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Efficacy of Vitrectomy in Improving the Outcome of *Bacillus cereus* Endophthalmitis

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Abstract

Purpose—To evaluate the efficacy of vitrectomy with vancomycin for the treatment of experimental *Bacillus cereus* endophthalmitis.

Methods—Endophthalmitis was initiated in rabbits via intravitreal injection of 100 CFU *B. cereus*. Treatment groups included included 25-gauge transconjunctival sutureless vitrectomy with intravitreal vancomycin (1 mg) or vancomycin alone. Groups were treated at 4 h, 5 h, or 6 h postinfection. At 48 h (for 4 h and 5 h groups) or 36 h (for the 6 h group) postinfection, eyes were analyzed by electroretinography, histology, and inflammatory cell counts.

Results—Treatment with vitrectomy/vancomycin at 4 h resulted in significantly greater retinal function compared to that of vancomycin alone. Intraocular inflammation following treatment at 4 h was minimal for both treatment groups. Treatment with vitrectomy/vancomycin or vancomycin alone at 5 h or 6 h postinfection resulted in similar levels of retinal function loss (i.e. >90%) and significant intraocular inflammation.

Conclusions—These results demonstrate that vitrectomy may be of therapeutic benefit in the treatment of *B. cereus* endophthalmitis, but only during the early stages of infection.

Keywords

Bacillus; infection; eye; vancomycin; vitrectomy

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INTRODUCTION

Bacterial endophthalmitis is an infection that can result from contamination of the posterior segment following eye surgery, a penetrating eye injury, or from septic spread of infection into the eye from the bloodstream (1, 2). The incidence of post-traumatic endophthalmitis varies from 3 to 17% following a penetrating injury (3, 4) *Bacillus cereus* is a common bacterial cause of post-traumatic bacterial endophthalmitis. Patients with *B. cereus* endophthalmitis often present with severe ocular pain, periorbital swelling, proptosis of the globe, polymorphonuclear leukocytosis, and a fever (5–7). Only 9% of *B. cereus* endophthalmitis patients retain 20/70 vision or better, and nearly half require enucleation or evisceration of the infected eye (6). *B. cereus* is susceptible to commonly-used antibiotics aminoglycosides, fluoroquinolones, and vancomycin (8–10). However, the regularity of treament failures necessitates improvements in therapy of this blinding infection.

Vitrectomy is widely used to remove intravitreal contents following ocular injuries, inflammation, and other diseases. Microincision vitrectomy surgery, such as 23-and 25gauge vitrectomy, is often described as being minimally invasive (11, 12). The main concept of sutureless vitrectomy is to decrease postoperative inflammation, potentially resulting in early recovery and improvement of patient comfort. In rabbits, inflammation induced by 25gauge vitrectomy was less than that induced by 20- or 23-gauge vitrectomy (13). Vancomycin is a commonly administered intravitreal antibiotic used for treating intraocular infections, with reported 100% effectiveness against the most common Gram-positive ocular pathogens, including B. cereus (8-10). In an experimental B. cereus endophthalmitis model, vancomycin sterilized infected eyes when intravitreally administered as late as 6 h postinfection (14, 15). However, significant vision was lost if vancomycin was administered after 4 h postinfection in this model, indicating that early treatment was critical for salvaging vision. We recently reported potential vancomycin-based anti-inflammatory activity in this model (15). Vitrectomy and injection of intravitreal vancomycin sterilized the vitreous cavity following experimental staphylococcal endophthalmitis, resulting in minimal inflammation (16). Previous efficacy studies suggested that vancomycin (17–19) can be effective against experimental *B. cereus* endophthalmitis, but time courses of these infections may not have been clinically similar to that seen in rapid human infections.

