelet and leukocyte binding to ECs.^{39, 55, 56} Such interactions may provoke endothelial injury,^{36, 37} endothelial expression of tissue factor,⁵⁷ and endothelial release of procoagulant factors such as vWF.^{38–40} Furthermore, activated leukocytes, platelets, and ECs secrete microparticles,⁴⁶ which express procoagulant factors important for forming the fibrin clot.^{46, 58} Finally, inflammation in PV may exacerbate the thrombotic state through varied mechanisms that include endothelial damage resulting in EC expression of tissue factor, the inhibition of EC-derived natural anticoagulant pathways, and the secretion of procoagulant factors, particularly vWF.⁵⁹

3.4 Treatment Options

Current treatment recommendations for patients with PV are supported by significant reductions in the risk of cardiovascular death and nonfatal thrombotic events observed in

patients receiving antiplatelet therapy (HR, 0.72; 95% CI, 0.53–0.97; $P=0.0315$) and significantly reduced risk of death from cardiovascular or thrombotic events (HR, 3.91; 95% CI, 1.45–10.53; $P=0.007$) among patients who maintained hematocrit $<45\%$ with phlebotomy, aspirin, and/or cytoreductive agents.²⁸ Hydroxyurea and IFN- α are considered first-line therapeutic agents for patients in a need of cytoreduction.⁶⁰

Second-line chemotherapeutic agents, including busulfan or radioactive phosphorus, are rarely used because of their leukemogenic potential.^{61, 62} Consensus recommendations should also be considered to support aggressive control of conventional thrombotic risk factors like diabetes, hypertension, hyperlipidemia and, of course, abstinence from smoking.^{7, 25} In women, avoiding hormone-based contraception likely reduces the rate of venous thromboembolism,²⁵ although clinical trials would be needed to address this hypothesis prospectively. Although these interventions address the conventional thrombosis risk factors, current treatment options do not target the underlying molecular cause of PV (ie, dysregulation of the JAK/STAT pathway) or PV-related inflammation.

The JAK1/JAK2 inhibitor ruxolitinib is currently in phase 3 trials. The RESPONSE trial compared ruxolitinib with best available therapy in patients with PV who were intolerant of or resistant to hydroxyurea. Initial results of the study suggest reduced thrombotic risk in patients randomized to ruxolitinib compared with best available therapy (0.9% of patients randomized to ruxolitinib had a thrombotic event after 32 weeks vs 5.4% randomized to best available therapy).⁶³ The JAK1/JAK2 inhibitor momelotinib ([ClinicalTrials.gov](https://clinicaltrials.gov/ct2/show/study/NCT01998828) identifier, NCT01998828) is currently in phase 2 clinical trials for patients with ET or PV. Finally, pegylated (PEG)-IFN- α 2a and PEG-IFN- α 2b have been evaluated in phase 2 clinical trials for patients with PV. One study ($n=37$) reported no thrombotic events after a median follow-up of 31 months⁶⁴; two other studies ($n=40$ or 43) indicated that 70% to 76% of patients achieved complete response (a composite endpoint that included absence of thrombosis) after a median follow-up time of 21 or 42 months, respectively.^{65, 66} Further studies will be required to confirm if/how these developing treatment options affect thrombotic-related outcomes in patients with PV.

4 Future Directions

Although many risk factors for thrombosis have been identified for PV, several mechanistic questions remain regarding the causal relationship. Of particular interest is the role of the JAK/STAT pathway and understanding the molecular mechanisms of activated JAK2 on blood cell and vascular activation. It will be important to clarify how thrombogenesis is affected by blood and vascular cell activation, and how serum microparticles, inflammation, and elevated blood cell counts interact with these factors to promote thrombosis. Such data could have important implications for guiding future treatment recommendations and for determining if targeting activated JAK2 is useful for preventing and/or treating PV-related arterial and venous thrombosis. If JAK2-dependent vascular cell activation or inflammation is the trigger of thrombogenesis in patients with PV, inhibition of JAK2 activity could be associated with greater thrombotic benefit than nontargeted treatment options.

5 Conclusions

Thrombogenesis in patients with PV is a complex process that likely involves synergistic interactions between several blood and vascular cells. Elevated blood cell counts; *JAK2*^{V617F} allele burden; high serum concentration of microparticles; and the activation of platelets, leukocytes, and endothelial cells have all been associated with increased risk of thrombotic events. Clarifying mechanisms of thrombogenesis in the PV setting is likely to offer opportunities to improve clinical outcomes for patients with PV and other myeloproliferative disorders.

