

Optimal combination of antiangiogenic therapy for hepatocellular carcinoma

Hui-Ju Ch'ang

Hui-Ju Ch'ang, National Institute of Cancer Research, National Health Research Institutes, Miaoli 35053, Taiwan

Hui-Ju Ch'ang, Department of Radiation Oncology, Taipei Medical University Hospital, Taipei 100, Taiwan

Author contributions: Ch'ang HJ solely contributed to this manuscript.

Supported by National Health Research Institutes, Taiwan.

Conflict-of-interest statement: Nothing to declare.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Hui-Ju Ch'ang, MD, National Institute of Cancer Research, National Health Research Institutes, R1-2034, 35, Keyen Road, Miaoli 35053, Taiwan. hjmc@nhri.org.tw
Telephone: +886-37-246166-35105
Fax: +886-37-586463

Received: July 29, 2014

Peer-review started: July 30, 2014

First decision: December 17, 2014

Revised: July 21, 2015

Accepted: July 24, 2015

Article in press: July 27, 2015

Published online: August 8, 2015

Abstract

The success of sorafenib in prolonging survival of patients with hepatocellular carcinoma (HCC) makes therapeutic inhibition of angiogenesis a component of treatment for HCC. To enhance therapeutic efficacy, overcome

drug resistance and reduce toxicity, combination of antiangiogenic agents with chemotherapy, radiotherapy or other targeted agents were evaluated. Nevertheless, the use of antiangiogenic therapy remains suboptimal regarding dosage, schedule and duration of therapy. The issue is further complicated by combination antiangiogenesis to other cytotoxic or biologic agents. There is no way to determine which patients are most likely respond to a given form of antiangiogenic therapy. Activation of alternative pathways associated with disease progression in patients undergoing antiangiogenic therapy has also been recognized. There is increasing importance in identifying, validating and standardizing potential response biomarkers for antiangiogenesis therapy for HCC patients. In this review, biomarkers for antiangiogenesis therapy including systemic, circulating, tissue and imaging ones are summarized. The strength and deficit of circulating and imaging biomarkers were further demonstrated by a series of studies in HCC patients receiving radiotherapy with or without thalidomide.

Key words: Antiangiogenesis; Hepatocellular carcinoma; Biomarker; Cytokines; Dynamic contrast enhanced magnetic resonance imaging

© **The Author(s) 2015.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Antiangiogenic therapy has become an important component of treatment in hepatocellular carcinoma (HCC) patients. However, traditional anatomic imaging of tumor shrinkage is not appropriate to evaluate the efficacy of antiangiogenesis achieved by normalizing tumor vasculature and systemic suppression of angiogenic and inflammatory cytokines. To identify and validate potential response biomarkers, standardized systemic, circulating, tissue and imaging assays should be incorporated in to preclinical and clinical studies regarding the combination of antiangiogenic agents to cytotoxic or biologic agents. The optimal dosage, schedule and duration of antiangiogenic during com-

bination therapy for HCC patients should be titrated according to these response biomarkers.

Ch'ang HJ. Optimal combination of antiangiogenic therapy for hepatocellular carcinoma. *World J Hepatol* 2015; 7(16): 2029-2040 Available from: URL: <http://www.wjgnet.com/1948-5182/full/v7/i16/2029.htm> DOI: <http://dx.doi.org/10.4254/wjh.v7.i16.2029>

INTRODUCTION

Patients with advanced hepatocellular carcinoma (HCC) have a poor prognosis. Systemic therapy with cytotoxic agents provides marginal benefit^[1,2]. HCC, a highly vascularized tumor, requires angiogenesis to grow, invade and metastasize^[3]. The success of pharmacological inhibition of angiogenesis in HCC provided by sorafenib^[4,5], makes it the first and only systemic agent to notably improve survival in HCC patients.

Although the inhibition of angiogenesis in HCC is an established modality of cancer treatment, concerns regarding toxicity and drug resistance still constitute barriers to be overcome. Recent randomized studies comparing multikinase inhibitors - sunitinib^[6], brivanib^[7], and linifanib^[8] - and the combination of sorafenib plus erlotinib^[9] with sorafenib alone does not reveal better survival rates or tolerability. In this review, issues brought up by combining antiangiogenic agents with chemotherapy, or other targeted therapy will be summarized. A series of our study, incorporating thalidomide, an antiangiogenic agent, during radiotherapy for HCC patients will be introduced.

STRATEGIES TO IMPROVE

ANTIANGIOGENESIS THERAPY IN HCC

To improve the clinical outcomes of antiangiogenic therapy in HCC, the combination of antiangiogenic agents with cytotoxic chemotherapy or with other molecularly targeted therapies may act synergistically to generate additive effects. A better understanding of the mechanisms regarding the action of sorafenib on HCC plus the investigation for predictive biomarkers may allow us to select patients suitable for anti-angiogenesis therapy.

Combinations of anti-Vascular endothelial growth factor (VEGF) agents with chemotherapy in HCC are under evaluation. Treatment with bevacizumab/capecitabine/oxaliplatin or with bevacizumab/gemcitabine/oxaliplatin in HCC patients resulted in median survivals of less than 10 mo^[10,11]. Based on the encouraging data from sorafenib plus doxorubicin in HCC^[12], a phase III randomized study (CALGB80802) comparing sorafenib plus doxorubicin with sorafenib alone is underway in patients with advanced HCC. Studies combining sorafenib with gemcitabine/oxaliplatin, modified FOLFOX, or capecitabine/oxaliplatin are ongoing.