The majority of recent studies analyzing the potential benefits of vitrectomy for the treatment of endophthalmitis have utilized experimental fungal infection models (20–22). For bacterial or fungal endophthalmitis, vitrectomy can be utilized as an effective strategy to remove not only dead organisms and cellular debris, but also damaging toxins and other inflammogenic factors that may exacerbate infection (23). We therefore hypothesized that additional surgical measures to remove offending toxic contents in the vitreous may improve the visual outcome of infection, which is important if treatment is delayed. To this end, we analyzed the therapeutic effectiveness of vitrectomy and intravitreal vancomycin with that of vancomycin alone in a well-established *B. cereus* endophthalmitis rabbit model (14, 15) to determine whether vitrectomy and antibiotics offered an improve therapeutic benefit over that of antibiotics alone.

SUBJECTS AND METHODS

Subjects and Drugs

Specific pathogen-free New Zealand White rabbits (male, 2–4 kg, Myrtle's Rabbitry, Thompsons Station, TN) were used in this study and were maintained in accordance with institutional guidelines and the Association for Research in Vision and Ophthalmology Statement on the Use of Laboratory Animals in Ophthalmic Research (online). Prior to intravitreal injection and retinal function analysis (electroretinography [ERG]), rabbits were anesthetized with an intramuscular injection of ketamine (Ketamine HCl Injection, Bioniche Pharma, Lake Forest, IL; 35 mg/kg of body weight) and xylaxine (RompunTM; Bayer Corp., Shawnee Mission, KS; 5mg/kg of body weight). Proparacaine HCl (OphtheticTM; Allergan, Hormigueros, Puerto Rico; 0.5%) was used to topically anesthetize the eyes prior to paracentesis and intravitreal injection. Vancomycin (1 mg final concentration in 100 μL PBS, Hospira, Lake Forest IL) was administered by intravitreal injection or immediately following vitrectomy. Vancomycin (1 mg) was previously demonstrated to be effective against *B. cereus* and non-toxic in this model (14, 15).

Experimental Endophthalmitis

Rabbit eyes were intravitreally infected with *B. cereus* as previously described (14, 15, 24, 25). Briefly, an overnight culture of *B. cereus* strain ATCC 14579 (American Type Tissue Culture, Manassas, VA) was subcultured into brain heart infusion media (BHI; Difco Laboratories, Detroit, MI), and serially diluted to 100 CFU/0.1 ml for intravitreal injections. Contralateral eyes served as a non-injection control. The MIC of *B. cereus* ATCC 14579 for vancomycin was 1.95 µg/mL, as reported previously (14).

Vitrectomy/Vancomycin Therapy

The treatment regimens chosen for this study sought to mimic an elapsed time when a patient suffering from a penetrating injury may reasonably expect to receive treatment. Previous studies have demonstrated that at and prior to 4 h, intravitreal administration of vancomycin can reduce inflammation and salvage significant vision (14, 15). We sought to determine whether vitrectomy further reduced the inflammation and vision loss associated with delayed treatment.

A total of 44 NZW rabbits were randomized to 7 treatment groups. At 4 h, 5 h, or 6 h postinfection, one eye of each rabbit underwent either 25-gauge transconjunctival sutureless vitrectomy (TSV-25 Millennium System; Bausch & Lomb Inc., Rochester NY) with intravitreal instillation of vancomycin or intravitreal instillation of vancomycin alone. A non-infected vitrectomy/vancomycin group was included as a control.

Vitrectomy was performed as follows. After general and topical anesthesia, a standard 3ports pars plana vitrectomy was performed with removal of all visible vitreous gel and inflammatory debris. One-step transconjunctival cannula insertion was achieved using a beveled trochar at 2.0 mm from the corneoscleral limbus. Three incisions were made and the infusion cannula was inserted into the inferotemporal cannula. Core vitrectomies removed vitreous using the Millennium 25-gauge high speed vitrector for 10 min by a combination of

cutting (1500 cuts/min) and suction (400 mmHg), while continually supplying BSS irrigating solution at an ocular fluid pressure of 30 mmHg. Upon completion of the surgery, the vitreous was infused through the cannula with 0.1 ml of 1.0% vancomycin. The surgical time required from opening to closure was an average of 20 min. Surgery was completed by removal of the entry site cannulas without scleral suturing. An experienced vitreoretinal surgeon performed all vitrectomies and was masked to the identity of the groups. There were no complications during surgery.

