

RESEARCH ARTICLE

Open Access



Water extract of brewers' rice induces apoptosis in human colorectal cancer cells via activation of caspase-3 and caspase-8 and downregulates the Wnt/ β -catenin downstream signaling pathway in brewers' rice-treated rats with azoxymethane-induced colon carcinogenesis

Bee Ling Tan¹, Mohd Esa Norhaizan^{1,2,3*}, Ky Huynh⁴, Sulaiman Rahman Heshu⁵, Swee Keong Yeap⁴, Hamzah Hazilawati⁵ and Karim Roselina⁶

Abstract

Background: Brewers' rice, is locally known as *temukut*, is a mixture of broken rice, rice bran, and rice germ. The current study is an extension of our previous work, which demonstrated that water extract of brewers' rice (WBR) induced apoptosis in human colorectal cancer (HT-29) cells. We also identified that brewers' rice was effective in reducing the tumor incidence and multiplicity in azoxymethane (AOM)-injected colon cancer rats. Our present study was designed to identify whether WBR confers an inhibitory effect via the regulation of upstream components in the Wnt signaling pathway in HT-29 cells. To further determine whether the *in vitro* mechanisms of action observed in the HT-29 cells inhibit the downstream signaling target of the Wnt/ β -catenin pathway, we evaluated the mechanistic action of brewers' rice in regulating the expressions and key protein markers during colon carcinogenesis in male Sprague–Dawley rats.

Methods: The mRNA levels of several upstream-related genes in the Wnt signaling pathway in HT-29 cells treated with WBR were determined by quantitative real-time PCR analyses. Caspase-3 and -8 were evaluated using a colorimetric assay. Male Sprague–Dawley rats were administered two intraperitoneal injections of AOM in saline (15 mg/kg body weight) over a two-week period and received with 10, 20, and 40 % (w/w) brewers' rice. The expressions and protein levels of cyclin D1 and c-myc were evaluated by immunohistochemical staining and western blotting, respectively.

Results: The overall analyses revealed that the treatment of HT-29 cells with WBR inhibited Wnt signaling activity through upregulation of the casein kinase 1 (*CK1*) and adenomatous polyposis coli (*APC*) mRNA levels. We discovered that the treatment of HT-29 cells with WBR resulted in the induction of apoptosis by the significant activation of caspase-3 and -8 activities compared with the control ($P < 0.05$). *In vivo* analyses indicated that brewers' rice diminished the β -catenin, cyclin D1, and c-myc protein levels.

(Continued on next page)

* Correspondence: nhaizan@upm.edu.my

¹Department of Nutrition and Dietetics, Faculty of Medicine and Health Sciences, Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia

²Research Centre of Excellent, Nutrition and Non-Communicable Diseases (NNCD), Faculty of Medicine and Health Sciences, Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia

Full list of author information is available at the end of the article

(Continued from previous page)

Conclusions: We provide evidence that brewers' rice can induce apoptosis and inhibit the proliferation of HT-29 cells through regulation of caspase-dependent pathways and inhibit the Wnt/ β -catenin downstream signaling pathway *in vivo*. We suggest that brewers' rice may be a useful dietary agent for colorectal cancer.

Keywords: Brewers' rice, Colorectal cancer, Wnt signaling, Cyclin D1, c-myc

Background

Colorectal cancer has become a global health problem, represents the third most common cancer after lung and breast cancers and contributes for nearly 10 % of the total cases of cancer and approximately 8 % of the total cancer deaths [1]. Chemoprevention using natural products is considered a practical strategy for reducing the ever-increasing cancer incidence [2]. The intervention of multistage carcinogenesis through the modulation of intracellular signaling pathways may provide the molecular basis of chemoprevention.

Emerging evidence reveals that Wnt/ β -catenin signaling plays critical roles in human biology [3]. Inappropriate stimulation of Wnt pathway is linked with many cancers [4]. The amount of cytoplasmic β -catenin is maintained at a low amount via ubiquitin-proteasome-mediated phosphorylation and is regulated by a destruction complex contained of AXIN1, adenomatous polyposis coli (APC), casein kinase 1 (CK1), and glycogen synthase kinase 3 β (GSK3 β) [5]. Cyclin D1 is a Wnt target gene [6], and mutations of this signaling pathway contribute to nearly 90 % of colorectal cancer [7]. Gene mutation in the Wnt pathway, including inactivating mutations of the APC gene and stimulating mutations in β -catenin, lead to the nuclear accumulation of β -catenin and subsequently result in the formation of a complex with T-cell factor (TCF)/lymphoid enhancing factor (LEF) to stimulate gene transcription [8]. The TCF/LEF binding sites on promoters of cell proliferation genes, including cyclin D1 and c-myc [6], result in the stimulation of aberrant mutations in the tumorigenic signals of the colonic crypts.

Rice is one of the main staple foods in approximately half of the world's population [9]. The milling of paddy rice results in an approximately 70 % yield of rice as its major product, with some of the unconsumed portions of the rice produced include rice husk (20 %), rice bran (8 %), and rice germ (2 %) [10, 11]. Brewers' rice, which is known locally as *temukut*, is typically used as an animal feed and brewing material [12]. Several studies as reported by Esa et al. [13] have demonstrated that rice by-products can inhibit human colon cancer cell

proliferation and reduce the AOM-induced colon tumors in rats. Therefore, the purpose of this study was to investigate whether water extract of brewers' rice (WBR) confers an inhibitory effect via the regulation of upstream components in the Wnt signaling pathway in colorectal cancer (HT-29) cells. To further determine whether the *in vitro* mechanisms of action observed in HT-29 cells inhibit the downstream signaling target of the Wnt/ β -catenin pathway, we evaluated the mechanistic action of brewers' rice in regulating the expressions and key protein markers during colon tumorigenesis in male Sprague–Dawley rats.

We observed that the treatment of HT-29 cells with WBR inhibited Wnt signaling activity through upregulation of the *CK1* and *APC* mRNA levels. In addition, the treatment of HT-29 cells with WBR caused in the induction of apoptosis by the significant activation of caspase-3 and -8 activities compared with the control ($P < 0.05$). *In vivo* analyses also showed that brewers' rice diminished the β -catenin, cyclin D1, and c-myc protein levels.

Materials and methods

Chemicals and reagents

Azoxymethane (AOM), 10 % (v/v) neutral buffered formalin, TRI Reagent[®], and specific primers were bought from Sigma (St. Louis, MO, USA). Dulbecco's Modified Eagle Medium (DMEM), RPMI-1640 medium, Mycoflex[™] fetal bovine serum (FBS), penicillin and streptomycin (100 \times), and trypsin EDTA (1 \times) were obtained from PAA Laboratories GmbH (Pasching, Austria). Colorimetric assay kit for caspase was bought from GenScript Corporation Inc (Piscataway, NJ, USA). High capacity RNA-to-cDNA Kit and SYBR[®] Select Master Mix (CFX) were purchased from Applied Biosystems (Foster City, CA, USA). Western blotting reagents were purchased from Bio-Rad (Hercules, CA, USA). All other chemicals and reagents used were of analytical grade and bought from Sigma-Aldrich (St. Louis, MO, USA).

Stabilization of brewers' rice

Freshly milled brewers' rice samples (MR 219) were obtained from BERNAS Milling Plant (Seri Tiram Jaya,

Selangor, Malaysia). Stabilized samples were carried out as previously described by Tan et al. [14]. Brewers' rice consists of broken rice (95.16 ± 4.62 %), rice bran (3.60 ± 0.39 %), and rice germ (1.11 ± 0.07 %), while WBR is the extract of brewers' rice. WBR contains the bioactive compounds from these three major compositions in brewers' rice (broken rice, rice bran, and rice germ).

Preparation of water extract of brewers' rice

Stabilized brewers' rice was extracted with water following Yu et al. [15].

Cell culture and treatment

The human colorectal cancer (HT-29) cells were bought from American Type Culture Collection (ATCC; Rockville, MD, USA), and the cells were cultured in DMEM supplemented with 10 % (v/v) FBS, 100 IU/mL penicillin, and 100 μ g/mL streptomycin. HT-29 cells were maintained and incubated at 37 °C in a humidified atmosphere with 5 % CO₂ atmosphere. The cells were treated with three different concentrations of WBR (16, 32, and 64 μ g/mL) for 72 h.

Gene expression

The TRI Reagent[®] was used to isolate the total RNA, according to the manufacturer's protocol. Two micrograms of RNA per 20 μ L was reverse-transcribed using the High-Capacity RNA-to-cDNA Kit according to the manufacturer's protocol. The reverse-transcription reaction was performed using an Authorized Thermal Cycler (Eppendorf, NY, USA). The cDNA was then ready for use as a template for amplifications through real-time PCR reactions. Quantitative real-time PCR was carried out using SYBR[®] Select Master Mix (CFX). The nucleotide primer sequences originating from human cell lines are shown in Table 1. All of the samples and controls were determined in triplicate using the BioRAD-iQ[™] 5 Multicolor Real-Time PCR Detection System (Hercules, CA, USA). The quantitative real-time PCR data were analyzed using the CFX Manager[™] software (version 1.6, Bio-Rad, Hercules, CA, USA). Beta-actin (*ACTB*), glyceraldehyde-3-phosphate dehydrogenase (*GAPDH*), and 18S rRNA were used as housekeeping genes to normalize the target genes expression.