ACKNOWLEDGMENTS

Writing assistance was provided by Cory Pfeiffenberger, PhD (Complete Healthcare Communications, Inc.), whose work was funded by Incyte Corporation.

CONFLICT OF INTEREST STATEMENT

MHK served as a consultant for Boehringer Ingelheim and has participated on a scientific advisory board for Aplagon Therapeutics. LCM received research support from Celgene, Millenium, Incyte, Pfizer and Seattle Genetics. She has participated in patient education programs funded by Incyte Corporation. SV has received research support from Astrazeneca, Bristol Myers Squibb, Celgene, Cell Therapeutics Inc., Galena BioPharma, Geron, Gilead, Incyte Corporation, Infinity Pharmaceuticals, Lilly Oncology, NS Pharma, Pfizer, Promedior, Roche, and Seattle Genetics.

REFERENCES

1. Gruppo Italiano Studio Policitemia. Polycythemia vera: the natural history of 1213 patients followed for 20 years. Gruppo Italiano Studio Policitemia. *Ann Intern Med.* 1995; 123:656–664. [PubMed: 7574220]
2. Tefferi A, Rumi E, Finazzi G, Gisslinger H, Vannucchi AM, Rodeghiero F, et al. Survival and prognosis among 1545 patients with contemporary polycythemia vera: an international study. *Leukemia.* 2013; 27:1874–1881. [PubMed: 23739289]
3. Kiladjian JJ, Chevret S, Dosquet C, Chomienne C, Rain JD. Treatment of polycythemia vera with hydroxyurea and pipobroman: final results of a randomized trial initiated in 1980. *J Clin Oncol.* 2011; 29:3907–3913. [PubMed: 21911721]
4. Hultcrantz M, Kristinsson SY, Andersson TM, Landgren O, Eloranta S, Derolf AR, et al. Patterns of survival among patients with myeloproliferative neoplasms diagnosed in Sweden from 1973 to 2008: a population-based study. *J Clin Oncol.* 2012; 30:2995–3001. [PubMed: 22802311]
5. Mehta J, Wang H, Fryzek JP, Iqbal SU, Mesa R. Health resource utilization and cost associated with myeloproliferative neoplasms in a large United States health plan. *Leuk Lymphoma.* 2014; 55:2368–2374. [PubMed: 24450579]
6. Tefferi A. Polycythemia vera and essential thrombocythemia: 2013 update on diagnosis, risk-stratification, and management. *Am J Hematol.* 2013; 88:507–516. [PubMed: 23695894]
7. Passamonti F. How I treat polycythemia vera. *Blood.* 2012; 120:275–284. [PubMed: 22611155]
8. Alvarez-Larran A, Pereira A, Cervantes F, Arellano-Rodrigo E, Hernandez-Boluda JC, Ferrer-Marin F, et al. Assessment and prognostic value of the European LeukemiaNet criteria for clinicohematologic response, resistance, and intolerance to hydroxyurea in polycythemia vera. *Blood.* 2012; 119:1363–1369. [PubMed: 22160617]
9. Barbui T, Finazzi MC, Finazzi G. Front-line therapy in polycythemia vera and essential thrombocythemia. *Blood Rev.* 2012; 26:205–211. [PubMed: 22784966]
10. Levine RL, Wadleigh M, Cools J, Ebert BL, Wernig G, Huntly BJ, et al. Activating mutation in the tyrosine kinase *JAK2* in polycythemia vera, essential thrombocythemia, and myeloid metaplasia with myelofibrosis. *Cancer Cell.* 2005; 7:387–397. [PubMed: 15837627]

