



REVIEW

Corneal Regeneration After Photorefractive Keratectomy: A Review[☆]



Javier Tomás-Juan^{a,*}, Ane Murueta-Goyena Larrañaga^b, Ludger Hanneken^a

^a Department of Visual Science, VallmedicVision International Eye Clinic, Andorra

^b Collaborator Researcher at Department of Neuroscience, University of Basque Country, Leioa, Spain

Received 28 June 2014 ; received in revised form 1 August 2014

Available online 23 October 2014

KEYWORDS

Photorefractive keratectomy;
Cornea;
Wound healing;
Contact lenses

Abstract Photorefractive keratectomy (PRK) remodels corneal stroma to compensate refractive errors. The removal of epithelium and the ablation of stroma provoke the disruption of corneal nerves and a release of several peptides from tears, epithelium, stroma and nerves. A myriad of cytokines, growth factors, and matrix metalloproteases participate in the process of corneal wound healing. Their balance will determine if reepithelization and stromal remodeling are appropriate. The final aim is to achieve corneal transparency for restoring corneal function, and a proper visual quality. Therefore, wound-healing response is critical for a successful refractive surgery. Our goal is to provide an overview into how corneal wounding develops following PRK. We will also review the influence of intraoperative application of mitomycin C, bandage contact lenses, anti-inflammatory and other drugs in preventing corneal haze and post-PRK pain.

© 2014 Spanish General Council of Optometry. Published by Elsevier España, S.L.U. All rights reserved.

PALABRAS CLAVE

Queratectomía fotorrefractiva;
Córnea;
Curación de heridas;
Lentes de contacto

Regeneración de la córnea tras queratectomía fotorrefractiva: revisión bibliográfica

Resumen La queratectomía fotorrefractiva (PRK) remodela el estroma de la córnea para compensar los errores refractivos. La eliminación del epitelio y la ablación del estroma provoca la alteración de los nervios corneales y la liberación de diversos péptidos de la lágrima, epitelio, estroma y nervios. Innumerables citoquinas, factores de crecimiento y metaloproteasas de la matriz participan en el proceso de regeneración y cicatrización corneal. Su equilibrio determinará si la re-epitelización y la remodelación del estroma son adecuados. El objetivo final

[☆] The authors have not proprietary or commercial interest in the medical devices that are involved in this manuscript.

* Corresponding author at: VallmedicVision Andorra Avinguda Naciones Unides 17. AD700, Escaldes-Engordany, Andorra.
E-mail address: javier.tomas@vallmedicvision.com (J. Tomás-Juan).

es el logro de la transparencia corneal para restablecer la función de la córnea, así como la calidad visual adecuada. Por tanto, la respuesta de regeneración y cicatrización corneal es esencial para el éxito de la cirugía refractiva. Nuestro objetivo es aportar una visión general sobre el modo en que se desarrolla dicho proceso tras la PRK. Revisaremos también la influencia de la aplicación intraoperatoria de mitomicina C, lentes de contacto terapéuticas, y otros fármacos para prevenir el haze y el dolor tras la PRK.

© 2014 Spanish General Council of Optometry. Publicado por Elsevier España, S.L.U. Todos los derechos reservados.

The ablation surgery of the corneal surface for the correction of refractive errors began with the development of the excimer laser. The acronym laser means ‘‘Light Amplification by the Stimulated Emission of Radiation’’. Photorefractive keratectomy (PRK), developed by Trokel and colleagues in 1983, uses an excimer laser that emits ultraviolet light of 193 nanometers (nm), a combination of Argon and Fluor (ArF) to remodel the cornea.^{1–6} It was not until 1996 when the Food and Drug Administration (FDA) approved PRK as a refractive surgery technique.⁷ In PRK the excimer laser acts on the anterior corneal stroma,^{2,8,9} producing a stromal remodeling, and, consequently, inducing a change in corneal refraction.^{10,11} It corrects mild to moderate myopia, hyperopia and astigmatism, with high level of safety and efficacy.^{3,11–20} However, the use of PRK has been reduced over the past years by the introduction of the Laser In Situ Keratomileusis (LASIK).^{12,21} Although LASIK provides less postoperative pain, less inflammation, and faster corneal wound healing and visual recovery,^{8,17,19,22–25} PRK may be a useful alternative in post-radial keratotomy,^{26–28} post-penetrating keratoplasty,²⁹ in thin corneas, irregular topographies, alterations of the basal membrane, treatment of some LASIK flap complications or residual refractive errors after LASIK.^{11,12,19,30–32} It is also indicated in military pilots, professional athletes, or patients that have a high risk for traumatic postoperative flap dislocation.^{12,31} In addition to the above-mentioned advantages, the PRK has gained popularity with the recent wave front guided laser ablation, which reduces postoperative high order aberrations (HOA), improving the optical quality.³⁰

The visual quality might not be optimal if some complications take place, like subepithelial corneal haze, epithelial hypertrophy, regression of refractive error, deposition of subepithelial extracellular matrix or fibrosis. Other adverse effects include postoperative pain, abnormal corneal nerve regeneration, and night vision symptoms like halos and glare.^{3,10,11,14,18,22,33–40}

The purpose of this review is to explain the main cellular changes and complications that occur in different corneal layers after PRK, and to explain how they affect the visual quality. We discuss the role of mitomycin C and bandage contact lenses in corneal regeneration, and the role of different drugs in postoperative corneal pain management.

Corneal Wound Healing

Corneal wound healing is a complex process that, in normal conditions, culminates in the restoration of the tissue, without scar formation or vascularization. The aim is to

maintain transparency to recover a proper visual function. After epithelial injury, the corneal healing starts with the removal of necrotic cells.⁴¹ Fibronectin provides a transient matrix for the adhesion of migrating cells, until an epithelial monolayer covers the injured area.⁴² Fibronectin also stimulates the production of plasminogen activator (PAA), and by a cascade of events, cell-subepithelial matrix adhesions break down.⁴² In the next step, limbal stem cells undergo mitosis to reestablish lost cells, and with the anchoring of hemidesmosomes to the underlying stroma, the epithelial regeneration process completes.⁴¹ Stromal wound healing depends on epithelial cells, and on their interaction with keratocytes.⁴³ Following stromal injury, released cytokines induce the apoptosis of keratocytes under the wound, and stimulate the proliferation and migration of neighboring keratocytes.⁴⁴ These active keratocytes synthesize matrix metalloproteases (MMP) to remodel the stroma. At later stages, a number of them take the repair phenotype, the so-called myofibroblasts,⁴⁵ and produce collagen and extracellular matrix (ECM), until the basement membrane prevents the inflow of cytokines in the stroma, and myofibroblast, presumably, commit apoptosis (Fig. 1).^{46,47}

Epithelial Wound Healing Following PRK

The corneal epithelium is formed by superficial, wing and basal cells.^{48,49} In order to facilitate the stromal ablation in PRK, the corneal epithelium is removed. The absence of the epithelium will condition corneal repair. Corneal epithelial cells are the first cells involved in the corneal regeneration process after PRK.⁵⁰ Epithelial cells proliferate and migrate from the limbus and the basal epithelial layer to reestablish corneal layers.^{8,51} Corneal regeneration after PRK can be better understood using current, non-invasive, confocal microscopy. It has been used on animals and on humans for corneal cellular structure visualization in real time.^{2,5,10,22,25,40,48} Esquenazi et al.²² proved using a new generation high-resolution in vivo confocal microscope that environmental conditions influenced the regeneration of the corneal epithelium. They showed that the number of the superficial cells was reduced in desiccating environments compared with normal conditions, and the number of basal epithelial cells was increased. Histological studies conducted in animals and in humans, have found that corneal epithelium is thicker after PRK,^{2,52} caused by an elongation of the basal epithelial cells and an increased number of superficial cell layers.²⁵ The corneal flattening in myopic PRK may result in postoperative epithelial thickening due to

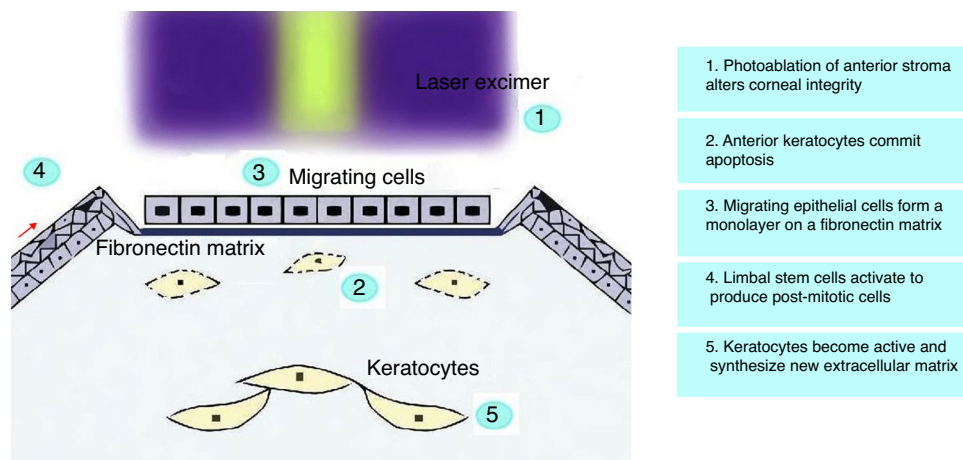


Figure 1 Corneal alterations and first steps of wound healing following PRK.

the lack of mechanical influences of the upper eyelid that polishes the corneal surface with blinking.² Epithelial hyperplasia in PRK is associated with deep stromal ablation depths and with small ablation zones (4.00–4.5 mm) because there is a marked curvature change in the edges of the ablated area. When ablation zones are large (6.00 mm), they have less demarcated contours, and thus, the change in epithelial thickness is minimal.^{53–56} Table 1 shows the variation of central corneal thickness with different surgery techniques published in the scientific literature. Erie² proved that, after PRK, the central epithelial thickness returned to preoperative levels at 1 month. However, it continued to progressively increase during the first year, being 21% thicker at that time. This result is similar to the 22% thickness increase seen in LASIK by Erie et al.⁵⁷ However, the time required for thickness stabilization differs between the two techniques, due to the complex interaction of epithelial cells and activated keratocytes in PRK.² According to Patel et al.,²⁵ central corneal epithelium in LASIK increased 24% during the first year after surgery and remained stable during the next 7 years. In PRK, corneal thickness continued to increase at 1 month, 1 year and 7 years ($442 \pm 39 \mu\text{m}$, $464 \pm 44 \mu\text{m}$, $471 \pm 45 \mu\text{m}$; respectively).²⁵ Recently, Ivarsen et al.⁵² have concluded that in PRK and LASIK, the epithelial thickness increases 15%–20% after surgery, but the epithelial changes in LASIK occur during the first week and remain unaltered during the following 3 years. It has been suggested that epithelial hyperplasia can induce a reduction of post-operative refractive effect. Erie showed myopic regression significantly associated with epithelial thickness increase.² Nevertheless, Ivarsen et al.⁵² did not find any correlation between changes in epithelial thickness and changes in refraction after PRK or LASIK, probably because of the small size of their sample.

Stromal Wound Healing Following PRK

Stroma occupies approximately the 90% of corneal thickness,⁵⁸ and it can be subdivided into three continuous layers: anterior, middle and posterior.⁴⁹ The corneal stroma is built up from collagen fibers, ground substance, kera-

toocytes and nerve fibers.^{5,49} Keratocytes – corneal stromal cells – play a major role in maintaining corneal transparency, and synthesizing the components of the extracellular matrix (ECM).⁵⁸ Active keratocytes produce collagen and proteoglycans to form the ECM after stromal injury. The human stromal cornea contains collagen type I, V and VI.^{59,60} Type I is predominant (75%), followed by type VI (approximately, 17%).⁶⁰ Type III collagen appears in inflammatory events or during wound healing. Proteoglycans participate in collagen fibrillogenesis and matrix assembly.⁶¹ After corneal injury, newly produced collagen fibers tend to have larger diameters, as they contain high levels of dermatan sulphate (a type of proteoglycan) that lasts up to 6 months.⁶²

Stromal keratocytes are normally quiescent or inactive, and are the second cells involved in the process of corneal regeneration, just after corneal epithelial cells. After PRK, keratocytes underlying the wound disappear by apoptosis due to a stress exposure.^{2,24,37,40,50,63} During the first 24 h after injury, macrophages, monocytes, T cells and polymorphonuclear cells infiltrate the area and remove damaged cells.^{44,64} Metalloproteinases (MMPs) and the plasminogen activator system remove the affected extracellular matrix.^{9,65–67} The MMPs are proteolytic enzymes secreted by active keratocytes or fibroblasts, and degrade complex molecules of the extracellular matrix. Although nine types of MMPs exist, in the cornea only four MMPs are important, being MMP-1 the most relevant.⁶⁷ MMP-8 concentration has been observed to be significantly elevated in the second day after PRK ($P = .001$).⁶⁸ The remaining keratocytes, adjacent to wound borders, are activated in response to various cytokines released by cells in upper layers, such as interleukin (IL)-1, and growth factors like tumor necrosis factor (TNF), fibroblast growth factor (FGF), platelet-derived growth factor (PDGF), epithelial growth factor (EGF), and transforming growth factor (TGF).^{2,8,14,22,24,35,51,58,69–74} These growth factors are normal components of the tear and corneal cells, produced by the lacrimal gland,³⁵ and regulate a variety of processes involved in homeostasis and corneal wound healing, including migration, mitosis and cell differentiation.⁷⁵ Particularly, transformation growth factor beta (TGF-beta) seems to transform active keratocytes into myofibroblasts that appear at later stages of stromal heal-

Table 1 Variation of Central Corneal Thickness, Mean±SD or Range (µm).

Study	Technique	Preop	1 week	1 month	3 months	6 months	1 year	2 years	3 years	5 years	7 years
Ivarsen et al. (2009) ⁵²	LASIK	529±49	460±47				463±32		477±43		
Wallau and Campos (2008) ¹⁵⁹	PRK	522±32	423±35								
Patel et al. (2007) ²⁵	LASIK	543±25		473±28	473±27	473±27					
	PRK with MMC ^a	545±26		456±30	458±30	460±29					
Kozak et al. (2003) ⁵⁶	LASIK	514±37		452±38			456±42	456±39	462±37	456±30	455±31
	PRK	(437-576)		(411-563)			(386-576)	(402-564)	(411-577)	(388-524)	(371-509)
Kozak et al. (2003) ⁵⁶	LASIK	493±36		442±40			464±44	469±44	472±59	468±48	471±45
	PRK	(427-560)		(388-526)			(396-534)	(411-544)	(393-573)	(406-555)	(400-557)
Kozak et al. (2003) ⁵⁶	LASIK	549±37	467±29		474±30	481±23					
	PRK	(515-620)	473±39		477±35	482±35					

LASIK, laser in situ keratomileusis; PRK, photorefractive keratectomy; MMC, mitomycin C.

