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Heatshock protein vaccines against Glioblastoma: From bench to bedside

Leonel Ampie¹, Winward Choy¹, Jonathan B Lamano¹, Shayan Fakurnejad¹, Orin Bloch^{1,*}, and Andrew T. Parsa¹

¹Department of Neurosurgery, Northwestern University Feinberg School of Medicine, Chicago, IL

Abstract

Current adjuvant treatment regimens available for the treatment of glioblastoma are widely ineffective and offer a dismal prognosis. Advancements in conventional treatment strategies have only yielded modest improvements in overall survival. Immunotherapy remains a promising adjuvant in the treatment of GBM through eliciting tumor specific immune responses capable of producing sustained antitumor response while minimizing systemic toxicity. Heat Shock Proteins (HSP) function as intracellular chaperones and have been implicated in the activation of both innate and adaptive immune systems. Vaccines formulated from HSP-peptide complexes, derived from autologous tumor, have been applied to the field of immunotherapy for glioblastoma. The results from the phase I and II clinical trials have been promising. Here we review the role of HSP in cellular function and immunity, and its application in the treatment of glioblastoma.

Keywords

vaccine; heat shock protein; glioblastoma; glioma; immunotherapy; clinical trial

Introduction

A diagnosis of glioblastoma (GBM) portends a bleak prognosis due to the malignant properties intrinsic to the neoplasm and the limited therapeutic options available for treatment. The current standard-of-care, acknowledged as the “Stupp protocol”, remains relatively poor, resulting in a median overall survival (OS) of 14.6 months with adjuvant temozolomide-based chemoradiation after optimal resection[1]. Concurrent with the disheartening GBM statistics over the last decade were advances in immunotherapy treatments for metastatic systemic-based cancers. These advances, along with our increased knowledge of the altered genomic landscape of GBM in predicting potential antigens, have sparked and propelled great interest in not only harnessing the immune system to target glioblastoma, but also investigating how GBM modulates the immune system.

As opposed to lower grade glial neoplasms, GBM is highly antigenic and is enriched with lymphocytic infiltrates. A large contributor to the infiltrative cohort of lymphocytes

*Corresponding Author: Department of Neurological Surgery, Feinberg School of Medicine, Northwestern University, 676 N. St. Clair Street, Suite 20-250, Chicago, IL 60611-2911, USA, orin.bloch@northwestern.edu.

phenotypically express CD4+ CD25+ FOXP3+ markers, thus identifying them as a regulatory T-cell (T_{Reg}) population which may serve to limit the immunogenic response of infiltrative cytotoxic lymphocytes (CTLs)[2]. Direct cellular inhibition of lymphocytes is also attributed to glioma cells as they acquire a higher expression of B7-H1 (PD-L1) with the loss of the PTEN tumor suppressor gene [3]. Additional immune-limiting barriers that patients face are attributed to systemic immunosuppression propagated by the neoplasm itself as well as the suppression imposed by the cytotoxic nature of the chemotherapeutic agent administered after resection[4]. Thus, current and future immunotherapy agents are tasked with the challenge of eliciting an immunogenic response towards GBM that can overcome both the local and systemic state of immunosuppression.

Immunotherapy can be stratified into either an active or passive form in terms of function. An example of a passive intervention would be monoclonal antibodies directed at aberrant neoplastic proteins or even immune checkpoint proteins which restrain immunogenic attack on a specific target. This method is considered passive because it does not directly stimulate the host immune system and relies on continuous exogenous introduction in order to gain benefit from the therapy. In contrast, the active arm of immunotherapy aims to educate the host immune system so it can autonomously train naïve immune cells against antigenic targets. An example of this would consist of vaccines which may be taken up and presented by resident antigen presenting cells (APCs) to lymphocytes.