At 12, 24, and 48 h (for eyes treated at 4 h or 5 h) or 12, 24, and 36 h (for eyes treated at 6 h) postinfection, eyes were analyzed as described below.

Analysis of Therapeutic Efficacy

Retinal Function Analysis—Retinal function was measured and recorded (UTAS3000; LKC Technologies, Inc., Gaithersburg, MD) for both eyes of each rabbit by scotopic electroretinography (ERG) as previously described (14, 24–27). Prior to ERG, eyes were dilated with phenylephrine HCl (AK-Dilate[®]; Akorn, Inc., Buffalo Grove, IL) and dark adapted for 10 min. A-wave amplitudes were measured from the pre-stimulus baseline to the A-wave trough, while B-wave amplitudes were measured from the trough of the A-wave to the peak of the B-wave. The following equations were used to calculate the percentage of retinal function retained (experimental = infected; absolute control = uninjected): (i) 100 – {[1-(experimental A-wave amplitude/absolute control A-wave amplitude)] × 100} or (ii) 100 – {[1-(experimental B-wave amplitude/absolute control B-wave amplitude)] × 100} (14, 24–27).

Biomicroscopy and Histology—An operating biomicroscope (Zeiss S7; Zeiss Inc, Thornwood, NY) was used to visualize and photograph rabbit eyes. Changes in anterior and posterior segment inflammation and retinal architecture were scored in a masked independent fashion based on a scale from 0 (no change) to 4+ (significant inflammation and retinal architecture damage) (28). Eyes used for histology were enucleated, fixed in 10% formalin for 24 h, placed in 70% alcohol for 48 h, paraffin sectioned, and stained with hematoxylin and eosin by standard procedures.

Bacterial Quantitation—Viable bacteria in aspirated vitreous were quantified, as described previously (14, 24–27). Briefly, eyes were harvested after euthanasia, vitreal contents were aspirated and homogenized, aspirates were serially diluted in PBS, and aliquots were plated out in triplicate on BHI agar for quantitation.

PMN Quantitation—Quantitation of infiltrating PMN into the aqueous humor is a direct measure of the progression of intraocular inflammation throughout the eye (25–27). Prior to harvest, eyes underwent paracentesis, aqueous samples were loaded onto a hemocytometer, and PMN were manually counted following trypan blue staining.

Antibiotic Penetration into the Eye—Antibiotic diffusion assays were used to quantify the concentration of antibiotics in the vitreous and aqueous humor, as previously described (14). Indicator strains (10^5 CFU/ml of *S. aureus* or *K. pneumoniae*) were inoculated onto BHI agar for vancomycin or gatifloxacin bioassays, respectively. Sterile filter discs loaded

with 10 μ l of aqueous humor, vitreous, or antibiotic standards were placed onto the inoculated agar. Standards and samples were prepared and analyzed in triplicate. Assays were incubated at 37°C for 24 h, and zones of inhibition were measured to the nearest 0.1mm. A standard curve of zone of inhibition size versus log₁₀ concentration was plotted and used to determine antibiotic concentrations, the slope of which was determined from a best-fit curve by least-square means method.

Statistics—For the PMN quantitation and antibiotic penetration assays, all values represent the mean \pm standard error of the mean (SEM) for 4 replicate samples per time point. All other reported values represent the mean \pm SEM for 5 eyes per time point. Two-tailed, two-sample Student's *t*-tests were used to statistically compare groups. A P-value of 0.05 was considered significant.