11. Kralovics R, Passamonti F, Buser AS, Teo SS, Tiedt R, Passweg JR, et al. A gain-of-function mutation of JAK2 in myeloproliferative disorders. *N Engl J Med*. 2005; 352:1779–1790. [PubMed: 15858187]
12. James C, Ugo V, Le Couedic JP, Staerk J, Delhommeau F, Lacout C, et al. A unique clonal JAK2 mutation leading to constitutive signalling causes polycythaemia vera. *Nature*. 2005; 434:1144–1148. [PubMed: 15793561]
13. Baxter EJ, Scott LM, Campbell PJ, East C, Fourouclas N, Swanton S, et al. Acquired mutation of the tyrosine kinase JAK2 in human myeloproliferative disorders. *Lancet*. 2005; 365:1054–1061. [PubMed: 15781101]
14. Passamonti F, Rumi E, Pietra D, Elena C, Boveri E, Arcaini L, et al. A prospective study of 338 patients with polycythemia vera: the impact of JAK2 (V617F) allele burden and leukocytosis on fibrotic or leukemic disease transformation and vascular complications. *Leukemia*. 2010; 24:1574–1579. [PubMed: 20631743]
15. Quintas-Cardama A, Kantarjian H, Cortes J, Verstovsek S. Janus kinase inhibitors for the treatment of myeloproliferative neoplasias and beyond. *Nat Rev Drug Discov*. 2011; 10:127–140. [PubMed: 21283107]
16. Barbui T, Carobbio A, Finazzi G, Vannucchi AM, Barosi G, Antonioli E, et al. Inflammation and thrombosis in essential thrombocythemia and polycythemia vera: different role of C-reactive protein and pentraxin 3. *Haematologica*. 2011; 96:315–318. [PubMed: 21173097]
17. Chung I, Lip GY. Virchow's triad revisited: blood constituents. *Pathophysiol Haemost Thromb*. 2003; 33:449–454. [PubMed: 15692259]
18. Virchow, R. *Thrombosis and emboli (1846–1856)*. Canton, Mass: Science History Publications; 1998.
19. Nieswandt B, Pleines I, Bender M. Platelet adhesion and activation mechanisms in arterial thrombosis and ischaemic stroke. *J Thromb Haemost*. 2011; 9(suppl 1):92–104. [PubMed: 21781245]
20. Wakefield TW, Myers DD, Henke PK. Mechanisms of venous thrombosis and resolution. *Arterioscler Thromb Vasc Biol*. 2008; 28:387–391. [PubMed: 18296594]
21. Mackman N. New insights into the mechanisms of venous thrombosis. *J Clin Invest*. 2012; 122:2331–2336. [PubMed: 22751108]
22. Rumbaut, RE.; Thiagarajan, P. *Platelet-Vessel Wall Interactions in Hemostasis and Thrombosis*. San Rafael (CA): Morgan & Claypool Life Sciences; 2010. Arterial, Venous, and Microvascular Hemostasis/Thrombosis.
23. Marchioli R, Finazzi G, Landolfi R, Kutti J, Gisslinger H, Patrono C, et al. Vascular and neoplastic risk in a large cohort of patients with polycythemia vera. *J Clin Oncol*. 2005; 23:2224–2232. [PubMed: 15710945]
24. Smalberg JH, Arends LR, Valla DC, Kiladjian JJ, Janssen HL, Leebeek FW. Myeloproliferative neoplasms in Budd-Chiari syndrome and portal vein thrombosis: a meta-analysis. *Blood*. 2012; 120:4921–4928. [PubMed: 23043069]
25. Previtali E, Bucciarelli P, Passamonti SM, Martinelli I. Risk factors for venous and arterial thrombosis. *Blood Transfus*. 2011; 9:120–138. [PubMed: 21084000]
26. Landolfi R, Di Gennaro L, Barbui T, De Stefano V, Finazzi G, Marfisi R, et al. Leukocytosis as a major thrombotic risk factor in patients with polycythemia vera. *Blood*. 2007; 109:2446–2452. [PubMed: 17105814]
27. Gangat N, Strand J, Li CY, Wu W, Pardananani A, Tefferi A. Leucocytosis in polycythaemia vera predicts both inferior survival and leukaemic transformation. *Br J Haematol*. 2007; 138:354–358. [PubMed: 17614822]
28. Marchioli R, Finazzi G, Specchia G, Cacciola R, Cavazzina R, Cilloni D, et al. Cardiovascular events and intensity of treatment in polycythemia vera. *N Engl J Med*. 2013; 368:22–33. [PubMed: 23216616]
29. Di Nisio M, Barbui T, Di Gennaro L, Borrelli G, Finazzi G, Landolfi R, et al. The haematocrit and platelet target in polycythemia vera. *Br J Haematol*. 2007; 136:249–259. [PubMed: 17156406]
30. Cinar Y, Demir G, Pac M, Cinar AB. Effect of hematocrit on blood pressure via hyperviscosity. *Am J Hypertens*. 1999; 12:739–743. [PubMed: 10411372]

