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Alterations of Mitochondrial DNA Copy Number and Telomere Length with Early Adversity and Psychopathology

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Abstract

Background—Telomere shortening and alterations of mitochondrial biogenesis are involved in cellular aging. Childhood adversity is associated with telomere shortening, and several investigations have shown short telomeres in psychiatric disorders. Recent studies have examined whether mitochondria might be involved in neuropsychiatric conditions; findings are limited and no prior work has examined this in relation to stress exposure.

Methods—Two-hundred and ninety healthy adults provided information on childhood parental loss and maltreatment and completed diagnostic interviews. Participants were categorized into four groups based upon the presence or absence of childhood adversity and the presence or absence of lifetime psychopathology (depressive, anxiety, and substance use disorders). Telomere length and mtDNA copy number were measured from leukocyte DNA by qPCR.

Results—Childhood adversity and lifetime psychopathology were each associated with shorter telomeres ($p < .01$) and higher mtDNA copy numbers ($p < .001$). Significantly higher mtDNA copy numbers and shorter telomeres were seen in individuals with major depression, depressive disorders, and anxiety disorders, as well as those with parental loss and childhood maltreatment. A

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history of substance disorders was also associated with significantly higher mtDNA copy numbers.

Conclusion—This study provides the first evidence of an alteration of mitochondrial biogenesis with early life stress and with anxiety and substance use disorders. We replicate prior work on telomere length and psychopathology, and show that this effect is not secondary to medication use or comorbid medical illness. Finally, we show that early life stress and psychopathology are each associated with these markers of cellular aging.

Keywords

mitochondria; telomere; childhood; early life stress; depressive disorder; anxiety disorder

Introduction

Many decades of research have documented robust associations of stress exposure and psychopathology with somatic conditions associated with aging, including cardiovascular disease, inflammatory conditions, and obesity (1–4). This may be due to effects of inflammation, oxidative stress, and glucocorticoid exposure on cellular aging (5, 6). Two key processes in cellular aging are telomere shortening and mitochondrial dysfunction (7–9). New research links telomere biology and mitochondrial biogenesis and function, and there is increasing interest in a role for telomere shortening and mitochondrial dysfunction in psychiatric disorders (10–14).

Mitochondrial Dysfunction, Cellular Aging, and Psychopathology

Mitochondrial respiration provides most of the ATP needed for cellular functions. Mitochondria also play an integral role in cell growth, differentiation, replication, and death. Reactive oxygen species (ROS) are generated during mitochondrial respiration. Aging is characterized by increases in mitochondrial ROS production, declines in mitochondrial function, accumulation of mutations in mitochondrial DNA (mtDNA), and alterations of mitochondrial biogenesis (7, 15, 16). Mitochondrial dysfunction and alterations of mitochondrial biogenesis have been documented in age-related diseases, including cardiovascular disease, diabetes, and various forms of cancer (7, 16, 17).

The possibility that mitochondrial dysfunction might be a pathophysiologic mechanism in psychiatric disorders is intriguing. The brain has high energy demands, and ATP is vital for such neural processes as ion transport, neurotransmitter production and release, and receptor function, (14, 18) as well as neural differentiation, dendritic branching, and synaptic plasticity (14, 19). Several studies support a role for abnormalities of mitochondrial function in psychiatric disorders, in particular bipolar disorder, schizophrenia, and major depressive disorder (MDD) (14, 19). Stress exposure may account for some of this association. Gluococorticoids act on coordinated physiological systems that can induce metabolic stress (20) and exert effects on mitochondria by altering expression of mitochondrial genes and nuclear genes that influence mitochondrial density and function (20, 21). Glucocorticoid receptors bind to mitochondrial membranes to regulate membrane potential (20, 21). Early

life stress also enhances glutamate transmission which increases oxidative stress and results in mitochondrial damage (22, 23).

