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The Cause of Follicular Spicules in Multiple Myeloma

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To The Editor

Hyperkeratotic spicules may develop as a paraneoplastic syndrome of multiple myeloma. These follicular spicules in multiple myeloma (FSMM) are composed of precipitates of monoclonal dysproteins identical to the serum protein found in these patients¹. Light microscopy of the affected follicular epithelium shows intercellular deposits of eosinophilic material that has been identified as IgG by direct immunofluorescence¹. Trichodysplasia spinulosa (TS) is a distinct disorder that also presents with follicular spicules in the setting of immunosuppression². There is increasing evidence that the TS polyomavirus (TSPyV) is involved in the pathogenesis of TS³.

Based on the clinical similarity between FSMM and TS, van Boheemen and colleagues assessed whether FSMM had a viral cause⁴. After the findings of TSPyV-specific polymerase chain reaction (PCR) and rolling circle amplification proved negative, a sensitive deep-sequencing approach identified sequences from Merkel Cell Polyomavirus (MCPyV); PCR confirmed that the virus was present at low copy numbers (about 0.1 copies/cell). Based on these findings, the researchers concluded that MCPyV might have a role in the disease. However, MCPyV can frequently be detected in skin swabs from healthy individuals. Thus, the detection of MCPyV at low copy numbers in this case of FSMM could be the result of its presence as a part of the normal skin flora rather than as a specific driver of disease.

In contrast to the low copy numbers of MCPyV detected by van Boheemen et al⁴ in their FSMM case, TSPyV is found at an average of about 10⁶ copies/cell in cases of TS³. Moreover, while ultrastructural analyses in cases of FSMM have consistently failed to identify viral particles in affected tissue, the overwhelming majority of cases of TS in which electron microscopy has been attempted do show evidence of viral particles².

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Van Boheemen et al⁴ also argue that the responsiveness of the eruption to cidofovir gel supports a viral role for FSMM. However, the concurrent treatment of the patient's multiple myeloma with systemic agents mitigates any conclusions regarding cidofovir's direct effects.

Finally, in a distinct case of FSMM⁵, our research group was unable to identify the presence of TSPyV or MCPyV using multiple primer sets. Thus, the majority of existing literature supports the follicular accumulation of an immunoglobulin dysprotein rather than a virus as the cause of FSMM. In the absence of compelling evidence for a virus in FSMM, patients with FSMM should be treated systemically for their multiple myeloma and topically with keratolytics.

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References

1. Bork K, Bockers M, Pfeifle J. Pathogenesis of paraneoplastic follicular hyperkeratotic spicules in multiple myeloma. Follicular and epidermal accumulation of IgG dysprotein and cryoglobulin. *Arch Dermatol.* 1990 Apr; 126(4):509–513. [PubMed: 2108615]
2. Matthews MR, Wang RC, Reddick RL, Saldivar VA, Browning JC. Viral-associated trichodysplasia spinulosa: a case with electron microscopic and molecular detection of the trichodysplasia spinulosa-associated human polyomavirus. *J Cutan Pathol.* 2011 Jan 19.
3. Kazem S, van der Meijden E, Kooijman S, et al. Trichodysplasia spinulosa is characterized by active polyomavirus infection. *Journal of clinical virology : the official publication of the Pan American Society for Clinical Virology.* 2012 Mar; 53(3):225–230. [PubMed: 22196870]
4. van Boheemen S, Jones T, Muhlemann B, Feltkamp MC, Fouchier RA, Hajdarbegovic E. Cidofovir Gel as Treatment of Follicular Spicules in Multiple Myeloma. *JAMA dermatology.* 2014 Sep 3.
5. Weibel L, Berger M, Regenass S, Kamarashev J, Hafner J, French LE. Follicular spicules of the nose and ears--quiz case. *Arch Dermatol.* 2009 Apr; 145(4):479–484. [PubMed: 19380676]