31. De Grandis M, Cambot M, Wautier MP, Cassinat B, Chomienne C, Colin Y, et al. JAK2V617F activates Lu/BCAM-mediated red cell adhesion in polycythemia vera through an EpoR-independent Rap1/Akt pathway. *Blood*. 2013; 121:658–665. [PubMed: 23160466]
32. Caramazza D, Caracciolo C, Barone R, Malato A, Saccullo G, Cigna V, et al. Correlation between leukocytosis and thrombosis in Philadelphia-negative chronic myeloproliferative neoplasms. *Ann Hematol*. 2009; 88:967–971. [PubMed: 19214510]
33. De Stefano V, Za T, Rossi E, Vannucchi AM, Ruggeri M, Elli E, et al. Recurrent thrombosis in patients with polycythemia vera and essential thrombocythemia: incidence, risk factors, and effect of treatments. *Haematologica*. 2008; 93:372–380. [PubMed: 18268279]
34. Vannucchi AM, Antonioli E, Guglielmelli P, Longo G, Pancrazzi A, Ponziani V, et al. Prospective identification of high-risk polycythemia vera patients based on JAK2(V617F) allele burden. *Leukemia*. 2007; 21:1952–1959. [PubMed: 17625606]
35. Levine RL, Belisle C, Wadleigh M, Zahrieh D, Lee S, Chagnon P, et al. X-inactivation-based clonality analysis and quantitative JAK2V617F assessment reveal a strong association between clonality and JAK2V617F in PV but not ET/MMM, and identifies a subset of JAK2V617F-negative ET and MMM patients with clonal hematopoiesis. *Blood*. 2006; 107:4139–4141. [PubMed: 16434490]
36. Harlan JM, Killen PD, Harker LA, Striker GE, Wright DG. Neutrophil-mediated endothelial injury in vitro mechanisms of cell detachment. *J Clin Invest*. 1981; 68:1394–1403. [PubMed: 7033282]
37. Weksler BB, Jaffe EA, Brower MS, Cole OF. Human leukocyte cathepsin G and elastase specifically suppress thrombin-induced prostacyclin production in human endothelial cells. *Blood*. 1989; 74:1627–1634. [PubMed: 2477083]
38. Celi A, Pellegrini G, Lorenzet R, De Blasi A, Ready N, Furie BC, et al. P-selectin induces the expression of tissue factor on monocytes. *Proc Natl Acad Sci U S A*. 1994; 91:8767–8771. [PubMed: 7522321]
39. Falanga A, Marchetti M, Evangelista V, Manarini S, Oldani E, Giovanelli S, et al. Neutrophil activation and hemostatic changes in healthy donors receiving granulocyte colony-stimulating factor. *Blood*. 1999; 93:2506–2514. [PubMed: 10194429]
40. Lip GY, Blann A. von Willebrand factor: a marker of endothelial dysfunction in vascular disorders? *Cardiovasc Res*. 1997; 34:255–265. [PubMed: 9205537]
41. Evangelista V, Rajtar G, de Gaetano G, White JG, Cerletti C. Platelet activation by fMLP-stimulated polymorphonuclear leukocytes: the activity of cathepsin G is not prevented by antiproteinases. *Blood*. 1991; 77:2379–2388. [PubMed: 1645603]
42. Falanga A, Marchetti M, Evangelista V, Vignoli A, Licini M, Balicco M, et al. Polymorphonuclear leukocyte activation and hemostasis in patients with essential thrombocythemia and polycythemia vera. *Blood*. 2000; 96:4261–4216. [PubMed: 11110700]
43. Falanga A, Marchetti M, Vignoli A, Balducci D, Barbui T. Leukocyte-platelet interaction in patients with essential thrombocythemia and polycythemia vera. *Exp Hematol*. 2005; 33:523–530. [PubMed: 15850829]
44. Cerletti C, Tamburrelli C, Izzi B, Gianfagna F, de Gaetano G. Platelet-leukocyte interactions in thrombosis. *Thromb Res*. 2012; 129:263–266. [PubMed: 22075180]
45. Hobbs CM, Manning H, Bennett C, Vasquez L, Severin S, Brain L, et al. JAK2V617F leads to intrinsic changes in platelet formation and reactivity in a knock-in mouse model of essential thrombocythemia. *Blood*. 2013; 122:3787–3797. [PubMed: 24085768]
46. Owens AP 3rd, Mackman N. Microparticles in hemostasis and thrombosis. *Circ Res*. 2011; 108:1284–1297. [PubMed: 21566224]
47. Tan X, Shi J, Fu Y, Gao C, Yang X, Li J, et al. Role of erythrocytes and platelets in the hypercoagulable status in polycythemia vera through phosphatidylserine exposure and microparticle generation. *Thromb Haemost*. 2013; 109:1025–1032. [PubMed: 23571603]
48. Han Y, Zhao S, Zhang W, Cen J, Zhang W, Qiu H, et al. Clinical significance of circulating microparticles in Ph-myeloproliferative neoplasms (MPN). *Blood (ASH Annual Meeting Abstracts)*. 2013; 122 abstract 2368.