RESULTS

Bacterial Killing

All eyes treated with vancomycin, with or without vitrectomy, were sterile at 12 h postinfection. These results are consistent with previous reports of vancomycin treatment in this infection model (14, 15)

Retinal Function

Retinal function retained following each treatment is summarized in Figure 1. In uninfected eyes, vitrectomy with vancomycin resulted in an approximate 40% decrease in A-wave amplitude and 35% decrease in B-wave amplitude at 48 h post-surgery.

Treatment at 4 h postinfection with vitrectomy and vancomycin resulted in a 45% reduction in A-wave amplitude and a 50% reduction in B-wave amplitude at 48 h. These values are less than but similar to those of the uninfected vitrectomy/vancomycin-treated group (P=0.07 A-wave, P=0.13 B-wave). These values are significantly greater than that of the vancomycin alone group, which resulted in70% and 85% reductions in A-wave and B-wave amplitudes, respectively (P=0.02 A-wave, P=0.004 B-wave). Five-h or 6-h treatment resulted in significant loss of vision (i.e. >95% loss) regardless of whether vitrectomy was part of the vancomycin treatment regimen. These results demonstrate that vitrectomy can reduce the vision loss associated with experimental *B. cereus* endophthalmitis, but only if treatment is begun no later than 4-h postinfection in this model.

Biomicroscopy and Histology—Representative biomicroscopy and histology data is presented in Figure 2. Prior to surgery, infected eyes at 4, 5, and 6 h postinfection demonstrated mild iritis and mild to moderate vitritis (scores of 1+ to 2+). Uninfected eyes were normal. Immediately after surgery, these observations were unchanged except for minimal conjunctival injection caused by speculum placement.

Uninfected eyes treated with vitrectomy and vancomycin demonstrated mild iritis and vitritis (scores of 1+) at 48 h postinfection. The histology data corroborate this observation, with fibrin and cellular infiltrate seen in the posterior segment and intact retinas. Infected eyes treated with vitrectomy and vancomycin appeared similar to that of uninfected treated

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eyes at 4 h, with mild iritis and mild to moderate vitritis (scores of 1+ to 2+). The histology data demonstrated significant infiltrate into the anterior and posterior segments, but retinas remained relatively intact. Vitrectomy/vancomycin treatment at 6 h resulted in eyes with moderate cell and flare and vitritis (scores of 3+), with the occasional hypopyon. The histology results of the vitrectomy/vancomycin 6-h treatment directly corroborate these findings. Biomicroscopy and histology of vancomycin-treated eyes at 4 and 6-h postinfection has been reported elsewhere (14, 15), with similarities in biomicroscopy scores and histology findings between the two treatment groups at each time point. As with the 6-h treatment groups, eyes treated with vitrectomy/vancomycin or vancomycin alone at 5 h postinfection were similar in both the rate of evolution and severity of endophthalmitis signs. These findings were corroborated by histology data.

Inflammation—Intraocular inflammation was estimated by counting PMN in harvested aqueous. The results are summarized in Figure 3. Vitrectomy/vancomycin treatment of uninfected eyes resulted in infiltration of PMN detected at 48 h post-surgery. The numbers of PMN in this group were similar to that of infected eyes treated with vitrectomy/ vancomycin at 4 h postinfection (P=0.22). The numbers of PMN in infected eyes treated with vitrectomy/vancomycin were significantly less than that of infected eyes treated with vancomycin alone at 4 h postinfection (P=0.02). The numbers of PMN recovered from infected eyes treated at 5 h or 6 h postinfection with vitrectomy/vancomycin or vancomycin alone were similar (P 0.08). Recall that eyes treated at 5 h were recovered at 48 h postinfection, while eyes treated at 6 h were recovered at 36 h postinfection. These results further corroborate that either type of treatment after 4 h postinfection leads to significant inflammation in this model.