Mitochondrial biogenesis serves the energy demands of the cell and compensates for cell damage (24). A small number of studies have examined mtDNA copy number, an index of mitochondrial biogenesis, in patients with psychiatric disorders. mtDNA copy number has been reported to be high in autism (25), and reduced in a recent study of bipolar disorder (26) but not others (27–31). A few studies have assessed mtDNA copy number in patients with MDD: no difference was found between MDD patients and controls in a small postmortem study of brain tissue (31) or in a large study in outpatients with MDD and matched controls (32), but there is some evidence of reduced leukocyte mtDNA copy number in association with depression in women over age 60 (33). We are not aware of any investigations of mtDNA copy number in relation to anxiety disorders, alcohol or substance disorder, or psychosocial stress exposure.

Telomere Length, Cellular Aging, and Psychopathology

Telomeres are repetitive nucleotide sequences at the ends of chromosomes that are critical to maintaining chromosomal stability. DNA sequences are lost with each replication cycle, and telomeres serve as a buffer that protects the chromosome from losing critical DNA (34, 35). Telomeres shorten with each cell division, and on average older individuals have shorter telomeres than younger individuals (36). Telomere shortening beyond a critical threshold initiates the DNA damage response pathway and induces replicative senescence or apoptosis (9, 34, 37).

Telomere shortening occurs in several diseases associated with aging, including cardiovascular disease, diabetes, stroke and autoimmune conditions (34, 38, 39). Reduced telomere length has also been documented in MDD (10, 40–44), although some studies have been negative (45, 46). A few investigations also found reduced leukocyte telomere length in anxiety disorders (45, 47), post-traumatic stress disorder (PTSD) (48–51), bipolar II disorder (52), schizophrenia (53), and substance use disorders (54, 55). Chronicity of MDD is linked to leukocyte telomere shortening (43, 47, 56), suggesting that loss of telomere length may be a consequence of other aspects of the disease process.

Psychiatric disorders are associated with high levels of perceived and chronic stress exposure (57, 58), and reduced telomere length is associated with chronic stress (10, 11). Stress-induced activation of glucocorticoids, pro-inflammatory cytokines, and oxidative stress have been implicated in this association (59). Telomeres undergo substantial shortening during early development, and exposure to early adversity is associated with short telomeres in leukocytes and buccal cells in cross-sectional (10, 11), (60–62) and prospective designs (47, 63), however not all studies have shown this effect (64, 65). This suggests that stress exposure may account for the association of psychopathology and telomere shortening. Although prior work has statistically controlled for early life stress exposure in testing for effects of psychiatric conditions (45, 47, 50, 66), we are not aware of any previous studies that have directly compared telomere length in subjects with both early life stress and psychopathology to those with psychopathology or early life stress alone.

Telomere Length and the Regulation of Mitochondrial Biogenesis and Function

Work in animal models and cell culture has recently uncovered shared pathways that regulate telomere length and mitochondrial biogenesis and function. Telomere dysfunction induces expression of p53, which suppresses PGC1α and PGC1β, proteins that regulate mitochondrial biogenesis and function (9, 67). Telomerase reverse transcriptase (TERT) may also regulate mitochondrial function. Under oxidative stress, TERT translocates from the nucleus to the mitochondria, where it appears to modulate respiratory chain function, ROS production, mtDNA damage, and apoptosis (68, 69).

The Present Study

Given this shared regulation of telomere and mitochondrial function and the emerging body of literature implicating these mechanisms of cellular aging in psychiatric disorders, we hypothesized alterations of mtDNA copy number and shorter telomere length in relation to early adversity as well as lifetime depressive, anxiety, and substance use disorders.

Methods and Materials

Subjects

Two hundred ninety participants, 177 women and 113 men, aged $18-61$ (31.0 \pm 10.7) years, were recruited using separate local, newspaper, and Internet advertisements directed toward healthy adults and individuals with depression, childhood parental loss, or a history of early life stress. Participants were white $(n=241)$, black $(n=26)$, Asian $(n=9)$, Hispanic $(n=4)$ and "other" (n=10). Subjects from our prior study of adversity and telomere length (70) were not included. The study was approved by the Butler Hospital Institutional Review Board. After complete description of the study, voluntary written informed consent was obtained.

