

RESEARCH ARTICLE

Open Access



Evaluation of anthelmintic potential of the Ethiopian medicinal plant *Embelia schimperi* Vatke *in vivo* and *in vitro* against some intestinal parasites

Yared Debebe^{1*}, Mesfin Tefera², Walegn Mekonnen¹, Dawit Abebe³, Samuel Woldekidan¹, Abiy Abebe¹, Yehualashet Belete³, Temesgen Menberu¹, Bethelhem Belayneh¹, Berhanu Tesfaye³, Ibrahim Nasir³, Kidist Yirsaw¹, Hirut Basha^{1,4}, Asrat Dawit^{1,4} and Asfaw Debella³

Abstract

Background: *Embelia schimperi* has been used for the treatment of intestinal parasites especially tapeworm infestations for centuries in Ethiopia. However, there is lack of scientific based evidences regarding the efficacy, safety and phytochemical analysis of this plant despite its frequent use as an anthelmintic. This study has therefore evaluated the efficacy and acute toxicity of *E. schimperi* thereby generating relevant preclinical information.

Methods: The anthelmintic activities of the crude hydroalcoholic extract of *E. schimperi* and the isolated compound, embelin, were conducted using *in vivo* and *in vitro* models against the dwarf tapeworm, *Hymenolepis nana*, and the hookworm, *Necator americanus*, respectively. LD₅₀ of the crude hydroalcoholic extract was determined using Swiss albino mice following the OECD guidelines. Chemical characterization of the isolated embelin was conducted using UV-spectroscopy, HPLC and NMR.

Results: In the acute toxicity study no prominent signs of toxicity and mortality were recorded among the experimental animals at the highest administered dose. Hence the LD₅₀ of the plant was found to be higher than 5000 mg/kg. *In vivo* cestocidal activity of the crude hydroalcoholic extract of *E. schimperi* showed 100 % parasite clearance at 1000 mg/kg, while the diammonium salt of embelin showed 85.3 % parasite clearance at 750 mg/kg. The *in vitro* anthelmintic activity study revealed that the LC₅₀ value of the crude extract and albendazole were 228.7 and 51.33 µg/mL, respectively.

Conclusion: The results clearly indicated that the hydroalcoholic extract of *E. schimperi* and the diammonium salt of the isolated compound embelin had anthelmintic activity against hookworm larva *in vitro* and *H. nana in vivo*. Hence the findings of this study showed *Embelia schimperi* appears to possess some anthelmintic activity that may support the usage of these plants by local traditional healers to treat helminthic infestations.

Keywords: Anthelmintics, Cestocidal, Embelin, *Hymenolepis nana*

Background

Helminthiasis have been affecting human beings throughout the history of mankind and still continue to be major cause of mortality and morbidity to over a billion people in the world, particularly in developing regions like the sub-Saharan Africa (SSA), Asia and the

Americas. These regions of the world are characterized by marginalized societies with resource constraints making the burden more vicious exacerbating other health and socioeconomic problems like malaria, HIV/AIDS and decreased productivity [1]. The Ethiopian scenario is not different from other SSA countries as the Federal Ministry of Health of Ethiopia (FMOH) [2] reported that annual visits of more than half a million cases in the outpatient departments of health facilities are as a result of intestinal parasitic infections including helminthiasis. This number might not represent the actual burden as

* Correspondence: papioan@gmail.com

¹Ethiopian Public Health Institute, Biomedical and Clinical Research Team, Traditional and Modern Medicine Research Directorate, P.O.Box 1242, Addis Ababa, Ethiopia

Full list of author information is available at the end of the article

some of the health facilities lack the appropriate diagnostic methods as well as failure of detecting lower parasite burden [3].

The Ethiopian custom of using medicinal plants for the treatment of intestinal parasites has existed for many generations. Quite many numbers of herbal remedies are prescribed in the traditional health care system in this regard. *Embelia schimperi*, a plant which belongs to family Myrsinaceae, is among the most widely used anthelmintic medicinal plant in Ethiopian folk medicine. Various ethnomedical studies revealed that the fruit is widely consumed to expel the adult stage of beef tapeworm and other intestinal parasites from the body [4–7]. Despite this frequent utilization of the plant as an alternative anthelmintic medicine, scientific evidences are less abundant supporting the traditional claims in Ethiopia. This study was therefore evaluated the acute toxicity and anthelmintic properties of the most frequently used medicinal plant, *Embelia schimperi* Vatke.

