

Draft Genome Sequences of Human-Pathogenic *Escherichia coli* O26:H11 Strains Carrying the *stx*₂ Gene Only and Circulating in France

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Shiga toxin-producing *Escherichia coli* (STEC) O26:H11 is one of the most frequent pathogens associated with diarrhea and hemolytic-uremic syndrome (HUS). In this report, we present the draft genome sequences of seven strains of STEC O26:H11 carrying the *stx*_{2a} or *stx*_{2d} gene only and isolated in France from HUS patients.

Received 22 June 2015 Accepted 25 June 2015 Published 30 July 2015

Citation Delannoy S, Mariani-Kurkdjian P, Bonacorsi S, Liguori S, Ison SA, Fach P. 2015. Draft genome sequences of human-pathogenic *Escherichia coli* O26:H11 strains carrying the *stx*₂ gene only and circulating in France. *Genome Announc* 3(4):e00852-15. doi:10.1128/genomeA.00852-15.

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Among Shiga toxin-producing *Escherichia coli* (STEC) isolates, serotype O26:H11 is the second most frequently associated with severe human diseases worldwide, after O157:H7 (1, 2). Strains of STEC O26:H11 usually harbor the *stx*_{1a} gene only or in combination with *stx*_{2a}. However, in the 1990s, a new clonal subgroup of STEC O26 emerged that carries the *stx*_{2a} gene alone (3). This new clone, first described in Germany, has spread over Europe and has recently been described on the American continent (4, 5).

In a previous study (6), we analyzed 23 STEC O26 strains carrying the *stx*₂ gene only, which were isolated in France between 2010 and 2013 from hemolytic-uremic syndrome (HUS) patients. Although most of the strains appeared to correspond to this new clone, 12 of them exhibited significantly different characteristics.

In order to investigate more thoroughly the genetic relationship between the different clones, we sequenced seven of these strains covering the different combination observed between sequence types (ST), *stx* gene subtype, and virulence gene profile.

Genomic DNA was extracted from an overnight culture in tryptic soy broth (TSB) medium using the DNeasy blood and tissue kit (Qiagen), with an additional RNase A (Roche) treatment. Libraries were prepared using the Nextera XT kit (Illumina). Whole-genome sequencing was performed using an Illumina MiSeq platform (Illumina), according to the manufacturer's

instructions. Two MiSeq runs were carried out, one with paired-end 150-nucleotide (nt) reads on MiSeq V2 microchemistry and another with paired-end 300-nt reads on V3 chemistry. The raw reads were trimmed (minimum length, 35 bp; quality score, 0.03) and assembled in CLC Genomics Workbench 7.5.1 by *de novo* assembly (minimum contig length, 1,000 bp), producing 192 to 223 contigs (Table 1). The median read depth of the assemblies ranged from 38× for 34827 and 34870 to 84× for 36708, with an *N*₅₀ between 65 kbp and 114 kbp (Table 1). The sequences were annotated using the National Center for Biotechnology Information (NCBI) Prokaryotic Genomes Automatic Annotation Pipeline (PGAAP) at <http://www.ncbi.nlm.nih.gov/genomes/static/Pipeline.html>.

The average size of the genomes in this study is 5.47 Mb, with 5.23 Mb being the smallest genome size (isolate 36084, Table 1) and 5.60 Mb the largest genome size (isolate 34827, Table 1). On average, 5,487 coding sequences were identified in the genomes (Table 1). BLAST analysis of the *stx* genes confirmed the previously identified subtypes (Table 1).

A detailed report on further analyses of the draft genome sequences will be released in a future publication.

Nucleotide sequence accession numbers. The annotated draft whole-genome sequences of these O26:H11 strains were deposited in DDBJ/ENA/GenBank under the accession numbers

TABLE 1 NCBI accession numbers and assembly metrics of the O26:H11 *E. coli* draft genomes

Isolate	ST	<i>stx</i> subtype	No. of contigs	Genome size (bp)	<i>N</i> ₅₀ (bp)	Median read depth (×)	No. of coding sequences (per PGAAP)	NCBI accession no.
36084	ST21	<i>stx</i> _{2a}	205	5,235,007	96,218	45	5,209	LDXI00000000
36708	ST29	<i>stx</i> _{2a}	192	5,526,827	114,644	84	5,579	LDXG00000000
34827	ST29	<i>stx</i> _{2a}	223	5,604,044	73,150	38	5,674	LDXF00000000
34870	ST29	<i>stx</i> _{2a}	196	5,498,010	65,024	38	5,524	LDXE00000000
36348	ST29	<i>stx</i> _{2d}	208	5,570,467	94,008	71	5,573	LDXD00000000
36293	ST29	<i>stx</i> _{2d}	207	5,458,923	75,621	45	5,460	LDXC00000000
36493	ST29	<i>stx</i> _{2d}	204	5,422,929	78,448	47	5,389	LDXB00000000

LDXB00000000 to LDXG00000000 and LDXI00000000 (Table 1). The versions described in this paper are the first versions, LDXB01000000 to LDXG01000000 and LDXI01000000.

ACKNOWLEDGMENT

This project was partly funded by the French Joint ministerial program of R&D against CBRNE risks (grant C17609-2).

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