Caspase-3 and caspase-8 assay

The caspase-3 and -8 activities were assessed spectrophotometrically using a commercial colorimetric assay kit, based on spectrophotometric detection of the chromophore *p*nitroanilide (*p*NA) after cleavage from the specific substrates DEVD-*p*NA (for caspase-3) and IETD-*p*NA (for caspase-8). Initially, HT-29 cells were treated with three different concentrations (16, 32, and 64 μ g/mL) of WBR for 72 h. The cells were centrifuged

Table 1 Nucleotide sequence of PCR primers for amplification and sequence-specific detection of cDNA (obtained from GenBank database)

Primer name [accession number]	Oligonucleotides (5'-3') Sequence
<i>AXIN1</i> [XM_005255610.2]	F- TTTACCGAAGATGCTCCCC R- CACTGCCCTCAGGCTCATA
<i>CK1</i> [NM_001271741.1]	F- GAGATCCCTTTCCAGAGTGC R- TTTGTGAAGGGCTTCTCGGC
<i>GSK3β</i> [BC012760]	F- CGAATGGGGAACAGTCGAGG R- TCGGAAATGCGACGGGAAAC
<i>APC</i> [NM_000038.5]	F- AGCAAGTTGAGGCACTGAAGA R- TCCCGGCTTCCATAAGAACG
<i>LRP6</i> [NM_002336.2]	F- ACATGACAGGTCGAGAGGGT R- CCAAGCCACAGGGATACAGT
<i>ACTB</i> ^a [NM_001101.3]	F- AGAGTACGAGCTGCCTGAC R- AGCACTGTGTTGGCGTACAG
<i>GAPDH</i> ^a [NM_002046.4]	F- GGATTTGGTCTGATTGGGC R- TGGAAGATGGTGATGGGATT
18S rRNA ^a [HQ387008.1]	F- GTAACCCGTTGAACCCATT R- CCATCCAATCGGTAGTAGCG

ACTB- beta-actin, *APC*- adenomatous polyposis coli, *CK1*- casein kinase 1, *GAPDH*- glyceraldehyde-3-phosphate dehydrogenase, *GSK3 β* - glycogen synthase kinase 3 β , *LRP6*- low density lipoprotein receptor-related protein 6
^aHousekeeping gene

at 370 \times g for 5 min to remove the medium and washed twice with phosphate-buffered saline (PBS) (pH 7.4). The cell pellets were then lysed with 50 μ L of cold lysis buffer, followed by 1 h incubation period on ice. The resulting cell lysates were centrifuged at 9300 \times g and 4 °C for 1 min, and the supernatants were collected. The protein concentration was quantified using a Bradford protein assay kit, according to the manufacturer's protocols. Fifty μ L of 2 \times reaction buffers were mixed with 50 μ L supernatant containing 200 μ g of proteins in each tube, followed by 5 μ L of caspase-3 or caspase-8 substrate. The suspension was collected and transferred to a 96-well microtiter plate and incubated at 37 °C for 4 h before measured at 405 nm using ELISA microplate reader.

Animals, diet, and *in vivo* experimental procedures

The animal use protocol was approved by the Animal Care and Use Committee (ACUC) of the Faculty of Medicine and Health Sciences, Universiti Putra Malaysia (UPM) Serdang, Selangor (Reference number: UPM/FPSK/PADS/BR-UUH/00461). A total of 30 four-week-old male Sprague-Dawley rats were housed in plastic cages (two rats per cage) and maintained in an animal facility at a controlled temperature of approximately 25–27 °C and a relative humidity of 50 \pm 10 %

with 12-hour light/dark cycles. Hygienic condition was maintained by weekly changes of the woodchip beds. The animals were acclimatized for seven days prior to receiving the following defined diets for twenty weeks ($n=6$ rats for each group): (G1), normal; (G2), AOM alone; (G3), AOM + 10 % (w/w) brewers' rice; (G4), AOM + 20 % (w/w) brewers' rice; and (G5), AOM + 40 % (w/w) brewers' rice. During the acclimatization period, the animals were fed an American Institute of Nutrition (AIN)-93G diet and tap water *ad libitum*. Beginning at six weeks of age, the rats in G2-G5 were received intraperitoneal injections of azoxymethane (AOM) at a dose of 15 mg/kg body weight once weekly over a 2-week period. The rats in the normal group (G1) received an equal volume of normal saline (vehicle control). The animals were administered fresh food and tap water every day. The experimental diets were prepared weekly and kept at 4 °C. The components of an AIN-93G diet (Table 2) were adjusted based on the nutrient composition of brewers' rice as previously described by Tan et al. [16].

Tissue sample collection

The animals were sacrificed after 20 weeks of treatment by anesthesia with diethyl ether. The whole colon tissues were removed, rinsed with PBS, opened and cut longitudinally, and fixed in 10 % (v/v) neutral buffered formalin. The colon tissue was processed for further analyses.

Table 2 Composition of experimental diets

Ingredients (g/1000 g diet)	Group				
	G1	G2	G3	G4	G5
Brewers' rice	-	-	100.0	200.0	400.0
Corn starch	397.5	397.5	315.3	233.2	68.9
Casein	200.0	200.0	191.0	182.0	164.0
Maltodextrin	132.0	132.0	132.0	132.0	132.0
Sucrose	100.0	100.0	100.0	100.0	100.0
Soybean oil	70.0	70.0	68.1	66.1	62.2
Powdered cellulose	50.0	50.0	44.7	39.4	28.7
AIN-93G mineral mix	35.0	35.0	33.4	31.9	28.8
AIN-93G vitamin mix	10.0	10.0	10.0	10.0	10.0
L-cystine	3.0	3.0	3.0	3.0	3.0
Choline bitartrate	2.5	2.5	2.5	2.5	2.5
tert-butylhydroquinone	0.014	0.014	0.014	0.014	0.014

G1 and G2, AIN-93G diet; G3, AIN-93G diet containing 10 % (w/w) of brewers' rice; G4, AIN-93G diet containing 20 % (w/w) of brewers' rice; G5, AIN-93G diet containing 40 % (w/w) of brewers' rice

Immunohistochemical staining of cyclin D1 and c-myc antigen

The colonic tissue sections were dehydrated, embedded in paraffin, and sectioned at 4-6- μ m thickness. The detailed analyses for immunohistochemical evaluation were described previously [16]. The primary antibodies against cyclin D1 (diluted 1:100) and c-myc (diluted 1:50) (Abcam, UK) were incubated with the sections. The whole colon was divided into three sections, including distal, middle, and proximal colon. The scoring was done in three sections for each rat for both normal colon mucosa and the lesion (tumor). Cyclin D1 and c-myc localization were evaluated, recording whether the staining was localized at the cytoplasm (the cell was classified as positive at the cytoplasm) or at the nucleus (positive at the nucleus). A semi-quantitative scoring system was performed following Kohno et al. [17] to examine the antibody staining against cyclin D1 and c-myc. The total scoring was evaluated after summation of the extent and intensity of the staining (score = extent + intensity) in seven randomly selected fields from each section of the colon of each rat stained with the respective antibody under 100 \times magnification. Quantification of cyclin D1 and c-myc staining that represents the extent (number of stained (positive) cells) and the intensity of the staining across 21 fields for each rat. The percentages of positive cells were evaluated using the following scale: 0 = no stained cells in any field; 1 = positive staining in 1 to 25 % of the cells; 2 = positive staining in 26 to 50 % of the cells; 3 = positive staining in 51 to 75 % of the cells; and 4 = positive staining in 76 to 100 % of the cells. The strength of the staining intensity was examined using the following range: 0, no cell staining; 1+, mild staining; 2+, moderate staining; and 3+, strong staining. Hence, after summation, the maximum score was 7, and the minimum score was 0.

Western blotting analysis

The proteins from rat colon tissue (100 mg) were extracted with 300 μ L of radio immune precipitation assay (RIPA) lysis buffer and 3 μ L of protease inhibitor cocktail. The colon tissue was homogenized for 30 s prior to incubation at 4 °C for 1 h with agitation. The protein concentration was quantified using a Bradford protein assay kit according to the manufacturer's protocol using bovine serum albumin as the standard. The protein suspensions were aliquoted into PCR tubes and stored at -80 °C prior to sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) analysis. Equal amounts (50 μ g) of protein were separated by electrophoresis with 1 \times running buffer (0.025 M Tris base, 0.192 M glycine, and 0.1 % SDS; pH 8.3). The protein lysates and protein ladder were loaded onto gels, and the

gels were run at 75 V for 15 min and then at 120 V for 45 min. After the protein samples were run on the SDS-PAGE gel, the gel was removed from the glass plate and rinsed in 1× Towbins transfer buffer (25 mM Tris-base, 190 mM glycine, 20 % (v/v) methanol; pH 8.3) at room temperature for 10 min. Protein transfer from the SDS-PAGE gel to a PVDF membrane was achieved by running the transfer apparatus at 100 V and 400 mA for 2 h. The PVDF membrane was blocked with blocking buffer (5 % skim milk solution) at room temperature for 1 h. The membranes were washed three times with phosphate-buffered saline in 0.1 % Tween-20 (PBST) for 10 min each before incubation with specific primary antibodies in 5 % skim milk PBST. Rabbit monoclonal antibody (E247) to β -catenin, rabbit monoclonal antibody (EPR2241) to cyclin D1, and rabbit monoclonal antibody (Y69) to c-myc were used at a 1:10,000 dilution (Abcam, UK) overnight at 4 °C on a roller mixer. After overnight incubation with the primary antibodies, the membranes were washed three times with PBST for 10 min each before incubation with goat polyclonal secondary antibody to rabbit IgG conjugated to horseradish peroxidase (HRP) (Abcam, UK) at a 1:10,000 dilution at room temperature for 1 h. The protein band was developed and detected using a chemiluminescence blotting substrate kit (Clarity western ECL substrate), and a chemiluminescence imager (Bio-Rad, USA) was used to view the protein bands on the membranes. Beta-actin (Abcam, UK) was used as a loading control to confirm that the protein concentrations in all of the loaded samples were equal. The densitometric analyses of the band intensities obtained were performed using the Image J Software (National Institute of Health, NIH, Bethesda, MD, USA).