Intraocular Antibiotic Levels

Significant bactericidal levels must be achieved in an infected eye following antibiotic treatment to prevent the infection from progressing. In vitrectomy-treated infected eyes, intraocular contents were removed and replaced with balanced salt solution containing vancomycin, and vancomycin levels were later quantified (Figure 4). No vancomycin was detected in the aqueous humor at 48 h postinfection regardless of the time of treatment or whether vitrectomy was involved in the regimen. Vancomycin was detected in the vitreous of infected eyes treated at 4 h or 5 h postinfection, but not in the vitreous of infected eyes treated at 4 h or 5 h postinfection, but not in the vitreous of infected eyes treated at 5 h postinfection, these values were not significantly different from those following treatment at 4 h postinfection (P 0.08). Vancomycin levels were also similar at these time points regardless of whether vitrectomy was involved in the regimen (P 0.3). When vancomycin was detected, these concentrations were well above the MIC for this particular *B. cereus* strain.

DISCUSSION

Vitrectomy is designed to remove potentially harmful contents and pathogens from the inside of the eye in an effort to minimize inflammation and salvage vision during many types of ocular infections. This study demonstrates the efficacy of vitrectomy and

vancomycin over that of vancomycin alone for the salvaging vision and limiting inflammation in experimental *B. cereus* endophthalmitis. However, vitrectomy and vancomycin were more effective than vancomycin alone only when treatment was initiated prior to 4 h postinfection. After that time, all treatments were relatively ineffective, resulting in significant inflammation and vision loss.

In this model, 4 h postinfection is the critical time within which intravitreal treatment must be initiated to salvage useful vision. *B. cereus* endophthalmitis is unique in its rapid course and invariably devastating outcome. The experimental rabbit model used herein reproducibly mimicks that course. *B. cereus* synthesizes multiple toxins in the eye during experimental infection (23), and we have demonstrated that toxins are involved in vision loss in this disease (24, 25). Which toxins are involved, the kinetics of their synthesis, and the specific activities on the retina are all open questions. However, considering the *in vitro* and *in vivo* findings of the importance of quorum sensing to *B. cereus* toxin production and virulence (25, 29–32), one can envision a scenario in which *B. cereus* reaches a threshold quorum in the eye between 2 and 4 h postinfection, begins to synthesize toxins, the retina is affected, and vision loss occurs. Once treatment is begun, the eye is sterilized, toxins are no longer produced, and the retinal damage and vision loss is limited to that already done.

In uninfected control eyes, vitrectomy with vancomycin caused inflammation and retinal function loss. Vitrectomy has been associated with blood-retinal barrier breakdown and other physiological changes (33–35). Inoue *et al.* (13) recently reported that vitreal protein concentrations decreased from 1 to 7 days following 20-, 23-, or 25-gauge vitrectomy, indicating that surgically-induced inflammation began to resolve within a week in this model. Wallenten *et al.* (36) demonstrated significant reductions in retinal function that were detected up to 28 days post-vitrectomy. In that study, upregulation of retinal GFAP, an indicator of vitrectomy-related retinal trauma, was also detected in vitrectomy-treated rabbit eyes. The majority of recent studies on retinal function following vitrectomy have been short-term toxicity or clearance experiments (37–39). Intravitreal injection of PBS, bacterial media, or vancomycin alone can cause a transient posterior segment inflammation that, in the rabbit eye, resolves within 4 h (data not shown). Therefore, any changes observed in uninfected eyes treated with vitrectomy and vancomycin are likely attributed to vitrectomy alone.

In comparing our results in this study with that of our previous therapeutic studies, we noted that retinal function loss was greater, eyes appeared to be clinically worse, and antibiotic concentrations were less in comparable treatment groups. The difference between this study and the previous two therapeutic studies (14, 15) was the animal. All three studies used New Zealand White rabbits of similar age. However, the present study used specific pathogen-free rabbits, while the previous studies used rabbits raised in conventional conditions. It is therefore possible that the difference in pathogenicity and clinical outcome seen in this study lie in the overall immune status of the rabbit. An animal raised in a pathogen-free environment may be more susceptible to infections with organisms that have not previously been encountered and may not be able to quickly mount the degree of immune response needed to fight the infection (40). This may account for the elevated severity of infection at an earlier time point in the present study compared to our previous studies. Although our

results draw clear comparisons among efficacies of the treatment regimens tested, careful interpretation of potential model-to-model variations must be considered when comparing the effectiveness of a particular experimental regimen. Nevertheless, these results reinforces the critical necessity for early antibiotic treatment of *B. cereus* endophthalmitis and indicates that timely vitrectomy in addition to antibiotics may result in a better therapeutic outcome than the use of antibiotics alone.