There are a number of vaccines which have been recruited for the treatment against glioblastoma[5]. Peptide vaccines introduce short protein sequences of known antigenic entities within glioblastoma (e.g. EGFRviii) in order to elicit an immune response against the neoplastic cells harboring the mutant proteins [6]. Autologous vaccines are based on retrieving a patient's peripheral blood cells, modifying them (e.g. stimulating with known tumor antigens or altering the autologous tumor cells with viruses) and re-infusing the primed immune cells back into the host[7]. Dendritic-cell-based vaccines pulse dendritic cells, isolated from peripheral blood mononuclear cells, with glioma antigens retrieved from resected tumors so they can stimulate naïve lymphocytes when reintroduced into the host[8]. In this review, we will focus primarily on heat shock protein (HSP)-peptide based vaccines. This vaccination method deals with the isolation and purification of HSPs from resected GBM patients with subsequent reinfusion of the complex to allow the chaperone to interact with APCs, thus priming the lymphocytes with a varied cohort of antigenic peptides.

Cellular function of heatshock proteins

Heat shock proteins (HSPs), also known as chaperone proteins, are abundant across mammalian cell types, in which they play a vital role in the stress response to cellular insults including hyperthermia, inflammation, hypoxia, oxidative stress, and radiation.[9] The assembly and transport of nascent proteins within the cell relies on the activity of HSPs, especially in adverse intracellular situations where HSPs function to stabilize proteins and prevent aggregation.[10] In addition, HSPs also act to resolve protein aggregates, reassemble salvageable misfolded proteins, and guide the degradation of unsalvageable misfolded proteins following the resolution of cellular insults.[11] As a result, it is believed that HSPs are transcriptionally upregulated in cancer where there exists increased translation

of abnormal protein products.[12] Analyses based on molecular weight and phylogenetics have distinguished five major HSP families, however only HSP gp 96, HSP 90, HSP 70, HSP 110, and HSP 170 have demonstrated immunogenic interactions as membrane-bound and extracellular components.[13, 14]

Specifically, in GBM, elevated constitutive and inducible expression of HSP27, α B-crystallin, HSP72, HSP73, and HSP90 has been reported both *in vitro* and *in vivo*. [15, 16] Moreover, HSP27, HSP60, HSP70, and HSP90 have been shown to be present in GBM released exosomes.[16] Of particular interest to GBM, however, are HSP70 and HSP90. The HSP70 family functions to inhibit cell stress induced apoptotic pathways, facilitate protein folding, and guide protein transport across membranes.[11] Recently, increased transcription of HSP70 mRNA was shown to correlate with glioma grade.[17] Moreover, HSPs within the HSP70 family of chaperone proteins were the first HSPs shown to bind antigenic peptides. [18] The family of HSPs to which HSP90 belongs is largely responsible for protein folding, protein stabilization, and peptide loading onto MHC class I molecules. Importantly, HSP90 substrates (including EGFRvIII, FAK, AKT, hTERT, p53, cdk4, MAPK, and PI3K) are involved in key tumor initiation and proliferation signaling pathways.[11] Similar to HSP70, HSP90 is also associated with the binding of tumor antigens that can elicit a tumor rejection response.[19, 20] As a result, HSPs have been targeted as potential vehicles by which to present tumor specific antigens in glioblastoma to elicit an antitumor immune response.

Heatshock proteins in immunity

A strength possessed by the HSP-vaccine is the ability to stimulate both the innate and adaptive immune responses. Alone, neither the HSP or the isolated peptides are immunogenic; only when complexed they are able to elicit an MHC-class I CD8+ cytotoxic T-lymphocyte (CTL) response[24]. Classically, exogenous antigens have been known to primarily be presented via MHC class II on APCs which promotes the interaction with T-helper cells (CD4+). HSP complexes have the added benefit of being able to undergo the endogenous MHC class I pathway to induce a CD8+ response. The CD91 receptor allocated on APCs is responsible for the uptake of the HSP complex into the cell [25]. Upon internalization, the complex undergoes processing via proteasomes, gets transported into the ER, and is ultimately loaded onto to MHC-class I for presentation to CD8+ CTLs[26]. In addition to this cytosolic pathway, an endosomal method which is proteasome-independent is also possible for loading the peptides onto MHC class I[27]. A small portion of the internalized HSP complex can also enter an acidic compartment which leads to MHC-class II loading of the peptide for the stimulation of CD4+ cells[28]. There are also other potential receptors which the HSP complex may interact, leading to a non-CD91 presentation of chaperoned proteins[29].