^a 0.002%, 1 min.

ing. Myofibroblasts can be identified through the expression of α -smooth muscle actin (SMA).⁴⁵

PRK produces oxygen free radicals, secondary to the exposure of ultraviolet radiation, thermal increase, and polymorphonuclear cell infiltration.^{76,77} Free oxygen radicals may interact with lipid components, nucleotic acids, and sulphur contained in enzymes,^{72,78} and particularly with reactive oxygen species (ROE) that are considered to produce the most reactive and cytotoxic damage. In fact, they have been described as a partial cause of keratocyte apoptosis.⁷⁷ Among the antioxidant enzymes that protect the cornea from radicals, superoxide dismutase (SOD), glutathione peroxidase (Gpx) and catalase are the most relevant.^{75,78} Ascorbic acid and DL-alpha-tocopherol (Vitamin E) also prevents from the effects of free radicals.⁷⁸ Corneal epithelial ascorbic acid absorbs ultraviolet radiation, protecting keratocytes, but high or altered corneal ascorbate levels in the human cornea after PRK, may produce accelerated keratocyte death.⁵ In rabbit corneas, decreased activity of SOD and Gpx enzymes has been proved after refractive surgery.⁷⁸ For this reason, additional antioxidant enzymes seem to be involved in reducing corneal oxidative stress following PRK. 1-cys peroxiredoxin (1-cys Prx) may be an important enzyme involved in the differentiation, migration and proliferation of epithelial cells. 1-cysPrx increases 4 h after PRK and remains in high levels until 7 days after PRK.⁷⁵

The density of keratocytes varies across the stroma. It is estimated that in the anterior stroma the density is 5%–10% greater than in middle and posterior stroma.^{79,80} It has been documented that a corneal stroma rich in keratocytes prevents the epithelial corneal infection or, at least, minimizes the extension of the infection.^{2,5,10} After PRK, the anterior keratocyte population drastically diminishes, and the distribution and shape is greatly altered.^{2,10,34,40,81,82} In confocal microscopy, high reflectance, hyperplasticity, hypertrophy and a decrease in the contrast of the anterior stromal keratocytes can be observed.^{34,63,83} Human histological studies confirm that the decrease of anterior stromal keratocytes in humans and animal respond similarly.^{84,85} Table 2 shows the variation of anterior, posterior and total keratocyte density in the different studies published in the scientific literature. Erié et al.⁵ confirmed that after 5 years of PRK there were evidences of keratocyte density loss in middle and posterior stroma. They observed a reduction of 20%–24% in the posterior stroma ($P<.05$) although they claimed that this loss was not completely evident. Keratocyte density in the anterior 10% of the stroma continues to decrease 5% per year between 1 and 3 years after PRK.² Erié² reported a progressive decline in anterior stromal keratocytes, becoming significant at 36 months after PRK ($P=.02$). In contrast, middle and posterior keratocyte densities remained unchanged between 1 and 3 years after PRK.² In another study, Erié et al.¹⁰ proved that the keratocyte density in the anterior 10% of the stroma, decreased at 6, 12, 24 and 36 months (41%, 40%, 43%, 45%; respectively) after PRK, compared to pre-PRK. In a posterior longer-term study, Erié et al.⁵ demonstrated a similar decreasing pace in anterior keratocyte density: 40%, 42%, 45%, and 47% at 6 months, 2 years, 3 years, and 5 years ($P<.001$). Amoozadeh et al.⁴⁰ found a reduction in keratocyte density 6 months after surgery, but the loss was similar for LASIK and PRK interventions: in

Table 2 Variation of Keratocyte Density After Surgery, Mean±SD (cell/mm²).

Study	Technique	Preop	3 months	6 months	1 year	2 years	3 years	5 years
Einollahi et al. (2011) ⁶³	PRK with MD	902 ± 107	704 ± 119	643 ± 134				
		653 ± 72	622 ± 53	609 ± 60				
	PRK with AAD	943 ± 100	734.3 ± 103.7	696.7 ± 129.6				
Amoozadeh et al. (2009) ⁴⁰		665 ± 69	617 ± 70	621 ± 72				
	LASIK	1058 ± 95		690 ± 55				
		708 ± 40		699 ± 57				
	PRK	1027 ± 80		707 ± 63				
Midena et al. (2007) ⁸²		719 ± 51		719 ± 45				
	PRK with MMC ^a	449 ± 58						305 ± 59
		363 ± 53						392 ± 53
Erie et al. (2006) ⁵	PRK+corticosteroid	473 ± 58						317 ± 66
		365 ± 53						397 ± 46
	LASIK ^b	31,108 ± 4984		28,337 ± 2863	27,533 ± 2757	27,491 ± 2693	26,320 ± 1973	22,987 ± 1829
	26,220 ± 2897		24,756 ± 3302	23,301 ± 3564	23,776 ± 3596	23,459 ± 3394	23,459 ± 3394	21,017 ± 4534
	23,524 ± 5003		24,779 ± 4234	22,564 ± 4065	23,049 ± 4362	22,337 ± 3894	22,337 ± 3894	17,935 ± 6668

PRK, photorefractive keratectomy; MD, mechanical debridement; AAD, alcohol-assisted debridement; LASIK, laser in situ keratomileusis; MMC, mitomycin C; AA, anterior stroma; PS, posterior stroma; FT, full-thickness.

^a 0.02%, 2 min.

^b Results in volumetric values (cell/mm³).

anterior stroma, 34.7% versus 31.13% ($P>.05$) and posterior stroma 0.31% versus 0.02%, ($P>.05$), respectively. However, other studies have seen differences between PRK and LASIK, probably associated with the more superficial ablation in PRK.^{5,25,40} The consequences of keratocyte density loss after PRK are still unknown, but the visual acuity and corneal clarity seem to be preserved.⁵

After the initial depletion of anterior stromal keratocytes, an increase in the keratocyte density is observed over time, probably secondary to mitosis, cellular migration, or reproduction of keratocytes and myofibroblasts.^{5,9,10,34,51,86} Following apoptotic keratocyte loss, the first morphological changes of remaining keratocytes that can be histologically observed, are an increase in cell size and an increase in the size and the number of nucleoli, rough endoplasmatic reticula, mitochondria, free ribosomes and Golgi complexes, indicating an active state.⁸⁷ These keratocytes quickly repopulate the anterior stroma, and return to similar preoperative levels.^{2,5,10,34} Several studies using confocal microscopy have analyzed the keratocyte density after PRK. Corbett et al.⁸⁸ found that at 2 days after PRK the anterior keratocyte density was increased 50%, 100% at 1 month, and returned to preoperative levels at 6 months. Frueh et al.⁸⁵ concluded that the anterior keratocyte density increased 15% at 1 and 4 months after PRK, and returned to preoperative levels 1 year after PRK. Similarly, Erie et al. found an increase of 20% in the anterior stroma at 3 months after PRK.^{2,89} According to the results of Corbett et al.⁸⁸ and Erie et al.,¹⁰ anterior keratocytes proliferation begins 1 month after PRK, with a pick at 3 months, and return to preoperative levels at 6 months.

Corneal Haze

Corneal haze reduces corneal transparency at variable degrees.^{90,91} Subepithelial haze occurs in all patients 1 month after PRK, reaching the greatest intensity at 3–6 months, and gradually decreases from then on.^{2,8,34,92} Yet, some authors affirm that it begins to decrease at 12–24 months after PRK.^{8,92} Corneal haze is more common after correction of high myopia (>−6.00D), and it is rarely seen after correction of <−6.00D of myopia or <+4.00D of hyperopia.^{43,71,91} Besides the ablation depth, the severity of corneal haze is correlated with excessive ocular UV-B radiation, duration of the epithelial defect, postoperative steroid treatment, male sex and with certain population with brown iris.^{2,16,19,21,24,28,71,81,86,93–95} PRK presents higher corneal haze incidence than LASIK, probably because of the destruction of the basement membrane.^{8,45,52} In the presence of damaged epithelial cells and basement membrane, cytokines and growth factors can easily flow from epithelium to anterior stroma.^{45,96} Cytokines released from epithelial cells activate keratocytes, as mentioned in a previous section, which synthesize large diameter collagen fibrils.^{8,11,33,73,81} Abnormally deposited extracellular matrix implies the development of corneal opacity.⁶⁹ Moreover, active keratocytes present a high reflectance that also contributes to the decrease in corneal transparency. In addition, subepithelial vacuolation, deposit materials like proteoglycans, hyaluronic acid and collagen Type IV are involved in the formation of the corneal haze in advanced stages.^{33,77} Plasminogen activator–plasmin

system degrades the damaged ECM, and extended low levels beyond the third day after PRK causes corneal haze formation.⁹⁷ Guerriero et al.⁵⁸ affirm that the loss of collagen type IV is related to the activation of keratocytes *in vivo* and *in vitro*, and Winkler et al.³³ and Mohrenfels et al.⁹⁸ emphasize on the role of type IV collagen in the development of corneal cloudiness. Secondary ultraviolet B (UV-B) exposure, originating from sun or solarium is a causal factor for aforementioned abnormal proteoglycan deposition and associated augmented corneal thickness.⁹⁹

On the other hand, myofibroblast, derivatives of TGF-beta responding keratocytes, are thought to be the first biological event for corneal haze formation.^{51,93,94} Myofibroblasts play an essential role in the recovery of the corneal integrity after penetrating injury, mainly in advanced stages.²² They secrete extracellular matrix, contract wounds and have the ability to generate adhesion structures with the surrounding substrate.⁷¹ TGF-beta also induces the expression of connective tissue growth factor (CTGF), which mediates collagen synthesis, and along with myofibroblasts regulates the corneal wound healing, and may promote scar formation.¹⁰⁰ After PRK, myofibroblasts appear as a pathological response to injury,⁷¹ and their decreased transparency roots in the low intracellular content of crystalline.¹⁰¹ Irregular surface has also been related to high incidence of corneal haze,^{94,102} and higher irregularity is seen with increasing dioptric corrections in PRK.¹⁰³ Interestingly, surface irregularity is positively correlated with myofibroblast density in the anterior stroma.⁴³ In normal corneal wound healing, complete regeneration of the basal membrane after PRK occurs within 6–8 weeks in rabbits,¹⁰⁴ which limits the access of growth factors to the stroma⁶⁹ and, consequently, myofibroblasts commit apoptosis⁴⁶ modulated by IL-1.⁴⁷ Therefore, the presence of myofibroblast, and subsequent corneal haze, is largely dependent upon the restoration of the basement membrane.^{43,105}

Corneal haze has been traditionally measured in the slit-lamp, and graded with diverse scales, like Hanna's scale. The new technology leads us to use automated instruments for corneal haze measurement. *In vivo* confocal microscopy is a reliable tool, as far as standardized methods are used.¹⁰⁶ It is the most widely used objective method in clinical setting for haze measurement. In the last years, alternative techniques have come out. Confocal imaging of second harmonic-generated (SHG) signals has been shown to be sensitive in measuring corneal fibrosis after refractive surgery.¹⁰⁷ Recently, the densitometry program of Pentacam Scheimpflug imaging system (Oculus Optikgeräte GmbH) has been proved to be a useful method for measuring corneal haze.¹⁰⁸

Visual Disturbances of Corneal Haze

The corneal haze produces a reduction of low contrast visual acuity and night vision symptoms that, in the vast majority of situations, improve with time.⁶⁷ It is possible to see corneal haze formation after PRK by means of confocal microscopy, observed as a decrease in the contrast of the image and an increase in reflectivity.⁸¹ Böhnke et al.⁸¹ using a Tandem scanning confocal microscopy, correlated corneal haze and

anterior stromal reflectivity. However, the tandem scanning confocal microscopy is not able to detect acellular regions of the anterior stroma early after PRK when epithelium and sub-basal plexus are not formed.¹⁰ Although corneal haze in humans is less pronounced than in animal models, if corneal haze persists and affects significantly to the corneal transparency, it causes light scatter.^{4,94,109} For this reason, corneal haze may be described and analyzed through back light scattering (backscatter).^{81,110} It also causes irregular astigmatism,^{2,34,93} and subsequent loss of corrected distance visual acuity (CDVA).⁸⁶

The regression of the refractive error may be produced by epithelial irregularity, alterations in the keratocyte density or subepithelial deposits. Myopic regression occurs in 78% of eyes in the first 12 months after PRK.² Table 3 shows the mean spherical equivalent changes reported in different scientific studies. In the first week after PRK, epithelial irregularity causes a reduction in visual quality.⁸⁸ During the first month, altered keratocytes decrease contrast sensitivity, mainly in high frequencies, and cause glare. During the next 2 months, subepithelial deposits produce a decrease in contrast sensitivity, especially in low frequencies.^{4,88} Ginis et al.⁴ reported that subepithelial deposits are the first factor that contributes to the development of corneal scatter. The visual quality is affected temporarily, although there is evidence that in some cases it persists for more than 1 year.^{43,91} In order to avoid a decrease in the visual quality, all postoperative efforts must go oriented to control the subepithelial matter.⁸⁸ The corneal epithelium does not seem to contribute significantly to the refractive change after PRK, although some studies suggest that epithelial thickening may produce myopic regression,² even 5 years after PRK.⁹⁰ Moller-Pedersen et al.⁵⁵ and Cua and Pepose⁹² suggested that new keratocytes growth in central cornea or postoperative corneal scarring is likely to be the main causes of myopic regression in ablations of 6 mm. In agreement with this hypothesis, Moller-Pedersen et al.⁵⁵ demonstrated that hyperopic changes were the direct result of a stromal thinning. Erie² found an increase of 12 μm of epithelial thickness at 12 months after PRK that was associated with a myopic regression of -0.41 diopters but no correlation was found between stromal thickening and myopic regression; however, the combined effect of epithelial and stromal thickening was correlated with myopic regression.

Regeneration of Corneal Innervation

The cornea is the most innervated tissue of the human body,⁷ and these sensory nerves are derived from the ophthalmic branch of the trigeminal nerve fibers.^{2,111} Corneal sensory nerves penetrate the limbus and form nerve bundles in the anterior third of the stroma. Once there, they run perpendicularly to cross Bowman's membrane, and form the sub-basal nerve plexus as a network between the basal epithelial cells and Bowman's layer (Fig. 2).^{49,111} Corneal nerve fibers, if visualized using confocal microscopy in normal conditions, show high reflectivity across the corneal stroma with a rectilinear pattern. Subepithelial nerve fibers, on the other hand, are thinner than stromal nerve fibers. Corneal fibers are considered primarily nociceptive (70%), followed by mechanosensitive fibers (20%).¹¹² In PRK, pho-

Table 3 Mean Spherical Equivalent After Surgery, Mean±SD or Range (Diopters).