Methods

Plant collection and authentication

Fruits of *Embelia schimperi* were collected from the localities in Gonder, Northwest Ethiopia. A voucher specimen (ES-2175) was collected and deposited at the herbarium of Traditional and Modern Medicine Research Directorate (TMMRD), Ethiopian Public Health Institute (EPHI) following its authentication by the taxonomist of the research directorate.

Experimental animals

The animals used in this study were the Swiss albino mice, *Mus musculus albinus*. The animals were obtained from the animal breeding unit of EPHI and kept in standard cages in the animal house of TMMRD. They were fed with standard pellet diet and tap water *ad libitum* and maintained at temperature of 21 ± 2 °C and humidity of 65 ± 0.5 % with 12 h light/dark cycles until the end of the experiment. All experimental animals were acclimatized for 10 days prior to the experimental procedures.

Preparation of crude extract

The air dried fruits of *E. schimperi* were pulverized into powder form using an electronic grinding mill. The fruit powder (200 g) was extracted in 80 % ethanol by maceration. The extract was then filtered with Whatman filter paper No 3 and the organic solvent was removed from the filtrate using rotary evaporator (BUCHI B-205, Switzerland). The aqueous residue was further lyophilized (Labconco, USA) resulting in 15.79 g of crude extract.

Isolation of embelin from *E. schimperi* and preparation of diammonium salt of embelin

Isolation of the reference compound, embelin, was conducted following the procedure described by Belete et al. [8]. Fruit powder of *E. schimperi* weighing 1.1 Kg was extracted using ethyl-acetate for 25 min and then transferred to a shaker for 4 h and filtered with Whatman filter paper No. 41. The marc was extracted one more time with the same solvent and followed by evaporation under vacuum yielding 119.71 g. The resulting extract was successively washed using hexane which resulted 5 g of an orange crystal of pure embelin having a melting point of 142–143 °C whose identity was fully elucidated by HPLC, ¹H, ¹³C, DEPT-135, HMQC, HMBC and UV spectra. The pure isolated embelin was then dissolved with methanol and ammonium was added until the solution turned into pink followed by evaporation to produce diammonium salt of embelin. The salt was completely soluble in water and easy for administration to the experimental animals as opposed to the pure embelin which was totally insoluble in water.

Acute toxicity study and determination of LD₅₀

The acute toxicity of the crude extract was determined following Lorke's [9] method. The crude fruit extract was administered orally at increasing doses of 625, 1250, 2500, and 5000 mg/kg, p.o. to six animals in each group. The general signs and symptoms of toxicity such as death, changes in physical appearance or behavioral changes were observed for 24 h post administration of the extract. The median lethal dose (LD₅₀) was calculated as geometric mean of the dose that resulted in 100 % lethality and the maximum dose with no lethality at all.

In vitro parasite culturing of hookworm eggs

The filter paper test tube culture technique following Harada and Mori [10] was used for the development of larvae from egg infested stool. Stool samples were collected in Wolkite health center after verbal consent was obtained from the patients. About 1 g of fecal matter was applied on a filter paper which was then inserted in a test tube containing water. The samples were then incubated under room temperature for approximately ten days. Monitoring of larval development of *N. americanus* was conducted until the desired larval stage was achieved for the efficacy study. After obtaining the infective larval stage the samples were centrifuged at a speed of $402 \times g$ for ten minutes and concentrated. The number of larvae present was adjusted to achieve a concentration of one larva per microliter.