Another approach has been to combine antiangiogenic therapy with inhibitors of other angiogenesis or molecular targets. Tivantinib, a c-MET inhibitor, was compared with placebo in a randomized phase II study in advanced HCC. Improved time to progression with tivantinib, especially for patients with tumors of high MET expression was noted^[13]. Cabozantinib, a receptor tyrosine kinase inhibitor of c-MET/VEGF receptor 2 (VEGFR2), is undergoing phase III evaluation in HCC patients failed or could not tolerate sorafenib^[14]. The mTOR inhibitor everolimus was compared with best supportive care alone in a randomized phase III trial (EVOLVE-1) in the second-line treatment of HCC patients. No significant survival benefit was noted using everolimus in HCC patients relapsed from sorafenib^[15]. Everolimus was also combined with sorafenib in a phase I trial of HCC, and 43% of the patients developed grade 3/4 thrombocytopenia^[16].

Other inhibitors of genetic or epigenetic targets of HCC including basic fibroblast growth factor (bFGF) inhibitors, heat-shock protein inhibitors, histone deacetylase inhibitors, MEK inhibitors, insulin growth factor (IGF)/IGF receptor inhibitors, Wnt signal inhibitors, immunotherapy with or without the combination of antiangiogenesis are under clinical investigation in advanced HCC^[17].

With all the efforts in improving clinical outcomes of antiangiogenesis, the optimal dosing schedule of antiangiogenic agents alone or in combination for HCC patients is largely unclear. Along with the development of new therapies, a parallel effort must be made to identify biomarkers of response, and toxicity in order to provide HCC patients with safe and effective therapies.

RESPONSE BIOMARKER

Anatomic imaging biomarkers that quantify liver tumor response to cytotoxic therapy are based on temporal change in the size of the tumors. Objective response by size-based decrease in tumor, may translate to an early clinical endpoint, in substitution for overall survival^[18]. Anti-VEGF therapy has primarily cytostatic effects, may prune and normalize the tumor vasculature, and can have substantial systemic effects such as modulation of circulating proangiogenic and proinflammatory cytokines and cells^[19-24]. These effects may not shrink but rather stabilize the tumor size and prolong survival^[25,26]. Unlike conventional chemotherapy, an effective dose of an antiangiogenic agent can be less than the maximum tolerated dose, whereas certain toxicities may be dose-related^[27]. The development of antiangiogenic therapy or other biologic therapy requires new methods for measuring response to therapy.

Blood pressure as a biomarker

Hypertension has been observed in patients with cancer treated with anti-VEGF antibodies or tyrosine kinase inhibitors (TKIs) and is clinically manageable in most cases with medication. There is evidence that patients

Table 1 Circulating biomarker of hepatocellular carcinoma patients receiving antiangiogenic therapy

Treatment	Patient enrolled	Patient n	Predictive value	Prognostic value	Ref.
Sorafenib, bevacizumab or thalidomide/oral 5FU	Elevated AFP	72	AFP responder (AFP decline > 20% in 4 wk) correlate with response	Early AFP responder: associate with PFS, OS	Shao <i>et al</i> ^[103]
Sorafenib	Advanced	30		High baseline IL-8 correlates with PD; high Ang2, G-CSF, HGF, leptin correlate with shorter PFS	Miyahara <i>et al</i> ^[104]
Sorafenib	Advanced	491	High baseline sc-KIT, low HGF correlate with sorafenib response	Baseline Ang2, VEGF, AFP correlate with survival	Llovet <i>et al</i> ^[106]
Sorafenib	Post-operative	29	High phosphor MET correlate with sorafenib resistance		Xiang <i>et al</i> ^[105]
Sunitinib	Advanced	34		High baseline AFP, IL-8, IL-6, SDF1, TNF correlate with PFS and OS; decreased IL-6, sc-KIT at day 14 correlate with improved PFS and OS	Zhu <i>et al</i> ^[24]
Sunitinib	Advanced	37	High baseline VEGFC correlates with response	High base VEGFC correlates with TTP; change in VEGFA, sVEGFR2 correlate with OS	Harmon <i>et al</i> ^[106]
Sunitinib	Advanced	23	Decrease sVEGFR2 or TNF correlate, with decrease in K^{trans} , K_{ep} ; Decrease K^{trans} , K_{ep} at week 2 correlate with response		Sahani <i>et al</i> ^[107]
Bevacizumab	Advanced	43	Increase CEC on day 15, low IL-8 correlate with disease control	High baseline IL-8, IL-6 correlate with short PFS, OS	Boige <i>et al</i> ^[108]
Bevacizumab	Advanced	59	High Ang2, EGFR, endothelin 1, no acneiform rash correlate with poor outcome		Kaseb <i>et al</i> ^[109]
Thalidomide	Advanced	47	No predictive value of VEGF, bFGF, PlGF		Hsu <i>et al</i> ^[110]
Thalidomide/tegafur/uracil	Advanced	43		High IL-6, IL-8 correlate with short survival	Shao <i>et al</i> ^[111]
Thalidomide/radiotherapy	Advanced	24	No predictive value of VEGF, bFGF, IL-6, SDF1, TNF	Baseline IL-6, SDF1 at week 2 correlate with PFS. SDF1 at 1 mo post radiotherapy correlates with OS	Ch'ang <i>et al</i> ^[89]
TSU68	Advanced	35	High sVCAM1 correlates with response		Kanai <i>et al</i> ^[112]