Study	Technique	Preop	1 month	6 months	1 year	2 years	3 years	5 years	7 years
Einollahi et al. (2011) ⁶³	PRK+MD	-2.42±0.75 (-4.13 to -1.13)		-0.34±1.00					
	PRK+AAD	-2.38±0.72 (-4.00 to -1.25)		-0.28±0.91					
Wallau and Campos (2008) ¹⁵⁹	LASIK	-3.99±1.20 (-1.46 to -6.96)	0.49±0.52 (-0.50 to 1.50)						
	PRK+MMC ^a	-3.85±1.12 (-1.95 to -6.40)	0.61±0.61 (-0.50 to 2.88)						
Ghirlando et al. (2007) ³	PRK	-4.37±1.35	-0.37±0.61		-0.27±0.31				
	LASEK	-3.95±1.29	+0.22±0.79		-0.17±0.35				
Nassaralla et al. (2007) ²⁸	PRK+MMC ^b	-2.72±0.76 (-1.50 to -4.00)	-0.08±0.38 (-0.75 to -0.75)		-0.18±0.35 (-0.75 to -0.50)				
	LASIK	-6.5±2.5 (-11.0 to -2.0)	-0.1±0.5 (-1.0 to +1.0)		-0.2±0.5 (-1.25 to +0.75)	-0.2±0.4 (-1.0 to +0.62)	-0.2±0.4 (-1.0 to +0.75)	-0.2±0.5 (-1.37 to +0.37)	-0.4±0.5 (-1.25 to +0.25)
Patel et al. (2007) ²⁵	PRK	-3.7±1.4 (-5.75 to -1.25)	-0.1±0.3 (-0.5 to +1.0)		-0.3±0.3 (-0.87 to plano)	-0.4±0.4 (-1.25 to +0.25)	-0.3±0.2 (-0.75 to plano)	-0.6±0.4 (-1.25 to plano)	-0.5±0.4 (-1.0 to plano)
	LASIK								
Lee et al. (2005) ¹⁸	PRK	-5.17±1.53 (-2.00 to -9.13)	0.24±0.61	-0.46±1.01					
	tPRK	-5.11±.51 (-1.87 to -9.50)	0.59±0.78	0.18±0.91					
	LASEK	-5.26±2.58 (-1.50 to -9.50)	0.13±0.62	-0.82±1.18					
Kozak et al. (2003) ⁵⁶	LASIK	-6.00		-0.48±0.30 (-0.16 to -1.10)					
	PRK	-6.00		-0.67±0.35 (-0.21 to -1.21)					

PRK, photorefractive keratectomy; MD, mechanical debridement; AAD, alcohol-assisted debridement; LASIK, laser in situ keratomileusis; MMC, mitomycin C; LASEK, laser-assisted subepithelial keratectomy; tPRK, trans-PRK.

^a (0.002%, 1 min).

^b (0.02%, 2 min).

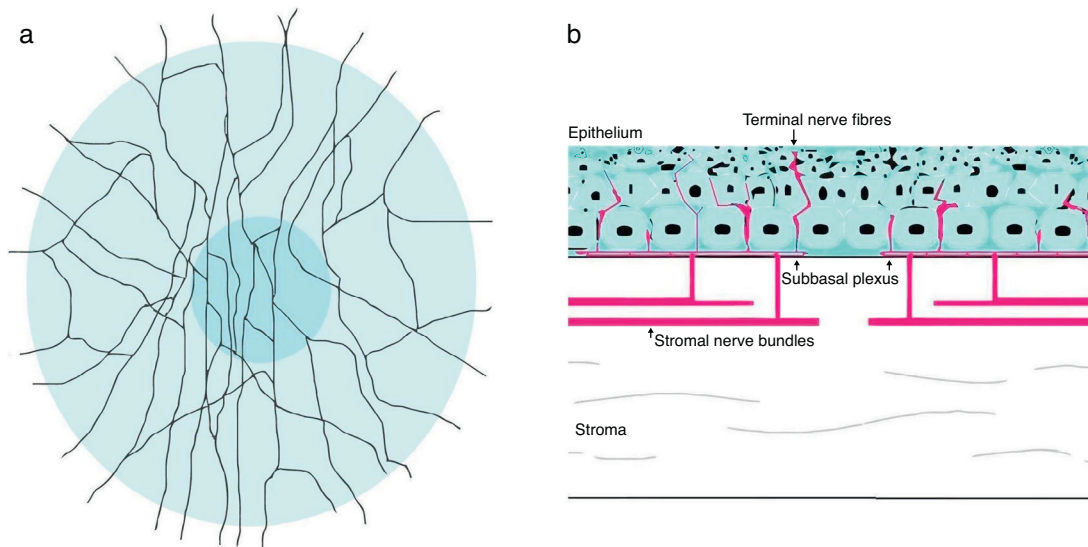


Figure 2 Schematic representation of corneal stromal nerves and subbasal plexus in human cornea. (A) Frontal view. (B) Cross section.

toablation severs nerves of the subbasal plexus and anterior stroma.^{2,83} It has been suggested that axotomy of corneal nerves might cause the decrease in keratocyte density after PRK,^{113,114} corneal nerves directly innervate keratocytes and provide trophic support in normal conditions.¹¹⁵

Animal studies have proved that the regeneration of the corneal nerves after PRK occurs as a biphasic process. In the first stage, a subbasal plexus originates from the cut end of subepithelial plexus, and the fine neurites run centrally with migrating cells.^{116,117} In the next phase, this transient plexus degenerates, and stromal originated nerves take place.¹¹⁷ Subbasal nervous plexus can have a significant influence on the regulation of epithelial healing.¹⁴ Substances like chemokines, proteases and neuropeptides are released after corneal injury,^{8,24,25,50,51,118} and it is postulated that neuropeptides like substance P (SP) and calcitonin-gene related peptide (CGRP) contribute to corneal wound healing.¹¹² Corneal nerves also influence the production of collagen type VII, necessary for the anchoring of the epithelium to the stroma.¹¹⁹ Conversely, injured epithelial cells release nerve growth factor (NGF) that stimulates nerve regeneration.

Approximately, at 8 weeks after PRK, sub-epithelial nerve fibers are visible on the edges. Erie,² using tandem scanning confocal microscope, visualized subbasal nerve fiber bundles in 17% of the corneas at 1 month after PRK. However, he noted that the density of these nerve fibers was 98% less than preoperatively.² After about 3 months of the surgery, no branched nerve fibers can be visualized in the center of the zone of ablation. Changes in subepithelial plexus and stromal trunks begin to appear 2–4 months postoperatively.⁸ At 6–8 months after the intervention of PRK subepithelial nerve regeneration is almost complete,^{8,111,120} although changes in the structure of the corneal nerves can be appreciated by confocal microscopy up to 12 months postoperatively.¹²⁰ However, nerve density continues to improve until 12 months after surgery, and returns to the preoperative values at 2 years.¹²¹ According

to Moilanen et al.¹²² in 71% of cases the central branching postoperatively was comparable to control subjects at 5 years ($P=.56$). Erie² proved that subbasal nerve density was reduced at 3, 6 and 12 months (87%, 75%, 60%, respectively) after PRK, and returned to preoperative levels at 24 and 36 months postoperatively. Subsequently, Erie et al.⁸³ in a prospective 5-year longitudinal clinical trial, proved with confocal microscopy that the recovery of subbasal nerve density in central cornea was faster in PRK than in LASIK. The authors observed that subbasal corneal density was reduced by 59% at 1 year after PRK ($2764 \pm 1321 \mu\text{m}/\text{mm}^2$) compared to preoperatively ($6786 \pm 1948 \mu\text{m}/\text{mm}^2$; $P < .001$). Sub-basal nervous plexus was almost recovered 2 years after PRK ($6242 \pm 1763 \mu\text{m}/\text{mm}^2$), and remained unchanged at 3 years ($6358 \pm 2447 \mu\text{m}/\text{mm}^2$) and 5 years ($5903 \pm 3086 \mu\text{m}/\text{mm}^2$).⁸³ In LASIK, they observed that subbasal corneal density was reduced by 34% at 3 years ($P < .001$),⁸³ with values at 5 years postoperatively comparable to those obtained preoperatively ($5903 \pm 3086 \mu\text{m}/\text{mm}^2$).⁸³ It is worth to note that in this study, the corneal flap was created using a mechanical microkeratome.⁸³ As the new technology allows making corneal flaps with laser instead of with a mechanical microkeratome, it is possible that studies in the near future report a faster corneal nerve recovery after LASIK.

When the process of corneal nerve regeneration finalizes, morphological abnormalities are often observed.^{8,83,122} According to Erie,² in the first 6 months after PRK the central subbasal nerves are organized in horizontal or oblique orientation. However, between 6 and 12 months, the subbasal nerve orientation rotates and comes to vertical orientation. In dry eye conditions, Esquenazi et al.²² observed active keratocytes, and they expressed nerve growth factor (NGF). NGF stimulates the proliferation of basal epithelial cells in normal conditions. Active keratocytes provoke an overexpression of NGF, which leads to abnormal findings in corneal nerves, such as hypertrophy.²² They also found higher nerve tortuosity, higher number of nerve beads, and the presence of nerve sprouts in desiccating environment group,²² which

means there is a high metabolic activity to repair the alterations in the corneal epithelium.

Corneal Pain and Sensitivity

Photoablation severs corneal nerves, disrupting the lacrimal functional unit (LFU). LFU is constituted by the lacrimal gland, ocular surface and innervation. It regulates tear secretion, and affects its composition.¹²³ Thereby, photoablation produces transitory dry eye, deterioration of corneal barrier function and alteration in corneal sensitivity.^{83,111,124} A reduction of the tear flow after PRK has been proved using Schirmer test.¹²⁴ According to Erie et al.,⁸³ LASIK presents higher prevalence of postoperative dry eye, altered corneal epithelium and tear film than PRK. Dry eye has been associated with low corneal sensitivity.^{125,126} Different devices are available to measure corneal sensitivity, as Cochet–Bonnet esthesiometry or non-contact gas esthesiometer. Cochet–Bonnet esthesiometer only stimulates mechanosensory fibers, whereas non-contact gas esthesiometer measures activation thresholds of nociceptors using controlled chemical, thermal and mechanical pulses. Non-contact gas esthesiometer is, therefore, a more sensitive device for measuring alterations in corneal sensitivity. Still, Coche–Bonnet esthesiometry is more widely used, and controversy remains about the time course of the corneal sensitivity recovery after PRK with this device. Kauffmann et al.¹²⁰ affirm that the recovery of corneal sensitivity usually starts at 4–6 weeks, completing approximately within 6–12 months following PRK. However, Erie et al.⁸³ claim that the recovery of corneal sensitivity is completed from 3 months to 1 year after PRK. Hypoesthesia is often expected until 3 months after surgery, due to the loss of corneal nerves.⁸ On the other hand, Gallar et al.¹²⁷ measured corneal mechanical and chemical sensitivity following PRK with non-contact gas esthesiometer, and found that both types of sensitivities were reduced even 5 years postoperatively, achieving normal values in 10 years. Despite the diminished corneal sensitivity, intense pain is usually present hours after PRK.¹²⁸ Gallar et al.¹²⁹ attributed corneal pain and discomfort sensations to the altered functionality of corneal nerves. They recorded spontaneous activity and modified responsiveness in corneal fibers of cats that underwent PRK.¹²⁹ Experimental evidences support the idea that ongoing activity evokes spontaneous pain sensations.^{130,131}

Acceleration of Corneal Regeneration Process, Reduction of Corneal Haze and Corneal Pain Management

Nowadays there are several alternatives to speed up the process of epithelial regeneration, like epithelial removal techniques, amniotic membrane, or bandage contact lenses. In PRK, agents like mytomicin-C (MMC) or fluoroquinones that reduce the corneal haze formation are used, and drugs to reduce the corneal pain and inflammation are also prescribed.

Epithelial Removal Techniques

In PRK, previous to the impact of laser energy over the cornea, the corneal epithelium has to be removed. The removal of the corneal epithelium is carried out mainly with epithelial mechanical scraping using chemical agents like diluted ethanol solution,⁹ through a rotary brush or using the laser itself – known as transepithelial ablation (Fig. 3).^{18,21,22,39,63,128,132,133} The epithelial scraping has post-operative adverse effects like pain, myopic regression or corneal haze. Some modification in PRK technique can alter the wound healing response with the aim of minimizing the adverse effects.²⁵ The exposition to agents such as ethanol can produce an increase in the inflammatory response and more damage to the anterior stromal keratocytes that could increase the haze formation.^{21,94} Yet, controversy remains in the scientific literature because other authors affirm that alcohol-assisted epithelial removal produces less inflammation, favoring epithelial regeneration and preventing corneal haze or keratocyte apoptosis.^{9,63} Esque-nazi et al.²² proved that the epithelial scraping might be associated with an increase in the number of reflective structures in the stroma, mainly in corneas with ocular dryness after PRK. The laser-scrape epithelial removal decreases the degree of keratocyte apoptosis, producing a less pronounced loss of superficial keratocytes.² However, the irrigation with cold balanced salt solution (BSS) may alter the keratocyte apoptosis in the retroablation zone.² The time necessary for mechanical debridement is greater than the time required for laser or alcohol scrape techniques, even for expert surgeons.¹⁸ Mechanical debridement is related to stromal dehydration and disappearance of anterior stromal keratocytes.^{18,63} This loss provokes an increase of cells in the underlying stroma, causing stromal hyperplasia and haze formation.¹³⁴ Einollahi et al.⁶³ found faster mean epithelial healing time in the alcohol-assisted group than in the mechanical group (3.0 ± 0.3 versus 3.2 ± 0.4 days, $P = .001$). They observed greater anterior retroablation stromal keratocyte density in the mechanical group than in the alcohol-assisted groups at 3 months (704.3 ± 119.9 cells/mm² versus 743.3 ± 103.7 cells/mm², $P = .05$) and at 6 months (643.8 ± 134.4 cells/mm² versus 696.7 ± 129.6 cells/mm², $P = .02$).⁶³ In the same study, Bahram et al. did not find statistically significant differences in middle and posterior keratocyte density between the mechanical and alcohol-assisted groups.⁶³ They also proved that mechanical and alcohol-assisted epithelial debridement after PRK present similar visual and refractive outcomes in patients with mild myopia,⁶³ in agreement with the results of Goreishi et al.¹³⁵ They reported similar safety and efficacy with alcohol-assisted and mechanical debridement in a 1250 eye sample, but anterior keratocyte density was not assessed in this study.¹³⁵

Laser-assisted subepithelial keratomileusis (LASEK) was developed in order to reduce corneal pain and haze formation associated with PRK, and to accelerate visual recovery. Epithelial delamination with diluted alcohol showed in an electron microscope study that was able to leave a smooth surface, ideal for LASEK intervention.¹³⁶ It seems that a regular surface before laser application helps corneal healing and prevents haze.¹³⁷ Chen et al.¹³⁸ contrasted these findings in a later study, and showed a high variability



Figure 3 TransPRK (transepithelial photorefractive keratectomy) on the Schwind Amaris 1050 RS laser platform (with permission of Schwind Eye-Tech-Solutions).

in morphological changes after diluted alcohol treatment, dependent upon concentration and time. Cell viability was affected when alcohol exceeded its concentration by 25% or 25-s exposure.¹³⁸ Yet, these studies have been conducted in vitro, and the complex interactions of tear film and corneal surface were not considered. In vivo studies do not show any difference between LASEK and PRK.¹³⁹ Lee et al.¹⁸ evaluated epithelial healing, postoperative pain and visual outcomes using epithelial mechanical (conventional PRK), transepithelial PRK and 20% diluted alcohol laser-assisted subepithelial keratomileusis (LASEK) with flap repositioning. After 6 months, they found little differences in clinical outcomes between the 3 techniques, noting a slight overcorrection in the transepithelial PRK and slight undercorrection in LASEK. Corneal pain and subepithelial haze results were similar.¹⁸ Subsequently, Ghanem et al.¹³⁹ proved in a prospective randomized double-masked study that the reepithelialization was faster in a PRK group compared with a butterfly LASEK group, even though epithelial semi-discs were repositioned intraoperatively in LASEK group. (4.35 ± 0.48 days versus 4.75 ± 0.72 days, $P = .002$). They also found lower pain level in PRK group, but pain scores and ocular discomfort were not statistically different from butterfly LASEK (3.31 ± 4.09 versus 4.43 ± 4.27 ; $P = .18$).¹³⁹