In vitro anthelmintic activity

The 96-well microtiter plate assay described by Gill and others [11] was followed to evaluate the effects of the crude extract on the third stage larvae of *N. americanus*. Stock solutions of the crude extract of *E. schimperi* and albendazole (Sigma-Aldrich), the standard drug serving as a positive control, were prepared at 100 and 20 mg/mL, respectively, in 1 % dimethylsulfoxide (DMSO) and were serially diluted by two-fold to produce a series of dilutions. Aliquots were added at a dilution of 1 % to molten nutrient agar in a total volume of 200 μ L in individual wells of a 96-well microtiter plate. The final concentrations in the assay plates consisted of two-fold serial dilutions starting 1, 0.5, 0.25, 0.125 and 0.0625 mg/mL for the plant extract and 0.2, 0.1, 0.05, 0.025 and 0.0125 mg/mL for the standard drug, albendazole. Approximately 30 larvae in 30 μ L of distilled water were added to each well, and the plate was incubated in the dark at 25 °C for 48 h.

The effect of the drugs on worm viability was assessed by counting the numbers of motile larvae after the 48 h of incubation period. The larvae were stimulated to move by addition of 40 μ L of water warmed at 50 °C to each well, control larvae and those unaffected by the extract and the standard drug were observed to move in a rapid sinusoidal motion. In contrast, extract and drug affected larvae showed a twitching motion or remained motionless. Individual larva moving with sinusoidal motion was counted. All assays were performed using triplicate assay wells at each drug concentrations.

In vivo anticestodal activity

The anticestodal activities of the test extracts were conducted on experimentally infected Swiss albino mice which were maintained in the laboratory. Adult worms of *H. nana* were collected from the intestines of infected mice. Eggs were collected from the worm through dissection and the dose of infection was adjusted to be 800 eggs in 0.1 ml of normal saline. Each experimental mouse was then infected orally by stomach tube. Two weeks post infection, fresh fecal samples from each infected mice were collected and examined for shedding of ova. Mice not shedding ova of *H. nana* were discarded from the experiments.

The mice were then divided randomly into five groups of six mice each for each test extract. The three groups of animals were assigned as the test groups whereas the other two groups were used as control (positive and negative). Animals in the positive control group were treated with the standard drug Praziquantel at 25 mg/kg body weight whereas the negative control received equal volume of the vehicle. The other three groups were used for evaluation of the Anticestodal activities of the test

extracts. 48 h post extract and drug administration; all the mice were sacrificed to determine percent deparasitization [12].

$$\% \text{ deparasitization} = (N-n)/N \times 100$$

N = numbers of worms counted in the negative control group

n = number of worms counted in the plant extract or Praziquantel treated mice

Percent host clearance was determined using the following formula

$$\% \text{ host clearance} = A/B \times 100$$

A = number of hosts found devoid of worms

B = number of hosts originally infected with worms

Statistical analysis

Determination of LC_{50} of a sigmoidal concentration response curve was performed using GraphPad Prism version 6.04 for Windows (GraphPad Prism®; GraphPad Software, Inc., San Diego, California, USA). Statistical significance for *in vivo* anthelmintic activity effect of the extracts was determined using one way ANOVA and LSD multiple comparison tests. P value <0.05 was considered significant.

