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# Caffeine in floral nectar enhances a pollinator's memory of reward

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#### Abstract

Plant defence compounds occur in floral nectar, but their ecological role is not well-understood. We provide the first evidence that plant compounds pharmacologically alter pollinator behaviour by enhancing their memory of reward. Honeybees rewarded with caffeine, which occurs naturally in nectar of *Coffea* and *Citrus* species, were three times more likely to remember a learned floral scent than those rewarded with sucrose alone. Caffeine potentiated responses of mushroom body neurons involved in olfactory learning and memory by acting as an adenosine receptor antagonist. Caffeine concentrations in nectar never exceeded the bees' bitter taste threshold, implying that pollinators impose selection for nectar that is pharmacologically active but not repellent. By using a drug to enhance memories of reward, plants secure pollinator fidelity and improve reproductive success.

Many drugs commonly consumed by humans are produced by plants as a form of toxic defence against herbivores (1, 2). While plant-derived drugs like caffeine or nicotine are lethal in high doses (3-5), they have pharmacological effects at low doses that affect mammalian behaviour. For example, low doses of caffeine are mildly rewarding and enhance cognitive performance and memory retention (6). Interestingly, caffeine has been detected in low doses in the floral nectar and pollen of *Citrus* (7), but whether it has an ecological function is unknown.

Two caffeine-producing plant genera, *Citrus* and *Coffea*, have large floral displays with strong scents and produce more fruits and seeds when pollinated by bees (8, 9). If caffeine confers a selective advantage when these plants interact with pollinators, we might expect it to be commonly encountered in nectar. We measured caffeine in the nectar of 3 species of *Coffea* (*C. canephora*, *C. arabica*, and *C. liberica*) and 4 species of *Citrus* (*C. paradisi*, *C. maxima*, *C. sinesis*, *C. reticulata*) using liquid chromatography-mass spectrometry (10, Fig.

Wright et al.

S1A). When caffeine was present, its concentration ranged from 0.003 - 0.253 mM. The median caffeine concentration in both genera was not significantly different (Fig. 1A, Mann-Whitney, Z = -1.09, P = 0.272). Caffeine was more common in the nectar of *C. canephora* than in *C. arabica* or *C. liberica* (*Coffea*: logistic regression  $\chi_2^2 = 11.1$ , P = 0.004); it was always present in *Citrus* nectar. The mean total nectar sugar concentration ranged from 0.338-0.843 M (Fig. 1B, see Fig S1B for individual sugars). Caffeine concentration in nectar did not correlate with total sugar concentration (Pearson's r = 0.063, P = 0.596).

We hypothesized that caffeine could affect the learning and memory of foraging pollinators. To test this, we trained individual honeybees to associate floral scent with 0.7 M sucrose and 7 different concentrations of caffeine and tested their olfactory memory. Using a method for classical conditioning of feeding responses (proboscis extension reflex, 11), bees were trained for 6 trials with 30 s between each pairing of odour with reward. This inter-trial interval approximated the rate of floral visitation exhibited by honeybees foraging from multiple flowers on a single Citrus tree (see methods). The presence of low doses of caffeine in reward had a weak effect on the rate of learning (Fig. 2A), but it had a profound effect on long-term memory. When rewarded with solutions containing nectar-levels of caffeine, three times as many bees remembered the conditioned scent 24 h later and to responded as if it predicted reward (Fig. 2B, logistic regression,  $\chi_7^2 = 41.9$ , P < 0.001). Twice as many bees remembered it 72 h later (Fig. 2C). This improvement in memory performance was not due to a general increase in olfactory sensitivity resulting from caffeine consumption (Fig. S2A). In fact, the effect of caffeine on long-term olfactory memory in bees was greater than that produced by high concentrations of sucrose when the same experimental methods were used (e.g. 2.0 M, Fig. S2B).

Caffeine's influence on cognition in mammals is in part mediated by its action as an adenosine receptor antagonist (11). In the hippocampal CA2 region, inhibition of adenosine receptors by caffeine induces long-term potentiation (12), a key mechanism of memory formation (13). The Kenyon cells (KCs) in mushroom bodies of the insect brain are similar in function to hippocampal neurons: they integrate sensory input during associative learning, exhibit long-term potentiation and are involved in memory formation (14-16). To test whether nectar-caffeine doses affect mushroom body function, we made whole-KC recordings in the intact honeybee brain. Caffeine (100  $\mu$ M) evoked a small increase in the holding current (I<sub>M</sub>) and depolarized KC membrane potential (V<sub>M</sub>) towards the action potential firing threshold, by increasing nicotinic ACh receptor (nAChR) activation (Fig. 3A-D). To test whether the observed effects of caffeine were due to interactions with adenosine receptors, we applied the adenosine receptor antagonist, DPCPX, and observed that it similarly increased I<sub>M</sub> and depolarized V<sub>M</sub> but to a lesser extent (Fig. 3E,F). Both caffeine and DPCPX affected KC response kinetics evoked by brief, local application of ACh, increasing the activation rate and slowing the decay (Fig.3G,H). Our data show that caffeine modulates cholinergic input via a postsynaptic action, but could act via presynaptic adenosine receptors to potentiate ACh release (17). The resulting increase in KC excitability should lead to an increased probability of action potential firing in response to sensory stimulation (18), thereby facilitating the induction of associative synaptic plasticity in KCs (19). The enhanced activation of KCs may also facilitate plasticity at synapses with