AFP: α -fetal protein; PFS: Progression free survival; OS: Overall survival; IL: Interleukin; PD: Progressive disease; Ang2: Angiopoietin 2; G-CSF: Granulocyte colony stimulating factor; HGF: Hepatocyte growth factor; VEGF: Vascular endothelial growth factor; SDF: Stem cell derived factor; TNF: Tumor necrosis factor; TTP: Time to progression; sVEGFR: Soluble VEGF receptor; K^{trans} : Transfer constant; K_{ep} : Redistribution rate constant; CEC: Circulating endothelial cells; EGFR: Epidermal growth factor receptor; bFGF: Basic fibroblast growth factor; PlGF: Placental growth factor; sVCAM1: Soluble vascular cell adhesion molecule 1.

with hypertension have better survival outcomes^[28,29]. A significantly improved progression-free survival (PFS) for patients with grade 2/3 hypertension after receiving bevacizumab is noted compared to those who did not develop hypertension on bevacizumab treatment ($P = 0.04$). These findings suggest the possibility of titrating the dose of anti-VEGF therapy by hypertension for efficacy optimization.

VEGF as a biomarker

The most extensively studied biomarker in antiangiogenic therapy has been VEGF (Table 1). Free VEGFA is rapidly cleared from the circulation, and a wide variation in plasma VEGF concentrations has been reported due to different assay sensitivities^[30]. After immune-depletion of VEGF bound to bevacizumab, Loupakis *et al*^[31] reported that the concentrations of free and active VEGF decreased significantly from day 0 to day 14 after bevacizumab treatment.

Associations between outcomes of antiangiogenic therapy with VEGF levels in the circulation have been reported in clinical trials of breast cancer and

HCC^[32,33]. However, in other cancers neither the intratumoral nor the circulating VEGF was associated with the outcome of bevacizumab treatment^[34,35]. Baseline VEGF and angiopoietin-2 concentrations were found to be independent prognostic markers in the sorafenib HCC assessment randomized protocol trial. However, these biomarkers were not predictive of response to sorafenib^[36].

Many studies have shown a lack of correlation between VEGF levels at baseline and the outcome of antiangiogenic therapy^[35,37]. Intriguingly, the circulating levels of VEGF seemed to be significantly elevated after most antiangiogenic therapies targeting this pathway^[38]. Similar phenomenon was noted after therapy with anti-VEGFR TKIs^[39-44]. Preclinical data indicate that this increase in VEGF may be induced by a host-response to hypoxia in tumors.

On the other hand, the VEGF genotype has emerged as a predictive biomarker from the phase III study of bevacizumab in metastatic breast cancer (ECOG 2100). VEGF-2578AA genotype was associated with a superior overall survival in the combination arm^[45].

Placental growth factor and soluble VEGFRs as biomarkers

Circulating plasma levels of placental growth factor (PIGF) increase consistently in response to anti-VEGF treatment. Thus, plasma PIGF is now being considered as a potential biomarker of anti-VEGF therapy^[41,43,46]. Of interest, the increase in PIGF may be due to systemic effects, as tumor-derived PIGF may actually be decreased after bevacizumab treatment^[46]. Ziv-aflibercept, a recombinant fusion protein that blocks multiple factors in the angiogenesis network by binding VEGFA, VEGFB and PIGF. Results of the EFC10262-VELOUR study revealed a significant improvement in the primary endpoint of overall survival (OS) with ziv-aflibercept and FOLFIRI compared to FOLFIRI alone^[47], despite approximately one-third of the patients having received prior bevacizumab treatment. These findings underscore the potential role of other VEGF family members in tumor angiogenesis.

Circulating levels of soluble VEGFR2 and VEGFR3 proteins are decreased by TKIs that directly target these receptors, but not by bevacizumab. Studies showed that patients with higher plasma levels of sVEGFR1 had a poor outcome after treatment with bevacizumab, sunitinib, vandetanib, and cediranib^[24,48-51]. Polymorphisms in the *FLT1* gene that are associated with higher VEGFR1 expression have been associated with poor outcome of bevacizumab containing regimens in phase III studies^[52]. The mechanisms by which these changes occur, their biological significance, and their utility as predictive biomarkers are not understood.

Other proteins as biomarkers

Collagen IV is one of the main constituents of tumor vascular basement membranes. Proteolytic degradation of the basement membrane during vascular normalization by antiangiogenic agents can release soluble collagen IV in blood circulation. In recurrent glioblastoma patients, increase in plasma collagen IV levels after anti-VEGF therapy was associated with an increase in PFS^[53]. In patients with metastatic colorectal cancer, responses to vatalanib plus chemotherapy correlated positively with tissue mRNA levels of VEGFR1, lactate dehydrogenase (LDH) A and glucose transporter (Glut) 1 and inversely with hypoxia inducible factor 1- α ^[54]. In addition, patients with high baseline serum LDH levels had longer PFS and OS after treatment with vatalanib and chemoradiation^[55]. Baseline soluble intracellular adhesion molecule (ICAM)-1 was shown to be an independent prognostic factor of OS in patients treated with bevacizumab and chemotherapy or chemotherapy alone in metastatic non-small cell lung cancer (NSCLC)^[37].