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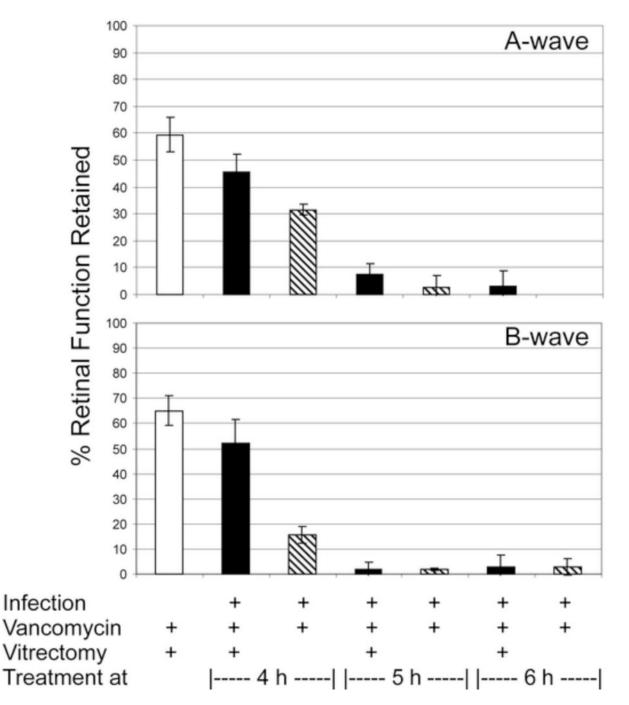


Figure 1.

Retinal function analysis following treatment of experimental *B. cereus* endophthalmitis with vitrectomy and vancomycin. Eyes were infected with *B. cereus* and treated with vancomycin (1%) and vitrectomy or vancomycin alone at various times postinfection. The control group included uninfected eyes treated with vitrectomy and vancomycin. Eyes were analyzed by electroretinography at 36 h or 48 h postinfection postinfection. Values represent the mean \pm SEM of N 5 eyes per group.

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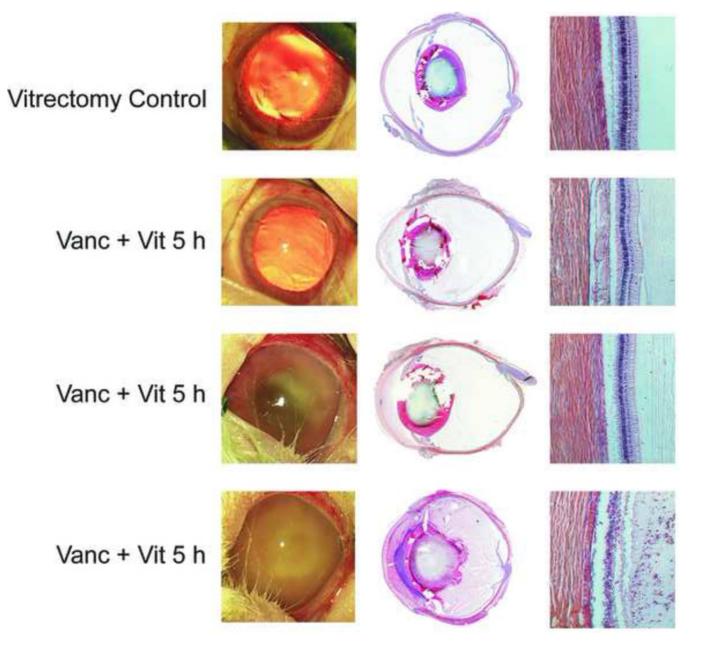


Figure 2.