49. Chen H, Angerer JJ, Napoleone M, Reininger AJ, Schneider SW, Wixforth A, et al. Hematocrit and flow rate regulate the adhesion of platelets to von Willebrand factor. *Biomicrofluidics*. 2013; 7:64113. [PubMed: 24396547]
50. Nuytens BP, Thijs T, Deckmyn H, Broos K. Platelet adhesion to collagen. *Thromb Res*. 2011; 127(suppl 2):S26–S29. [PubMed: 21193111]
51. Holme PA, Orvim U, Hamers MJ, Solum NO, Brosstad FR, Barstad RM, et al. Shear-induced platelet activation and platelet microparticle formation at blood flow conditions as in arteries with a severe stenosis. *Arterioscler Thromb Vasc Biol*. 1997; 17:646–653. [PubMed: 9108776]
52. Evangelista V, Piccardoni P, White JG, de Gaetano G, Cerletti C. Cathepsin G-dependent platelet stimulation by activated polymorphonuclear leukocytes and its inhibition by antiproteases: role of P-selectin-mediated cell-cell adhesion. *Blood*. 1993; 81:2947–2957. [PubMed: 7684621]
53. Lu WJ, Lin KC, Huang SY, Thomas PA, Wu YH, Wu HC, et al. Role of a Janus kinase 2-dependent signaling pathway in platelet activation. *Thromb Res*. 2014; 133:1088–1096. [PubMed: 24731555]
54. Passamonti F, Rumi E, Pietra D, Della Porta MG, Boveri E, Pascutto C, et al. Relation between JAK2 (V617F) mutation status, granulocyte activation, and constitutive mobilization of CD34+ cells into peripheral blood in myeloproliferative disorders. *Blood*. 2006; 107:3676–3682. [PubMed: 16373657]
55. Henn V, Slupsky JR, Grafe M, Anagnostopoulos I, Forster R, Muller-Berghaus G, et al. CD40 ligand on activated platelets triggers an inflammatory reaction of endothelial cells. *Nature*. 1998; 391:591–594. [PubMed: 9468137]
56. Dole VS, Bergmeier W, Mitchell HA, Eichenberger SC, Wagner DD. Activated platelets induce Weibel-Palade-body secretion and leukocyte rolling in vivo: role of P-selectin. *Blood*. 2005; 106:2334–2339. [PubMed: 15956287]
57. Slupsky JR, Kalbas M, Willuweit A, Henn V, Kroczeck RA, Muller-Berghaus G. Activated platelets induce tissue factor expression on human umbilical vein endothelial cells by ligation of CD40. *Thromb Haemost*. 1998; 80:1008–1014. [PubMed: 9869175]
58. De Caterina R, Husted S, Wallentin L, Andreotti F, Arnesen H, Bachmann F, et al. General mechanisms of coagulation and targets of anticoagulants (Section I). Position Paper of the ESC Working Group on Thrombosis--Task Force on Anticoagulants in Heart Disease. *Thromb Haemost*. 2013; 109:569–579. [PubMed: 23447024]
59. Aksu K, Donmez A, Keser G. Inflammation-induced thrombosis: mechanisms, disease associations and management. *Curr Pharm Des*. 2012; 18:1478–1493. [PubMed: 22364132]
60. Barbui T, Barosi G, Birgegard G, Cervantes F, Finazzi G, Griesshammer M, et al. Philadelphia-negative classical myeloproliferative neoplasms: critical concepts and management recommendations from European LeukemiaNet. *J Clin Oncol*. 2011; 29:761–770. [PubMed: 21205761]
61. Berk PD, Goldberg JD, Silverstein MN, Weinfeld A, Donovan PB, Ellis JT, et al. Increased incidence of acute leukemia in polycythemia vera associated with chlorambucil therapy. *N Engl J Med*. 1981; 304:441–447. [PubMed: 7005681]
62. Haanen C, Mathé G, Hayat M, Peetermans N, Cauchie C, Michel J, et al. Treatment of polycythaemia vera by radiophosphorus or busulphan: a randomized trial. "Leukemia and Hematosarcoma" Cooperative Group, European Organization for Research on Treatment of Cancer (E.O.R.T.C.). *Br J Cancer*. 1981; 44:75–80. [PubMed: 7020738]
63. Verstovsek S, Kiladjian JJ, Griesshammer M, Masszi T, Durrant S, Passamonti F, et al. Results of a prospective, randomized, open-label phase 3 study of ruxolitinib (RUX) in polycythemia vera (PV) patients resistant to or intolerant of hydroxyurea (HU): the RESPONSE trial. *J Clin Oncol*. 2014; 32 abstract 7026.
64. Kiladjian JJ, Cassinat B, Chevret S, Turlure P, Cambier N, Roussel M, et al. Pegylated interferon- α -2a induces complete hematologic and molecular responses with low toxicity in polycythemia vera. *Blood*. 2008; 112:3065–3072. [PubMed: 18650451]
65. Quintas-Cardama A, Kantarjian H, Manshoury T, Luthra R, Estrov Z, Pierce S, et al. Pegylated interferon α -2a yields high rates of hematologic and molecular response in patients with