Statistical analyses

The statistical data were expressed as the mean \pm standard deviation (SD) and analyzed using a one-way analysis of variance (ANOVA). Statistical analyses were conducted following the Statistical Package for Social Science (SPSS) version 19.0 (SPSS Inc., Chicago, IL, USA). A P-value of < 0.05 was considered significant.

Results and discussion

WBR upregulates the mRNA level of *CK1* in HT-29 cells

According to published guidelines, any extract that exerts potentially cytotoxic effects should have an IC_{50} less than 100 $\mu\text{g}/\text{mL}$ [18]. We previously reported that WBR and methanol extract of brewers' rice (MBR) is cytotoxicity towards colorectal cancer (HT-29) cells with IC_{50} values of $38.33 \pm 6.51 \mu\text{g}/\text{mL}$ and $54.00 \pm 5.29 \mu\text{g}/\text{mL}$, respectively [14], as evaluated through a 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) proliferation assay, whereas a lactate dehydrogenase

(LDH) assay showed IC_{50} values of $21.88 \pm 12.43 \mu\text{g}/\text{mL}$ and $34.50 \pm 5.92 \mu\text{g}/\text{mL}$ for WBR and MBR, respectively [19]. The results showed that both WBR and MBR are sensitive to HT-29 cells, indicating that WBR and MBR can inhibit the proliferation of HT-29 cells. However, HT-29 cell line was more sensitive to WBR because the median inhibition concentration values were lower than that of MBR, as confirmed by MTT proliferation and LDH assays. The proliferation of HT-29 cells after treated with WBR is shown in Fig. 1. Therefore, we tested whether WBR can affect the Wnt signaling pathway in HT-29 cells.

To obtain further insights into the growth inhibitory effects of WBR in HT-29 cells are caused by the mechanistic action of WBR, the mRNA levels of several upstream-related genes in the Wnt signaling pathway were subsequently determined by quantitative real-time PCR analyses. In general, the Wnt/ β -catenin signaling activity depends on the amount of β -catenin in the cytoplasm, and β -catenin is targeted for degradation by a destruction complex consisting of AXIN1, APC, CK1, and GSK3 β [5]. Thus, the expression levels of *AXIN1*, *CK1*, *GSK3 β* , and *APC* in response to WBR were examined using quantitative real-time PCR. Furthermore, to understand how WBR regulates the Wnt signaling, Wnt coreceptor low-density lipoprotein receptor-related protein 6 (*LRP6*), which is essential in signal transduction and is more likely to stimulate Wnt signaling when over-expressed, was analyzed. Given the broad cytotoxicity range of WBR against HT-29 cell line determined by the MTT and LDH assays, as reported in our earlier study [14, 19], only three concentrations (16, 32, and 64 $\mu\text{g}/\text{mL}$) were selected.

The stabilization of β -catenin through inhibition of destruction complex formation in Wnt signaling has been proposed by Cselenyi et al. [20]. AXIN1 overexpression induces the phosphorylation of β -catenin in cells that express truncated *APC* [21]. It has been reported that AXIN1 expression in SW480 cells stimulates the phosphorylation of β -catenin [22]. Therefore, it is plausible that the *AXIN1* mRNA level needs to be modulated to ensure proper Wnt signaling pathway. In the present study, we found that no significant difference in *AXIN1* gene between the control cells and the cells treated with 32 $\mu\text{g}/\text{mL}$ WBR and between the control cells and the cells treated with 64 $\mu\text{g}/\text{mL}$ WBR ($P > 0.05$) (Fig. 2a). Therefore, *AXIN1* mRNA level may not play a critical role in negatively regulating the Wnt signaling pathway to induce the phosphorylation and degradation of β -catenin.

In addition to the effects observed on *AXIN1*, β -catenin is targeted for phosphorylation, in part via the regulation by *CK1 γ* . β -catenin is targeted for degradation via ubiquitin-mediated proteolysis through the

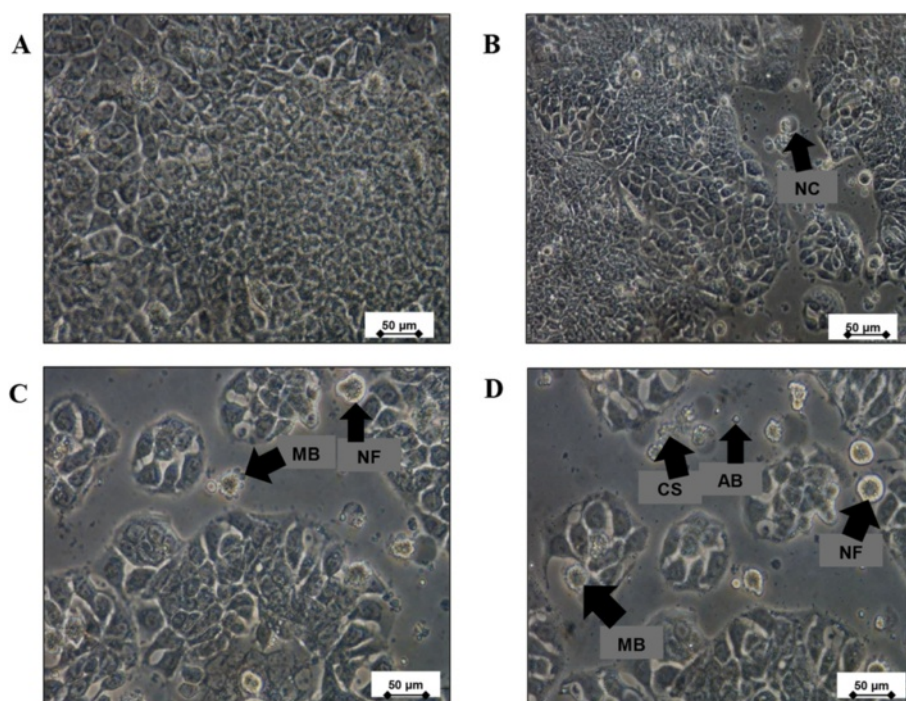


Fig. 1 Morphological changes of HT-29 cells observed under an inverted light microscope. **a** Untreated HT-29 cells, HT-29 cells treated with water extract of brewers' rice (WBR) at concentrations of **b** 16 µg/mL, **c** 32 µg/mL, and **d** 64 µg/mL. The cells showed typical characteristics of apoptosis, such as nuclear compaction (NC), apoptotic bodies (AB), membrane blebbing (MB), cellular shrinkage (CS), and nuclear fragmentation (NF) (magnification 400×). Scale bars in the figures indicate 50 µm (Source: Tan et al. [19])

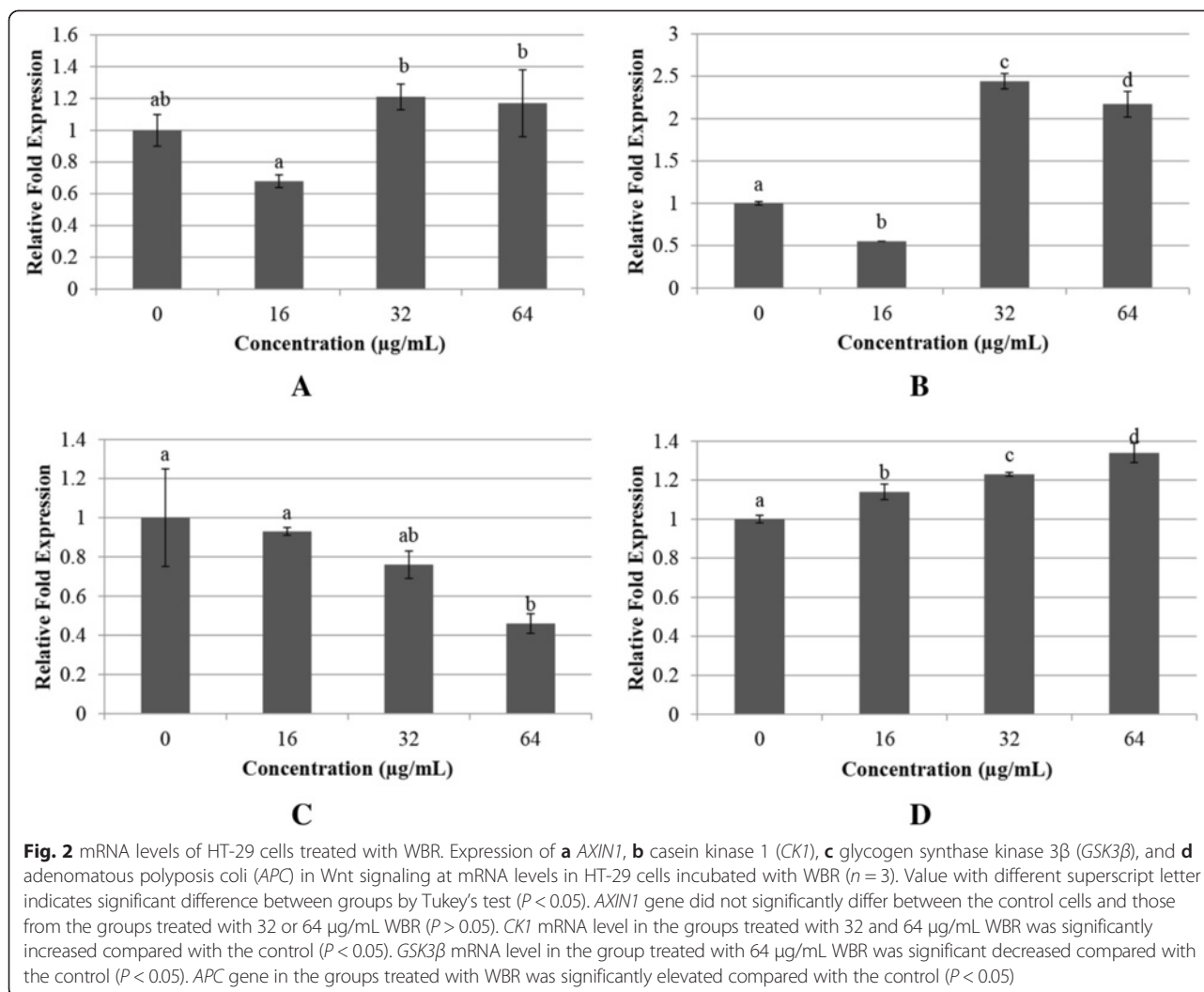
action of CK1 α [23]. As shown in Fig. 2b, the *CK1* mRNA level in HT-29 cells treated with 32 and 64 µg/mL WBR for 72 h was significantly increased compared with the control ($P < 0.05$). Thus, this finding suggested that treatment with WBR at concentrations of 32 and 64 µg/mL elevated the *CK1* mRNA level, and this increase may be associated with the induction of apoptosis, which may result to the phosphorylation of β -catenin in a Wnt pathway. Because WBR promotes the *CK1* mRNA level, the susceptibility of HT-29 cells to WBR may also be due to *GSK3 β* . A previous study found that *GSK3 β* may promote cancer development and tumorigenesis [24]. Thus, the effect of *GSK3 β* in response to WBR was analyzed in HT-29 cells.