Photography and histology of eyes following treatment of experimental *B. cereus* endophthalmitis with vitrectomy and vancomycin (1%) at various times postinfection. The control group included uninfected eyes treated with vitrectomy + vancomycin. Eyes were photographed, then harvested for histology and hematoxylin and eosin staining. Figures are representative of N=3 eyes per group. Vanc = vancomycin, Vit = vitrectomy. Magnification of retina sections, $100\times$.

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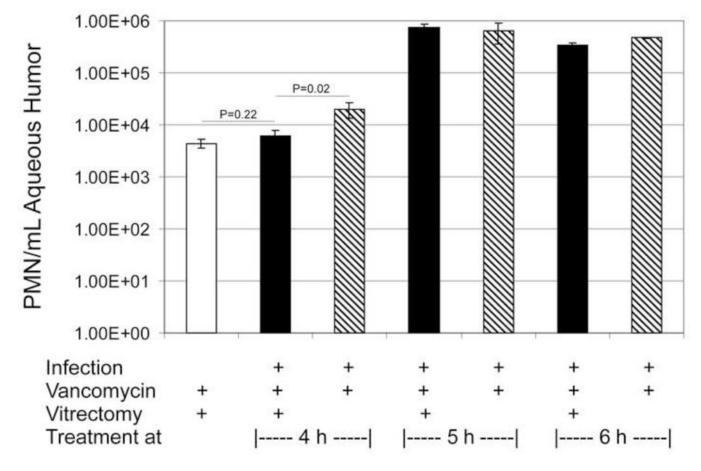


Figure 3.

PMN infiltration into the anterior segment following treatment of experimental *B. cereus* endophthalmitis with vitrectomy and vancomycin. PMN were quantified from aqueous humor samples at 48 h postinfection for eyes treated and 4 and 5 h or at 36 h postinfection for eyes treated at 6 h postinfection. The values represent the mean \pm SEM of N 3 eyes per group.

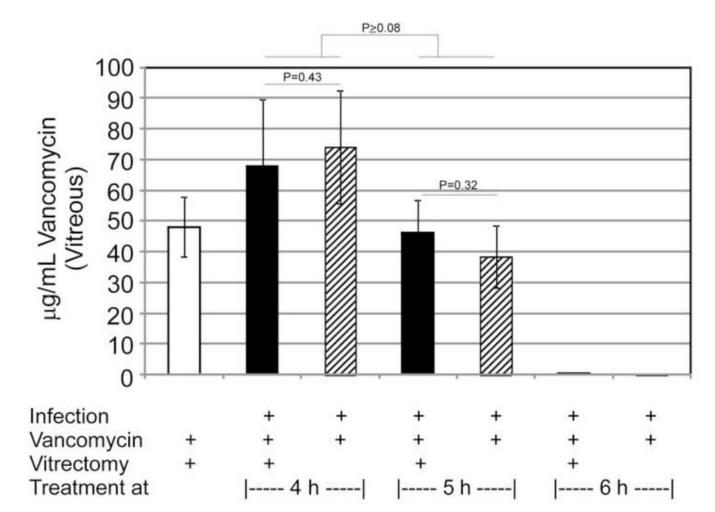


Figure 4.

Ocular vancomycin concentrations following treatment of experimental *B. cereus* endophthalmitis with vitrectomy and vancomycin. Aqueous and vitreous were harvested at 48 h postinfection for eyes treated and 4 and 5 h or at 36 h postinfection for eyes treated at 6 h postinfection. Vancomycin concentrations were quantified using a standard bioassay. The values represent the mean \pm SEM of N 3 eyes per group. Vancomycin was not detected in any aqueous samples, so only vitreous results are shown.