advanced essential thrombocythemia and polycythemia vera. *J Clin Oncol.* 2009; 27:5418–5424. [PubMed: 19826111]

66. Quintas-Cardama A, Abdel-Wahab O, Manshouri T, Kilpivaara O, Cortes J, Roupie AL, et al. Molecular analysis of patients with polycythemia vera or essential thrombocythemia receiving pegylated interferon alpha-2a. *Blood.* 2013; 122:893–901. [PubMed: 23782935]

Practice Points

- Cardiovascular and thrombotic events contribute to the increased mortality rate among patients with PV compared with the general population.
- Advanced age, a history of thrombosis, elevated hematocrit, and leukocytosis are risk factors for cardiovascular and thrombotic events in patients with PV.
- To reduce thrombotic risk, patients with PV should receive aspirin with phlebotomy and/or cytoreductive therapy to maintain hematocrit <45%.

Research Agenda

- Characterization of mechanistic links between thrombotic risk factors and hematologic proliferation in PV
- Determination of the prognostic and predictive value of nonconventional risk factors for thrombosis in patients with PV
- Determination of the clinical benefit associated with PV treatment options that target the underlying causes of PV and thrombosis

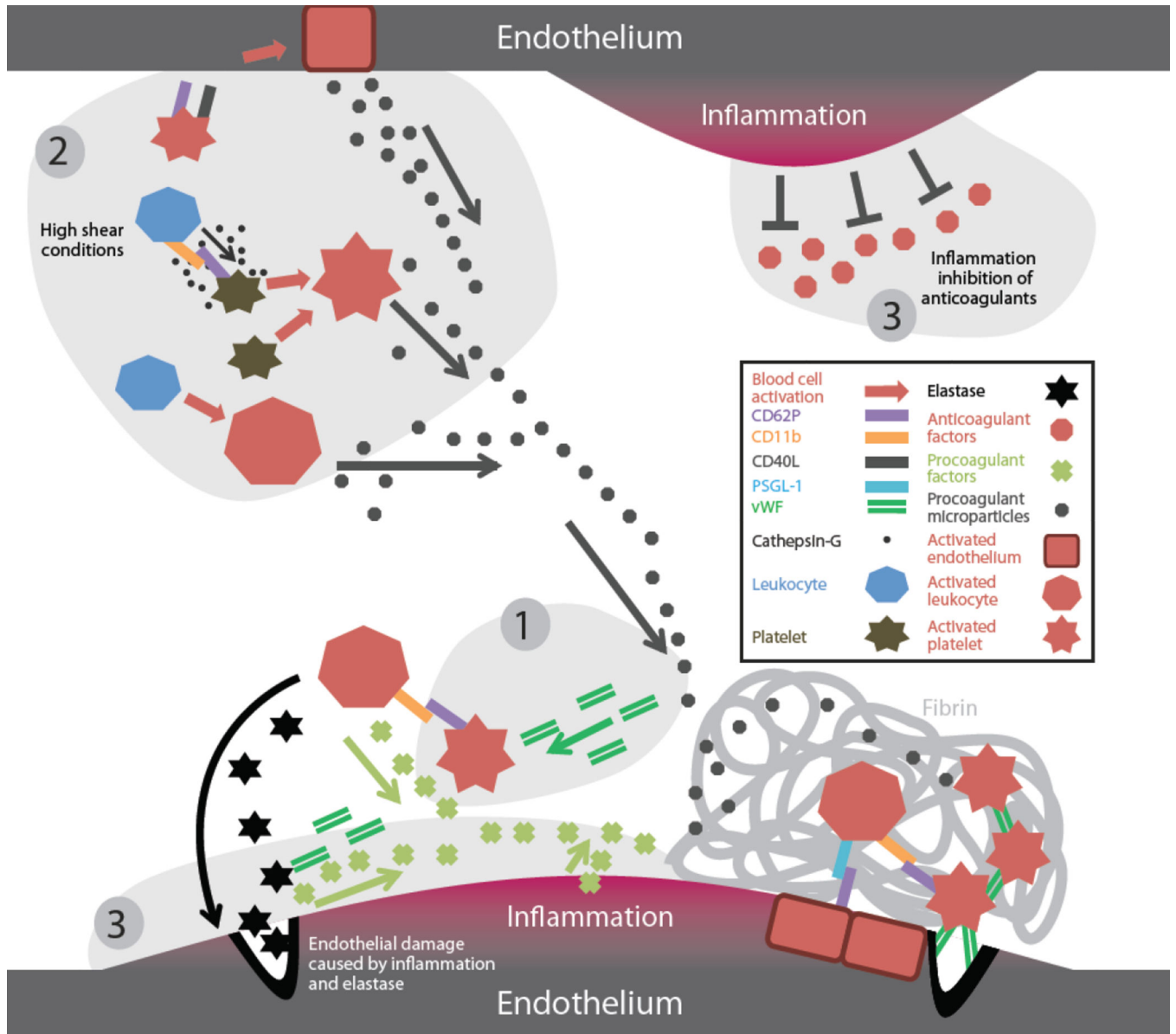


Figure 1. Proposed Mechanism of Thrombogenesis in Patients With PV. Thrombogenesis in patients with PV is a complex process that involves many components. Future research will be necessary to confirm and refine this proposed mechanism; however, several drivers of thrombogenesis are depicted in this figure that may be unique to patients with PV. (1) Under high hematocrit conditions common in patients with PV, platelets are pushed centrifugally and are more likely to adhere to vessel wall collagen and vWF, thus initiating thrombus formation. (2) High shear conditions resulting from vascular damage and elevated blood cell counts directly enhance platelet adhesion and activation, and thereby promote leukocyte-platelet interactions. These interactions, which are principally mediated by platelet CD62P binding to leukocyte CD11b, result in neutrophil and monocyte activation. Neutrophil activation amplifies platelet activation through the release of cathepsin G, and monocyte