Treatment with WBR inhibits the mRNA level of *GSK3 β* in HT-29 cells

Because *GSK3 β* modulates many substrates and signaling pathways, the mechanisms underlying its tumor suppressor or promoter activities are complex [25]. One of the critical roles of *GSK3 β* on neoplastic transformation tumor development is more likely via the modulation of Wnt/ β -catenin signaling. Active *GSK3 β* induces the phosphorylation of β -catenin via ubiquitin-mediated proteasomal degradation, and the cytoplasmic β -catenin level thus remains low.

Our data revealed that *GSK3 β* mRNA level was mainly present in untreated HT-29 cells (Fig. 2c). These findings were consistent with the results reported by Shakoori et al. [26], who found high levels of *GSK3 β* in colon cancer cells and colorectal cancer patients compared with the normal counterparts. It is recognized that tumors depend on anaerobic pathways to shift glucose to ATP in the presence of ample oxygen, and tumor cells maintain the production of ATP via elevation of glucose influx to fuel the energy requirements for uncontrolled proliferation [27]. In the control (untreated HT-29 cells) group, the high *GSK3 β* level observed in HT-29 cells may subsequently contribute to the high consumption of glucose by cancer cells via the inhibition of glycogen synthesis. Importantly, this finding indicates that high *GSK3 β* levels may contribute to colorectal cancer, which contrasts its predicted role [28] as a tumor suppressor.

However, the *GSK3 β* mRNA level in WBR-treated HT-29 cells was reduced, with a maximum reduction observed at a concentration of 64 µg/mL (Fig. 2c). These findings were further supported by Ougolkov et al. [29] and Cao et al. [30], who found that the suppression of *GSK3 β* reduces pancreatic cancer cell survival and proliferation, suppresses the growth of ovarian cancer cells *in vitro*, and inhibits the formation of tumors in nude mice. Furthermore, non-steroidal anti-inflammatory drugs



(NSAIDs), which are cancer chemopreventive agents, have also been reported to increase the phosphorylation of *GSK3 β* and thereby induce the downregulation of β -catenin/TCF signaling [31]. In contrast, no significant differences were found in *GSK3 β* between the control group and those from the groups treated with 16 $\mu\text{g/mL}$ WBR or 32 $\mu\text{g/mL}$ WBR ($P > 0.05$) (Fig. 2c). Taken together, these results suggested that treatment with WBR downregulated *GSK3 β* via mechanisms other than Wnt/ β -catenin signaling. Because we observed the upregulation of *CK1* and the downregulation of *GSK3 β* , we further investigated the mRNA level of *APC*.

Treatment with WBR upregulates the mRNA level of *APC* in HT-29 cells

APC is a large protein that binds to both β -catenin and *AXIN1*. It plays an essential role as a destruction complex in colorectal cancer [32]. As shown in Fig. 2d, the untreated cells (control) presented the lowest *APC* mRNA level compared with HT-29 cells treated with

WBR. A previous study reported that most of the colorectal cancer cases present inactivating *APC* mutations [33]. Mutations in *APC* gene are observed in 70 % of colorectal cancer cases and are regarded as an initiating event due to an inability of the destruction complex to stimulate the degradation of β -catenin [34, 35]. When the destruction complexes contained of *AXIN1*, *APC*, *CK1*, and *GSK3 β* are inhibited, subsequently resulted in the accumulation of cytoplasmic β -catenin [36] in the mutated HT-29 cell line. Our current data revealed that treatment with various concentrations (16, 32, and 64 $\mu\text{g/mL}$) of WBR increase the *APC* mRNA level in a dose-dependent manner. This finding indicated that the *APC* gene in HT-29 cells was significantly elevated after 72 h of incubation with WBR compared with the untreated HT-29 cells (control) ($P < 0.05$) (Fig. 2d).

The interaction of a Wnt ligand with transmembrane co-receptors, LRP6, suppresses the phosphorylation of the transcriptional coactivator β -catenin, which allows the translocation of β -catenin to the nucleus and

activates the Wnt target genes [20]. Thus, the expression of *LRP6* in response of HT-29 cells was investigated to determine whether WBR can modulate *LRP6* at the mRNA level.

Treatment with WBR downregulates the mRNA level of *LRP6* in HT-29 cells

The *LRP6* is a crucial co-receptor in Wnt signaling [37]. Wnt ligands stabilize cytoplasmic β -catenin through the binding of the seven-transmembrane-domain receptor frizzled (*Fz*) and *LRP5/LRP6* [37]. It has been reported that *LRP6* is more potent than *LRP5* in certain analyses; however, the qualitative differences in their mechanisms of action remain to be fully defined [38].

In addition to the destruction complex consisting of *AXIN1*, *CK1*, *GSK3 β* , and *APC*, our data revealed that treatment with various concentrations (16, 32, and 64 $\mu\text{g/mL}$) of WBR significantly reduced the mRNA level of *LRP6* in HT-29 cells in a dose-dependent manner compared to the untreated cells ($P < 0.05$) (Fig. 3). Ligand-induced receptor downregulation has been shown to play a critical role in the regulation of the propagation and duration of growth factor receptor signaling and prevents stimulation of aberrant cell [39]. Collectively, these results suggest that the treatment of HT-29 cells with WBR results in the upregulation of *CK1* and *APC* and the downregulation of *GSK3 β* and *LRP6* at the mRNA levels, which may be associated with a marked increase in apoptotic cell death, as reported in our previous study [19]. Apoptosis is another strategy used to eliminate cancerous cells [40]. It serves as a protective mechanism to destroy damaged cells prior to the manifestation of malignancy [41] without the activation of inflammatory responses [42]. To ascertain whether the growth inhibitory activity could be dependent on the

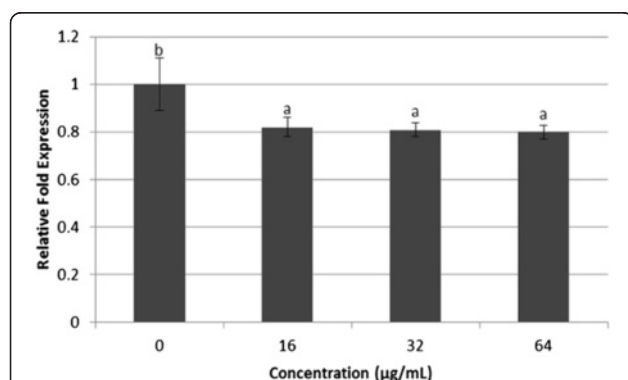


Fig. 3 mRNA levels of HT-29 cells treated with WBR. Expression of low density lipoprotein receptor-related protein 6 (*LRP6*) mRNA level in HT-29 cells incubated with water extract of brewers' rice ($n = 3$). Value with different superscript letter indicates significant difference between groups by Tukey's test ($P < 0.05$). WBR treatment significantly reduced the *LRP6* mRNA level compared with the control ($P < 0.05$)

activation of caspase-3 and caspase-8, which play vital roles in the modulation of apoptotic responses [43], the intracellular levels of caspase-3 and -8 in HT-29 cells after induction with WBR were investigated.

Treatment with WBR promotes activation of caspase-3 and -8 activities in HT-29 cells

Resistance against apoptosis is important in survival and causes a drug resistance in cancer cells. Thus, apoptosis induction is a vital approach in the cancer chemoprevention and chemotherapy [2]. To gain a better understanding of the mechanism underlying apoptosis induction in WBR-treated HT-29 cells, we evaluated the action of WBR on caspase-3 and -8 activities. An apoptotic program is executed through a family of highly conserved cysteinyl aspartate-specific proteases known as caspases that dismantle the cell by cleaving high amounts of cellular substrates. Caspase-8 is considered an initiator caspase that is responsible for stimulating the downstream effector caspase-3. Modulating the mechanism through the activation of caspases is a vital molecular target in chemoprevention because these processes contribute to apoptosis [2].