HSP-peptide complexes (HSPPCs) have the ability to interact with a number of cell surface receptors on APC's which induces downstream activation of the NF- κ B pathway (Figure 1). Some of these receptors are believed to include: CD36/CD91/CD40/CD14/Toll-like receptor 2 (TLR2) /Toll-like receptor 4(TLR4). There are potentially other cell surface receptors that interact with the HSP complex that have yet to be elucidated. In macrophages in particular, HSP stimulates the secretion of proinflammatory cytokines such as tumor necrosis factor α .

(TNF α), granulocyte macrophage colony-stimulating factor (GM-CSF), IL-12, and IL-1 β . IL-12 may serve to activate the cytotoxic activity of both lymphocytes and natural killer (NK) cells. Additionally, HSP complexes are able to augment the production and secretion of nitric oxide in both dendritic cells and macrophages[30]. Interestingly, the combination of these secretory products induced by the interaction of HSP with cell surface receptors on macrophages coincide with the proinflammatory phenotype of macrophages (M1). The HSPPC additionally induces immature dendritic cells to undergo maturation which is noted by the increased expressivity of MHC class II and CD86 as well as the increased secretion of IL-12 and TNF α which further potentiate a proinflammatory response[31].

One advantage possessed by HSP-based vaccinations is that they are not specific to one pre-defined antigen. While other vaccine modalities target one specific GBM antigen (e.g. EGFRviii), HSPPCs manage to present various types of potential antigenic proteins upon vaccination. This is a crucial facet to this vaccine methodology due to the intratumoral heterogeneity posed by glioblastoma. One of the hallmarks of cancer in general is that of immunoediting which selects for the non-immunogenic subset of cells within a tumor to survive and thrive. By vaccinating individuals with a patient-specific polyvalent HSPPC vaccine, it may provide an added advantage compared to vaccination against one specific antigen. An additional advantage of HSPPC vaccines is that they manifest their immunogenic benefit via multiple mechanisms which augment cytotoxic effects via other cell types in addition CTLs. A potential drawback to blindly vaccinating against unknown antigenic variants is that the immune system may be trained to target antigens which are not ultimately essential and expressed only in a minority of neoplastic cells. Although, this limitation may be hampered via epitope spreading whereby immune cells originally primed for a specific antigenic epitope can detect different unrelated epitopes, allowing for the detection of new antigens on the peptide[32]. This may theoretically further increase the antigenic repertoire of the induced immune response, thus allowing the immune system to target neoplastic cells with distinct antigenic epitopes differing from the initial epitope used for lymphocytic priming.

Heatshock protein vaccines

Following positive results acquired in the preclinical setting, phase I, II, and III clinical trials were conducted to investigate the safety and efficacy of vaccination with autologous tumor-derived HSP-peptide complex based antitumor vaccines in various tumor types.. The majority of HSP vaccine trials have utilized heat shock protein-peptide complex-96 (HSPPC-96), comprising autologous antigenic peptides chaperoned by HSP glycoprotein-96 (HSP gp-96) While other HSP families have share similar theoretical advantages for clinical translation, early pilot studies have demonstrated HSPPC-96 to be safe with minimal toxicity, and feasible with regards to purification and production as a clinical grade product. HSPPC-96 vaccines, developed from resected tumor specimens that are frozen and delivered to the vaccine manufacturer. Utilizing liquid chromatography, HSPs are isolated and further enriched by subsequent denaturing gel electrophoresis and anti-gp 96 western blot[33, 34]. Including sterility and endotoxin quality control screening, the manufacturing process takes three to four weeks from the date of tumor resection to vaccine release[35]. Typical

vaccination schedules utilizing HSPPC-96 require weekly vaccination for the initial four weeks of therapy, followed by biweekly administration until supply depletion.