Exclusions included prescription medication use other than oral contraceptives, acute or chronic medical illness, pregnancy, and a history of brain injury or seizure disorder. Also excluded were those with night-shift work, current alcohol or substance disorders, and lifetime history of bipolar disorder, psychotic disorder, and obsessive-compulsive disorder.

Demographics

Height and weight were measured without shoes, and body mass index (BMI) (weight $(kg)/ht$ (m)²) calculated. Participants answered a single item about childhood socioeconomic adversity ("My family was generally financially stable when I was growing up, and all of my basic needs (food, shelter, and clothing) were met during my childhood") and reported their highest level of education.

Diagnoses and Symptoms

Axis I psychiatric diagnoses were assessed using the Structured Clinical Interview for DSM-IV (SCID) (71). The Inventory for Depressive Symptoms, Self Report (IDS-SR) (72), the State-Trait Anxiety Inventory (STAI) (73), the Perceived Stress Scale (PSS) (74), and the Connor-Davidson Resilience Scale (CD-RISC) (75) were also administered.

Childhood Adversity

Parental Loss—Loss of a parent before age 18 (N=76) included death (N=36) and/or prolonged separation/desertion (i.e., parent deserted for at least six months with no attempts at contact or responses to child's attempts) (N=43).

The Childhood Trauma Questionnaire (CTQ), 28-item version (76)—This measure has high internal consistency, test-retest reliability, and convergent validity (76). Moderate-severe levels of one or more of the five types of maltreatment (physical, sexual, and emotional abuse, physical and emotional neglect) were assessed.

Measures of Cellular Aging

DNA was isolated from whole blood using standard techniques. Three parallel quantitative polymerase chain reactions (qPCRs) were performed to quantitate copy numbers for telomeres, mitochondrial genomes, and the beta-hemoglobin gene as a single-copy standard, as previously described (77). Data were acquired using the ABI Prism HT79000 DNA Sequence Detection System (Applied Biosystems, Grand Island, NY). qPCRs were performed on 384-well plates in a reaction volume of 10 μL containing approximately 25 ng genomic DNA, 300 nM of each primer, and 1x Sybr Select Master Mix (Life Technologies Corporation, Grand Island, NY). For telomere length, the reaction also included 2% DMSO. All reaction plates contained wells with serial dilutions of a cloned amplicon (i.e., cloned telomere amplicon, cloned mitochondrial amplicon, or cloned beta-hemaglobin amplicon) to permit absolute quantitation of telomere, mitochondrial DNA, and beta-hemoglobin copy number. A standard curve was used to directly determine copy number for telomere amplicons, mitochondria, or a single-copy gene in each PCR run. All analyses were performed in triplicate.

Single Copy Gene—The sequences of the forward and reverse primers for the betahemoglobin gene were: GCT TCT GAC ACA ACT GTG TTC ACT AGC and CAC CAA CTT CAT CCA CGT. An initial heating step of 95°C for 10 minutes was followed by 40 cycles of 95°C for 15 sec and 60°C for 1 min.

mtDNA Copy Number—The sequences of the forward and reverse mitochondrial primers (directed towards the D-loop region) were: CAT CTG GTT CCT ACT TCA GGG and TGA GTG GTT AAT AGG GTG ATA GA (78). An initial heating step of 95°C for 10 minutes was followed by 40 cycles of 95°C for 15 sec and 60°C for 1 min. To obtain an index of mitochondrial number, mtDNA copy number was divided by the copy number for the betahemoglobin gene.