As shown in Fig. 4, WBR clearly increases the caspase-3 and -8 activities in a dose-dependent manner. The caspase-3 activity after treatment with 16, 32, and 64 $\mu\text{g/mL}$ WBR was significantly increased compared with the control ($P < 0.05$). In addition to the upregulation of caspase-3, the caspase-8 activity in HT-29 cells was also significantly increased after treatment with WBR for 72 h ($P < 0.05$). Therefore, the findings may suggest that an increase in caspase-8 activity result in the stimulation

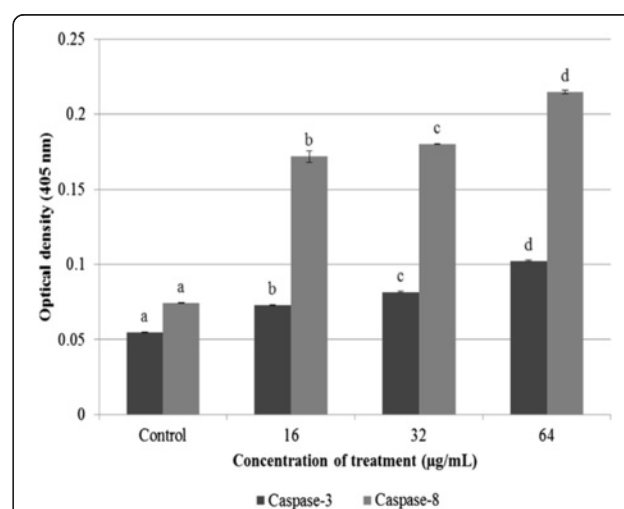


Fig. 4 Caspase-3 and -8 activities in HT-29 cells treated with WBR ($n = 3$). Value with different superscript letter indicates significant difference between groups by Tukey's test ($P < 0.05$). Both caspase-3 and -8 activities in the groups treated with WBR were significantly increased compared with the control ($P < 0.05$)

of the downstream apoptotic executioner caspase-3, which subsequently activates the molecular cascade of apoptosis in HT-29 cells. Taken together, our data indicate that the caspase-3 and -8 activities are induced by WBR in a dose-dependent manner. WBR markedly increase in apoptosis, which was accompanied by the activation of caspase-3 and -8 activities. Collectively, these results demonstrated that the activation of caspase activities in HT-29 cells after treatment with WBR increments initiator caspase-8 and executioner caspase-3. Our data presented in this study demonstrated that WBR inhibits the proliferation of colon cancer *in vitro*, as reported in our earlier study [19], leading to programmed cell death, which was confirmed to be through the regulation of caspase-dependent pathways. To further verify whether the *in vitro* mechanisms of action observed in HT-29 cells could suppress the downstream signaling target of the Wnt/ β -catenin pathway, we investigated whether brewers' rice can reduce the expressions and key protein markers during colon carcinogenesis in male Sprague–Dawley rats using immunohistochemical and western blot evaluations. Thus, the levels of the downstream signaling targets cyclin D1 and c-myc in response to brewers' rice were further evaluated in the colons of rats injected with AOM.

Brewers' rice downregulates the cyclin D1 and c-myc expression

The current study is an extension of our earlier work, which determined brewers' rice to be an effective dietary agent for the reduction of colon tumor incidence and multiplicity formation in AOM-treated rats [16]. We also identified that brewers' rice markedly reduces β -catenin expression in both the cytoplasm and the nucleus [16]. In the present study, male Sprague–Dawley rats received different doses (10, 20, and 40 % (w/w)) of brewers' rice. A dosage of a preparation of 10 % (w/w) brewers' rice was used as suggested by a previous study conducted by Boateng et al. [44] on rice bran and rice germ. This dosage has been reported to suppress tumor formation. In addition, higher concentrations (20 and 40 % (w/w) brewers' rice) were also tested to determine the dose-dependent effect of brewers' rice as a chemopreventive agent in a rat colon carcinogenesis experimental model. As shown in our previous study, feeding up to 40 % (w/w) brewers' rice is well-tolerated and did not inhibit the growth of rats [16].

Because cyclin D1 and c-myc are downstream signaling targets in Wnt signaling and the overexpression of cyclin D1 and c-myc are found in most patients with colorectal tumors, in whom lowering the expression of cyclin D1 has therapeutic significance [45], we investigated whether brewers' rice suppresses the cyclin D1 and c-myc expression in colon tumors. The positive

brownish staining of cyclin D1 and c-myc in the AOM-alone group and the brewers' rice-administered groups were mostly localized in the cytoplasm and nucleus. The colon tumors from the AOM-alone group demonstrated intense staining for cyclin D1 and c-myc compared with those from the brewers' rice-fed group. Reductions in the staining intensities for cyclin D1 and c-myc were also found in colon tumors from rats received with a diet containing brewers' rice (Figs. 5 and 6).

Immunohistochemical analysis showed that the colon mucosa of AOM treated rats, which presented the highest values of tumor formation among all of the groups [16], demonstrated the highest score from the evaluation of the immunohistochemical analysis of cyclin D1 compared with the other treatment groups (Table 3). Consistent with the high expression of cyclin D1 in colon tumors, we also found the highest score of c-myc expression in the AOM-alone group compared with the other treatment groups (Table 3). The dietary administration of brewers' rice has been demonstrated to diminish cyclin D1 expression in a dose-dependent manner. A similar trend was also observed in c-myc expression in AOM-injected rats administered brewers' rice. The heterogeneous expression of cyclin D1 and c-myc was observed in the treatment groups (G3, G4, and G5), but only two treatment groups (20 and 40 % (w/w) of brewers' rice) showed a significant reduction in cyclin D1 and c-myc expression compared with AOM alone ($P < 0.05$). However, no significant difference in both cyclin D1 and c-myc was found between the AOM alone and 10 % (w/w) brewers' rice treatments ($P > 0.05$) (Table 3). These findings showed that brewers' rice has the potential to decrease the downstream signaling targets cyclin D1 and c-myc. Overall, the data presented in this study suggest that brewers' rice may modulate colon tumor development through Wnt/ β -catenin signaling.

Treatment with brewers' rice reduces the protein level of β -catenin in colonic tumors

To investigate the inhibitory effects of brewers' rice in regulating the Wnt signaling pathway, we employed AOM-induced colonic tumors, which are known to exert the wild-type β -catenin and truncated *APC* gene [46]. We hypothesized that brewers' rice would downregulate the β -catenin, cyclin D1, and c-myc protein levels through the Wnt/ β -catenin signaling pathway. Activation of the Wnt/ β -catenin signaling pathway has been associated with colorectal tumors [47].

The protein level of β -catenin was diminished accordingly when the concentration of brewers' rice was increased, indicating that brewers' rice increased the degradation of β -catenin in a dose-dependent manner (Fig. 7a). Expectedly, we found no β -catenin protein expression in the normal colon tissue. This result may

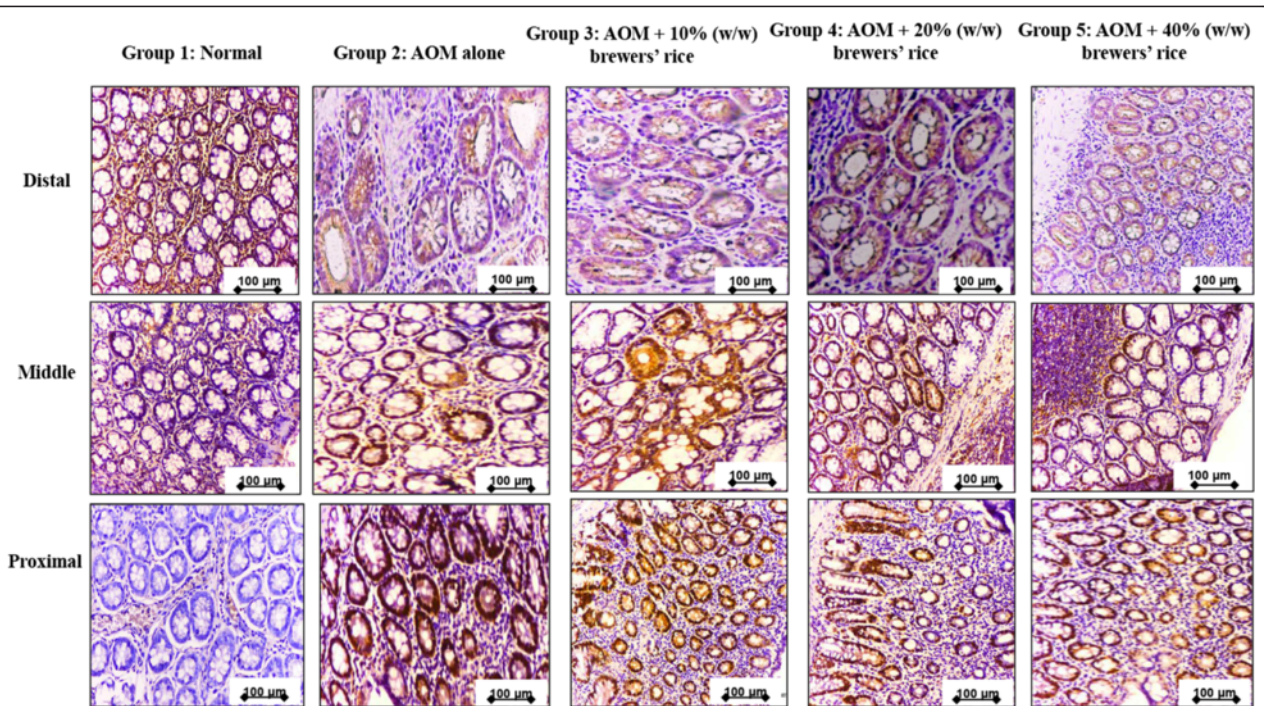


Fig. 5 Immunohistochemical staining of cyclin D1. The expression of cyclin D1 in three sections, including distal, middle, and proximal colon in normal (Group 1), AOM alone (untreated) (Group 2) compared to groups of rats treated with 10 % (Group 3), 20 % (Group 4), and 40 % (Group 5) (w/w) of brewers' rice ($n = 3$). Expression of cyclin D1 showed cytoplasm and nucleus staining: strong cyclin D1 in Group 2, weaker staining of cyclin D1 in Group 3 and Group 4, and the weakest staining of cyclin D1 in Group 5 (Magnification 200x). Scale bars in the figures indicate 100 μm