It has been proven in animal studies that transepithelial ablation produces a uniform surface for corneal regeneration, and prevents keratocyte apoptosis,³⁶ reducing the risk of corneal haze.^{21,72} Wang et al.¹⁴⁰ presented promising preliminary results of SCHWIND-ESIRIS excimer laser for transepithelial ablation, but the flawed design of the study makes difficult to assess the real value of this technique. Later, Aslanides et al.²¹ proved in humans that transepithelial ablation was safer than the epithelial

mechanical scraping using chemical agents as alcohol, as it provides a faster epithelial healing, less postoperative pain and less corneal haze at 1 week ($P = .07$), and at 1, 3, and 6 months after surgery ($P < .05$). In addition, they observed an improvement of 3 Snellen lines in visual acuity on day 3 in the modified transepithelial PRK (all-surface laser ablation) group compared to conventional alcohol-assisted PRK group (0.4 versus 0.2; $P < .05$).²¹ Transepithelial ablation also resulted in better corrected distance visual acuity (DCVA) than conventional alcohol-assisted PRK (33% versus 13%, respectively, $P > .05$),²¹ although differences in higher order aberrations were not statistically significant.²¹

Amniotic Membrane Transplantation

Apart from the above mentioned techniques, amniotic membrane transplantation reduces the inflammation after PRK, prevents polymorphonuclear cell infiltration, produces less peroxidation, avoids keratocyte apoptosis and stimulates corneal epithelialization.^{37,77} It is usually combined with PRK to treat corneal dystrophies, corneal degenerations, scars, keratopathies,^{141,142} or even to treat corneal haze secondary to PRK.¹⁴³ The amniotic membrane restricts the influx of polymorphonuclear cells (PMC) to the patch.^{144,145} PMCs adhere to the amniotic membrane and eventually commit apoptosis.¹⁴⁶ This is a physiological way of suppressing corneal inflammation.¹⁴⁷ In addition, amniotic membrane has intrinsic keratocyte growth factors, EGF and neurotrophins that promote epithelialization.^{148,149} It also suppresses TGF-beta1, collagen III and fibronectin.¹⁵⁰ Taken together, amniotic membrane has a potent anti-scarring effect that reduces corneal haze formation, as demonstrated in animal studies.^{144,151}

Agents to Enhance Wound Healing

The wound healing response may be altered by the prophylactic application of a topical solution of mitomycin-C (MMC) immediately after the laser ablation,²⁷ in order to avoid or minimize myofibroblast activation.^{8,25,40,71,86,90,133,152-154} MMC is an antineoplastic antibiotic agent of the family of anti-tumor quinolones and derived from *Streptomyces caespitosus*. It is a potent DNA crosslinker: it inhibits the replication of deoxyribonucleic acid (DNA).^{28,32,91,93,155-159} Thereby, MMC inhibit cell mitosis, including epithelial and stromal cells.^{8,34,86,93,133,155,157,160,161} Mitomycin-C decreases corneal haze compared to corticosteroid treatment,⁸² and, consequently, improves visual acuity.¹⁵² Its use is specially indicated in high myopia (≥ -6.00 D) and deeper ablation depths ($\geq 75 \mu\text{m}$).^{82,86,90,133,155,160} Wallau and Campos¹⁶² obtained better UCVA and BSCVA with the combination of PRK with MMC, than with LASIK ($P=.027$ and $P<.001$, respectively) at 3, 6 and 12 months after surgery. Goreishi et al.¹⁶³ reported an incidence of 4% of corneal haze at 1 year post-operatively with intraoperative application of 0.02% MMC, in a sample with a mean refractive error of -5 D. Fazel et al.¹⁶⁴ found that two-step administration of 0.02% MMC (45 s, followed by 15 s) further decreased corneal haze formation in high myopia, compared to a single dose of 45 s. The benefits of MMC have also been described once the haze has been established, where mechanical epithelial scraping and instillation of MMC restores corneal transparency.¹⁶⁵

Although the application of the mitomycin C is helpful for corneal recovery, it is necessary to control the doses and the time of exposure.²⁰ According to Thornton et al.¹³³ the concentration is a more important factor than the duration of MMC exposure in corneal haze prevention. Rajan et al.³⁴ analyzed the effects of MMC after correction of -9.00 diopters by PRK in 3 groups of human corneas: without MMC application, with MMC (0.2 mg/mL) application for 1 min and with MMC (0.2 mg/mL) application for 2 min. The 2 min MMC group (0.2 mg/mL) had thinner epithelium than the 1 min and without MMC application groups ($P<.0001$). The application of the intraoperative MMC lasts between 10 s and 120 s, depending on the surgeon.¹¹ According to Khoury et al.¹⁵⁶ the application of intraoperative MMC vary from 12 s to 5 min. Shojaei et al.¹⁵³ affirm that short-time MMC exposure prevents low-grade haze in low ablation depths. The MMC doses oscillate between 0.002% and 0.06%.¹⁵⁶ The intraoperative application of 0.02% MMC solution is the most recommended, as it produces less corneal haze, and provides better uncorrected visual acuity (UCVA) and best spectacle-corrected visual acuity (BSCVA).^{124,166,167} Still, Ramjoo et al.⁹⁰ found similar refractive and haze outcomes with 0.01% and 0.02% MMC for mild myopia, recommending the use of 0.01%. The lowest dose available is recommended to avoid side effects. Rajan et al.³⁴ observed a delay in keratocyte regeneration after MMC application ($P<.0005$). Midena et al.⁸² proved by means of confocal microscopy that the application of 0.02% MMC produced a considerable decrease of anterior stromal keratocytes, but there is no evidence of this decline in the posterior stromal keratocytes. Subsequently, Thornton et al.¹³³ observed a keratocyte loss in the anterior stroma 1 month and 6 months after PRK with standard MMC concentrations (0.02%). Razmjoo et al.⁹⁰ did not found significant

reduction in keratocyte density after application of 0.02% mitomycin C (MMC). The dose of MMC is associated with the grade of refractive error. Thornton et al.¹³³ believe that for high myopia corrections (>-6.00 D) standard concentration of topical MMC (0.02%) may be used, whereas for moderate myopia (-3.00 to -5.90 D) low dose of MMC (0.002%) may be considered, although it seems that intermediate dose of MMC (0.02%) is more effective than 0.002% for moderate myopia.

The cytotoxicity of MMC increases with cumulative doses,¹⁶¹ and when MMC is combined with ethanol, which increments the apoptosis of keratocytes.²⁰ Few complications have been associated with its use with the exception of a decrease in the short term of the keratocyte density.^{71,155,157} However, some complications have been documented at the time of instillation or after some weeks. Although unusual, scleral ulceration, non-healing conjunctivas and complications associated with high MMC doses (0.04%) or prolonged postoperative topical use may appear,⁸⁶ because high doses of MMC suppress cellular RNA replication and protein synthesis.⁹³ As MMC is applied in the stromal bed, it seems that it might penetrate into the anterior chamber, because cytotoxic effects on the ciliary body epithelium have been reported.^{11,161} There is controversy in the scientific literature, but MMC does not seem to cause any alteration in the ciliary body or intraocular pressure (IOP) after PRK.¹⁶¹ Kymionis et al.¹⁶¹ investigated the effects of MMC after PRK in 40 eyes of 20 rabbits. They applied 0.02% MMC for 2 min in one eye, and balanced salt solution (BSS) for 2 min in the contralateral eye. After 3 months, they did not found differences in the morphology of the ciliary body, and tonometric measurements remained stable ($P=.075$).

The endothelium is the inner layer of the cornea. Endothelial cells have a hexagonal or polygonal shape,⁴⁸ and they are homogeneously distributed, without signs of polymegatism and pleomorphism in normal conditions. Endothelial cells are not able to regenerate,^{40,157} and a reduction in the number of cells is seen with age. After PRK, endothelial structure, shape and density remain unaltered.^{81,85,168} Table 4 shows the variation of endothelial cells in the different studies published in the scientific literature. Polymegatism or pleomorphism, if present, may be secondary to still unknown corneal metabolism.¹⁶⁹ There is also controversy about the toxic effect of MMC in the overall morphology of the endothelium.^{8,11,86,153} Morales et al.¹⁵⁸ proved that intraoperative 0.02% MMC during 30 s after PRK induced corneal endothelial cell loss at 1 month and 3 months ($P=.0006$, $P=.002$; respectively). Diakonis et al.¹¹ applied Mitomycin C (MMC) for 15 s and the density of endothelial cells was not affected. Zare et al.¹⁷⁰ obtained similar results when 0.02% MMC was applied for 45 s. Subsequently, Shojaei et al.¹⁵³ found significant differences of mean endothelial cell densities in the MMC group and in the control group at 6 months after surgery (2878.79 ± 283.04 cells/mm² versus 2826.19 ± 286.25 cells/mm², $P=.25$). Undoubtedly, after the application of MMC the DNA of endothelial cells gets damaged.¹⁷¹ It remains to be determined the long-term effects of such event. According to Wilson,⁷¹ long-term studies (more than 10 years) are necessary to determine the adverse effects of MMC.

Table 4 Variation of Endothelial cell density After Surgery, Mean±SD or Range (cell/mm²).

Study	Technique	Preoperatively	1 month	3 months	6 months	12 months
Shojaei et al. (2013) ¹⁵³	PRK with MMC	2879±298		2849±296	2878 ± 283	
Einollahi et al. (2011) ⁶³	PRK with BSS	2819±303		2825±283	2826 ± 286	
	PRK with MD	3102±281 (2498–3823)		2996±259	2795 ± 764	
Amoozadeh et al. (2009) ⁴⁰	PRK with AAD	3125±299 (2610–4276)		3011±240	2946 ± 240	
	LASIK	3022±224			3030 ± 186	
Wallau and Campos (2008) ¹⁵⁹	PRK	2983±293			3025 ± 404	
	LASIK	2709±242			2667 ± 277	
Diakonis et al. (2007) ¹¹	PRK with MMC ^b	2709±246			2686 ± 253	
Nassaralla et al. (2007) ²⁸	PRK+MMC ^e	2757±117	2736 ± 144	2729±131	2716 ± 136	2721±113
	Epi-LASIK	2769±158	2727 ± 179	2741±177	2758 ± 176	2760±102
Morales et al. (2006) ¹⁵⁸	PRK with MMC ^{a,c}	2150±180 (1800–2650)		2100±205 (1680–2540)		2200±210 (1680–2500)
	PRK+MMC ^d	2835±395	2416 ± 291	2357±404		
	PRK+BSS	2779±492	2711 ± 555	2746±526		

PRK, photorefractive keratectomy; MMC, mitomycin C; BSS, balanced saline solution; MD, mechanical debridement; AAD, alcohol-assisted debridement; LASIK, laser in situ keratomileusis; Epi-LASIK, epipolis laser in situ keratomileusis.

^a After radial keratotomy.

^b 0.002%, 1 min

^c 0.02%, 2 min.

^d 0.02%, 30 s.

^e 0.02%, 15 s.

New generation quinolones, instead of preventing corneal haze, are used as prophylactic antibiotics to avoid corneal infections after refractive surgery.¹⁷² They also enhance the rate of corneal recovery. Fourth generation fluoroquinolones like gatifloxacin (Zymar, Allergan, Irvine, California) and moxifloxacin (Vigamox, Alcon Laboratories, Fort Worth, Texas) have been demonstrated to mediate faster corneal healing,¹⁷² without evident differences between both of them in terms of visual outcomes.¹¹⁸

Bandage Contact Lenses

After PRK, the corneal surface needs between 2 and 4 days to regenerate,² and the vision may fluctuate for several weeks to months. If epithelial regeneration delays, the subepithelial haze increases; for this reason, an appropriate corneal reepithelization is crucial.^{8,105} Reepithelization is the first step during corneal regeneration after PRK.⁵¹ If the reepithelialization is facilitated with the appropriate contact lenses, visual acuity improves.^{30,173} Although therapeutic contact lenses have been used for more than 40 years, PRK has increased their popularity.^{23,30} One of the major disadvantages of PRK is the pain and discomfort during 1–3 days after intervention.^{7,15,174} To ease off the postoperative pain and discomfort, and to promote

epithelial healing, bandage contact lenses are fitted for 3–5 days after surgery.^{12,23,30,31} Other techniques and medications has been proposed in order to reduce corneal pain like occlusive pressure patching, but the bandage contact lenses are still the gold standard.¹⁷³ Bandage contact lenses are used to protect the epithelium from the eyelid, to reduce the haze formation,^{31,173} to enhance epithelial healing, to control the sensation of pain, and to prevent epithelial erosions.^{12,23,30,128} Faster reepithelialization produces a reduction of discomfort, facilitates visual recovery, and restores the corneal barrier to prevent infections.¹²

Because of the prolonged use of therapeutic contact lenses, and to assure the proper corneal metabolism, a high oxygen permeability (Dk/t) contact lens are used.^{23,30,31,173} Silicone hydrogel contact lenses have a Dk/t coefficient 5–10-fold greater than conventional hydrogel lenses.¹² For this reason, silicone hydrogel bandage contact lenses are widely fitted,^{7,12,30,31} and are the ones approved by the FDA for prolonged use after PRK. Currently, a variety of contact lenses are used as therapeutic soft contact lenses after PRK like Lotrafilcon A (Focus Night & Day, Ciba Vision), Lotrafilcon B (O 2 Optix, Ciba Vision), Senofilcon A (Acuvue Oasys, Vistakon Inc.), Balafilcon A, Omafilcon A (Proclear, Cooper Vision) and Senofilcon A.^{12,23,30,31} Lotrafilcon B is approved by FDA for 6 days of continuous wear and Senofilcon A for 1 week of continuous wear, while Lotrafilcon A

is approved for 30 days of continuous wear and therapeutic use.^{12,23} The therapeutic efficacy of the Lotrafilcon A after PRK has been intensively studied,^{12,23,30,31} and reduction of discomfort and faster corneal reepithelialization in 48 h have been described.^{12,23} Edwards et al.³¹ proved that Lotrafilcon A showed better best spectacle-correction visual acuity (BSCVA) than Omafilcon A, without statistically significant differences in contrast sensitivity or uncorrected visual acuity (UVA). Omafilcon A reduced the BSCVA in 40.4% of patients at 1 month, whereas Lotrafilcon A reduced the BSCVA in 18.6% of the patients ($P=.002$). The corneal pain was greater with Omafilcon A than with Lotrafilcon A at 1 day ($P=.000$) and 4 days postoperatively ($P=.027$).³¹ In contrast, an increase in corneal infiltrates with Lotrafilcon A was observed compared to Omafilcon A, and there was not a statistically significant difference in reepithelialization.³¹ The authors suggested that corneal infiltrates might be a consequence of Lotrafilcon A's rigidity due to its reduced water content (24%) versus 59% of Omafilcon A.³¹ Subsequently, Razmjoo et al.³⁰ in a comparative study, found that the 58.3% of the eyes with Senofilcon A and 41.7% of the eyes with Lotrafilcon A completed the reepithelialization at day 5 ($P>.05$). Although there were not statistically significant differences in the rate of corneal reepithelialization between both contact lenses ($P>.05$), and the postoperative pain and discomfort index was significantly lower in Senofilcon A group ($P<.05$).³⁰ They also compared the visual acuity between Senofilcon A and Lotrafilcon A after PRK, and proved that in both groups the UCVA was worse at 3 days than at day 1. However, the UCVA improved at day 5, with 97.7% reaching UCVA of 20/40. A feasible explanation is that, at day 3, the epithelial healing process is located in the center of the cornea.³⁰ As only 44 patients were included, in future studies a larger size sample would be recommendable.