Telomere Length—Modified telomere primer sequences were provided by Richard Cawthon (Eccles Institute of Human Genetics, University of Utah): CGG TTT GTT TGG GTT TGG GTT TGG GTT TGG GTT TGG GTT (Tel1b) and GGC TTG CCT TAC CCT TAC CCT TAC CCT TAC CCT TAC CCT (Tel2b). An initial heating step of 95°C for 30 minutes was followed by 40 cycles of 95°C for 15 sec and 60°C for 1 min. The ratio of telomere copy number over beta-hemoglobin copy number was obtained, multiplied by the

length of the telomere amplicon to obtain telomere length per genome, and then divided by

Statistical Analysis

the number of chromosome ends.

Telomere length and mtDNA copy number were log transformed to adjust for skewed distributions. Age, gender, education, BMI, and childhood socioeconomic adversity were included as covariates in the statistical models given literature demonstrating their effects on mtDNA copy number and telomere length. As previously reported (79), mtDNA copy number and telomere length were highly correlated ($r = .33$, $p < .001$) and were therefore included as covariates. Means and standard deviations shown in figures represent log transformed data and were adjusted for the covariates; unadjusted raw means are provided in the supplementary materials. Subjects were categorized into four groups: no early adversity and no lifetime psychiatric diagnosis ($n = 113$, No Adversity/No Disorder group); early adversity but no lifetime diagnosis ($n = 66$, Adversity/No Disorder group); lifetime diagnosis but no history of early adversity ($n = 39$, No Adversity/Disorder group); and both early adversity and a lifetime diagnosis ($n = 72$, Adversity/Disorder group). Analyses that involved subjects with substance use disorders included only those with past disorders because those with current substance disorders (meeting criteria in the last month) were excluded from the study. General linear models (GLMs) were used to predict the grouping variable, controlling for covariates; the full models are shown in Supplementary tables S1 and S2. We then examined individual disorder types (MDD, any depressive disorder, PTSD, any anxiety disorder, and past alcohol or substance disorders), parental loss, and childhood maltreatment, in models that controlled for the covariates and used the No Adversity/No Disorder as the comparison group.

Results

Preliminary Analyses

Descriptive statistics are displayed in Table 1. Age and BMI were negatively associated with telomere length ($r = -.13$, $p = .026$ and $r = -.14$, $p = .019$, respectively), but not with mtDNA copy number. Race was not associated with telomere length or mtDNA copy number. Gender, education, and childhood socioeconomic adversity were also not associated with telomere length or mtDNA copy number. Telomere length and mtDNA copy number were significantly correlated ($r = .33$, $p < .001$).

mtDNA Copy Number

The adversity and diagnosis grouping variable was significantly associated with mtDNA copy number $(F(3, 280) = 8.40, p < .001$; Figure 1). The No Adversity/No Disorder group had significantly lower mtDNA copy numbers than the three other groups. The Adversity/No Disorder group was also significantly different from the Adversity/ Disorder group ($p = .040$) and the No Adversity/Disorder group at trend level ($p = .089$). As shown in Figure 2, in comparison with the No Adversity/No Disorder group, higher mtDNA copy numbers were seen in participants with lifetime MDD $(F(1,153) = 12.18, p = .001)$, any lifetime depressive disorder (including MDD; $F(1, 164) = 14.65$, p <.001), any lifetime anxiety disorder (including PTSD; $F(1, 133) = 14.05$, $p < .001$), and past substance disorders

 $(F(1, 160) = 12.36, p = .001)$. As shown in Figure 3, elevated mtDNA copy numbers were also seen in participants who experienced parental loss (n = 76; F(1, 181) = 15.85, p < .001) and those who reported any form of maltreatment on the CTQ $(n = 101; F(1, 206) = 11.60, p$ $= .001$).