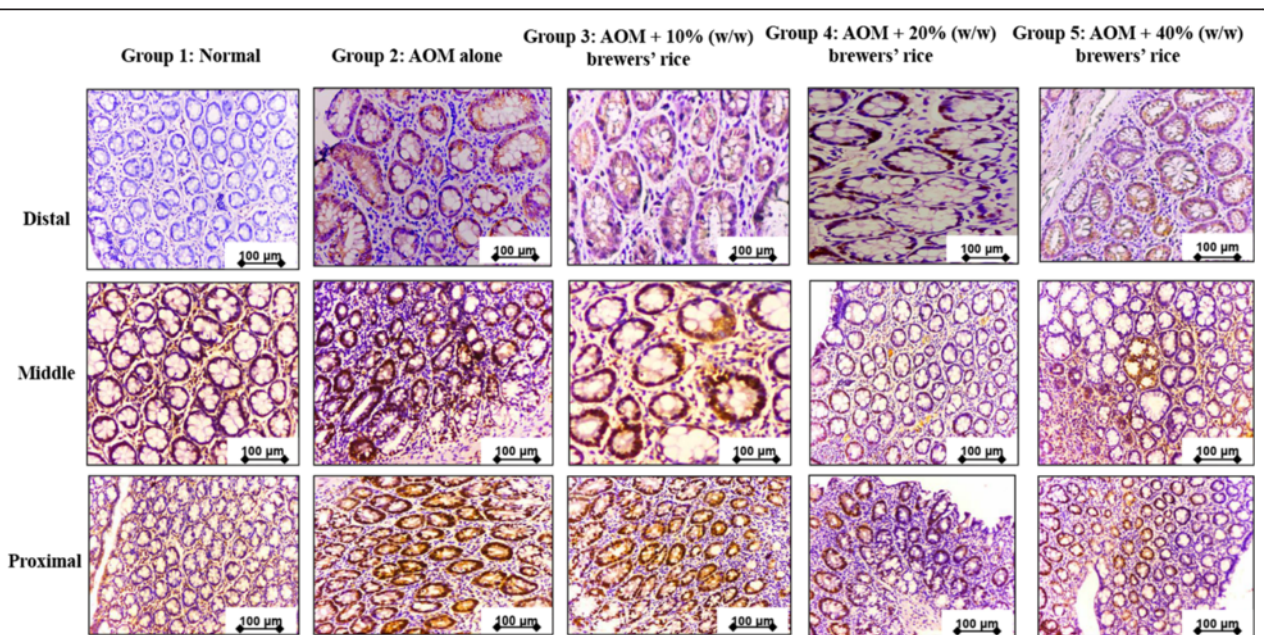


Fig. 6 Immunohistochemical staining of c-myc. The expression of c-myc in three sections, including distal, middle, and proximal colon in normal (Group 1), AOM alone (untreated) (Group 2) compared to groups of rats treated with 10 % (Group 3), 20 % (Group 4), and 40 % (Group 5) (w/w) of brewers' rice ($n = 3$). Expression of c-myc showed cytoplasm and nucleus staining: strong c-myc in Group 2, weaker staining of c-myc in Group 3 and Group 4, and the weakest staining of c-myc in Group 5 (Magnification 200x). Scale bars in the figures indicate 100 μm

Table 3 Total score of cyclin D1 and c-myc expression in colonic tissue

Groups	Normal	AOM alone	AOM + 10 % (w/w) of brewers' rice	AOM + 20 % (w/w) of brewers' rice	AOM + 40 % (w/w) of brewers' rice	P
Cyclin D1	0	123.67 ± 9.61 ^a	123.00 ± 9.54 ^a	94.33 ± 4.73 ^b	85.67 ± 8.14 ^b	0.000
c-myc	0	125.00 ± 5.57 ^a	123.00 ± 8.00 ^a	97.00 ± 4.58 ^b	93.00 ± 6.08 ^b	0.000

Each value expressed as mean ± SD of three determinations. Value in the same row with different superscript letter indicates significant difference by Tukey's test ($P < 0.05$)

reveal that the β -catenin amount in the normal colon tissue was too low to be detected through western blotting. In the AOM-alone group (G2), a carcinogen was injected, but no treatment with brewers' rice was administered, and prominent β -catenin protein expression was observed. The overall analysis showed that the colon mucosa of G2, which had the highest values of tumor incidence and tumor multiplicity compared with other groups [16], displayed the highest β -catenin protein expression in the western blotting evaluation compared with the other treatment groups.

A significant reduction in the protein expression of β -catenin was also found in the treatment groups (G3, G4, and G5) compared with G2 ($P < 0.05$) (Fig. 7a). This result was consistent with the findings obtained by Nurul Husna et al. [48], who reported that the protein level of β -catenin was diminished after treatment with rice bran phytic acid. These data implied that brewers' rice has the potential to reduce the active form of β -catenin.

Treatment with brewers' rice reduces the cyclin D1 and c-myc protein levels in colonic tumors

To evaluate whether the dietary administration of brewers' rice on AOM-injected colon carcinogenesis mediates the inhibition of the Wnt/ β -catenin downstream signaling pathway, we investigated the Wnt target genes at the protein level. Cyclin D1 is a well-recognized cell cycle protein targeted by β -catenin [6] that is usually overexpressed in colonic tumors [49]. c-myc is one of the crucial protein in the modulation of cell proliferation by β -catenin and the Wnt pathway [50].

Expectedly, we observed no cyclin D1 and c-myc protein expression in the normal colon mucosa (Fig. 7b and c). As shown in Fig. 7b and c, the AOM-alone group (G2) exhibited the highest protein expression of cyclin D1 and c-myc in the western blotting analyses compared with the other treatment groups. We also found reductions in the cyclin D1 and c-myc protein levels in the colon tumors of rats received with a brewers' rice diet. The dietary administration of brewers' rice has been

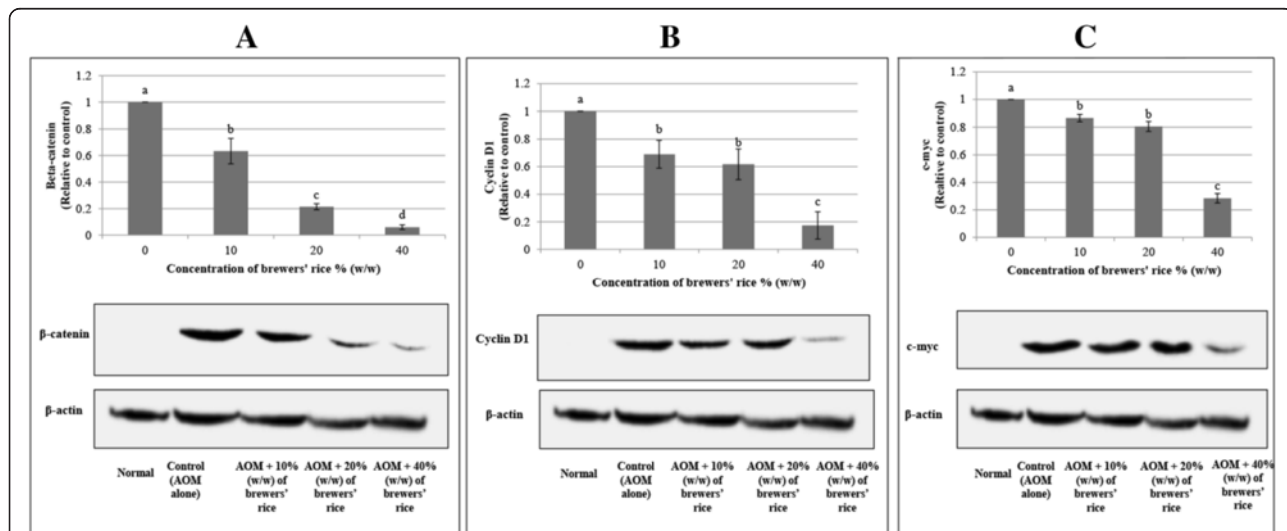


Fig. 7 Western blotting of β -catenin, cyclin D1, and c-myc in brewers' rice treated AOM-induced colon cancer. The protein levels of β -catenin, cyclin D1, and c-myc in normal (Group 1), AOM alone (untreated) (Group 2) compared to groups of rats treated with 10 % (Group 3), 20 % (Group 4), and 40 % (Group 5) (w/w) of brewers' rice. **a** Brewers' rice decreased protein expression of β -catenin, **b** brewers' rice reduced protein expression of cyclin D1, and **c** brewers' rice decreased protein expression of c-myc in a dose-dependent manner. Each value expressed as mean ± SD ($n = 3$). Value with different superscript letter indicates significant difference between groups by Tukey's test ($P < 0.05$). Administration of 10, 20, and 40 % (w/w) of brewers' rice significantly reduced β -catenin, cyclin D1, and c-myc protein expression compared to the AOM-alone group ($P < 0.05$). However, the protein levels of cyclin D1 and c-myc did not significantly differ between 10 % and 20 % (w/w) brewers' rice ($P > 0.05$)