Non-glioma malignancies

To date, the technique has been utilized in several cancer types in an attempt to exploit the distinctive, patient-specific immunogenic potential offered by HSPPC-96 vaccination with varying degrees of success. Janetzki et al was the first to investigate the application of autologous HSPPC-96 in human malignancies. [36] To establish the safety profile of HSPPC-96 and evaluate immune responses in this pilot study, patients with a variety of cancers refractory to standard therapies received HSPPC-96 vaccines prepared from resected tumor tissue. Results from the study demonstrated feasibility of vaccine production and lack of toxicity. While the limited number of patients and study design precluded a clear evaluation of clinical efficacy, robust immune responses following immunization were noted in a majority of patients characterized by increased levels of NK cells and expansion of tumor specific T cells, consistent with observations in preclinical murine studies. [37, 38] Rivoltini et al. similarly demonstrated that treatment with tumor derived HSPPC-96 in 10 patients with either melanoma or colon carcinoma led to activation and expansion of tumor antigen specific CD8+ T cells *in vitro* and *in vivo*. [39] Subsequently, a number of phase I/II studies further demonstrated the feasibility of vaccine production, lack of toxicity, and signs of clinical activity in a range of tumors including colorectal cancer [40], non-small cell lung cancer (NSCLC) [41], pancreatic adenocarcinoma [42], and melanoma [35, 43, 44]. These studies demonstrated feasibility and safety, while also noting evidence of post vaccination tumor specific immune responses.

Two phase III clinical trials followed these studies, comprising the largest studies on HSPPC-96 tumor vaccines to date. The first randomized, multicenter trial enrolled 322 patients with metastatic melanoma and compared autologous HSPPC-96 vaccine to controls who received physician's choice of therapy, comprised of Dacarbazine, Temozolomide (TMZ), IL-2, complete tumor resection either alone or in combination.[45] OS, via intention to treat analysis, did not differ between the vaccine and control arms. Consistent with limitations cited in phase I/II trials, vaccine production was constrained by the availability of adequate resected tumor tissue and technical challenges. Only 61.8% of patients assigned to the vaccine arm received one or more doses. For those who received vaccinations, the number of doses was also highly variable. Controlling for the bias in which patients living longer would be able to receive more vaccine doses through landmark analysis, subset analysis revealed that patients harboring M1a and M1b disease substages who received a greater number of immunizations survived longer than those who received fewer vaccinations (1+ vs 10+ doses). Thus, clinical efficacy was most evident in those with earlier stages of disease receiving higher number of doses. [45] The second a multicenter randomized phase III trial investigated efficacy of HSPPC-96 vaccine versus observation following nephrectomy in 728 patients with locally advanced renal cell carcinoma.[46] On median follow up of 1.9 years, there were no differences in recurrence (37.7% vs. 39.8%) or survival (19.4% vs. 19.6%) between the treatment and observation groups, respectively., Post-hoc subgroup analysis demonstrated a PFS benefit in intermediate risk patients as defined by ECOG risk stratification. [47] Rate of recurrence was lower in the treatment

versus observation group in intermediate risk patients (15.2% vs 26.4%, $p=0.026$). However, there were no difference in overall survival. While this suggested increased vaccine efficacy in patients with less advanced disease, this should be interpreted with caution given the limitations of post-hoc analysis [48] No clinical benefit was seen in patients who were at high risk. These large phase III trials highlighted a number of factors concerning HSPPC-94 vaccines. First, increased number of vaccine doses was correlated with improved clinical response. Second, vaccine was most effective in patients with less advanced disease, possibly secondary to increasing numbers of mechanisms by which more advanced staged malignancies evade an immune mediated antitumor response.

Glioblastoma

In a phase I dose escalation trial, Crane et al. investigated of the role of HSPPC-96 in the vaccination of patients with recurrent high grade glioma.[49] Twelve patients met postsurgical study criteria (table 1). Patients either received 25 μ g HSPPC-96 every 2 weeks totaling 4 vaccinations or 25 μ g HSPPC-96 weekly for a total of 4 vaccinations. After the first 4 vaccination treatments, all patients were placed on a biweekly dosing schedule. Overall, this vaccine strategy appeared safe and tolerable with no significant toxicities encountered. A tumor-specific peripheral immune response to vaccine administration was present in 11 of the 12 patients. Restimulation of peripheral blood leukocytes with autologous HSP *ex vivo* demonstrated increased T cell proliferation and significant increase in IFN- γ production. These peripheral immune assays correlated with the proinflammatory immunogenic response induced by the vaccine. This was demonstrated in the 7 patients that underwent subsequent tumor biopsies after receiving the vaccine; their tumors harbored IFN- γ positive NK and T-cells which demonstrated that immune effector cells were localizing to the tumor site. Immune response was associated with clinical outcome, with a median OS of 47 weeks in immune responders compared to 16 weeks in nonresponders. [49]