Ethical considerations

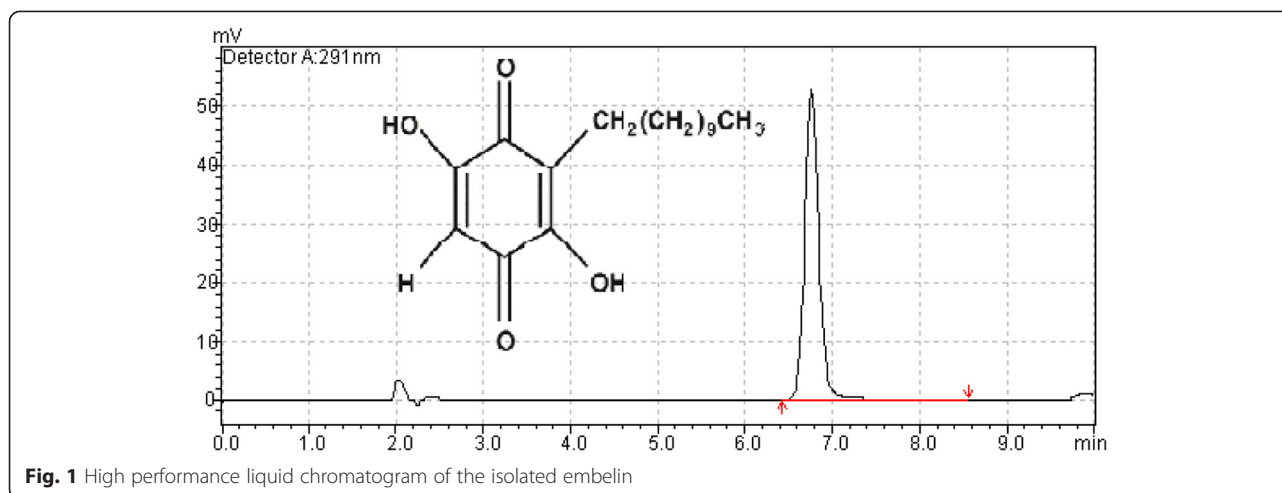
Ethical approval for collection of patient stool samples and use of experimental animals for the experimental procedures was obtained from the Scientific and Ethical Review Office, EPHI (006/2003). Patients' consent was sought in verbal form. Those people with confirmed hookworm infection were treated with albendazole. Animals were handled with care as per the guideline of EPHI for animal handling.

Result**Isolation of embelin from *E. schimperi***

The compound isolated was embelin confirmed by NMR spectroscopy. $^1\text{H-NMR}$ and $^{13}\text{C-NMR}$ spectra of the purified sample verified the identity of the compound to be embelin. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ : 7.68 (s, 2H, OH), 6.00 (s, 1H, H-6), 2.44 (t, 2H, H-1¹, J = 6.9 Hz), 1.47 (m, 2H, H-2¹) 1.25-1.30 (m, 16H, H-3¹ to 10¹), 0.88 (t, 3H, H-11¹, J = 6.9 Hz); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) (3, 116.99), (6, 102.15), (1¹-10¹, 22.51–31.90), (11¹, 14.09). The HPLC profile of the purified embelin is shown in Fig. 1.

Acute toxicity study and LD_{50} determination

No prominent sign of toxicity and mortality was recorded among experimental animals at all administered dose levels of the crude hydroalcoholic extract of *E.*



schimperi. Since no death was recorded at the maximum administered dose, the LD₅₀ of the plant was found to be higher than 5000 mg/kg.

In vitro larval mortality assay

The crude hydroalcoholic extract of *E. schimperi* showed larvicidal activity against the infective stage (L₃) of hookworms. The best-fit LC₅₀ values for the crude extract and the standard drug, albendazole are indicated in Table 1. The LC₅₀ value of the crude extract and albendazole were 228.7 and 51.33 µg/mL, respectively.

In vivo anticestodal activity

The result of *in vivo* anticestodal activity of the crude hydroalcoholic extract of *E. schimperi* and the diammonium salt of embelin is presented in Tables 2 and 3, respectively. The crude extract significantly ($p < 0.05$) reduced the worm count at necropsy as compared with the negative control. The percent deparasitization and percent parasite clearance of mice treated with 1000 mg/kg were both 100 %. The mice in the negative control group had mean parasite count of 2.00 ± 0.32 with no single mouse devoid of worms at necropsy.

Mice treated with 250, 500 and 750 mg/kg of diammonium salt of embelin had average worm count of 1.43 ± 0.30 , 1.00 ± 0.69 , 0.71 ± 0.36 , respectively, at necropsy. While the mean worm count for the negative control group was 4.86 ± 1.18 . The highest percent deparasitization

and host parasite clearance, 85.3 % and 57.14 %, respectively was recorded in mice treated with the highest dose, 750 mg/kg.

Discussion

In the present study single administration of the crude extract of *E. schimperi* up to a dose of 5,000 mg/kg to mice showed no behavioral changes like hypoactivity, prostration and piloerection. The following day after the treatment, all mice were as active as the controls and no death was observed in the treated animals. The LD₅₀, being greater than 5000 mg/kg, is thought to be safe as suggested by Lork (1983) and Schorderet (1992) [9, 13]. Again, the absence of death among mice in all the dose groups throughout the two weeks of the experimental follow up seems to support this claim. Thus, the crude extract of *E. schimperi* can be considered as a substance with low toxicity.