demonstrated to reduce cyclin D1 and c-myc protein expression in a dose-dependent manner. The protein expression of cyclin D1 and c-myc in the AOM-alone group was significantly higher than that of the brewers' rice-treated groups ($P < 0.05$) (Fig. 7b and c). However, no significant difference in the cyclin D1 and c-myc protein expression levels was observed between the groups treated with 10 % and 20 % (w/w) brewers' rice ($P > 0.05$). These results suggested that the effects of the dietary administration of brewers' rice on cyclin D1 and c-myc corroborated the potential of brewers' rice for regulating the β -catenin level in colon tumors. Taken together, this finding indicated that the dietary administration of brewers' rice inhibited the Wnt/ β -catenin downstream signaling pathway.

In the present study, the colon tumor samples in each group were randomly selected for analysis. Only three determinations conducted for each group in both immunohistochemistry and Western blotting analyses were based on several previous studies [48, 51–53].

Most studies have reported the additive and/or synergistic protective effects of several components [54]. Therefore, in the current study, brewers' rice rather than isolated compounds were fed to the rats. In our previous study, we identified a greater inhibition of tumor incidence and multiplicity in AOM-induced rats after treatment with brewers' rice [16]. We speculated that this was partially a result of the nutrient compositions and bioactive compounds, which synergistically contribute to this significant suppression of β -catenin, cyclin D1, and c-myc expression. Additionally, the synergistic/additive effects of the components and the presence of other active compounds in WBR such as phenolic compounds (mainly ferulic acid, gallic acid, and p-coumaric acid) [19], phytic acid, and antioxidant activity as evaluated using β -carotene bleaching test, 1,1-diphenyl-2-picryl-hydrazyl (DPPH) radical scavenging capacity, and ferric reducing antioxidant power (FRAP) assays [14] are more likely to cause the observed apoptotic effects in HT-29 cells, as confirmed by a colorimetric assay. A dietary chemoprevention study demonstrated that the magnitude of the anticancer effect in the whole food or whole food extract is greater compared to that of the individual phytochemical components [55]. Whole rice bran is recognized to be vital in providing a comprehensive protection against cancerous cells compared with the protection provided by the isolated compounds [56]. Therefore, the downregulation of the downstream signaling targets, including cyclin D1 and c-myc, are likely attributed to the synergistic/additive effects of the active constituents present in brewers' rice, such as phenolic compounds (mainly ferulic, gallic, and p-coumaric acids), phytic acid, antioxidant activity, vitamin E (mainly γ -tocotrienol and γ -tocopherol), and γ -oryzanol, as reported in our previous studies [14, 19].

Conclusions

This study provides clear and substantial evidence that brewers' rice offers great potential against colorectal cancer through the inhibition of colon carcinogenesis by the modulation of upstream and downstream components in the Wnt signaling pathway. Our findings demonstrated that brewers' rice downregulates β -catenin and the Wnt target proteins cyclin D1 and c-myc, which provides a good rationale for a further comprehensive study of dietary brewers' rice against colon cancer. Collectively, these findings provide evidence that WBR can induce apoptosis and inhibit the proliferation of HT-29 cells through regulation of caspase-dependent pathways and suppression of the Wnt/ β -catenin downstream signaling pathway in AOM-induced rats. We suggest that brewers' rice may be a useful dietary agent for colorectal cancer.

Abbreviations

AIN-93G: American institute of nutrition; AOM: Azoxymethane; APC: Adenomatous polyposis coli; CK1: Casein kinase 1; DMEM: Dulbecco's modified eagle medium; DPPH: 1,1-diphenyl-2-picryl-hydrazyl; FBS: Fetal bovine serum; FRAP: Ferric reducing antioxidant power; Fz: Frizzled; GAPDH: Glyceraldehyde-3-phosphate dehydrogenase; GSK3 β : Glycogen synthase kinase 3 β ; HRP: Horseradish Peroxidase; HT-29: Colorectal cancer; LDH: Lactate dehydrogenase; LEF: Lymphoid enhancing factor; LRP6: Low-density lipoprotein receptor-related protein 6; MBR: Methanol extract of brewers' rice; MTT: 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; NSAIDs: Non-steroidal anti-inflammatory drugs; PBS: Phosphate-buffered saline; PBST: Phosphate-buffered saline in 0.1 % Tween-20; pNA: p-nitroanilide; RIPA: Radio immune precipitation assay; SDS-PAGE: Sodium dodecyl sulfate-polyacrylamide gel electrophoresis; TCF: T-cell factor; WBR: Water extract of brewers' rice.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

BLT, MEN, SKY, and KR participated in the design of the study. BLT performed the experiments. BLT, KH, and SRH conducted acquisition of data. BLT, MEN, and HH contributed to the analysis and interpretation of data. BLT and MEN drafted the manuscript. All authors read and approved the final version of the manuscript.

Acknowledgements

We acknowledge the BERNAS, Seri Tiram Jaya, Selangor, Malaysia for supplying the brewers' rice sample. We also appreciate the Ministry of Education, Malaysia for providing the research funding to implement the project.

Author details

¹Department of Nutrition and Dietetics, Faculty of Medicine and Health Sciences, Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia.

²Research Centre of Excellent, Nutrition and Non-Communicable Diseases (NNCD), Faculty of Medicine and Health Sciences, Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia.

³Laboratory of Molecular Biomedicine, Institute of Bioscience, Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia.

⁴Department of Cell and Molecular Biology, Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia.

⁵Department of Veterinary Pathology and Microbiology, Faculty of Veterinary Medicine, Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia.

⁶Department of Food Technology, Faculty of Food Science and Technology, Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia.