In a subsequent open label phase II multicenter clinical trial, 68 adult patients with recurrent GBM were enrolled and underwent gross total resection. Only 41 patients met pre- and postoperative criteria (Table 2).[50] All patients received 25 μ g HSPPC-96 weekly for 4 weeks, followed by a biweekly dosing schedule. Only 3 patients failed to receive the protocol minimum of 4 doses. There were 17 vaccine attributable grade 3–4 adverse effects. Median and 6 month PFS were 19.1 weeks and 29.3%, respectively. Median and 6 months OS were 42.6 weeks and 29.3%, respectively. Evaluation of the prognostic impact of immunological status through subgroup analysis based on absolute lymphocyte count (ALC) demonstrated that an ALC above the median of the cohort was associated with improved survival on univariate (49.1 vs 37.1 weeks, $p = 0.39$) and multivariate analysis (HR 4.0, CI 1.4–11.8; $p = 0.012$). Results are promising in comparison to historical controls within similarly surgically focused trials for recurrent GBM. Examples of these include the PRECISE phase III Trial. Treatment in this study consisted of convection-enhanced delivery of a chimeric cytotoxin comprising human interleukin-13 fused to a truncated form of pseudomonas exotoxin (Cintredekin Besudotox) which was compared to implanted Gliadel wafers following resection in the management of recurrent GBM. Median OS was 36.4 weeks in patients receiving the chimeric cytotoxin and 35.3 weeks for the group receiving Gliadel Wafers. [51]

For recurrent GBM, the HSPPC-96 vaccination trial uniquely demonstrated both a peripheral and tumoral immune response which correlated with clinical outcome. A strong association between pre-vaccination lymphopenia and significantly worse outcomes further elaborates on the role of GBM mediated immunosuppression and possible benefit of addressing a patient's immune status prior to vaccination. One of the methods in which GBM exerts a state of immunosuppression is by inducing B7-H1 expression in both circulating and tumor-infiltrating macrophages. Patients that demonstrated monocytes with high expression of B7-H1 had significantly worse median PFS when compared to patients with low B7-H1 expressing monocytes (10 vs 17 months respectively).[52] Since vaccine efficacy is dependent on a viable immunological response, addressing these immunologic deterrents may yield promising results.

Additionally, there is a completed multicenter trial with data pending publication. This phase II single arm study investigated the application of autologous HSPPC-96 vaccine in newly diagnosed adult patients with GBM undergoing standard of therapy (NCT00905060). Patients received weekly intradermal injections of vaccine for 4 consecutive weeks following tumor resection and adjuvant radiation therapy and temozolomide.

Ongoing clinical trials

After the encouraging results demonstrated by the previous phase II trial of HSPPC-96 on recurrent glioblastomas, a subsequent multi-institutional trial sponsored by the Alliance for Clinical Trials in Oncology (ALLIANCE) is currently recruiting (NCT01814813). This trial will help provide evidence as to whether if HSPPC-96 can prolong overall survival in cases of recurrent GBM as an adjuvant therapeutic agent. The study will consist of three different arms which include: HSPPC-96 with concomitant bevacizumab, HSPPC-96 with administration of bevacizumab at tumor progression, and bevacizumab alone. In addition to the primary measure of OS, secondary outcomes evaluated will include PFS, in addition to the safety and tolerability of the combinatorial therapy. Samples collected throughout the trial will be utilized to correlate immune responders to HSPPC-96 with survival outcome as well as investigating whether lymphocytic infiltrates at tumor baseline correlate with the response to the vaccine. There is also a phase I trial in Beijing, China studying the safety and efficacy of autologous HSP gp96 in newly diagnosed supratentorial gliomas (NCT02122822).