mushroom body extrinsic neurons (20), which exhibit spike-timing-dependent plasticity (21). In this way, a 'memory trace' could be formed for the odour associated with reward during and after conditioning (22-23).

Caffeine is bitter tasting to mammals and is both toxic (24) and repellent to honeybees at high concentrations (25, 26). If bees can detect caffeine, they might learn to avoid flowers offering nectar containing it (27). We found that honeybees were deterred from drinking sucrose solutions containing caffeine at concentrations greater than 1 mM (Fig. 4); they also have neurons that detect caffeine in sensilla on their mouthparts (Fig. S3). However, nectar concentrations never exceeded 0.3mM (i.e. 0.058 mg/ml), even though levels of caffeine in vegetative and seed tissues of *Coffea* have been reported to be as great as 24mg/ml (28). This implies that pollinators drive selection towards concentrations of caffeine that are not repellent but still pharmacologically active.

Our data show that plant-produced alkaloids like caffeine have a role in addition to defence: they can pharmacologically manipulate a pollinator's behaviour. When bees and other pollinators learn to associate floral scent with food while foraging (29), they are more likely to visit flowers bearing the same scent signals. Such behaviour increases their foraging efficiency (30) while concomitantly leading to more effective pollination (31, 32). Our experiments suggest that by affecting a pollinator's memory, plants reap the reproductive benefits arising from enhanced pollinator fidelity.

## **Supplementary Material**

Refer to Web version on PubMed Central for supplementary material.

### Acknowledgments

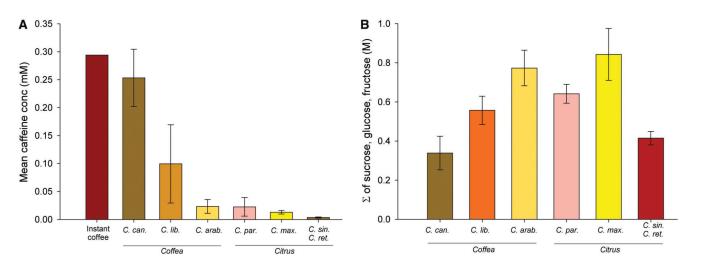
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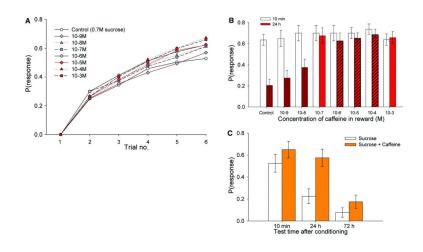
Wright et al.



#### Figure 1.

(A) Caffeine concentration in *Coffea* and *Citrus sp.* and a cup of instant coffee. Caffeine concentration depended on species within each genus (*Coffea*: Kruskal-Wallis,  $\chi_2^2 = 28.1$ , P < 0.001; *Citrus*: Kruskal-Wallis,  $\chi_2^2 = 6.98$ , P = 0.030); *C. canephora* had the highest mean concentration of all species sampled. (**B**) The sum of the concentration of sucrose, glucose and fructose (total nectar sugars) depended on species (1-way ANOVA: F<sub>5, 161</sub> = 4.64, P < 0.001) and was greatest in *Citrus maxima* and hybrids (citron, lemons, clementines). (*C. can.* = *Coffea canephora*, N = 34; *C. lib.* = *Coffea liberica*, N = 31; *C. arab.* = *Coffea arabica*, N = 27; *C. par.* = *Citrus paradisi* and hybrids, N<sub>cp</sub> = 17; *C. max.* = *Citrus maxima* and hybrids, N = 5; *C. sin.* and *C. ret.* = *Citrus sinensis* and *Citrus reticulata*, N<sub>CS</sub> = 7, N<sub>CR</sub> = 5 – data for these two species was pooled).