The crude hydroalcoholic extract of the fruits of *E. schimperi* and the diammonium salt of the isolated embelin exhibited significant anthelmintic activities against the dwarf tapeworm, *H. nana* and hookworm, *N. americanus*, *in vivo* and *in vitro*, respectively. Our finding is in agreement with various authors who

Table 1 LC₅₀ of the crude extract and albendazole using global sigmoidal model curve fitting

	Log LC ₅₀		LC ₅₀ (µg/mL)		R ²
	Best-fit	Std. error	Best-fit	95 % CI	
Crude extract of <i>E. schimperi</i>	2.359	0.0328	228.7	194.2 to 269	0.9402
Albendazole	1.710	0.0343	51.33	42.78 to 61.59	0.9583

Table 2 Worm count, percent deparasitization and host clearance after treatment with crude extract of *E. schimperi*

Groups	Dose administered (mg/kg)	Mean no of parasites	% deparasitization	% host clearance
Group I	750	1.2 ± 0.49^a	40	40
Group II	1000	$0.0 \pm 0.0^{*b}$	100	100
Group III	2000	$0.0 \pm 0.0^{*b}$	100	100
Group IV	25 (PQ)	$0.0 \pm 0.0^{*b}$	100	100
Group V	vehicle	2.00 ± 0.32	0	0

PQ: Praziquantel

* Mean number of parasite within row is significantly different at $p < 0.05$, while the letters ^a ^b show significant difference at $p < 0.01$ as determined by LSD multiple comparison test

Table 3 Worm count, percent deparasitization and host clearance after treatment with diammonium salt of Embelin

Groups	Dose administered (mg/kg)	Mean no of parasites	% deparasitization	% host clearance
Group I	250	1.43 ± 0.30*	70.6	0
Group II	500	1.00 ± 0.69*	79.4	57.14
Group III	750	0.71 ± 0.36*	85.3	57.14
Group IV	25 (PQ)	0.00 ± 0.00*	100	100
Group V	vehicle	4.86 ± 1.18	0	0

PQ: Praziquantel

* Mean number of parasites within row is significantly different at $p < 0.05$ determined by LSD multiple comparison test

reported that many plant species in family Myrsinaceae showed substantial anthelmintic properties. For instance, Choudhary [14] reported that the ethanolic extract of the seeds of *Embelia ribes* showed significant anthelmintic efficacy against the roundworm *Rhabditis pseudoelongata* (strain L. Lamy) *in vitro* when compared to the standard drugs, levamisole and ivermectin. Furthermore, Jalalpure *et al.* [15] evaluated the efficacy of the seed oil of *E. ribes* against *Pheritima posthuma* to determine the time of paralysis and time of death and found out that the seed oil was found to be more potent in both parameters than the standard drug reference, piperazine citrate. Two plant species in family Myrsinaceae, *Myrsine africana* and *Maesa lanceolata*, have been evaluated for their efficacy against *Haemonchus contortus* in Ethiopia. *In vitro* ovicidal and larvicidal activity studies of the two plants revealed that significant egg hatching inhibition and larvicidal effects were exhibited by different extracts of the plants [16, 17].

Embelin was isolated from the fruits of *E. schimperi* and its identity was confirmed by recording IR, MS and NMR spectra and by comparison of the spectral data with those reported by other authors [18, 19]. The diammonium salt of embelin showed significant anthelmintic activity *in vivo* against the flat worm *H. nana* with 85.3 and 57.14% deparasitization and host clearance, respectively. Similar finding have been reported regarding the anthelmintic properties of embelin. Gupta *et al.* [20] indicated that embelin had an anthelmintic activity *in vitro*. Mojumder and Mishra [21] reported the nematocidal activities of embelin and its two haloderivatives against root-knot nematode, *Meloidogyne incognita*. Embelin was efficacious at 48 and 72 h of exposure, and the haloderivatives were nematocidal even at 24 h post exposure. Bøgh *et al.* [22] evaluated the anthelmintic efficacy of diammonium salt of embelin *in vivo* and *in vitro* against the cestode parasites (*H. diminuta* and *H. microstoma*), the trematode *Echinostoma caproni* and the nematode *Heligmosomoides polygyrus*. It significantly reduced the parasite number and total worm biomass of *H. diminuta* in rats treated with 100 mg/kg. *In vitro* studies also revealed that all adult *H. diminuta*