Certain inflammatory cytokines might have potent proangiogenic effects. In patients with advanced NSCLC who were treated with vandetanib plus chemotherapy, vandetanib alone or chemotherapy alone, increase plasma VEGF levels for vandetanib monotherapy and increase in plasma interleukin (IL)-8 for combination

therapy were associated with increased risk of disease progression^[56]. A phase II study suggested that IL-8 A-251T polymorphism may be a molecular predictor of response to bevacizumab based chemotherapy in ovarian cancer^[57]. In phase II studies, the extent of increase in inflammatory cytokines such as IL-10 in the plasma during treatment was associated with an inferior outcome in patients with rectal and ovarian cancer after bevacizumab and chemoradiation treatment, and an inferior outcome in patients with advanced HCC after sunitinib therapy^[11,23,46]. Association between increased plasma stem cell derived factor (SDF)1 α after treatment and poor outcome in anti-VEGF studies in recurrent glioblastoma, sarcoma and breast cancer were reported^[19,58,59]. Increased plasma SDF1 α and plasma IL-6 have been associated with poor outcomes in locally advanced rectal cancer and HCC patients treated with bevacizumab, chemoradiation and sunitinib^[24,60]. In line with these findings, preclinical studies have shown that sunitinib can induce elevation of circulating inflammatory cytokines in mice, which might result in more aggressive recurrent or metastatic tumors^[61,62].

Other circulating factors reported to be associated with clinical outcomes after antiangiogenesis includes plasma angiopoietin-2, bFGF, platelet derived growth factor-BB, soluble Tie2, sICAM-1, and matrix metalloproteinases^[24,49,56,63-66] (Table 1).

Tissue biomarkers

Tissue based biomarkers are difficult to establish because of the invasive and costly nature of these procedures and the variations in immunohistochemical procedures and interpretations. Intratumoral levels of VEGF have not been shown to predict survival outcome of anti-VEGF therapy^[35,67], although correlations with response rates have been reported^[68]. On the other hand, increased SDF1 and CXCR4 were noted in rectal cancer patients after anti-VEGF treatment^[58,69]. High carbonic anhydrase IX expression was associated with better tumor shrinkage for metastatic renal cell carcinoma patients treated with sorafenib^[70]. Genetic studies of colorectal cancer did not associate *p53*, *KRAS* or *BRAF* mutations with bevacizumab treatment outcome^[71]. Single nucleotide polymorphisms (SNPs) in VEGF, VEGFR2 and VEGFR1 were associated with survival after treatment with bevacizumab based regimens^[45,52,72]. In line with the important role of inflammatory cytokines in angiogenesis, a consistent finding appeared to be the association between SNPs in *CXCR2* and *IL-8* genes and the outcome after anti-VEGF therapies^[57,73,74]. More extensive investigation and validation are warranted to determine a biomarker for antiangiogenesis therapy.

Circulating cells

In response to sunitinib, the number of circulating endothelial cells (CECs) and monocytes can be decreased in patients with HCC and gastrointestinal stromal tumors (GISTs)^[22,24]. However, In GISTs

Table 2 Imaging biomarkers of hepatocellular carcinoma patients receiving antiangiogenic therapy

Treatment	Imaging study	Patient n	Predictive value	Prognostic value	Ref.
Sorafenib or sunitinib	Perfusion CT, DCEUS	19	≤ 40% decrease in AUC at 1 mo correlates with PD		Frampas <i>et al</i> ^[113]
Sorafenib	CEUS	21	Reduction in enhancement correlates with response		Moschouris <i>et al</i> ^[114]
Sorafenib	Perfusion CT	10	Increase in MTT correlates with AFP response		Sacco <i>et al</i> ^[115]
Sunitinib	DCEMRI	24	Decreased K^{trans} or K_{ep} correlate PR/SD		Zhu <i>et al</i> ^[24]
Sunitinib	DWI, MRP	23	Decreased K^{trans} or K_{ep} at week 2 correlate with response	High baseline K^{trans} and decrease in EVF correlate with longer PFS	Sahani <i>et al</i> ^[107]
Bevacizumab	Perfusion CT	25	Low baseline MTT correlates with PD; increased MTT correlates with PR/SD		Zhu ^[2]
Bevacizumab	DCEUS	42	Decrease between day 0-3 of AUC, AUC during wash-in, AUC during wash-out, time to peak intensity correlate with tumor response	Time to peak intensity correlates with PFS; AUC and ACU during wash-out correlate with OS	Lassau <i>et al</i> ^[116]
Bevacizumab/ gemcitabine and oxaliplatin	Perfusion CT, dual-phase contrast enhanced CT	23	High baseline MTT correlates with PR/SD; high baseline K^{trans} correlates with responder	High baseline MTT correlates with better PFS	Jiang <i>et al</i> ^[117]
Bevacizumab	Perfusion CT	22		Reduction in percentage change of FD and low baseline FD correlate with longer OS	Hayano <i>et al</i> ^[118]
Thalidomide	Power Doppler US	47	High baseline vascular index in responder		Hsu <i>et al</i> ^[110]
Thalidomide	Perfusion CT	18	High baseline blood flow and blood volume correlates with progression		Petralia <i>et al</i> ^[119]
Thalidomide/ radiotherapy	DCEMRI	22	High baseline and week 2 Slope in responder	Perfusion parameters over liver parenchyma correlate with PFS and OS	Liang <i>et al</i> ^[87]
Pazopanib	DCEMRI	26	Reductions in IAUGC and K^{trans} not correlate with pharmacokinetic parameters		Yau <i>et al</i> ^[120]