activation results in its expression of tissue factor, leading to the activation of the soluble coagulation system. *JAK2* mutations also promote the direct activation of leukocytes and platelets. Activated platelets and leukocytes bind to and activate ECs, and activated leukocytes, platelets, and ECs secrete procoagulant microparticles that contribute to formation of the fibrin clot. (3) PV-related blood cell and EC activation, as well as PV-related inflammation, results in endothelial injury. EC injury results in a prothrombotic state through the inhibition of EC-derived natural anticoagulant pathways, and the stimulation of several EC-derived procoagulant factors, particularly tissue factor and vWF. EC=endothelial cell; PV=polycythemia vera; vWF= von Willebrand factor.

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

Table 1

Incidence of Thrombotic Events in Patients With PV

Reference	Patients, n	Thrombotic event	Incidence, %
Tefferi et al 2013 ²	1545	At or before diagnosis ^a	
		Arterial	16
		Venous	7.4
		After diagnosis ^b	
		Arterial	12
		Venous	9
Marchioli et al 2005 ²³	1638	Nonfatal thrombotic events during follow-up ^c	10.3
		Arterial thrombosis	5.3
		TIA	2.0
		Stroke	1.4
		Peripheral	1.2
		MI	0.9
		Venous thrombosis	5.4
		Superficial thrombophlebitis	2.8
		DVT	2.3
		Pulmonary embolism	0.8
		Fatal cardiovascular events during follow-up ^c	4.5
		Coronary heart disease	1.5
		Congestive heart failure	0.8
		Nonhemorrhagic stroke	0.8
		Pulmonary embolism	0.4
		Intracranial hemorrhage	0.1
		Other cardiac disease	0.3
		Other hemorrhage	0.3
Other vascular death	0.3		
Gruppo Italiano Studio Policitemia 1995 ¹	1213	Before diagnosis ^d	14
		At diagnosis	20
		During follow-up ^e	19

DVT=deep vein thrombosis; MI=myocardial infarction; PV=polycythemia vera; TIA=transient ischemic attack.