Limitations

Application of HSPPC-96 vaccine has demonstrated promise but is not without limitations. Acquisition of adequate tissue for vaccine production has been challenging in previous clinical trials. In the HSPPC-96 phase II trial for vaccination against GBM, the authors note that while vaccines were unable to be produced in 13 out of 63 patients, modifications and improvement in technique with patients enrolled later in the study led to improved rates of vaccine yield.[56] The inclusion criteria requiring near-complete tumor resection limiting patient eligibility may limit the generalizability of HSPPC-96 vaccine from the phase II trial. [56, 57] Progression free survival in the phase II study was not significantly improved compared to conventional salvage therapy for recurrent GBM. However distinguishing

tumor recurrence from pseudoprogression due to treatment-related changes on postoperative imaging can be difficult. The trial demonstrated promising median overall survival, which provides a more definitive measure of clinical efficacy.

Conclusion

Immunotherapy remains a promising adjuvant in the treatment of GBM through eliciting tumor specific immune responses capable of producing sustained antitumor response while minimizing systemic toxicity. Theoretically, HSP vaccines provides a number of advantages including direct interaction APCs for antigen internalization and presentation, stimulation of both innate and adaptive immune responses. HSP vaccines allow for the delivery of a patient specific polyvalent vaccine that does not require identification of specific immunogenic GBM antigens. Multiple antigens are used to minimize the risk of immune evasion, which may occur with vaccines that utilize a single antigen. Within clinical trials, HSPPC-96 was safe with minimal adverse effects in the treatment of a variety of cancer types. Efficacy has been variable and improvements in outcomes were not seen in a variety of cancer types. However, HSPPC-96 has been most promising in phase I and II trials with recurrent GBM, a devastating disease with limited treatment options, as demonstrated by superior outcomes compared to historical controls. Challenges include acquisition of sufficient tumor for vaccine production and requirement of gross total resection, which may not always be achievable. However, with increasing experience through past and ongoing trials, issues with vaccine yield, patient selection, and screening will be optimized. HSPPC-96 is a promising immunotherapeutic adjuvant for treatment of GBM, and pending results from completed and ongoing trials will help further elaborate on the role of HSPPC-96 in the treatment of this devastating disease with limited treatment options.