CT: Computed tomography; DCEUS: Dynamic contrast enhanced ultrasonography; AUC: Area under curve; PD: Progressive disease; CEUS: Contrast enhanced ultrasonography; MTT: Mean transit time; AFP: α -fetal protein; DCEMRI: Dynamic contrast enhanced magnetic resonance imaging; K^{trans} : Transfer constant; K_{ep} : Redistribution rate constant; PR: Partial response; SD: Stable disease; DWI: Diffusion weighted imaging; MRP: Magnetic resonance imaging derived perfusion parameter; EVF: Extracellular volume fraction; PFS: Progression free survival; FD: Fractal dimension; OS: Overall survival; Slope: Initial first-pass enhancement slope; IAUGC: Initial area under the tissue gadolinium concentration-time curve.

patients, clinical benefit was significantly associated with increases in CECs ($P = 0.007$) as compared with those with progressive disease^[22]. TKIs such as cediranib or bevacizumab combined with chemotherapy did not change the amount of circulating progenitor cells. One of the caveats of using CECs as a biomarker is the means of assessment, which needs to be more rigorously established and standardized.

Imaging biomarkers

Noninvasive imaging has been widely applied for monitoring antiangiogenesis therapy in cancer drug discovery (Table 2). The techniques used in molecular imaging include positive emission tomography, single-photon emission computed tomography, molecular magnetic resonance imaging (MRI), optical fluorescence, optical bioluminescence, and targeted contrast-enhanced ultrasound. For example, temporal change in dynamic MRI and computed tomography (CT)-based tissue vascular measures such as blood flow, blood volume, or permeability have been shown to occur after

treatment with bevacizumab or anti-VEGFR TKIs in clinical studies^[75]. In HCC patients successfully treated with bevacizumab, CT perfusion imaging demonstrated substantial reductions in hepatic tumor blood flow, blood volume and permeability, findings that may predict treatment response^[76]. In MRI perfusion studies, HCC nodules treated with sorafenib showed a higher decrease in K^{trans} , which represents the volume transfer constant between blood plasma and the extravascular extracellular space. This finding reflects a decrease in tumor permeability and correlates with longer PFS and OS^[77]. The extent of drop in K^{trans} at day 14 after sunitinib in advanced HCC was significantly associated with PFS^[24]. The wide spread incorporation of perfusion as a biomarker has been hampered by inconsistencies in quantification results from different software and acquisition methods, as well as the time intensive analysis of data^[78,79] (Table 2).

The validation of clinical imaging of angiogenesis will be a slow and costly process. Different types of clinical trials that include histologic analysis will be needed.

CHALLENGES IN IDENTIFYING AND VALIDATING BIOMARKERS

Despite numerous investigations of antiangiogenic biomarkers, no validated biomarkers currently exist for predicting response or identifying appropriate patients for antiangiogenic therapy. Several challenges need to be overcome. Since the mechanisms regarding the actions of the currently approved antiangiogenic agents are not fully understood, there are no adequate criteria of pharmacologic response^[80,81]. The development of toxicity or resistance due to the activation of VEGF-independent pathways should also be explored. Besides, the biopsy or blood sample before treatment may not reflect the biology before subsequent treatment. There is also regional heterogeneity with one part of a tumor not necessarily having the same vascularity as another part. A spatially resolved "dynamic biomarkers" are warranted. Furthermore, the measurement of candidate biomarkers should be optimized and standardized before independent validation.

INCORPORATING THALIDOMIDE INTO RADIOTHERAPY FOR HCC: DYNAMIC CONTRAST ENHANCED MRI STUDIES FOR HCC DURING RADIOTHERAPY

With the advancement of modern radiation and respiratory-gating technique, radical radiation to a portion of liver can achieve a high local control rate in patients with advanced HCC^[82-84]. However, slow tumor shrinkage and rapid recurrence compromise treatment outcomes. The development of surrogate markers to monitor the response of HCC to radiation is important^[85]. The maximal response to radiotherapy is often achieved 6 mo after completion treatment. This slow response makes it difficult to modify an ineffective regimen for HCC in a timely fashion, especially in patients with a low level of serum α -fetoprotein. Furthermore, intrahepatic recurrence outside the field of radiation is a common cause of treatment failure^[82,83]. Scattered radiation related tissue inflammation and damage may have a deleterious effect on tumor control because of the release of cytokines or angiogenic factors^[86].

We evaluated the signal parameters of dynamic contrast enhanced MRI (DCEMRI) over liver parenchyma as well as liver tumor in HCC patients before, during and after radiotherapy. Initial enhancement slope and peak enhancement ratio, representing microcirculation and permeability to contrast material were measured over an operator-defined region of interest. From nineteen patients with advanced HCC, we found that increased signal parameters of the tumor at week 2 during radiation were associated with an improved local response. In the parenchyma, increased signal parameters at week 2 were associated with recurrence

or progression^[85]. The observation was validated in another forty-three patients. Signal parameters of baseline as well as week 2 during radiotherapy were higher in patients with responsive tumor^[87]. Multivariate analysis, however, showed signal parameters over liver parenchyma, but not over tumor, independently predicted PFS and OS^[87]. In line with the observation, univariate analysis showed Child-Pugh classification B and poor liver function predicted shorter PFS. These observations emphasized that liver function reserve, but not tumor response, of these heavily pretreated HCC patients impacts the survival after radiotherapy^[88].