^aThe analysis was initiated in 2010 and included patients diagnosed with PV after 1970.

^bThe median (range) follow-up was 6.9 years (0–39.3 years).

^cThe mean (SD) follow-up was 2.7 (1.3) years.

^dPatient histories included 33 years before diagnosis.

^eThe median (range) follow-up was 5.3 years (0–32.9 years).

Table 2

Risk Factors for Thrombosis in Patients With PV

Reference	Analysis Type	Patients, n	Risk Factor for Thrombosis	Comparison	P Value and/or HR (95% CI) ^a
Gangat et al 2007 ²⁷	Retrospective	459	Prior thrombotic event	vs no prior thrombotic event	<i>P</i> <0.0001
Landolfi et al 2007 ²⁶	Retrospective	1638		vs no prior thrombotic event	1.69 (1.21–2.36)
Marchioli et al 2005 ²³	Prospective	1638		vs no prior thrombotic event	2.09 (1.55–2.81) ^b
Gruppo Italiano 1995 ¹	Retrospective	1213		vs no prior thrombotic event	<i>P</i> =0.001
Caramazza et al 2009 ³²	Prospective	99 ^c	Leukocytosis	>9.5 × 10 ⁹ /L vs 9.5 × 10 ⁹ /L at baseline	<i>P</i> =0.03 1.8 (upper 95% CI, 3.4)
				>9.5 × 10 ⁹ /L vs 9.5 × 10 ⁹ /L time-dependent analysis	No significant correlation 1.3 (upper 95% CI, 2.9)
De Stefano et al 2008 ³³	Retrospective	235 ^d		High vs low WBC count quartile in patients aged <60 years	3.55 (1.02–12.25)
Landolfi et al 2007 ²⁶	Retrospective	1638		>15 × 10 ⁹ /L vs 10.1 to 15 × 10 ⁹ /L	<i>P</i> =0.028 1.56 (1.05–2.30)
Gangat et al 2007 ²⁷	Retrospective	459		15 × 10 ⁹ /L vs <15 × 10 ⁹ /L	<i>P</i> =0.005
Marchioli et al 2013 ²⁸	Prospective, randomized	365	Elevated hematocrit	<45% vs 45% to 50%	<i>P</i> =0.02 ^e 2.69 (1.19–6.12)
Caramazza et al 2009 ³²	Prospective	99 ^c		45% vs >45% at baseline	No significant correlation 1.3 (upper 95% CI, 1.8)
				45% vs >45% time-dependent analysis	No significant correlation 1.4 (upper 95% CI, 3.3)
Gangat et al 2007 ²⁷	Retrospective	459		Not specified	No significant correlation
Di Nisio et al 2007 ²⁹	Retrospective	1638		45% vs 46% to 50%	No significant correlation
				45% vs >50%	<i>P</i> =0.5844 0.89 (0.60–1.34)
					No significant correlation
					<i>P</i> =0.8884 1.04 (0.61–1.78)
Passamonti et al 2010 ¹⁴	Prospective	338	JAK2 ^{V617F} allele burden	<50% vs 50%	No significant correlation
Vannucchi et al 2007 ³⁴	Prospective	173		25% vs >75%	<i>P</i> =0.8 1.0 (0.9–1.0)
					<i>P</i> =0.003 7.1 ^f (1.6–10.1)

HR=hazard ratio; PV=polycythemia vera; WBC=white blood cell.

^a *P* value and HR presented when available in the source material.

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

^b Risk of cardiovascular events, defined as cardiovascular death and nonfatal thrombotic events.

^c Analyses were conducted on the total patient population, which included 187 patients (PV, n=99; essential thrombocythemia, n=88).

^d Analyses were conducted on the total patient population, which included 494 patients (PV, n=235; essential thrombocythemia, n=259).

^e Total cardiovascular events.

^f Relative risk for thrombosis during a median follow-up of 24 months.