Bandage contact lenses also minimize corneal haze. Edwards et al.³¹ showed a minimum tendency to a high level of corneal haze with Omafilcon A compared with Lotrafilcon A ($P=.0064$). However, all efforts are made to minimize the corneal haze intraoperatively, using cold balanced saline (BSS) and MMC. Application of BSS in the stromal body reduces the corneal pain and corneal haze,¹²⁸ yet, the application of mitomycin-C (MMC) is more widely used.

Although bandage contact lenses have various advantages, the presence of silicone may produce irritation, increased protein and lipid deposits, and reduced wettability because of its hydrophobicity.³¹ A plasma treatment is given to enhance the hydrophilicity of Lotrafilcon A surface, but this technique is not completely effective.³¹ Bacterial keratitis and subepithelial infiltrates have been described with bandage contact lenses after PRK.¹⁷ The risk of infectious keratitis of soft contact lenses fitted for approximately 3 days is low, and antibiotics are prescribed to further minimize the risk.¹⁷⁵

Corticosteroids and Non-steroidal Anti-inflammatory Agents (NSAIDs) Therapy

It is necessary to distinguish between corneal haze that appears in the first weeks or months after PRK and pathological corneal haze that appears as a result of myofibroblasts.⁷¹

If the corneal haze persists over time, it may cause a corneal opacity and the thickening of the tissue that would result in a regression of the refractive error, decreased visual acuity and irregular astigmatism.^{19,34,67,71} Clinically significant corneal haze occurs in 0.5%–5% of the cases.¹⁰⁹ Corneal haze that most commonly occurs after PRK is not clinically significant, and is not attributed to myofibroblasts.^{71,121} According to Wilson,⁷¹ in human corneas that develop late corneal haze after PRK, the resolution of the opacity is slow, and the restoration of the refractive correction is produced between 1 and 3 years postoperatively. It has been postulated that the extinction of corneal haze can be influenced by the disappearance of myofibroblasts, reabsorption of abnormal extracellular matrix (ECM) and restoration of normal corneal structure.⁷¹

After surgery, a variety of drugs are prescribed to avoid corneal haze, for instance, corticosteroids – antiinflammatories to avoid the pain and inflammation-, plasmin inhibitors, growth factors or antimetabolites.^{13,176} Topical therapy after PRK prevents complications like keratitis, infections or corneal haze.¹⁷⁷ The most common treatment after PRK to avoid the corneal inflammation is the application of corticosteroids.^{109,158} Corticosteroids are not recommended for long periods because of their side effects, like intraocular pressure (IOP) rise and the risk of cataracts.^{109,178,179} Javadi et al.¹⁸⁰ reported a rise in the IOP using 0.1% betamethasone at 2 weeks post-PRK in a minority of patients. Furthermore, corticosteroids delay epithelial healing.¹⁷⁹ When corneal haze appears 2–3 months after PRK, the clinical observations confirm that haze is "corticosteroid-responsive" in 10%–15% of patients.⁷¹ Researchers disagree about the benefit of corticosteroids to reduce the corneal haze after PRK.^{2,177} According to Wilson,⁷¹ the topical administration of 1% prednisolone acetate (Pred Forte) quickly removes the corneal opacity and produces a change in refractive error. In the remaining 85% or 90% of cases, the corticosteroids do not exert any change.⁷¹ Corticosteroids could be replaced by non-steroidal anti-inflammatory agents (NSAIDs), tranilast, cysteine or antioxidants like Vitamine E.^{109,177} NSAIDs are effective in reducing corneal pain, postoperative photophobia and inflammation.^{7,128} The inflammatory response is mediated by prostaglandins synthesized from arachidonic acid by cyclooxygenase 1 (COX-1) or cyclooxygenase 2 (COX-2).¹²⁸ The antiinflammatory and analgesic properties of the nonsteroidal anti-inflammatory drugs (NSAIDs) are achieved by the inhibition of COXs activity.^{7,128}

The use of certain steroidal and non-steroidal anti-inflammatory drugs (NSAID) delay reepithelialization and increase the risk of haze formation,⁷ although the results are still contradictory. Vetrugno et al.¹⁷⁷ proved that 0.1% fluorometholone acetate administered in the first day after PRK reduced corneal haze and myopic regression, particularly in high myopic patients. NSAIDs like diclofenac and ketorolac have shown reduction in the pain sensation,^{7,128} but also a significant delay in corneal reepithelialization after PRK.¹⁸¹ Nepafenac (Nevanac; Alcon Laboratories Inc., Ft Worth, Tex) is a new topical NSAID with greater corneal permeability that has been approved for the treatment of inflammation after surgery.^{7,181} Jalali et al.¹³ found that 0.1% Nepafenac did not increase haze formation, neither hamper corneal epithelial healing, but they did

not found statistically significant differences in corneal reepithelialization between nepafenac and non-nepafenac groups ($P=.61$). Caldwell et al.,¹⁸¹ in a randomized double-masked study, demonstrated that 0.1% nepafenac was safe for corneal reepithelialization, and reduced the post-operative pain at day 1 (0.76 versus 1.68) and day 2 (1.26 versus 2.23) compared with the placebo group ($P<.0005$). Other NSAIDs for corneal pain reduction are also available, like Bromfenac, Flurbiprofen sodium and Indomethacin.⁷

Despite the presence of complications is low, non-steroidal anti-inflammatory drugs (NSAIDs) may produce conjunctival hyperemia, transient burning, stinging, superficial punctate keratitis, epithelial defects, subepithelial infiltrates, corneal melting and perforation.^{7,17} However, Caldwell et al.,¹⁸¹ in a randomized double-masked study, proved that 0.1% nepafenac did not have adverse effects. Postoperative oral analgesics, like NSAIDs, are able to produce gastrointestinal, cardiovascular, respiratory and central nervous system complications.^{17,181} Another treatment that is widespread for the inhibition of inflammation and for treatment of dry eye is the Cyclosporine A, with doses from 0.05% to 2.00%.¹⁰⁹ Nien et al.¹⁰⁹ used Cyclosporine A 0.05% and prednisolone acetate 0.01% to compare the effect in corneal haze prevention in rabbit corneas following PRK. They concluded that Cyclosporine A did not have any effect, whereas prednisolone acetate was effective in reducing short-term corneal haze, but did not prevent corneal fibrosis.¹⁰⁹

Alternative Therapies for Corneal Haze Prevention

The use of drugs does not completely suppress corneal haze formation after PRK. Research has focused on new therapies that could prevent corneal haze, like genetic evaluation of type IV collages synthesis.³³ Lumican and keratocan genes have also been evaluated for management of subepithelial persistent corneal haze after PRK, but without a consistent finding.¹⁸² It has been postulated that vitamin E, probucol or heparin may inhibit collagen type IV synthesis, but they have not been approved for topic use because of their adverse effects.³³ Vitamin E and amino acids play an important role in corneal reepithelialization and in the prevention of corneal haze and keratocyte apoptosis, especially in high myopia.^{9,77} A preliminary clinical trial concluded that oral supplementation with vitamin A and vitamin E accelerated the reepithelialization, and reduced corneal haze formation,¹⁸³ but it seems that the topical administration of vitamin A alone do not have any effect.¹⁸⁴ Alternative treatments to MMC that prevent corneal haze formation, but produce less damage to keratocyte are bevacizumab and rapamycin.¹⁸⁵ Subconjunctival injection of PRM-151 could presumably prevent corneal haze, as it inhibits the pro-fibrotic myofibroblast differentiation.¹⁸⁶ Trichostatin A, similarly, prevents myofibroblast formation by inhibiting TGF-beta1.¹⁸⁷ As cytokines and growth factors control the synthesis of collagen type IV, they might be also useful treatments for corneal haze prevention.³³ PRK increases the release of leukocytes, TGF- β 1, TNF- α and PDGF-BB in human tears during the first days of wound healing.^{14,50} TGF is a cytokine released by

the lacrimal gland, corneal epithelium and conjunctival cells.¹⁷⁹ Three forms of TGF- β exist (TGF- β 1, TGF- β 2 and TGF- β 3) and each one is involved in the wound healing process in a different way. TGF- β 1 is increased in early epithelial healing, and exerts an influence in the subepithelial fibrosis formation and activation of keratocytes after PRK.⁷³ Bühren et al.¹⁷⁹ proved that the application of anti-TGF- β in felines reduced the differentiation in vitro of keratocytes into myofibroblast, and corneal haze diminished. They suggested that this reduction in differentiation improved optical quality. The combination of the nerve growth factor (NGF) and docosahexanoic acid stimulated the regeneration of basal epithelial cells in rabbits after PRK,²² which is imperative for a proper wound healing. Medduri et al.³⁵ studied the effect of basic fibroblast growth factor (b-FGF) in circumstances of delayed healing after PRK.⁹ 50 patients were enrolled in b-FGF eye drop treatment group and 50 patients in saline drops (placebo) group. They observed greater corneal epithelial healing in the b-FGF group than in the placebo group at 4 days (98% versus 72%, respectively) and 5 days after surgery (100% versus 92%, respectively).³⁵

Artificial tears are the most widely used solution for corneal lubrication. However, they do not have biological components that promote corneal regeneration. In fact, they contain stabilizers, preservatives, or other additives that may induce toxic or allergic reactions.⁵¹ Blood derivatives as plasma rich in growth factors are an alternative to artificial tears, and have not possibility of rejection.⁵¹ Anitua et al.⁵¹ proved that plasma rich in growth factors obtained from patient's blood enhanced corneal healing, and reduced the formation of corneal haze. The difference between the plasma rich in growth factors group (PRGF-Endoret treatment) and control group was negligible at day 3. They attributed it to the increase of proliferative cells (Ki-67 β) in the control group. They suggested that the increase in proliferative cells could be associated with epithelial hyperplasia observed at day 3 and 7 after PRK in control group.⁵¹ They also found that the epithelium of the PRGF-Endoret group was formed by 5–6 layers.⁵¹

Corneal Nerve Regeneration and Neuropathic Corneal Pain Management

The regeneration of the corneal nerves after PRK is associated with the improvement of cellular integrity.²² To date, few therapeutic treatments have been developed for nerve regeneration. Javaloy et al.¹⁸⁸ investigated the benefits of topical platelet-rich plasma, but subbasal nerve density did not improve after 3 months of treatment compared to controls ($P=.66$). Studies in animal models have demonstrated more encouraging results. Esquenazi et al.²² studied the outcomes of the combination of nerve growth factor (NGF) and docosahexanoic acid (DHA) in rabbits in promoting corneal nerve regeneration. They observed that this combination increased corneal nerve regeneration, as well as epithelial proliferation and decreased rose bengal staining compared to the application of NGF or DHA alone. Cortina et al.¹⁸⁹ showed similar results with pigment epithelial-derived factor (PEDF) plus docosahexanoic acid (DHA).

Moreover, this combination proved to enhance corneal sensitivity. Recent evidences suggest that peripheral nervous system regeneration and inflammatory processes share common pathways, and some degree of inflammation is required for neuroregeneration.¹⁹⁰ Therefore, cyclosporine A and corticosteroid treatments could interfere in a proper nervous recovery.

When corneal nerve regeneration process fails, corneal neuropathic pain might take place. The up- and down-regulation of ion channels in axotomized nerves can change the excitability of fibers,¹⁹¹ and produce spontaneous discharges and altered sensibility to exogenous stimuli.¹⁹² This would result in corneal pain non-treatable with aforementioned drugs. Anticonvulsants, opiates and topical local anesthetics can manage corneal neuropathic pain. The anticonvulsant Gabapentin (Neurontin) is an analog of gamma-aminobutyric acid (GABA),¹⁹³ and its reliability in treating corneal pain is conflicting, mainly because of a lack of studies.⁷ Lichtinger et al.¹⁹³ compared in a prospective randomized, double-blind, placebo-controlled study the efficacy of Gabapentin in the reduction of the corneal pain. They administrated gabapentin capsules (300 mg) in 20 patients and additional 20 patients received identical placebo capsules. They demonstrated that gabapentin reduced corneal pain during the first 24 h ($P=.003$), at postoperative day 1 ($P=.002$), between 24 and 48 h ($P=.024$), at postoperative day 2 ($P=.018$) and between 48 and 72 h ($P=.001$). Faktorovich et al.¹⁷ trying to prove the efficacy of topical opioid in the treatment of pain, concluded in a double-blind randomized prospective study that the administration of 0.5% morphine drops was an effective and safe method to control of post-PRK pain, and did not hamper epithelial healing or refractive outcomes. Topical local anesthetics include tetracaine, proparacaine, lidocaine and bupivacaine can be also used. Topical tetracaine has been documented to be successful in pain control management and does not produce delayed corneal healing times.^{7,39} However, it produces keratocyte toxicity and keratitis.^{7,17,39} Topical anesthetics should be used cautiously and for short-term treatments. Antidepressants are prescribed for neuropathic pain management elsewhere in the body. To date, no study has been published evaluating the effect of antidepressants for treating post-PRK corneal pain.

Conclusions

Photorefractive keratectomy disrupts corneal structure affecting epithelium, Bowman's membrane, and anterior stroma. Corneal nerves are severed, which alters corneal integrity and function temporarily. The subsequent corneal wound healing is a complex process that is regulated by a variety of factors. A balance of peptides will determine the final outcome, and the presence of postoperative complications. Corneal wound healing process can be managed with several drugs to enhance regeneration and prevent corneal haze and pain after PRK. Further research in this field is required to completely understand post-PRK corneal regeneration in order to prevent complications, and provide outstanding visual outcomes.

Directions for Future Research

This review clearly states that corneal regeneration after PRK is not completely understood. The ongoing research in new drugs development, more efficient surgical techniques, and new imaging technologies are trying to answer some of the unresolved questions. Still, future research should be oriented to elucidate the following aspects:

- The long-term effects of keratocyte death and MMC application.
- Although corneal haze has been correlated to several factors, its origin is still unknown.
- The beneficial role of corticosteroid administration in corneal haze prevention.
- The causal factors of myopic regression.
- More studies using non-contact gas esthesiometer will help to better assess the time course of corneal sensitivity recovery.

Acknowledgements

Authors would like to thank Manuel Alejandro Amaya Alcaraz for the collaboration given in the revision of this article.