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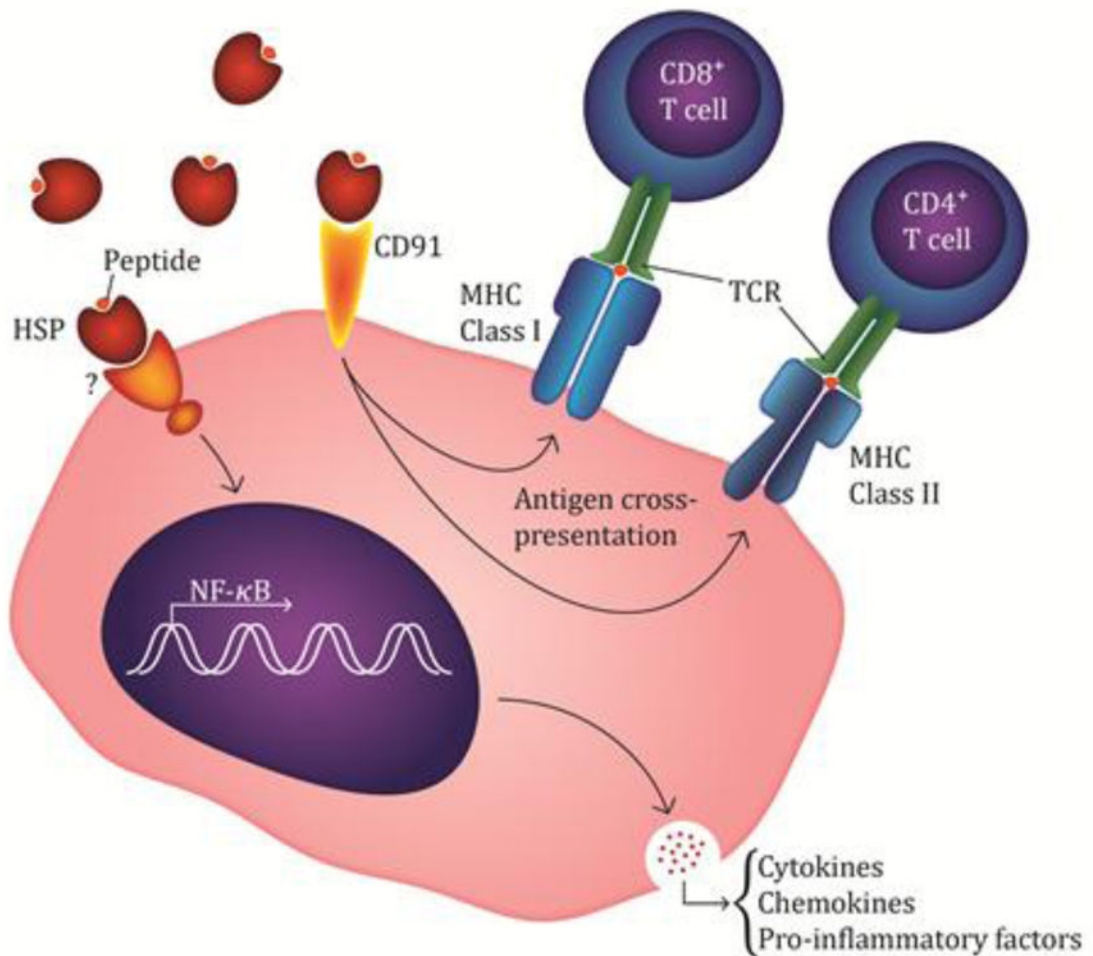


Figure 1. HSP-peptide complex interaction with antigen presenting cells

The proposed mechanism by which the HSP-peptide complex interacts with APC's consists of cell surface receptor interaction. Primarily, CD91 has been shown to endocytose the complex and via either proteasome dependent or independent pathways lead to the presentation via MHC-class I receptor. In addition, a portion of the internalized complex enters an acidic compartment which leads to its loading onto MHC-class II receptors. Additional cell surface receptors, such as TLR2/TLR4, and others that have not been elucidated are also involved in eliciting a downstream effect which leads to the activation of the NF- κ B pathway. Upon activation, proinflammatory cytokines and chemokines are generated and secreted in order to further augment a proinflammatory response.

Table 1

Inclusion/Exclusion criteria for the phase I trial of autologous HSPPC-96 in the recurrent setting of glioblastoma [49].

| Inclusion Criteria | Exclusion Criteria |
|--|---|
| <ul style="list-style-type: none">• Recurrent grade III or IV glioma• KPS \geq 60, life expectancy $>$ 8 weeks.• 4 vaccines available for use | <ul style="list-style-type: none">• Treatment with corticosteroids at time of resection• Hx of immunodeficiency, immunosuppressive drug use excluding corticosteroids, current malignancies at other sites or other cancers within 5 years• Uncontrolled active infection |

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Table 2

Inclusion/Exclusion criteria for the phase II trial of autologous HSPPC-96 in the recurrent setting of glioblastoma [50].

| Inclusion Criteria | Exclusion Criteria |
|---|---|
| <ul style="list-style-type: none"> • Age >18 years • Histologically confirmed recurrent GBM • Postoperative KPS 70% • Life expectancy >8 weeks • Extent of resection >90% | <ul style="list-style-type: none"> • Systemic autoimmune disease • Primary or secondary immunodeficiency • Other malignancy within past 5 years • Bleeding diathesis • Uncontrolled active infection • Serious medical comorbidity <p data-bbox="673 625 844 651">Postoperative criteria:</p> <ul style="list-style-type: none"> • Pseudoprogression without recurrent tumor • Documented tumor growth within 4 weeks of surgery • Insufficient tumor for 4 doses of vaccine |