were killed when incubated in culture medium with 0.08 mg/ml of diammonium salt of embelin. However, the *in vivo* studies on the other test parasites had no significant effect though the worms were inhibited *in vitro* suggesting that embelin might be more effective against cestodes than other intestinal parasites.

Conclusion

In this study the possible anthelmintic efficacy of the traditionally claimed medicinal plant, *E. schimperi*, was investigated. The results clearly indicated that both the crude extract and the diammonium salt of the isolated embelin were effective anthelmintics against the dwarf tapeworm, *H. nana*, on mice at higher doses. Hence the findings of this study showed *E. schimperi* appear to possess some anthelmintic activity that may support the usage of this plant by local traditional healers to treat helminthic infestations. However, these higher doses might not be directly extrapolated to human use for health related issues and therefore further studies to optimize the dose for human use need to be conducted. Additional studies also need to be conducted on the mechanism of action, long term toxicity, and efficacy on other experimental models including human and standardization before it can be recommended for human use.

Abbreviations

AIDS: Acquired immuno deficiency syndrome; ANOVA: Analysis of variance; DMSO: Dimethyl sulfoxide; EPHI: Ethiopian public health institute; FMOH: Federal ministry of health; HIV: Human immunodeficiency virus; HPLC: High performance liquid chromatography; LSD: Least significant difference; NMR: Nuclear magnetic resonance; OECD: Organization for economic co-operation and development; TLC: Thin layer chromatography; TMMRD: Traditional and modern medicine research directorate; UV: Ultraviolet.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

KU, DA, WM designed the study. MT, WM, YD, ADH, SW contributed to the study protocol. DA collected, identified and authenticated the plant. YB, ADH, BT, IN studied the chemistry. YD, MT, SW, AA, TM, BB, KY, HB conducted the *in vivo* and *in vitro* efficacy studies. AD, ADH, SW, MT conducted the acute toxicity study. YD coordinated the overall activities of the study, analyzed the data and drafted the manuscript. All authors critically revised the manuscript and approved the final manuscript.

Acknowledgement

The authors would like to thank the Ethiopian Public Health Institute and the Ministry of Finance and Economic Development of Ethiopia for financial support (Project No. 342/02/04/01/006). The effort of the laboratory technician and animal attendant of the Traditional and Modern Medicine Research Directorate is highly appreciated. The authors also would like to thank Mr. Eric Saunders for editing the final draft of the manuscript.

Author details

¹Ethiopian Public Health Institute, Biomedical and Clinical Research Team, Traditional and Modern Medicine Research Directorate, P.O.Box 1242, Addis Ababa, Ethiopia. ²Ethiopian Public Health Institute, National Polio Laboratory, P.O.Box 1242, Addis Ababa, Ethiopia. ³Ethiopian Public Health Institute, Natural products Research Team, Traditional and Modern Medicine Research Directorate, P.O.Box 1242, Addis Ababa, Ethiopia. ⁴College of Health Sciences, Addis Ababa University, P.O.Box 9086, Addis Ababa, Ethiopia.