INCORPORATING THALIDOMIDE INTO RADIOTHERAPY FOR HCC: CYTOKINES AND IMAGE STUDIES

With the a priori DECMRI study in HCC patients receiving radiotherapy, we evaluated the combination effect of thalidomide to radiotherapy within the same population of patients with identical image acquisition and analysis protocols^[87,89]. Thalidomide, an angiogenesis inhibitor, was noted to radio-sensitize tumors by reducing interstitial fluid pressure, increase perfusion and tumor reoxygenation^[90]. The anti-inflammatory effect of thalidomide could contribute to the radio-sensitization and disease control of HCC^[24,91]. Low dose thalidomide resulted in a response rate of less than 10% and a disease stabilizing rate of 50% in HCC patients^[92,93]. Twenty-four patients were enrolled and received concomitant thalidomide and radiation. Thalidomide was prescribed at a dose of 100 mg twice daily starting three days before radiotherapy to achieve a steady serum level^[94]. The clinical outcomes, cytokine and DCEMRI studies were compared with patients receiving radiotherapy alone. Thalidomide suppressed the serum bFGF significantly and to a lesser extent, the IL-6 and tumor necrosis factor α levels. Multivariate analysis revealed that baseline IL-6 and week 2 SDF1 α level independently predicted the PFS. A decreased SDF1 α at one month after radiotherapy complete was a significant prognostic factor of longer OS of HCC patients receiving radiotherapy. Patient with responsive or stabilized disease had significant longer OS (288 ± 51 d vs 203 ± 52 d, $P = 0.02$). However, none of the cytokines evaluated correlated significantly with tumor response after radiation. Despite acceptable toxicity and significant suppression of serum bFGF, thalidomide at current dosage and schedule did not correlate with tumor response and survival of HCC patients receiving radiotherapy^[89].

On the other hand, DECMRI studies of the 22 HCC patients receiving thalidomide and radiotherapy showed consistently that signal parameters at baseline and at week 2 during radiotherapy correlated with tumor response. However, the addition of thalidomide at current dosage and schedule did not change the signal

parameters significantly compared to the 22 patients receiving radiotherapy only^[87]. The inconsistency between serum biomarker and DCEMRI parameter was reported in a study using ribonucleotide reductase M2 inhibitor with radiation in pancreatic cancer patients^[95]. In our study, the significant suppression of bFGF by low dose thalidomide may be tumor-independent changes, nonetheless, reflect systemic exposure to thalidomide. They could serve as drug activity markers to determine optimal biological dose ranges, but not as predictive or prognostic biomarkers^[20,22]. Our studies indicate that daily dose of 200 mg thalidomide may induce a systemic suppression of angiogenic and inflammatory cytokines. However, the cytokine effect did not translate into vascular change within liver tumor or liver parenchyma. The optimal dosage and schedule of thalidomide during radiotherapy for HCC patients should be further explored.

The superior sensitivity and the lack of radiation put DCEMRI at the forefront of clinical translation as imaging biomarker. However, the analysis of abdominal and thoracic DCEMRI is often impaired by artifacts and mis-registration caused by physiologic motion. More recent reports suggested methods available to alleviate post-processing difficulties in DCEMRI for image analysis^[96]. DCEMRI parameters seemed to help to predict tumor angiogenesis measured by microvascular density and VEGF expression levels and discriminate malignant from normal tissue^[97-99]. A sufficient decrease in tumor vascular parameters was used to assign an appropriate dose for an additional phase II trial of an antiangiogenic therapy (AG-013736). The author showed that the day 2 vascular response measured using DCEMRI seemed to be a useful indicator of drug pharmacology^[100]. However, paradoxical negative correlation between K^{trans} and CD31 expression was reported as well^[101,102]. Continuing investigations are needed to accurately depict whether DCEMRI truly has a role in imaging tumor angiogenesis and evaluating response to antiangiogenesis therapy.

CONCLUSION

Recent preclinical and clinical data suggest the advantage of combining antiangiogenic agents with chemotherapy, radiotherapy or other biologic agents in numerous pathologies. However, in order to optimize the effectiveness of the combination, it is essential to study the mechanisms by which antiangiogenesis or strategies over molecular targets are obtained. Standardized systemic, tissue, circulating and imaging biomarkers should be incorporated into well run preclinical and clinical studies, in order to choose the optimal sequence and administration time of these drugs.

ACKNOWLEDGMENTS

We thank Jeffrey S Chang for English editing of this manuscript.