References

1. L'Esperance FA Jr, Taylor DM, Del Pero RA, Roberts A, Gigstad J, Stokes MT, et al. Human excimer laser corneal surgery: preliminary report. *Trans Am Ophthalmol Soc.* 1988;86:208–275.
2. Erie JC. Corneal wound healing after photorefractive keratectomy: a 3-year confocal microscopy study. *Trans Am Ophthalmol Soc.* 2003;101:293–333.
3. Ghirlando A, Gambato C, Midena E. LASEK and photorefractive keratectomy for myopia: clinical and confocal microscopy comparison. *J Refract Surg.* 2007;23:694–702.
4. Ginis H, Pentari I, de Brouwere D, Bouzoukis D, Naoumidis I, Pallikaris I. Narrow angle light scatter in rabbit corneas after excimer laser surface ablation. *Ophthalmic Physiol Opt.* 2009;29:357–362.
5. Erie JC, Patel SV, McLaren JW, Hodge DO, Bourne WM. Corneal keratocyte deficits after photorefractive keratectomy and laser in situ keratomileusis. *Am J Ophthalmol.* 2006;141:799–809.
6. Trokel SL, Srinivasan R, Braren B. Excimer laser surgery of the cornea. *Am J Ophthalmol.* 1983;96:710–715.
7. Woreta FA, Gupta A, Hochstetler B, Bower KS. Management of post-photorefractive keratectomy pain. *Surv Ophthalmol.* 2013;58:529–535.
8. Alio JL, Javaloy J. Corneal inflammation following corneal photoablative refractive surgery with excimer laser. *Surv Ophthalmol.* 2013;58:11–25.
9. Meduri A, Scorolli L, Scalinci SZ, Grenga PL, Lupo S, Rechichi M, et al. Effect of the combination of basic fibroblast growth factor and cysteine on corneal epithelial healing after photorefractive keratectomy in patients affected by myopia. *Indian J Ophthalmol.* 2014;62:424–428.

10. Erie JC, Patel SV, McLaren JW, Hodge DO, Bourne WM. Keratocyte density in the human cornea after photorefractive keratectomy. *Arch Ophthalmol.* 2003;121:770–776.
11. Diakonis VF, Pallikaris A, Kymionis GD, Markomanolakis MM. Alterations in endothelial cell density after photorefractive keratectomy with adjuvant mitomycin. *Am J Ophthalmol.* 2007;144:99–103.
12. Engle AT, Laurent JM, Schallhorn SC, Toman SD, Newacheck JS, Tanzer DJ, et al. Masked comparison of silicone hydrogel lotrafilcon A and etafilcon A extended-wear bandage contact lenses after photorefractive keratectomy. *J Cataract Refract Surg.* 2005;31:681–686.
13. Jalali S, Yuen LH, Boxer Wachler BS. Effect of nepafenac sodium 0.1% on delayed corneal epithelial healing and haze after photorefractive keratectomy retrospective comparative study. *J Cataract Refract Surg.* 2008;34:1542–1545.
14. Tuominen IS, Tervo TM, Teppo AM, Valle TU, Grönhagen-Riska C, Vesaluoma MH. Human tear fluid PDGF-BB, TNF-alpha and TGF-beta1 vs corneal haze and regeneration of corneal epithelium and subbasal nerve plexus after PRK. *Exp Eye Res.* 2001;72:631–641.
15. Kanitkar KD, Camp J, Humble H, Shen DJ, Wang MX. Pain after removal by ethanol-assisted mechanical versus transepithelial excimer laser debridement. *J Refract Surg.* 2000;16:519–522.
16. Gómez S, Herreras JM, Merayo J, García M, Argüeso P, Cuevas J. Effect of hyaluronic acid on corneal haze in a photorefractive keratectomy experimental model. *J Refract Surg.* 2001;17:549–554.
17. Faktorovich EG, Basbaum AI. Effect of topical 0.5% morphine on postoperative pain after photorefractive keratectomy. *J Refract Surg.* 2010;26:934–941.
18. Lee HK, Lee KS, Kim JK, Kim HC, Seo KR, Kim EK. Epithelial healing and clinical outcomes in excimer laser photorefractive surgery following three epithelial removal techniques: mechanical, alcohol, and excimer laser. *Am J Ophthalmol.* 2005;139:56–63.
19. Gamaly TO, El Danasoury A, El Maghraby A. A prospective, randomized, contralateral eye comparison of epithelial laser in situ keratomileusis and photorefractive keratectomy in eyes prone to haze. *J Refract Surg.* 2007;23(9 Suppl.):S1015–S1020.
20. Netto MV, Barreto J Jr, Santo R, Bechara S, Kara-Jose N, Wilson SE. Synergistic effect of ethanol and mitomycin C on corneal stroma. *J Refract Surg.* 2008;24:626–632.
21. Aslanides IM, Padroni S, Arba Mosquera S, Ioannides A, Mukherjee A. Comparison of single-step reverse transepithelial all-surface laser ablation (ASLA) to alcohol-assisted photorefractive keratectomy. *Clin Ophthalmol.* 2012;6:973–980.
22. Esquenazi S, Bazan HE, Bui V, He J, Kim DB, Bazan NG. Topical combination of NGF and DHA increases rabbit cornea nerve regeneration after photorefractive keratectomy. *Invest Ophthalmol Vis Sci.* 2005;46:3121–3127.
23. Grentzelos MA, Plainis S, Astyrakakis NI, Diakonis VF, Kymionis GD, Kallinikos P, et al. Efficacy of 2 types of silicone hydrogel bandage contact lenses after photorefractive keratectomy. *J Cataract Refract Surg.* 2009;35:2103–2108.
24. Mohan RR, Hutcheon AE, Choi R, Hong J, Lee J, Mohan RR, et al. Apoptosis, necrosis, proliferation, and myofibroblast generation in the stroma following LASIK and PRK. *Exp Eye Res.* 2003;76:71–87.
25. Patel SV, Erie JC, McLaren JW, Bourne WM. Confocal microscopy changes in epithelial and stromal thickness up to 7 years after LASIK and photorefractive keratectomy for myopia. *J Refract Surg.* 2007;23:385–392.
26. Ghanem RC, Ghanem VC, de Souza DC, Kara-José N, Ghanem EA. Customized topography-guided photorefractive keratectomy with the MEL-70 platform and mitomycin C to correct hyperopia after radial keratotomy. *J Refract Surg.* 2008;24:911–922.
27. Virasch VV, Majmudar PA, Epstein RJ, Vaidya NS, Dennis RF. Reduced application time for prophylactic mitomycin C in photorefractive keratectomy. *Ophthalmology.* 2010;117:885–889.
28. Nassaralla BA, McLeod SD, Nassaralla JJ Jr. Prophylactic mitomycin C to inhibit corneal haze after photorefractive keratectomy for residual myopia following radial keratotomy. *J Refract Surg.* 2007;23:226–232.
29. Barreto J Jr, Netto MV, Reis A, Nakano M, Alves MR, Bechara SJ. Topography-guided (NIDEK customized aspheric treatment zone) photorefractive keratectomy with mitomycin C after penetrating keratoplasty for keratoconus: case report. *J Refract Surg.* 2009;25(1 Suppl.):S131–S135.
30. Razmjoo H, Abdi E, Atashkadi S, Reza AM, Reza PA, Akbari M. Comparative study of two silicone hydrogel contact lenses used as bandage contact lenses after photorefractive keratectomy. *Int J Prev Med.* 2012;3:718–722.
31. Edwards JD, Bower KS, Sediq DA, Burka JM, Stutzman RD, Vanroekel CR, et al. Effects of lotrafilcon A and omafilcon A bandage contact lenses on visual outcomes after photorefractive keratectomy. *J Cataract Refract Surg.* 2008;34:1288–1294.
32. Srinivasan S, Drake A, Herzig S. Photorefractive keratectomy with 0.02% mitomycin C for treatment of residual refractive errors after LASIK. *J Refract Surg.* 2008;24:S64–S67.
33. Winkler von Mohrenfels C, Reischl U, Lohmann CP. Corneal haze after photorefractive keratectomy for myopia: role of collagen IV mRNA typing as a predictor of haze. *J Cataract Refract Surg.* 2002;28:1446–1451.
34. Rajan MS, O'Brart DP, Patmore A, Marshall J. Cellular effects of mitomycin-C on human corneas after photorefractive keratectomy. *J Cataract Refract Surg.* 2006;32:1741–1747.
35. Meduri A, Aragona P, Grenga PL, Roszkowska AM. Effect of basic fibroblast growth factor on corneal epithelial healing after photorefractive keratectomy. *J Refract Surg.* 2012;28:220–223.
36. Buzzonetti L, Petrocelli G, Laborante A, Mazzilli E, Gaspari M, Valente P, et al. A new transepithelial phototherapeutic keratectomy mode using the NIDEK CXIII excimer laser. *J Refract Surg.* 2009;25(1 Suppl.):S122–S124.
37. Kim TH, Lee DY, Rho JH, Rho SH, Yoo KW, Ahn HB, et al. Application of newly developed amniotic membrane ointment for photorefractive keratectomy in rabbits. *Ophthalmic Res.* 2006;38:58–61.
38. Chung SA, Kim EK, Ryu IH, Kim JK, Lee HK. Effectiveness of cultured human keratinocyte onlays on epithelial healing and clinical outcome after photorefractive keratectomy. *J Refract Surg.* 2008;24:826–832.
39. Magone MT, Engle AT, Easter TH, Stanley PF, Howells J, Pasternak JF. Flap-off epi-LASIK versus automated epithelial brush in PRK: a prospective comparison study of pain and reepithelialization times. *J Refract Surg.* 2012;28:682–689.
40. Amoozadeh J, Aliakbari S, Behesht-Nejad AH, Seyedian MA, Rezvan B, Hashemi H. Confocal microscopy of corneal stroma and endothelium after LASIK and PRK. *J Refract Surg.* 2009;25(10 Suppl.):S963–S967.
41. Dua HS, Gomes JA, Singh A. Corneal epithelial wound healing. *Br J Ophthalmol.* 1994;78:401–408.
42. Fujikawa LS, Foster CS, Harrist TJ, Lanigan JM, Colvin RB. Fibronectin in healing rabbit corneal wounds. *Lab Invest.* 1981;45:120–129.
43. Netto MV, Mohan RR, Sinha S, Sharma A, Dupps W, Wilson SE. Stromal haze, myofibroblasts, and surface irregularity after PRK. *Exp Eye Res.* 2006;82:788–797.
44. Martínez-García MC, Merayo-Llósés J, Blanco-Mezquita T, Mar-Sardaña S. Wound healing following

- refractive surgery in hens. *Exp Eye Res.* 2006;83:728–735.
45. Chaurasia SS, Kaur H, de Medeiros FW, Smith SD, Wilson SE. Dynamics of the expression of intermediate filaments vimentin and desmin during myofibroblast differentiation after corneal injury. *Exp Eye Res.* 2009;89:133–139.
 46. Wilson SE, Chaurasia SS, Madeiros FW. Apoptosis in the initiation. Modulation and termination of the corneal wound healing response. *Exp Eye Res.* 2007;85:305–311.
 47. Barbosa FL, Chaurasia SS, Kaur H, de Medeiros FW, Agrawal V, Wilson SE. Stromal interleukin-1 expression in the cornea after haze-associated injury. *Exp Eye Res.* 2010;91:456–461.
 48. Tavakoli M, Hossain P, Malik RA. Clinical applications of corneal confocal microscopy. *Clin Ophthalmol.* 2008;2:435–445.
 49. Efron N. Contact lens-induced changes in the anterior eye as observed in vivo with the confocal microscope. *Prog Retin Eye Res.* 2007;26:398–436.
 50. Gan L, Hamberg-Nyström H, Fagerholm P, Van Setten G. Cellular proliferation and leukocyte infiltration in the rabbit cornea after photorefractive keratectomy. *Acta Ophthalmol Scand.* 2001;79:488–492.
 51. Anitua E, Muruzabal F, Alcalde I, Merayo-Llodes J, Orive G. Plasma rich in growth factors (PRGF-Endoret) stimulates corneal wound healing and reduces haze formation after PRK surgery. *Exp Eye Res.* 2013;115:153–161.
 52. Ivarsen A, Fledelius W, Hjortdal JØ. Three-year changes in epithelial and stromal thickness after PRK or LASIK for high myopia. *Invest Ophthalmol Vis Sci.* 2009;50:2061–2066.
 53. Gauthier CA, Epstein D, Holden BA, Tengroth B, Fagerholm P, Hamberg-Nyström H, et al. Epithelial alterations following photorefractive keratectomy for myopia. *J Refract Surg.* 1995;11:113–118.
 54. Hamberg-Nyström H, Gauthier CA, Holden BA, Epstein D, Fagerholm P, Tengroth B. A comparative study of epithelial hypertrophy after PRK: summit versus VISX in the same patient. *Acta Ophthalmol Scand.* 1996;74:228–231.
 55. Moller-Pedersen T, Cavanagh HD, Petroll WM, Jester JV. Stromal wound healing explains refractive instability and haze development after photorefractive keratectomy: a 1-year confocal microscopic study. *Ophthalmology.* 2000;107:1235–1245.
 56. Kozak I, Hornak M, Juhas T, Shah A, Rawlings EF. Changes in central corneal thickness after laser in situ keratomileusis and photorefractive keratectomy. *J Refract Surg.* 2003;19:149–153.
 57. Erie JC, Patel SV, McLaren JW, Ramirez M, Hodge DO, Maguire LJ, et al. Effect of myopic laser in situ keratomileusis on epithelial and stromal thickness: a confocal microscopy study. *Ophthalmology.* 2002;109:1447–1452.
 58. Guerriero E, Chen J, Sado Y, Mohan RR, Wilson SE, Funderburgh JL, et al. Loss of alpha3 (IV) collagen expression associated with corneal keratocyte activation. *Invest Ophthalmol Vis Sci.* 2007;48:627–635.
 59. Lee RE, Davison PF. The collagens of the developing bovine cornea. *Exp Eye Res.* 1984;39:639–652.
 60. Zimmermann DR, Trueb B, Winterhalter KH, Witmer R, Fischer RW. Type VI collagen is a major component of the human cornea. *FEBS Lett.* 1986;197:55–58.
 61. Michelacci YM. Collagens and proteoglycans of the corneal extracellular matrix. *Braz J Med Biol Res.* 2003;36:1037–1046.
 62. Mutoh T, Nishio M, Matsumoto Y, Arai K, Chikuda M. Photorefractive keratectomy: measuring the matrix metalloproteinase activity and chondroitin sulfate concentration in tear fluid. *Clin Ophthalmol.* 2010;4:1015–1018.
 63. Einollahi B, Baradaran-Rafii A, Rezaei-Kanavi M, Eslani M, Parchegani MR, Zare M, et al. Mechanical versus alcohol-assisted epithelial debridement during photorefractive keratectomy: a confocal microscopic clinical trial. *J Refract Surg.* 2011;27:887–893.
 64. Dupps WJ Jr, Wilson SE. Biomechanics and wound healing in the cornea. *Exp Eye Res.* 2006;83:709–720.
 65. Berman M, Leary R, Gage J. Evidence for a role of the plasminogen activator–plasmin system in corneal ulceration. *Invest Ophthalmol Vis Sci.* 1980;19:1204–1221.
 66. Ye HQ, Maeda M, Yu FS, Azar DT. Differential expression of MT1-MMP (MMP-14) and collagenase III (MMP-13) genes in normal and wounded rat corneas. *Invest Ophthalmol Vis Sci.* 2000;41:2894–2899.
 67. Corbett MC, O'Brart DP, Patmore AL, Marshall J. Effect of collagenase inhibitors on corneal haze after PRK. *Exp Eye Res.* 2001;72:253–259.
 68. Holopainen JM, Moilanen JA, Sorsa T, Kivelä-Rajamäki M, Tervahartiala T, Vesaluoma MH, et al. Activation of matrix metalloproteinase-8 by membrane type 1-MMP and their expression in human tears after photorefractive keratectomy. *Invest Ophthalmol Vis Sci.* 2003;44:2550–2556.
 69. Torricelli AA, Singh V, Agrawal V, Santhiago MR, Wilson SE. Transmission electron microscopy analysis of epithelial basement membrane repair in rabbit corneas with haze. *Invest Ophthalmol Vis Sci.* 2013;54:4026–4033.
 70. Davies BW, Panday V, Caldwell M, Scribbick F, Reilly CD. Effect of topical immunomodulatory interleukin 1 receptor antagonist therapy on corneal healing in New Zealand white rabbits (*Oryctolagus cuniculus*) after photorefractive keratectomy. *Arch Ophthalmol.* 2011;129:909–913.
 71. Wilson SE. Corneal myofibroblast biology and pathobiology: generation, persistence, and transparency. *Exp Eye Res.* 2012;99:78–88.
 72. Bilgihan K, Adiguzel U, Sezer C, Akyol G, Hasanreisoglu B. Effects of topical vitamin E on keratocyte apoptosis after traditional photorefractive keratectomy. *Ophthalmologica.* 2001;215:192–196.
 73. Lee JB, Choe CM, Kim HS, Seo KY, Seong GJ, Kim EK. Comparison of TGF-beta1 in tears following laser subepithelial keratomileusis and photorefractive keratectomy. *J Refract Surg.* 2002;18:130–134.
 74. Kaur H, Chaurasia SS, Agrawal V, Wilson SE. Expression of PDGF receptor-alpha in corneal myofibroblasts in situ. *Exp Eye Res.* 2009;89:432–434.
 75. Tchah H, Kim MJ, Kim TI, Choi HJ, Kim JY, Kim MJ, et al. Regulation of 1-cys peroxiredoxin expression in the process of stromal wound healing after photorefractive keratectomy. *Invest Ophthalmol Vis Sci.* 2005;46:2396–2403.
 76. Hayashi S, Ishimoto S, Wu GS, Wee WR, Rao NA, McDonnell PJ. Oxygen free radical damage in the cornea after excimer laser therapy. *Br J Ophthalmol.* 1997;81:141–144.
 77. Brancato R, Fiore T, Papucci L, Schiavone N, Formigli L, Orlandini SZ, et al. Concomitant effect of topical ubiquinone Q10 and vitamin E to prevent keratocyte apoptosis after excimer laser photoablation in rabbits. *J Refract Surg.* 2002;18:135–139.
 78. Bilgihan A, Bilgihan K, Yis O, Sezer C, Akyol G, Hasanreisoglu B. Effects of topical vitamin E on corneal superoxide dismutase, glutathione peroxidase activities and polymorphonuclear leucocyte infiltration after photorefractive keratectomy. *Acta Ophthalmol Scand.* 2003;81:177–180.
 79. Patel S, McLaren J, Hodge D, Bourne W. Normal human keratocyte density and corneal thickness measurements by using confocal microscopy in vivo. *Invest Ophthalmol Vis Sci.* 2001;42:333–339.
 80. McLaren JW, Nau CB, Patel SV, Bourne WM. How precisely can we determine keratocyte density by confocal microscopy? *Invest Ophthalmol Vis Sci.* 2001;42:S281.