Received: 19 December 2014 Accepted: 8 June 2015

Published online: 18 June 2015

References

- World Health Organization (WHO). Control of tropical diseases. Geneva: WHO; 1998.
- Ministry of Health (MoH), MoH. Comprehensive health service directory. Addis Ababa: MoH; 1996.
- Haileyesus A. The prevalence of intestinal parasites and molecular characterization of *Cryptosporidium* species in Ethiopia. PhD thesis. Addis Ababa University, Department of Biology; 2010.
- Giday M, Asfaw Z, Woldu Z. Medicinal plants of the Meinit ethnic group of Ethiopia: an ethnobotanical study. *J Ethnopharmacol.* 2009;124:513–21.
- Wabe N, Mohammed M, Raju N. An ethnobotanical survey of medicinal plants in the Southeast Ethiopia used in traditional medicine. *Spat DD-Peer Rev J Complement Med Drug Discov.* 2011;1:153–9.
- Scantlebury CE, Peachey L, Hodgkinson J, Matthews JB, Trawford A, Mulugeta G, et al. Participatory study of medicinal plants used in the control of gastrointestinal parasites in donkeys in Eastern Shewa and Arsi zones of Oromia region, Ethiopia. *BMC Vet Res.* 2013;9:179.
- d'Avigdor E, Wohlmut H, Asfaw Z, Awas T. The current status of knowledge of herbal medicine and medicinal plants in Fiche, Ethiopia. *J Ethnobiol Ethnomed.* 2014;10:38.
- Belete Y, Debebe Y, Abebe A, Menberu T, Debella A. Quantitative determination and optimization of extraction conditions for embelin in *Embelia schimperi* by UV–vis spectrometry. *J Drug Deliv Ther.* 2014;4(4):10–3.
- Lorke D. A new approach to practical acute toxicity testing. *Arch Toxicol.* 1983;54:275–87.
- Harada U, Mori O. A new method for culturing hookworm. *Yonago Acta Med.* 1955;1:177–9.
- Gill JH, Redwin JM, van Wyk JA, Lacey E. Detection of resistance to ivermectin in *Haemonchus contortus*. *Int J Parasitol.* 1991;21:771–6.
- Tangpu TV, Yadav AK. Anticestodal efficacy of *Psidiumguajava* against experimental *Hymenolepis diminuta* infection in rats. *Indian J Pharmacol.* 2006;28:29–32.
- Schorderet M. Pharmacologie des concepts fondamentaux aux applications thérapeutiques. Edition Frison-Roche Paris: Editions Slatkine Genève; 1992.
- Choudhary GP. Anthelmintic activity of fruits of *Embelia ribes* Burm. *Int J Pharm ChemSci.* 2012;1:1336–7.
- Jalalpure SS, Alagawadi KR, Mahajanashetti CS, Shah BN, Salahuddin, Singh V, et al. *In vitro* anthelmintic property of various seed oils against *Pheritima posthuma*. *Indian J Pharm Sci.* 2007;69:158–60.
- Getachew S, Ibrahim N, Abebe B, Eguale T. *In vitro* evaluation of Anthelmintic activities of crude extracts of selected medicinal plants against *Haemonchus contortus* in AlemgenaWereda, Ethiopia. *Acta ParasitologicaGlobalis.* 2012;3:20–7.
- Tadesse D, Eguale T, Giday M, Mussa A. Ovicidal and larvicidal activity of extracts of *Maesa lanceolata* and *Plectranthus punctatus* against *Haemonchus contortus*. *J Ethnopharmacol.* 2009;122:240–4.
- Haq K, Ali M, Siddiqui AW. New compounds from the seeds of *Embelia ribes* Burm. *Pharmazie.* 2005;60:69–71.
- Swamy HMK, Krishna V, Shankarmurthy K, Abdul Rahiman B, Mankani KL, Mahadevan KM, et al. Wound healing activity of embelin isolated from the ethanol extract of leaves of *Embelia ribes* Burm. *Journal of Ethnopharmacol.* 2007;109:529–34.
- Gupta P, Anand KK, Ghatak BJ, Atal CK, Ali M. *In vitro* anthelmintic activity of disalts of embelin. *Indian J ExpBiol.* 1976;14:356–7.
- Mojumder V, Mishra SD. *In vitro* studies on toxicity of embelin and its haloderivatives to *Meloidogyne incognita* juveniles. *Curr Nematol.* 1990;1:47–8.
- Bogh HO, Andreassen J, Lemmich J. Anthelmintic usage of extracts of *Embelia schimperi* from Tanzania. *J Ethnopharmacol.* 1996;50:35–42.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