REFERENCES

- 1 **Thomas MB**, Abbruzzese JL. Opportunities for targeted therapies in hepatocellular carcinoma. *J Clin Oncol* 2005; **23**: 8093-8108 [PMID: 16258107 DOI: 10.1200/JCO.2004.00.1537]
- 2 **Zhu AX**. Development of sorafenib and other molecularly targeted agents in hepatocellular carcinoma. *Cancer* 2008; **112**: 250-259 [PMID: 18041064 DOI: 10.1002/cncr.23175]
- 3 **Yoshiji H**, Kuriyama S, Yoshii J, Ikenaka Y, Noguchi R, Hicklin DJ, Wu Y, Yanase K, Namisaki T, Kitade M, Yamazaki M, Tsujinoue H, Masaki T, Fukui H. Halting the interaction between vascular endothelial growth factor and its receptors attenuates liver carcinogenesis in mice. *Hepatology* 2004; **39**: 1517-1524 [PMID: 15185292 DOI: 10.1002/hep.20218]
- 4 **Llovet JM**, Ricci S, Mazzaferro V, Hilgard P, Gane E, Blanc JF, de Oliveira AC, Santoro A, Raoul JL, Forner A, Schwartz M, Porta C, Zeuzem S, Bolondi L, Greten TF, Galle PR, Seitz JF, Borbath I, Häussinger D, Giannaris T, Shan M, Moscovici M, Voliotis D, Bruix J. Sorafenib in advanced hepatocellular carcinoma. *N Engl J Med* 2008; **359**: 378-390 [PMID: 18650514 DOI: 10.1056/NEJMoa0708857]
- 5 **Cheng AL**, Kang YK, Chen Z, Tsao CJ, Qin S, Kim JS, Luo R, Feng J, Ye S, Yang TS, Xu J, Sun Y, Liang H, Liu J, Wang J, Tak WY, Pan H, Burock K, Zou J, Voliotis D, Guan Z. Efficacy and safety of sorafenib in patients in the Asia-Pacific region with advanced hepatocellular carcinoma: a phase III randomised, double-blind, placebo-controlled trial. *Lancet Oncol* 2009; **10**: 25-34 [PMID: 19095497 DOI: 10.1016/S1470-2045(08)70285-7]
- 6 **Cheng AL**, Kang YK, Lin DY, Park JW, Kudo M, Qin S, Chung HC, Song X, Xu J, Poggi G, Omata M, Pitman Lowenthal S, Lanzalone S, Yang L, Lechuga MJ, Raymond E. Sunitinib versus sorafenib in advanced hepatocellular cancer: results of a randomized phase III trial. *J Clin Oncol* 2013; **31**: 4067-4075 [PMID: 24081937 DOI: 10.1200/JCO.2012.45.8372]
- 7 **Johnson PJ**, Qin S, Park JW, Poon RT, Raoul JL, Philip PA, Hsu CH, Hu TH, Heo J, Xu J, Lu L, Chao Y, Boucher E, Han KH, Paik SW, Robles-Aviña J, Kudo M, Yan L, Sobhonslidsuk A, Komov D, Decaens T, Tak WY, Jeng LB, Liu D, Ezzeddine R, Walters I, Cheng AL. Brivanib versus sorafenib as first-line therapy in patients with unresectable, advanced hepatocellular carcinoma: results from the randomized phase III BRISK-FL study. *J Clin Oncol* 2013; **31**: 3517-3524 [PMID: 23980084 DOI: 10.1200/JCO.2012.48.4410]
- 8 **Cainap C**, Qin S, Huang WT, Chung KJ, Pan H, Yin C. Phase III trial of lenvatinib versus sorafenib in patients with advanced hepatocellular carcinoma (HCC). *J Clin Oncol* 2013; **31** Suppl 4: abstr 249
- 9 **Zhu AX**, Kudo M, Assenat E, Cattani S, Kang YK, Lim HY, Poon RT, Blanc JF, Vogel A, Chen CL, Dorval E, Peck-Radosavljevic M, Santoro A, Daniele B, Furuse J, Jappe A, Perraud K, Anak O, Sellami DB, Chen LT. Effect of everolimus on survival in advanced hepatocellular carcinoma after failure of sorafenib: the EVOLVE-1 randomized clinical trial. *JAMA* 2014; **312**: 57-67 [PMID: 25058218 DOI: 10.1001/jama.2014.7189]
- 10 **Sun W**, Sohal D, Haller DG, Mykulowycz K, Rosen M, Soulen MC, Caparro M, Teitelbaum UR, Giantonio B, O'Dwyer PJ, Shaked A, Reddy R, Olthoff K. Phase 2 trial of bevacizumab, capecitabine, and oxaliplatin in treatment of advanced hepatocellular carcinoma. *Cancer* 2011; **117**: 3187-3192 [PMID: 21264839 DOI: 10.1002/cncr.25889]
- 11 **Zhu AX**, Blaszkowsky LS, Ryan DP, Clark JW, Muzikansky A, Horgan K, Sheehan S, Hale KE, Enzinger PC, Bhargava P, Stuart K. Phase II study of gemcitabine and oxaliplatin in combination with bevacizumab in patients with advanced hepatocellular carcinoma. *J Clin Oncol* 2006; **24**: 1898-1903 [PMID: 16622265 DOI: 10.1200/JCO.2005.04.9130]
- 12 **Abou-Alfa GK**, Johnson P, Knox JJ, Capanu M, Davidenko I, Lacava J, Leung T, Gansukh B, Saltz LB. Doxorubicin plus sorafenib vs doxorubicin alone in patients with advanced