Telomere Length

The Adversity/Disorder grouping variable was associated with telomere length ($F(3, 280) =$ 6.39, $p < .001$; Figure 4). The No Adversity/No Disorder group had significantly longer telomeres on average than the No Adversity/Disorder group ($p = .002$) and the Adversity/ Disorder group ($p < .001$) but not the Adversity/No Disorder group ($p = .153$). The Adversity/No Disorder group was significantly different from the Adversity/Disorder group $(p = .019)$ and the No Adversity/Disorder group at trend level $(p = .085)$. Figure 5 shows that shorter telomere lengths were seen in participants with lifetime MDD ($F(1,153) = 21.09$, $p < .001$), any lifetime depressive disorder (F(1, 164) = 26.35, $p < .001$), any lifetime anxiety disorder F(1, 133) = 19.06, p <.001) and lifetime PTSD (F(1, 117 = 4.02, p = .047). Shorter telomere lengths were also seen in participants who experienced parental loss and those who experienced maltreatment $(F(1, 181) = 8.84, p = .003$ and $F(1, 206) = 7.07, p = .$ 008, respectively) (Figure 6).

Potential Confounds and Sensitivity Analyses

Race, smoking status, and oral contraceptive use were not associated with mtDNA copy number or telomere length, and the grouping variable remained a significant predictor of telomere length and mtDNA copy number when these variables were included in the statistical models. Additional analyses were conducted that did not control for the significant positive association of telomere length and mtDNA copy number. The grouping variable remained a significant predictor of mtDNA copy number and telomere length. Details of the individual contrasts are provided in the supplementary materials.

As shown in Table 1, the groups differed with respect to depressive symptoms on the IDS-SR, anxiety symptoms on the STAI, perceived stress in the last month on the PSS, and resilience on the CD-RISC; the Adversity/No Disorder group had intermediate scores compared to the two disorder groups and the No Adversity/No Disorder group, and the two disorder groups did not differ. This pattern resembles the differences between groups on telomere length and mtDNA copy number. However, when these variables were entered into the models predicting telomere length and mtDNA number, they did not account for the group effect. Additional analyses were aimed at further elucidating the effects in the two groups that had either childhood adversity or lifetime psychopathology. First, we sought to determine whether the effects of the No Adversity/Disorder group might be explained by an effect of recent perceived stress, low levels of resilience in the face of stress, or the symptom burden in these subjects. Within this group, telomere length was negatively correlated with scores on the PSS (r=−.46, p < .01), IDS-SR (r=−.40, p < .05), STAI-Trait (r=−.34, p < .05), and STAI-State (r=−.37, p < .05) after controlling for the covariates, suggesting that the burden of stress and symptoms may account for effects in this group. There was no effect of the CD-RISC. None of these variables was associated with mtDNA copy number in this group.

Similarly, we examined whether the intermediate scores in the Adversity/No Disorder group might due to an effect of subclinical symptoms, recent perceived stress, or resilience. There were no associations of telomere length or mtDNA number with depressive symptoms, state or trait anxiety, recent perceived stress, or resilience in the Adversity/No Disorder group.

The two other groups (No Adversity/No Disorder and Adversity/Disorder) also did not show any associations of telomere length or mtDNA copy number with any of these three domains.

Discussion

This study provides the first evidence of an association of early stress exposure and lifetime psychopathology with alterations of mtDNA copy number, a proposed measure of cellular aging. These findings are bolstered by our similar results for telomere length, another marker of cellular aging with robust links to stress and psychopathology. Although the specific mechanisms governing mtDNA copy number are poorly understood, mitochondrial replication and degradation can occur as a response to mtDNA dysfunction or other cellular signals (80). Glucocorticoids induce expression of nuclear genes involved in the production of new mitochondria (81, 82), and in skeletal and cardiac muscle, mitochondrial proliferation occurs following mechanical or oxidative stress (83). Consistent with evidence linking the regulation of telomeres and telomerase with mitochondrial function (9), there was a positive association between telomere length and mtDNA copy number in this study. However, early stress and psychopathology were associated with shorter telomeres but higher mtDNA numbers, suggesting that co-regulation of these processes might serve a compensatory role, particularly in healthy adults. Telomere dysfunction leads to activation of the tumor suppressor protein p53. Depending on levels of p53 and the degree of genotoxic stress, p53 can either promote the production of antioxidants and enhance mitochondrial function and biogenesis, or promote the expression of ROS and impair mitochondrial function (9). Glucocorticoids and inflammatory mediators likely also play a role in the associations of stress exposure and psychopathology with telomere length and mitochondrial biogenesis (20, 59).