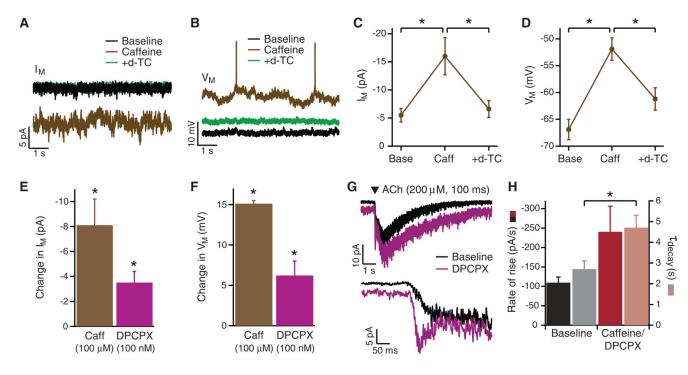
Wright et al.



#### Figure 2.

(A) The rate of learning of bees conditioned with an odour stimulus paired with a 0.7 M sucrose reward containing caffeine. The rate of learning was slightly greater for the bees fed caffeine in reward during conditioning (logistic regression,  $\chi_1^2 = 4.85$ , P = 0.028). N 79 for all groups. (B) Memory recall test for odours at 10 min (clear bars) or 24 h (red bars) after bees had been trained as in (A). Bright red bars indicate that the response at 24 h was significantly different from the control (0.7 M sucrose) (least-squares contrasts: P < 0.05); dark red bars were not significantly different. Nectar-levels of caffeine are indicated by hatching. N > 79 for each group. (C) Bees fed 0.1 mM caffeine in sucrose (red bars) were more likely to remember the conditioned odour sucrose alone (white bars) (logistic regression,  $\chi_1^2 = 9.04$ , P < 0.003) at 24 h and 72 h after conditioning. N = 40 per group.

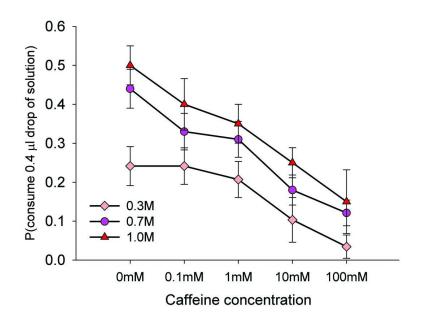
Wright et al.



#### Figure 3.

The effect of caffeine on Kenyon cells. (**A**, **B**) Example traces from a KC in intact honeybee brain recorded under voltage-clamp (**A**,  $V_H = -73 \text{ mV}$ ) and current-clamp (**B**; at resting  $V_M$ ), showing the increase in  $I_M$  and depolarization evoked by bath application of caffeine (100 µM), and subsequent reversal by the nAChR antagonist d-TC (500 µM). (**C**, **D**) Mean data showing the reversal by d-TC (500 µM) of the effect of caffeine (Caff; 100 µM) on  $I_M$  (**C**; N = 6, t<sub>5</sub> = 4.03, P = 0.010; t<sub>5</sub> = 4.07, P = 0.010) and  $V_M$  (**D**; N = 6, t<sub>5</sub> = 34.1, P < 0.001; t<sub>5</sub> = 12.0, P < 0.001). (**E**, **F**) Comparison of the mean effects of caffeine and DPCPX on  $I_M$  (**E**, Caff: N = 10, t<sub>9</sub> = 3.84, P = 0.004; DPCPX: N = 6, t<sub>5</sub> = 4.04, P = 0.010) and  $V_M$  (**F**, Caff: N = 6, t<sub>5</sub> = 34.1, P < 0.001; DPCPX: N = 6, t<sub>5</sub> = 3.39, P = 0.019). (**G**, **H**) Example traces (**G**; rising phase shown on an expanded time-scale below) and mean data (**H**, Rate of rise: N = 6, t<sub>5</sub> = 2.20, P = 0.079;  $\tau_{decay}$ : N = 9, t<sub>8</sub> = 3.54, P = 0.008) showing that DPCPX (100 nM) and caffeine (100 µM) slowed the decay and, in 6 of 9 KCs, potentiated the fast component of the response evoked by exogenous ACh. (Student's paired t-test used in all comparisons).

Wright et al.



#### Figure 4.

Bees are more likely to reject sucrose solutions containing caffeine of concentrations greater than 1 mM (logistic regression,  $\chi_4^2 = 23.4$ , P < 0.001; for 0.7M and 1.0M, 1 mM caffeine vs sucrose *post hoc*, P < 0.05; for 0.3M, 100 mM caffeine vs sucrose *post hoc*, P < 0.05). Bees were less likely to drink 0.3 M sucrose (pale pink diamonds) than 0.7M (pink circles) or 1.0M solutions (red triangles) (logistic regression,  $\chi_2^2 = 8.69$ , P = 0.013). Mean responses + SE. N<sub>0.3M</sub> = 29, N<sub>0.7M</sub> = 100, N<sub>1.0M</sub> = 20.