Received: 10 December 2014 Accepted: 17 June 2015

Published online: 30 June 2015

References

- American Cancer Society. Global Cancer Facts and Figures. Atlanta: American Cancer Society; 2011.
- Khan N, Afaq F, Mukhtar H. Apoptosis by dietary factors: the suicide solution for delaying cancer growth. *Carcinogenesis*. 2007;28:233–9.
- Thompson MD, Monga SPS. Wnt/ β -catenin signaling in liver health and disease. *Hepatology*. 2007;45:1298–305.
- Polakis P. The many ways of Wnt in cancer. *Curr Opin Genet Dev*. 2007;17:45–51.
- Takahashi-Yanaga F, Kahn M. Targeting Wnt signalling: can we safely eradicate cancer stem cells? *Clin Cancer Res*. 2010;16:3153–62.
- Tetsu O, McCormick F. β -Catenin regulates expression of cyclin D1 in colon carcinoma cells. *Nature*. 1999;398:422–6.
- Giles RH, van Es JH, Clevers H. Caught up in a Wnt storm: wnt signaling in cancer. *Biochim Biophys Acta*. 2003;1653:1–24.
- Klaus A, Birchmeier W. Wnt signalling and its impact on development and cancer. *Nat Rev Cancer*. 2008;8:387–98.
- Slayton T, Timmer CP. Japan, China and Thailand can solve the rice crisis—but US Leadership is needed. *CGD Notes*. Washington, DC: Center for Global Development; 2008. p. 1–6.
- Van Hoed V, Depaemelaere G, Vila Ayala J, Santiwattana P, Verhé R, Greyt WD. Influence of chemical refining on the major and minor components of rice bran oil. *J Am Oil Chem Soc*. 2006;83:315–21.
- de Deckere EA, Korver O. Minor constituents of rice bran oil as functional foods. *Nutr Rev*. 1996;54:120–6.
- Glatthar J, Heinisch J, Senn T. The use of unmalted triticale in brewing and its effect on wort and beer quality. *J Am Soc Brew Chem*. 2003;61:182–90.
- Esa NM, Ling TB, Peng LS. By-products of rice processing: An overview of health benefits and applications. *J Rice Res*. 2013;1:107.
- Tan BL, Norhaizan ME, Suhaniza HJ, Lai CC, Norazalina S, Roselina K. Antioxidant properties and antiproliferative effect of brewers' rice extract (*temukut*) on selected cancer cell lines. *Int Food Res J*. 2013;20:2117–24.
- Yu S, Nehus ZT, Badger TM, Fang N. Quantification of vitamin E and gamma-oryzanol components in rice germ and bran. *J Agric Food Chem*. 2007;55:7308–13.
- Tan BL, Norhaizan ME, Heshu SR, Hazilawati H, Roselina K. Brewers' rice induces apoptosis in azoxymethane-induced colon carcinogenesis in rats via suppression of cell proliferation and the Wnt signaling pathway. *BMC Complement Altern Med*. 2014;14:304.
- Kohno H, Suzuki R, Sugie S, Tanaka T. Suppression of colitis-related mouse colon carcinogenesis by a COX-2 inhibitor and PPAR ligands. *BMC Cancer*. 2005;5:46–58.
- Prayong P, Barusrux S, Weerapreeyakul N. Cytotoxic activity screening of some indigenous Thai plants. *Fitoterapia*. 2008;79:598–601.
- Tan BL, Norhaizan ME, Yeap SK, Roselina K. Water extract of brewers' rice induces antiproliferation of human colorectal cancer (HT-29) cell lines via the induction of apoptosis. *Eur Rev Med Pharmacol Sci*. 2015;19:1022–9.
- Cselenyi CS, Jernigan KK, Tahinci E, Thorne CA, Lee LA, Lee E. LRP6 transduces a canonical Wnt signal independently of Axin degradation by inhibiting GSK3's phosphorylation of β -catenin. *Proc Natl Acad Sci USA*. 2008;105:8032–7.
- Kishida M, Koyama S, Kishida S, Matsubara K, Nakashima S, Higano K, Takada R, Takada S, Kikuchi A. Axin prevents Wnt-3a-induced accumulation of β -catenin. *Oncogene*. 1999;18:979–85.
- Kishida S, Yamamoto H, Hino S, Ikeda S, Kishida M, Kikuchi A. DIX domains of Dvl and Axin are necessary for protein interactions and their ability to regulate β -catenin stability. *Mol Cell Biol*. 1999;19:4414–22.
- Nelson WJ, Nusse R. Convergence of Wnt, beta-catenin, and cadherin pathways. *Science*. 2004;303:1483–7.
- Ougolkov AV, Billadeau DD. Targeting GSK-3: a promising approach for cancer therapy? *Future Oncol*. 2006;2:91–100.
- Luo J. Glycogen synthase kinase 3 β (GSK3 β) in tumorigenesis and cancer chemotherapy. *Cancer Lett*. 2009;273:194–200.
- Shakoori A, Ougolkov A, Yu ZW, Zhang B, Modarressi MH, Billadeau DD, Mai M, Takahashi Y, Minamoto T. Deregulated GSK3 β activity in colorectal cancer: its association with tumor cell survival and proliferation. *Biochem Biophys Res Commun*. 2005;334:1365–73.
- Gatenby RA, Gawlinski ET. The glycolytic phenotype in carcinogenesis and tumor invasion: insights through mathematical models. *Cancer Res*. 2003;63:3847–54.
- Jope RS, Johnson GW. The glamour and gloom of glycogen synthase kinase-3. *Trends Biochem Sci*. 2004;29:95–102.
- Ougolkov AV, Fernandez-Zapico ME, Bilim VN, Smyrk TC, Chari ST, Billadeau DD. Aberrant nuclear accumulation of glycogen synthase kinase-3 β in human pancreatic cancer: association with kinase activity and tumor dedifferentiation. *Clin Cancer Res*. 2006;12:5074–81.
- Cao Q, Lu X, Feng YJ. Glycogen synthase kinase-3 β positively regulates the proliferation of human ovarian cancer cells. *Cell Res*. 2006;16:671–7.
- Dihlmann S, Klein S, Doeberitz MVK. Reduction of β -catenin/T-cell transcription factor signaling by aspirin and indomethacin is caused by an increased stabilization of phosphorylated β -catenin. *Mol Cancer Ther*. 2003;2:509–16.
- Clevers H, Nusse R. Wnt/ β -catenin signaling and disease. *Cell*. 2012;149:1192–205.
- Wood LD, Parsons DW, Jones S, Lin J, Sjöblom T, Leary RJ, Shen D, Boca SM, Barber T, Ptak J, Silliman N, Szabo S, Dezso Z, Ustyanksky V, Nikolskaya T, Nikolsky Y, Karchin R, Wilson PA, Kaminker JS, Zhang Z, Croshaw R, Willis J, Dawson D, Shiptsin M, Willson JKV, Sukumar S, Polyak K, Park BH, Pethiyagoda CL, Krishna Pant PV et al. The genomic landscapes of human breast and colorectal cancers. *Science*. 2007;318:1108–13.
- Nagase H, Nakamura Y. Mutations of the APC (adenomatous polyposis coli) gene. *Hum Mutat*. 1993;2:425–34.
- Moon RT, Miller JR. The APC tumor suppressor protein in development and cancer. *Trends Genet*. 1997;13:256–8.
- Verheyen EM, Gottardi CJ. Regulation of Wnt/ β -catenin signaling by protein kinases. *Dev Dyn*. 2010;239:34–44.
- He X, Semenov M, Tamai K, Zeng X. LDL receptor-related proteins 5 and 6 in Wnt/ β -catenin signaling: arrows point the way. *Development*. 2004;131:1663–77.
- Mi K, Johnson GV. Role of the intracellular domains of LRP5 and LRP6 in activating the Wnt canonical pathway. *J Cell Biochem*. 2005;95:328–38.
- Bache KG, Slagsvold T, Stenmark H. Defective downregulation of receptor tyrosine kinases in cancer. *EMBO J*. 2004;23:2707–12.
- Shankar S, Ganapathy S, Hingorani SR, Srivastava RK. EGCG inhibits growth, invasion, angiogenesis and metastasis of pancreatic cancer. *Front Biosci*. 2008;13:440–52.
- Singh BN, Shankar S, Srivastava RK. Green tea catechin, Epigallocatechin-3-gallate (EGCG): Mechanisms, perspectives and clinical applications. *Biochem Pharmacol*. 2011;82:1807–21.
- Majno G, Joris I. Apoptosis, oncosis, and necrosis. An overview of cell death. *Am J Pathol*. 1995;146:3–15.
- Nieves-Neira W, Pommier Y. Apoptotic response to camptothecin and 7-hydroxystaurosporine (UCN-01) in the 8 human breast cancer cell lines of the NCI anticancer drug screen: Multifactorial relationships with topoisomerase I, protein kinase C, Bcl-2, p53, MDM-2 and caspase pathways. *Int J Cancer*. 1999;82:396–404.
- Boateng J, Verghese M, Panala V, Walker LT, Shackelford L. Protective effects of rice bran on chemically induced colon tumorigenesis may be due to synergistic/additive properties of bioactive components. *Int J Cancer Res*. 2009;5:153–66.
- Alao JP. The regulation of cyclin D1 degradation: roles in cancer development and the potential for therapeutic invention. *Mol Cancer*. 2007;6:24.
- Yang J, Zhang W, Evans PM, Chen X, He X, Liu C. Adenomatous polyposis coli (APC) differentially regulates beta-catenin phosphorylation and ubiquitination in colon cancer cells. *J Biol Chem*. 2006;281:17751–7.
- van de Wetering M, Sancho E, Verweij C, de Lau W, Oving I, Hurlstone A, van der Horn K, Batlle E, Coudreuse D, Haramis A-H, Tjon-Pon-Fong M, Moerer P, van der Born M, Soete G, Pals S, Eilers M, Medema R, Clevers H. The β -catenin/TCF-4 complex imposes a crypt progenitor phenotype on colorectal cancer cells. *Cell*. 2002;111:241–50.
- Nurul Husna S, Norhaizan ME, Hairuszah I, Abdah Md A, Norazalina S, Ashok KP. Preventive Inositol Hexaphosphate extracted from rice bran inhibits colorectal cancer through involvement of Wnt/ β -catenin and COX-2 pathways. *BioMed Res Int*. 2013;2013:681027.
- Zhang T, Nanney LB, Luongo C, Lamps L, Heppner KJ, DuBois RN, Beauchamp RD. Concurrent overexpression of cyclin D1 and cyclin dependent kinase 4 (Cdk4) in intestinal adenomas from multiple intestinal neoplasia (Min) mice and human familial adenomatous polyposis patients. *Cancer Res*. 1997;57:169–75.

50. He TC, Sparks AB, Rago C, Hermeking H, Zawel L, da Costa LT, Morin PJ, Vogelstein B, Kinzler KW. Identification of c-MYC as a target of the APC pathway. *Science*. 1998;281:1509–12.
51. Paul S, DeCastro AJ, Lee HJ, Smolarek AK, So JY, Simi B, Wang CX, Zhou R, Rimando AM, Suh N. Dietary intake of pterostilbene, a constituent of blueberries, inhibits the β -catenin/p65 downstream signaling pathway and colon carcinogenesis in rats. *Carcinogenesis*. 2010;31:1272–8.
52. Li S-C, Chou T-C, Shih C-K. Effects of brown rice, rice bran, and polished rice on colon carcinogenesis in rats. *Food Res Int*. 2011;44:209–16.
53. Kunnumakkara AB, Diagaradjane P, Anand P, Kuzhuvilil HB, Deorukhkar A, Gelovani J, Guha S, Krishnan S, Aggarwal BB. Curcumin sensitizes human colorectal cancer to capecitabine by modulation of cyclin D1, COX-2, MMP-9, VEGF and CXCR4 expression in an orthotopic mouse model. *Int J Cancer*. 2009;125:2187–97.
54. Tsuda H, Ohshima Y, Nomoto H, Fujita K, Matsuda E, Iigo M, Takasuka N, Moore MA. Cancer prevention by natural compounds. *Drug Metab Pharmacokinet*. 2004;19:245–63.
55. Ricciardiello L, Bazzoli F, Fogliano V. Phytochemicals and colorectal cancer prevention—myth or reality? *Nature Reviews. Gastroenterol Hepatol*. 2011;8:592–6.
56. Phutthaphadoong S, Yamada Y, Hirata A, Tomita H, Taguchi A, Hara A, Limtrakul P-N, Iwasaki T, Kobayashi H, Mori H. Chemopreventive effects of fermented brown rice and rice bran against 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone-induced lung tumorigenesis in female A/J mice. *Oncol Rep*. 2009;21:321–7.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