In a study of peripheral blood mononuclear cells (PBMCs) from depressed patients compared to healthy controls, mitochondrial respiration was impaired and the intracellular density of mitochondria, as measured by citrate synthase activity, was increased. In addition, mitochondrial respiration was negatively correlated with early trauma exposure (84). Another study found impaired mitochondrial respiration in platelets of patients with depression (84, 85). With regard to mtDNA copy number, one study did not show alterations in 17–45 year-old Chinese outpatients with MDD compared with matched healthy controls (32). However, in a study of mtDNA copy number in community-dwelling Korean women over age 60, lower mtDNA copy numbers were seen in a subgroup on antidepressants or with high levels of depressive symptoms (33). In another analysis of this sample, the negative association of depressive symptoms with mtDNA copy number was not significant after controlling for effects of current smoking, hypertension, telomere length and other covariates (79). We speculate that in our study of medically-healthy adults, increased mtDNA copy numbers might be an early compensatory response that could be limited by

time and disease burden. We observed a negative association of age and BMI with telomere length, but not with mtDNA copy number. However, when analyzed by group, there were significant negative correlations of mtDNA number with age ($r = -0.275$, p = .018) and BMI $(r = -0.291, p = 0.012)$ only in the Adversity/Diagnosis group. This group had the greatest spread of age and BMI, and the greatest burden of early adversity and psychopathology.

This study is unique in assessing discrete groups of adults with and without childhood adversity and lifetime psychopathology; this allowed examination of whether telomere length and mtDNA copy number might be related to risk or resilience for psychopathology. For both measures, associations with the Adversity/No Disorder group were not as robust as those for the two disorder groups. Within this group, there were no associations of telomere length or mtDNA with subclinical symptoms, perceived stress, or resilience, arguing against the possibility that subgroups of subjects who were at greater risk or more resilient were responsible for the intermediate association. The Adversity/No Disorder group had a lower burden of adversity than the Adversity/Disorder group (Table 1), which could explain why the effects were less robust.

It is of note that lifetime psychopathology without childhood adversity was also linked to telomere length and mtDNA copy number, suggesting that lifetime psychopathology and early adversity share some mechanisms of cellular aging. Within the No Adversity/Disorder group, telomere length was associated with perceived stress and with depressive and anxiety symptoms. Psychological and physiological stress associated with the disorders themselves or other adverse experiences could account for the effect.

It is notable that the Adversity/Disorder group did not differ from the groups with early stress alone or lifetime disorder alone, particularly given that these subjects had a higher burden of illness and stress exposure (Table 1). This could be due to "ceiling" or "floor" effects for mtDNA copy number and telomere length, respectively. There is a critical threshold for telomere shortening beyond which point the cell enters senescence or undergoes apoptosis, and production of excess ROS or other mechanisms might limit compensatory mitochondrial proliferation.

Because we used a sample of mixed leukocytes, it is possible that our findings are influenced by a difference in cell type that might covary with early stress and psychopathology. We excluded individuals with acute and chronic illness and pregnancy, but it remains possible that the groups differed with respect to leukocyte distribution. Few studies have examined these measures in leukocyte subtypes. There is evidence that telomere length is similar in lymphocytes and granulocytes in younger adults, but that lymphocyte telomere length declines at a faster rate with age (86). Given the large differences seen between our groups and the fact that we controlled for age, this would appear unlikely to explain the findings. Moreover, a recent study found that subjects with current depression and those with past episodes both had significantly shorter telomere length than healthy controls in each of three lymphocyte subpopulations (CD4+ helper T cells, CD8+ cytotoxic T cells, and CD20+ B cells) (84). For mtDNA copy number, there is evidence that whole blood is comparable to measurements in isolated lymphocytes (87).