81. Böhnke M, Thaeer A, Schipper I. Confocal microscopy reveals persisting stromal changes after myopic photorefractive keratectomy in zero haze corneas. *Br J Ophthalmol*. 1998;82:1393–1400.
82. Midena E, Gambato C, Miotto S, Cortese M, Salvi R, Ghirlando A. Long-term effects on corneal keratocytes of mitomycin C during photorefractive keratectomy: a randomized contralateral eye confocal microscopy study. *J Refract Surg*. 2007;23(9 Suppl.):S1011–S1014.
83. Erie JC, McLaren JW, Hodge DO, Bourne WM. Recovery of corneal subbasal nerve density after PRK and LASIK. *Am J Ophthalmol*. 2005;140:1059–1064.
84. Ambrósio R Jr, Kalina R, Mohan RR, Mohan RR, Possin DD, Huang J, et al. Early wound healing response to epithelial scrape injury in the human cornea. *Invest Ophthalmol Vis Sci*. 2002;43. E-Abstract 4206.
85. Frueh BE, Cadez R, Böhnke M. In vivo confocal microscopy after photorefractive keratectomy in humans. A prospective, long-term study. *Arch Ophthalmol*. 1998;116:1425–1431.
86. Hofmeister EM, Bishop FM, Kaupp SE, Schallhorn SC. Randomized dose–response analysis of mitomycin-C to prevent haze after photorefractive keratectomy for high myopia. *J Cataract Refract Surg*. 2013;39:1358–1365.
87. Fini EM. Keratocyte and fibroblast phenotypes in the repairing cornea. *Prog Retin Eye Res*. 1999;18:529–551.
88. Corbett MC, Prydal JI, Verma S, Oliver KM, Pande M, Marshall J. An in vivo investigation of the structures responsible for corneal haze after photorefractive keratectomy and their effect on visual function. *Ophthalmology*. 1996;103:1366–1380.
89. Erie JC, Patel SV, McLaren JW, Maguire LJ, Ramirez M, Bourne WM. Keratocyte density in vivo after photorefractive keratectomy in humans. *Trans Am Ophthalmol Soc*. 1999;97:221–236.
90. Razmjoo H, Kooshanmehr MR, Peyman A, Kor Z, Mohammadesmaeil E. Comparison of standard and low dose intraoperative mitomycin C in prevention of corneal haze after photorefractive keratectomy. *Int J Prev Med*. 2013;4:204–207.
91. Netto MV, Mohan RR, Sinha S, Sharma A, Gupta PC, Wilson SE. Effect of prophylactic and therapeutic mitomycin C on corneal apoptosis, cellular proliferation, haze, and long-term keratocyte density in rabbits. *J Refract Surg*. 2006;22:562–574.
92. Cua IY, Pepose JS. Late corneal scarring after photorefractive keratectomy concurrent with development of systemic lupus erythematosus. *J Refract Surg*. 2002;18:750–752.
93. Kremer I, Ehrenberg M, Levinger S. Delayed epithelial healing following photorefractive keratectomy with mitomycin C treatment. *Acta Ophthalmol*. 2012;90:271–276.
94. de Medeiros FW, Mohan RR, Suto C, Sinhá S, Bonilha VL, Chaurasia SS, et al. Haze development after photorefractive keratectomy: mechanical vs ethanol epithelial removal in rabbits. *J Refract Surg*. 2008;24:923–927.
95. Resch MD, Nagy ZZ, Szentmáry N, Máthé M, Kovalszky I, Süveges I. Spatial distribution of keratin sulfate in the rabbit cornea following photorefractive keratectomy. *J Refract Surg*. 2005;21:485–493.
96. Meltendorf C, Burbach GJ, Bühren J, Bug R, Ohrloff C, Deller T. Corneal femtosecond laser keratotomy results in isolated stromal injury and favorable wound-healing response. *Invest Ophthalmol Vis Sci*. 2007;48:2068–2075.
97. Csutak A, Tözsér J, Békési L, Hassan Z, Berta A, Silver DM. Plasminogen activator activity in tears after excimer laser photorefractive keratectomy. *Invest Ophthalmol Vis Sci*. 2000;41:3743–3747.
98. von Mohrenfels CW, Reischl U, Gabler B, Lohmann CP. Corneal haze after photorefractive keratectomy. Role of individual collagen type IV synthesis on postoperative corneal opacity. *Ophthalmology*. 2002;99:532–537.
99. Nagy ZZ, Hiscott P, Seitz B, Shlötzer-Schrehardt U, Simon M Jr, Süveges I, et al. Ultraviolet-B enhances stromal response to 193-nm excimer laser treatment. *Ophthalmology*. 1997;104:375–380.
100. Blalock TD, Duncan MR, Varela JC, Goldstein MH, Tuli SS, Grotenhorst GR, et al. Connective tissue growth factor expression and action in human corneal fibroblast cultures and rat corneas after photorefractive keratectomy. *Invest Ophthalmol Vis Sci*. 2003;44:1879–1887.
101. Jester JV, Petroll WM, Cavanagh HD. Corneal stromal wound healing in refractive surgery: the role of myofibroblasts. *Prog Retin Eye Res*. 1999;18:311–356.
102. Vinciguerra P, Azzolini M, Airaghi P, Radice P, De Molfetta V. Effect of decreasing surface and interface irregularities after photorefractive keratectomy and laser in situ keratomileusis on optical and functional outcomes. *J Refract Surg*. 1998;14:S199–S203.
103. Taylor SM, Fields CR, Barker FM, Sanzo CJ. Effect of depth upon the smoothness of excimer laser corneal ablation. *Optom Vis Sci*. 1994;71:104–108.
104. Javier JA, Lee JB, Oliveira HB, Chang JH, Azar DT. Basement membrane and collagen deposition after laser subepithelial keratomileusis and photorefractive keratectomy in the leghorn chick eye. *Arch Ophthalmol*. 2006;124:703–709.
105. Nakamura K, Kurosaka D, Bissen-Miyajima H, Tsubota K. Intact corneal epithelium is essential for the prevention of stromal haze after laser assisted in situ keratomileusis. *Br J Ophthalmol*. 2001;85:209–213.
106. McLaren JW, Bourne WM, Patel SV. Standardization of corneal haze measurement in confocal microscopy. *Invest Ophthalmol Vis Sci*. 2010;51:5610–5616.
107. Farid M, Morishige N, Lam L, Wahlert A, Steinert RF, Jester JV. Detection of corneal fibrosis by imaging second harmonic-generated signals in rabbit corneas treated with mitomycin C after excimer laser surface ablation. *Invest Ophthalmol Vis Sci*. 2008;49:4377–4383.
108. Takacs AI, Mihaltz K, Nagy ZZ. Corneal density with the Pentacam after photorefractive keratectomy. *J Refract Surg*. 2011;27:269–277.
109. Nien CJ, Flynn KJ, Chang M, Brown D, Jester JV. Reducing peak corneal haze after photorefractive keratectomy in rabbits: prednisolone acetate 1.00% versus cyclosporine A 0.05%. *J Cataract Refract Surg*. 2011;37:937–944.
110. Møller-Pedersen T, Cavanagh HD, Petroll WM, Jester JV. Corneal haze development after PRK is regulated by volume of stromal tissue removal. *Cornea*. 1998;17:627–639.
111. Nejima R, Miyata K, Tanabe T, Okamoto F, Hiraoka T, Kiuchi T, et al. Corneal barrier function, tear film stability, and corneal sensation after photorefractive keratectomy and laser in situ keratomileusis. *Am J Ophthalmol*. 2005;139:64–71.
112. Belmonte C, Acosta MC, Gallar J. Neural basis of sensation in intact and injured corneas. *Exp Eye Res*. 2004;78:513–525.
113. Prydal JI, Kerr Muir MG, Dilly PN, Corbett MC, Verma S, Marshall J. Confocal microscopy using oblique sections for measurement of corneal epithelial thickness in conscious humans. *Acta Ophthalmol Scand*. 1997;75:624–628.
114. Vesaluoma M, Perez-Santonja J, Petroll WM, Linna T, Alió J, Tervo T. Corneal stromal changes induced by myopic LASIK. *Invest Ophthalmol Vis Sci*. 2000;41:369–376.
115. Müller LJ, Pels L, Vrensen GF. Ultrastructural organization of human corneal nerves. *Ophthalmol Vis Sci*. 1996;37:476–488.
116. Tervo K, Latvala TM, Tervo TM. Recovery of corneal innervation following photorefractive keratoablation. *Arch Ophthalmol*. 1994;112:1466–1470.