- hepatocellular carcinoma: a randomized trial. *JAMA* 2010; **304**: 2154-2160 [PMID: 21081728 DOI: 10.1001/jama.2010.1672]
- 13 **Santoro A**, Rimassa L, Borbath I, Daniele B, Salvagni S, Van Laethem JL, Van Vlierberghe H, Trojan J, Kolligs FT, Weiss A, Miles S, Gasbarrini A, Lencioni M, Cicalese L, Sherman M, Gridelli C, Buggisch P, Gerken G, Schmid RM, Boni C, Personeni N, Hassoun Z, Abbadessa G, Schwartz B, Von Roemeling R, Lamar ME, Chen Y, Porta C. Tivantinib for second-line treatment of advanced hepatocellular carcinoma: a randomised, placebo-controlled phase 2 study. *Lancet Oncol* 2013; **14**: 55-63 [PMID: 23182627 DOI: 10.1016/S1470-2045(12)70490-4]
 - 14 **Verslype C**, Cohn AL, Kelley RK, Yang TS, Su WC, Ramies DA, Lee Y, Shen X, Cutsem EV. Activity of cabozantinib (XL184) in hepatocellular carcinoma: Results from a phase II randomized discontinuation trial (RDT). *J Clin Oncol* 2012; **30** Suppl: abstr 4007
 - 15 **Zhu AX**, Kudo M, Assenat E, Caltan S, Kang YK, Lim Y, Poon TP, Blanc JF, Vogel A, Chen CL, Doival EP, Santoro A, Daniele B, Furuse J, Jappe A, Perraud K, Anak O, Sellami DB, Chen LT. EVOLVE-1: Phase 3 study of everolimus for advanced HCC that progressed during or after sorafenib. *J Clin Oncol* 2014; **32** Suppl 3: abstr 172
 - 16 **Finn RS**, Poon RT, Yau T, Klumpen HJ, Chen LT, Kang YK, Kim TY, Gomez-Martin C, Rodriguez-Lope C, Kunz T, Paquet T, Brandt U, Sellami D, Bruix J. Phase I study investigating everolimus combined with sorafenib in patients with advanced hepatocellular carcinoma. *J Hepatol* 2013; **59**: 1271-1277 [PMID: 23928403 DOI: 10.1016/j.jhep.2013.07.029]
 - 17 **Goyal L**, Zhu AX. Beyond sorafenib: Finding new effective targets in hepatocellular carcinoma. GI cancer symposium 2014. Daily News, 2014
 - 18 **El-Maraghi RH**, Eisenhauer EA. Review of phase II trial designs used in studies of molecular targeted agents: outcomes and predictors of success in phase III. *J Clin Oncol* 2008; **26**: 1346-1354 [PMID: 18285606 DOI: 10.1200/JCO.2007.13.5913]
 - 19 **Batchelor TT**, Sorensen AG, di Tomaso E, Zhang WT, Duda DG, Cohen KS, Kozak KR, Cahill DP, Chen PJ, Zhu M, Ancukiewicz M, Mrugala MM, Plotkin S, Drappatz J, Louis DN, Ivy P, Scadden DT, Benner T, Loeffler JS, Wen PY, Jain RK. AZD2171, a pan-VEGF receptor tyrosine kinase inhibitor, normalizes tumor vasculature and alleviates edema in glioblastoma patients. *Cancer Cell* 2007; **11**: 83-95 [PMID: 17222792 DOI: 10.1016/j.ccr.2006.11.021]
 - 20 **Ebos JM**, Lee CR, Christensen JG, Mutsaers AJ, Kerbel RS. Multiple circulating proangiogenic factors induced by sunitinib malate are tumor-independent and correlate with antitumor efficacy. *Proc Natl Acad Sci USA* 2007; **104**: 17069-17074 [PMID: 17942672 DOI: 10.1073/pnas.0708148104]
 - 21 **Kamoun WS**, Ley CD, Farrar CT, Duyverman AM, Lahdenranta J, Lacombe DA, Batchelor TT, di Tomaso E, Duda DG, Munn LL, Fukumura D, Sorensen AG, Jain RK. Edema control by cediranib, a vascular endothelial growth factor receptor-targeted kinase inhibitor, prolongs survival despite persistent brain tumor growth in mice. *J Clin Oncol* 2009; **27**: 2542-2552 [PMID: 19332720 DOI: 10.1200/JCO.2008.19.9356]
 - 22 **Norden-Zfoni A**, Desai J, Manola J, Beaudry P, Force J, Maki R, Folkman J, Bello C, Baum C, DePrimo SE, Shalinsky DR, Demetri GD, Heymach JV. Blood-based biomarkers of SU11248 activity and clinical outcome in patients with metastatic imatinib-resistant gastrointestinal stromal tumor. *Clin Cancer Res* 2007; **13**: 2643-2650 [PMID: 17473195 DOI: 10.1158/1078-0432.CCR-06-0919]
 - 23 **Willett CG**, Duda DG, di Tomaso E, Boucher Y, Ancukiewicz M, Sahani DV, Lahdenranta J, Chung DC, Fischman AJ, Lauwers GY, Shellito P, Czito BG, Wong TZ, Paulson E, Poleski M, Vujaskovic Z, Bentley R, Chen HX, Clark JW, Jain RK. Efficacy, safety, and biomarkers of neoadjuvant bevacizumab, radiation therapy, and fluorouracil in rectal cancer: a multidisciplinary phase II study. *J Clin Oncol* 2009; **27**: 3020-3026 [PMID: 19470921 DOI: 10.1200/JCO.2008.21.1771]
 - 24 **Zhu AX**, Sahani DV, Duda DG, di Tomaso E, Ancukiewicz M, Catalano OA, Sindhwani V, Blaszkowsky LS, Yoon SS, Lahdenranta J, Bhargava P, Meyerhardt J, Clark JW, Kwak EL, Hezel AF, Miksad R, Abrams TA, Enzinger PC, Fuchs CS, Ryan DP, Jain RK. Efficacy, safety, and potential biomarkers of sunitinib monotherapy in advanced hepatocellular carcinoma: a phase II study. *J Clin Oncol* 2009; **27**: 3027-3035 [PMID: 19470923 DOI: 10.1200/JCO.2008.20.9908]
 - 25 **Crabb SJ**, Patsios D, Sauerbrei E, Ellis PM, Arnold A, Goss G, Leigh NB, Shepherd FA, Powers J, Seymour L, Laurie SA. Tumor cavitation: impact on objective response evaluation in trials of angiogenesis inhibitors in non-small-cell lung cancer. *J Clin Oncol* 2009; **27**: 404-410 [PMID: 19047292 DOI: 10.1200/JCO.2008.16.2545]
 - 26 **Ferrara N**, