

Genome Sequence of *Geobacillus* sp. Strain ZGt-1, an Antibacterial Peptide-Producing Bacterium from Hot Springs in Jordan

Rawana N. Alkhalili,^a Rajni Hatti-Kaul,^a Björn Canbäck^b

Department of Biotechnology, Center for Chemistry and Chemical Engineering, Lund University, Lund, Sweden^a; Department of Biology, Microbial Ecology Group, Lund University, Lund, Sweden^b

This paper reports the draft genome sequence of the firmicute *Geobacillus* sp. strain ZGt-1, an antibacterial peptide producer isolated from the Zara hot spring in Jordan. This study is the first report on genomic data from a thermophilic bacterial strain isolated in Jordan.

Received 22 June 2015 Accepted 22 June 2015 Published 23 July 2015

Citation Alkhalili RN, Hatti-Kaul R, Canbäck B. 2015. Genome sequence of *Geobacillus* sp. strain ZGt-1, an antibacterial peptide-producing bacterium from hot springs in Jordan. *Genome Announc* 3(4):e00799-15. doi:10.1128/genomeA.00799-15.

Copyright © 2015 Alkhalili et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to Rawana N. Alkhalili, rawana.alkhalili@biotek.lu.se.

Lately, the species of the genus *Geobacillus* have been gaining interest as antimicrobial peptide producers (1, 2). *Geobacillus* sp. strain ZGt-1, isolated from the Zara hot spring in Jordan, has been shown to produce an as-yet-uncharacterized antimicrobial peptide (3). In order to screen for the antibacterial protein-encoding genes and to identify potential novel genes associated with antibacterial peptide biosynthesis, we performed a whole-genome sequencing of the bacterium that was already identified by sequencing the PCR-amplified 16S rRNA gene (GenBank accession no. KT026965).

Here, we report the genome sequence of *Geobacillus* sp. strain ZGt-1. Total genomic DNA was extracted from pure cultures of the isolate using ZR Fungal/Bacterial DNA MiniPrep (Zymo Research). A DNA library was constructed using the Nextera protocol with modifications as described earlier (4). Input to the assembly consisted of 680,000 single-end Illumina reads with a length of 151 nucleotides. Quality control was performed by the FastQC version 0.11.2 software (<http://www.bioinformatics.bbsrc.ac.uk/projects/fastqc>). Reads were assembled using Velvet and Velvetg, both with version 1.2.10 (5). This resulted in an assembly containing 9,625 contigs with a total length of 3.7 million bp. Taking into account that genome sequences from closely related strains were available, it was decided to produce ZGt-1 scaffolds based on the genome sequence of *Geobacillus kaustophilus* HTA426 (GenBank accession number NC_006510.1). This was conducted online with the Scaffold_builder tool (6) using default settings. This resulted in a new assembly with 241 scaffolds and a total length 3,483,107 bp. On this final assembly, gene prediction was carried out with Prodigal version 2_60 using default settings (7). The predicted number of protein-encoding genes was 3,546, which is close to the reported number of genes from *G. kaustophilus* HTA426 (3,397 protein-encoding genes). The GC content was calculated to 52.2% and gene density to 88%.

Genome analysis using antiSMASH version 3.0 software (8) revealed that strain ZGt-1 harbors a lantipeptide biosynthetic gene cluster, where one of the genes encodes for a lantipeptide similar to geobacillin I. The presence of this cluster was also con-

firmed using BAGEL version 3.0 software (9). The antiSMASH also revealed that the strain harbors another cluster containing a gene encoding for a bacteriocin similar to Linocin M18. A number of putative genes found in the lantipeptide and bacteriocin clusters showed low percentage identity with already described genes. This indicates that the ZGt-1 strain possibly possesses novel genes related to antibacterial peptide production.

Combining the *in silico* analysis of the draft genome of strain ZGt-1 with *in vitro* experimentation is likely to lead to the discovery of novel bioactive compounds.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited in DDBJ/EMBL/GenBank under the accession number [LDPD00000000](https://www.ncbi.nlm.nih.gov/nuccore/LDPD00000000). The version described in this paper is the first version, [LDPD01000000](https://www.ncbi.nlm.nih.gov/nuccore/LDPD01000000).

ACKNOWLEDGMENTS

This work was supported by Erasmus Mundus Partnership (JOSYLEEN).

We thank Gerton Lunter and the High-Throughput Genomics Group at the Wellcome Trust Centre for Human Genetics (funded by Wellcome Trust grant reference 090532/Z/09/Z), Oxford, United Kingdom, for the generation of sequencing data.

REFERENCES

- Garg N, Tang W, Goto Y, Nair SK, van der Donk WA. 2012. Lantibiotics from *Geobacillus thermodenitrificans*. *Proc Natl Acad Sci USA* 109: 5241–5246. <http://dx.doi.org/10.1073/pnas.1116815109>.
- Pokusaeva K, Kuisiene N, Jasinskyte D, Rutiene K, Saleikiene J, Chitavichius D. 2009. Novel bacteriocins produced by *Geobacillus stearothermophilus*. *Open Life Sciences* 4:196–203. <http://dx.doi.org/10.2478/s11535-009-0009-1>.
- Alkhalili R, Dishisha T, Mamo G, Hatti-Kaul R. Abstr 667, Abstr 3rd Int Conf Antimicrob Res, 1–3 October 2014, Madrid, Spain.
- Lamble S, Batty E, Attar M, Buck D, Bowden R, Lunter G, Crook D, El-Fahmawi B, Piazza P. 2013. Improved workflows for high throughput library preparation using the transposome-based nextera system. *BMC Biotechnol* 13:104. <http://dx.doi.org/10.1186/1472-6750-13-104>.
- Zerbino DR, Birney E. 2008. Velvet: algorithms for *de novo* short read assembly using de Bruijn graphs. *Genome Res* 18:821–829. <http://dx.doi.org/10.1101/gr.074492.107>.
- Silva GG, Dutilh BE, Matthews TD, Elkins K, Schmieder R, Dinsdale EA,

- Edwards RA. 2013. Combining *de novo* and reference-guided assembly with scaffold_builder. Source Code Biol Med 8:23. <http://dx.doi.org/10.1186/1751-0473-8-23>.
7. Hyatt D, Chen GL, Locascio PF, Land ML, Larimer FW, Hauser LJ. 2010. Prodigal: prokaryotic gene recognition and translation initiation site identification. BMC Bioinformatics 11:119. <http://dx.doi.org/10.1186/1471-2105-11-119>.
8. Weber T, Blin K, Duddela S, Krug D, Kim HU, Brucoleri R, Lee SY, Fischbach MA, Müller R, Wohlleben W, Breitling R, Takano E, Medema MH. 6 May 2015. antiSMASH 3.0—a comprehensive resource for the genome mining of biosynthetic gene clusters. Nucleic Acids Res. <http://dx.doi.org/10.1093/nar/gkv437>.
9. van Heel AJ, de Jong A, Montalbán-López M, Kok J, Kuipers OP. 2013. BAGEL3: automated identification of genes encoding bacteriocins and (non-)bactericidal posttranslationally modified peptides. Nucleic Acids Res 41:W448–W453. <http://dx.doi.org/10.1093/nar/gkt391>.