117. Rozsa AJ, Guss RB, Beuerman RW. Neural remodeling following experimental surgery of the rabbit cornea. *Invest Ophthalmol Vis Sci.* 1983;24:1033–1051.
118. Burka JM, Bower KS, Vanroekel RC, Stutzman RD, Kuzmowycz CP, Howard RS. The effect of moxifloxacin and gatifloxacin on long-term visual outcomes following photorefractive keratectomy. *J Refract Surg.* 2007;23:414–417.
119. Baker KS, Anderson SC, Romanowski EG, Thoft RA, Sundar-Raj N. Trigeminal ganglion neurons affect corneal epithelial phenotype. Influence on type VII collagen expression in vitro. *Invest Ophthalmol Vis Sci.* 1993;34:137–144.
120. Kauffmann T, Bodanowitz S, Hesse L, Kroll P. Corneal reinnervation after photorefractive keratectomy and laser in situ keratomileusis: an in vivo study with a confocal videomicroscope. *Ger J Ophthalmol.* 1996;5:508–512.
121. Neira-Zalentein W, Moilanen JA, Tuisku IS, Holopainen JM, Tervo TM. Photorefractive keratectomy retreatment after LASIK. *J Refract Surg.* 2008;24:710–712.
122. Moilanen JA, Vesaluoma MH, Müller LJ, Tervo TM. Long-term corneal morphology after PRK by in vivo confocal microscopy. *Invest Ophthalmol Vis Sci.* 2003;44:1064–1069.
123. Stern ME, Gao J, Siemasko KF, Beuerman RW, Pflugfelder SC. The role of the lacrimal functional unit in the pathophysiology of dry eye. *Exp Eye Res.* 2004;78:409–416.
124. Kymionis GD, Tsiklis NS, Ginis H, Diakonis VF, Pallikaris I. Dry eye after photorefractive keratectomy with adjuvant mitomycin C. *J Refract Surg.* 2006;22:511–513.
125. Hoşal BM, Ornek N, Zilelioğlu G, Elhan AH. Morphology of corneal nerves and corneal sensation in dry eye: a preliminary study. *Eye (Lond).* 2005;19:1276–1279.
126. Bourcier T, Acosta MC, Borderie V, Borrás F, Gallar J, Bury T, et al. Decreased corneal sensitivity in patients with dry eye. *Invest Ophthalmol Vis Sci.* 2005;46:2341–2345.
127. Gallar J, Moilanen J, Acosta C, Holopainen J, Belmonte C, Tervo T, et al. Long-term corneal sensitivity after PRK determined by non-contact gas esthesiometry. *Invest Ophthalmol Vis Sci.* 2013;54 [E-Abstract 904].
128. Sher NA, Golben MR, Bond W, Trattler WB, Tauber S, Voirin TG. Topical bromfenac 0.09% vs. ketorolac 0.4% for the control of pain, photophobia, and discomfort following PRK. *J Refract Surg.* 2009;25:214–220.
129. Gallar J, Acosta MC, Gutiérrez AR, Belmonte C. Impulse activity in corneal sensory nerve fibers after photorefractive keratectomy. *Invest Ophthalmol Vis Sci.* 2007;48:4033–4037.
130. Campero M, Serra J, Marchettini P, Ochoa JL. Ectopic impulse generation and autoexcitation in single myelinated afferent fibers in patients with peripheral neuropathy and positive sensory symptoms. *Muscle Nerve.* 1998;21:1661–1667.
131. Ji RR, Kohno T, Moore KA, Woolf CJ. Central sensitization and LTP: do pain and memory share similar mechanisms? *Trends Neurosci.* 2003;26:696–705.
132. Sia RK, Ryan DS, Stutzman RD, Psolka M, Mines MJ, Wagner ME, et al. Alcohol versus brush PRK: visual outcomes and adverse effects. *Lasers Surg Med.* 2012;44:475–481.
133. Thornton I, Xu M, Krueger RR. Comparison of standard (0.02%) and low dose (0.002%) mitomycin C in the prevention of corneal haze following surface ablation for myopia. *J Refract Surg.* 2008;24:S68–S76.
134. Hanna KD, Pouliquen YM, Waring GO III, Savoldelli M, Fantes F, Thompson KP. Corneal wound healing in monkeys after repeated excimer laser photorefractive keratectomy. *Arch Ophthalmol.* 1992;110:1286–1291.
135. Ghoreishi M, Attarzadeh H, Tavakoli M, Moini HA, Zandi A, Masjedi A. Alcohol-assisted versus mechanical epithelium removal in photorefractive keratectomy. *J Ophthalmic Vis Res.* 2010;5:223–227.
136. Browning AC, Shah S, Dua HS, Maharajan SV, Gray T, Bragheeth MA. Alcohol debridement of the corneal epithelium in PRK and LASEK: an electron microscopic study. *Invest Ophthalmol Vis Sci.* 2003;44:510–513.
137. Shah S, Sebai Sarhan AR, Doyle SJ, Pillai CT, Dua HS. The epithelial flap for photorefractive keratectomy. *Br J Ophthalmol.* 2001;85:393–396.
138. Chen CC, Chang JH, Lee JB, Javier J, Azar DT. Human corneal epithelial cell viability and morphology after dilute alcohol exposure. *Invest Ophthalmol Vis Sci.* 2002;43:2593–2602.
139. Ghanem VC, Souza GC, Souza DC, Viese JM, Weber SL, Kara-José N. PRK and butterfly LASEK: prospective, randomized, contralateral eye comparison of epithelial healing and ocular discomfort. *J Refract Surg.* 2008;24:591–599.
140. Wang DM, Du Y, Chen GS, Tang LS, He JF. Transepithelial photorefractive keratectomy mode using SCHWIND-ESIRIS excimer laser: initial clinical results. *Int J Ophthalmol.* 2012;5:334–337.
141. Mannan R, Pruthi A, Rampal U. Combined phototherapeutic keratectomy and amniotic membrane grafts for symptomatic bullous keratopathy. *Cornea.* 2010;29:1207–1208.
142. Rathi VM, Vyas SP, Sangwan VS. Phototherapeutic keratectomy. *Indian J Ophthalmol.* 2012;60:5–14.
143. Lee HK, Kim JK, Kim EK, Kim GO, Lee IS. Phototherapeutic keratectomy with amniotic membrane for severe subepithelial fibrosis following excimer laser refractive surgery. *J Cataract Refract Surg.* 2003;29:1430–1435.
144. Choi YS, Kim JY, Wee WR, Lee JH. Effect of the application of human amniotic membrane on rabbit corneal wound healing after excimer laser photorefractive keratectomy. *Cornea.* 1998;17:389–395.
145. Wang MX, Gray T, Parks WC, Prabhasawat P, Culbertson W, Forster R, et al. Reduction in corneal haze and apoptosis by amniotic membrane matrix in excimer laser photoablation in rabbits. *J Cataract Refract Surg.* 2001;27:310–319.
146. Park WC, Tseng SC. Modulation of acute inflammation and keratocyte death by suturing, blood, and amniotic membrane in PRK. *Invest Ophthalmol Vis Sci.* 2000;41:2906–2914.
147. Savill J, Haslett C. Granulocyte clearance by apoptosis in the resolution of inflammation. *Semin Cell Biol.* 1995;6:385–393.
148. Khokhar S, Natung T, Sony P, Sharma N, Agarwal N, Vajpayee RB. Amniotic membrane transplantation in refractory neurotrophic corneal ulcers: a randomized, controlled clinical trial. *Cornea.* 2005;24:654–660.
149. Meller D, Pauklin M, Thomasen H, Westekemper H, Steuhl KP. Amniotic membrane transplantation in the human eye. *Dtsch Arztebl Int.* 2011;108:243e8.
150. Zhong Y, Zhai Z, Zhou Y, Ye W, Wang K. Effect of amniotic membrane on expressions of TGF-beta 1, collagens I, III and fibronectin in rabbit corneal healing after photorefractive keratectomy. *Yan Ke Xue Bao.* 2000;16:239–242.
151. Woo HM, Kim MS, Kweon OK, Kim DY, Nam TC, Kim JH. Effects of amniotic membrane on epithelial wound healing and stromal remodelling after excimer laser keratectomy in rabbit cornea. *Br J Ophthalmol.* 2001;85:345–349.
152. Farahi A, Hashemi H, Mehravaran S. The effects of mitomycin C on tear function after photorefractive keratectomy: a contralateral comparative study. *J Refract Surg.* 2013;29:260–264.
153. Shojaei A, Ramezanzadeh M, Soleyman-Jahi S, Almasi-Nasrabadi M, Rezazadeh P, Eslani M. Short-time mitomycin-C application during photorefractive keratectomy in patients with low myopia. *J Cataract Refract Surg.* 2013;39:197–203.
154. Dawson DG, Grossniklaus HE, McCarey BE, Edelhauser HF. Biomechanical and wound healing characteristics of corneas after excimer laser keratorefractive surgery: is there a difference between advanced surface ablation and subBowman's keratomileusis? *J Refract Surg.* 2008;24:S90–S96.

155. Netto MV, Chalita MR, Krueger RR. Corneal haze following PRK with mitomycin C as a retreatment versus prophylactic use in the contralateral eye. *J Refract Surg.* 2007;23:96–98.
156. Khoury JM, Farah T, El-Haibi CP, Noureddin BN. Corneal light shield as a delivery system for standardized application of mitomycin C in excimer surface ablation. *J Refract Surg.* 2007;23:716–719.
157. Goldsberry DH, Epstein RJ, Majmudar PA, Epstein RH, Dennis RF, Holley G, et al. Effect of mitomycin C on the corneal endothelium when used for corneal subepithelial haze prophylaxis following photorefractive keratectomy. *J Refract Surg.* 2007;23:724–727.
158. Morales AJ, Zadok D, Mora-Retana R, Martínez-Gama E, Robledo NE, Chayet AS. Intraoperative mitomycin and corneal endothelium after photorefractive keratectomy. *Am J Ophthalmol.* 2006;142:400–404.
159. Wallau AD, Campos M. Photorefractive keratectomy with mitomycin C versus LASIK in custom surgeries for myopia: a bilateral prospective randomized clinical trial. *J Refract Surg.* 2008;24:326–336.
160. Sia RK, Ryan DS, Edwards JD, Stutzman RD, Bower KS. The U.S. Army Surface Ablation Study: comparison of PRK, MMC-PRK, and LASEK in moderate to high myopia. *J Refract Surg.* 2014;30:256–264.
161. Kymionis GD, Diakonou VF, Charisis S, Pallikaris AI, Bouzoukis DI, Yoo SH, et al. Effects of topical mitomycin C on the ciliary body and intraocular pressure after PRK: an experimental study. *J Refract Surg.* 2008;24:633–638.
162. Wallau AD, Campos M. One-year outcomes of a bilateral randomised prospective clinical trial comparing PRK with mitomycin C and LASIK. *Br J Ophthalmol.* 2009;93:1634–1638.
163. Ghoreishi M, Attarzadeh H, Zandi A, Moini HA, Tavakoli M, Fesharaki H. Outcomes of photorefractive keratectomy with intraoperative mitomycin-C. *J Ophthalmic Vis Res.* 2009;4:142–146.
164. Fazel F, Roshani L, Rezaei L. Two-step versus single application of mitomycin-C in photorefractive keratectomy for high myopia. *J Ophthalmic Vis Res.* 2012;7:17–23.
165. Spadea L, Verrecchia V. Effectiveness of scraping and mitomycin C to treat haze after myopic photorefractive keratectomy. *Open Ophthalmol J.* 2011;5:63–65.
166. Majmudar PA, Forstot SL, Dennis RF, Nirankari VS, Damiano RE, Brenart R, et al. Topical mitomycin-C for subepithelial fibrosis after refractive corneal surgery. *Ophthalmology.* 2000;107:89–94.
167. Xu H, Liu S, Xia X, Huang P, Wang P, Wu X. Mitomycin C reduces haze formation in rabbits after excimer laser photorefractive keratectomy. *J Refract Surg.* 2001;17:342–349.
168. Patel SV, Bourne WM. Corneal endothelial cell loss 9 years after excimer laser keratorefractive surgery. *Arch Ophthalmol.* 2009;127:1423–1427.
169. Mardelli PG, Piebenga LW, Matta CS, Hyde LL, Gira J. Corneal endothelial status 12 to 55 months after excimer laser photorefractive keratectomy. *Ophthalmology.* 1995;102:544–549.
170. Zare M, Jafarinasab MR, Feizi S, Zamani M. The effect of mitomycin-C on corneal endothelial cells after photorefractive keratectomy. *J Ophthalmic Vis Res.* 2011;6:8–12.
171. Roh DS, Funderburgh JL. Impact on the corneal endothelium of mitomycin C during photorefractive keratectomy. *J Refract Surg.* 2009;25:894–897.
172. Burka JM, Bower KS, Vanroekel RC, Stutzman RD, Kuzmowych CP, Howard RS. The effect of fourth-generation fluoroquinolones gatifloxacin and moxifloxacin on epithelial healing following photorefractive keratectomy. *Am J Ophthalmol.* 2005;140:83–87.
173. Plaka A, Grentzelos MA, Astyrakakis NI, Kymionis GD, Pallikaris IG, Plainis S. Efficacy of two silicone-hydrogel contact lenses for bandage use after photorefractive keratectomy. *Cont Lens Anterior Eye.* 2013;36:243–246.
174. Brilakis HS, Deutsch TA. Topical tetracaine with bandage soft contact lens pain control after photorefractive keratectomy. *J Refract Surg.* 2000;16:444–447.
175. Dantas PE, Nishiwaki-Dantas MC, Ojeda VH, Holzchuh N, Mimica LJ. Microbiological study of disposable soft contact lenses after photorefractive keratectomy. *CLAO J.* 2000;26:26–29.
176. Cherry PM. The treatment of pain following excimer laser photorefractive keratectomy: additive effect of local anesthetic drops, topical diclofenac, and bandage soft contact. *Ophthalmic Surg Lasers.* 1996;27(5 Suppl.):S477–S480.
177. Vetrugno M, Maino A, Quaranta GM, Cardia L. The effect of early steroid treatment after PRK on clinical and refractive outcomes. *Acta Ophthalmol Scand.* 2001;79:23–27.
178. Vigo L, Scandola E, Carones F. Scraping and mitomycin C to treat haze and regression after photorefractive keratectomy for myopia. *J Refract Surg.* 2003;19:449–454.
179. Bühren J, Nagy L, Swanton JN, Kenner S, MacRae S, Phipps RP, et al. Optical effects of anti-TGFbeta treatment after photorefractive keratectomy in a cat model. *Invest Ophthalmol Vis Sci.* 2009;50:634–643.
180. Javadi MA, Mirbabaee-Ghafghazi F, Mirzade M, Yazdani S, Yaseri M. Steroid induced ocular hypertension following myopic photorefractive keratectomy. *J Ophthalmic Vis Res.* 2008;3:42–46.
181. Caldwell M, Reilly C. Effects of topical nepafenac on corneal epithelial healing time and postoperative pain after PRK: a bilateral, prospective, randomized, masked trial. *J Refract Surg.* 2008;24:377–382.
182. Szabó V, Balogh K, Süveges I, Rácz K, Hunyady L, Nagy ZZ. The role of lumican and keratocan genes in persistent subepithelial corneal haze following excimer laser photorefractive keratectomy. *Mol Vis.* 2006;12:597–605.
183. Vetrugno M, Maino A, Cardia G, Quaranta GM, Cardia L. A randomised, double masked, clinical trial of high dose vitamin A and vitamin E supplementation after photorefractive keratectomy. *Br J Ophthalmol.* 2001;85:537–539.
184. Chelala E, Dirani A, Fadlallah A, Fahd S. The role of topical vitamin A in promoting healing in surface refractive procedures: a prospective randomized controlled study. *Clin Ophthalmol.* 2013;7:1913–1918.
185. Lee KS, Ko DA, Kim ES, Kim MJ, Tchah H, Kim JY. Bevacizumab and rapamycin can decrease corneal opacity and apoptotic keratocyte number following photorefractive keratectomy. *Invest Ophthalmol Vis Sci.* 2012;53:7645–7653.
186. Santhiagar MR, Singh V, Barbosa FL, Agrawal V, Wilson SE. Monocyte development inhibitor PRM-151 decreases corneal myofibroblast generation in rabbits. *Exp Eye Res.* 2011;93:786–789.
187. Sharma A, Mehan MM, Sinha S, Cowden JW, Mohan RR. Trichostatin A inhibits corneal haze in vitro and in vivo. *Invest Ophthalmol Vis Sci.* 2009;50:2695–2701.
188. Javaloy J, Alió JL, Rodríguez AE, Vega A, Muñoz G. Effect of platelet-rich plasma in nerve regeneration after LASIK. *J Refract Surg.* 2013;29:213–219.
189. Cortina MS, He J, Li N, Bazan NG, Bazan HE. Recovery of corneal sensitivity, calcitonin gene-related peptide-positive nerves, and increased wound healing induced by pigment epithelial-derived factor plus docosahexaenoic acid after experimental surgery. *Arch Ophthalmol.* 2012;130:76–83.
190. Shaheen BS, Bakir M, Jain S. Corneal nerves in health and disease. *Surv Ophthalmol.* 2014;59:263–285.
191. Devor M. Sodium channels and mechanisms of neuropathic pain. *J Pain.* 2006;7(Suppl. 1):S3–S12.

192. Rivera L, Gallar J, Pozo MA, Belmonte C. Responses of nerve fibres of the rat saphenous nerve neuroma to mechanical and chemical stimulation: an in vitro study. *J Physiol.* 2000;527:305–313.
193. Lichtinger A, Purcell TL, Schanzlin DJ, Chayet AS. Gabapentin for postoperative pain after photorefractive keratectomy: a prospective, randomized, double-blind, placebo-controlled trial. *J Refract Surg.* 2011;27:613–617.