The mechanisms by which mtDNA is elevated in conditions where telomere length is shortened remain to be elucidated. We do not have data on mitochondrial function, oxidative stress or telomerase levels so we cannot assess these factors. Future research should also examine mtDNA mutations; there is some evidence for a role of mtDNA variants, deletions, and mutations in various psychiatric disorders (12, 13) and it has been posited that some mutations might confer a replicative advantage (88). It is also possible that our findings were due to shared heritable influences on psychopathology, early stress, telomere length, and mtDNA copy number, although this is unlikely to fully account for the consistent pattern of findings in these different conditions and distinct markers of cellular aging. Future work could address such influences using multi-generation or twin designs. We did assess for effects of BMI, age, and smoking, but did not investigate quality of sleep, diet, and exercise (19, 89). A major strength of this study is that subjects did not have acute or chronic medical conditions and were not taking psychotropic medications which can alter mitochondrial biogenesis, gene expression, and function (12, 90) and telomere length (91).

In conclusion, we report new evidence of an alteration of mitochondrial biogenesis in association with early life stress and psychopathology. This study replicates prior findings of shortened telomere length in association with depressive and anxiety disorders and shows that this effect is not due to medical comorbidity or medication effects. Early adversity and lifetime psychopathology were uniquely linked to markers of cellular aging, and we suggest that this may be due to shared biological sequelae of these conditions.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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Figure 1.

Mitochondrial Copy Number by Adversity/Disorder Group Note. $* p < .05$, $*** p < .001$ in comparison with the No Adversity/No Disorder group. In addition, the Adversity/Disorder group was significantly different from the Adversity/No Disorder group ($p = .040$).

Figure 2.

Mitochondrial Copy Number by Disorder Type

Note. ** $p < .01$, *** $p < .001$ in comparison with the No Adversity/No Disorder group. Lifetime Depressive Disorder includes major depressive disorder (MDD), dysthymia, and depressive disorder not otherwise specified (NOS). Lifetime anxiety disorder includes posttraumatic stress disorder (PTSD), panic disorder, generalized anxiety disorder, social phobia, and anxiety disorder not otherwise specified (NOS).

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Mitochondrial Copy Number by Adversity Type

Note. $** p < .01$, $*** p < .001$ in comparison with the No Adversity/No Disorder group.

Figure 4.

Telomere Length by Adversity/Disorder Group

Note. ** $p < .01$, *** $p < .001$ in comparison with the No Adversity/No Disorder group.

Figure 5.

Telomere Length by Disorder Type

Note. * $p < .05$, *** $p < .001$ in comparison with the No Adversity/No Disorder group. Lifetime Depressive Disorder includes MDD, dysthymia, and depressive disorder NOS. Lifetime anxiety disorder includes PTSD, panic disorder, generalized anxiety disorder, social phobia, and anxiety disorder NOS.

Telomere Length by Adversity Type

Note. ** p < .01 in comparison with the No Adversity/No Disorder group.

Table 1

Demographics, disorders, and symptoms. Demographics, disorders, and symptoms.

Note: In four group analyses, p values indicate ANOVA significance level. In two group analyses, p values indicate t-test or chi-square significance level. For those four group analyses with p values < .05,
same superscrip Note: In four group analyses, p values indicate ANOVA significance level. In two group analyses, p values indicate t-test or chi-square significance level. For those four group analyses with p values < 05, same superscripts within the same line indicate groups are not significantly different from one another. Alcohol/Substance refers to alcohol abuse or dependence. Depressive disorders include MDD, dysthymia, and depression not otherwise specified. Anxiety disorders include PTSD, generalized anxiety disorder, social phobia, panic disorder, and anxiety disorder not otherwise specified. dysthymia, and depression not otherwise specified. Anxiety disorders include PTSD, generalized anxiety disorder, social phobia, panic disorder, and anxiety disorder not otherwise specified.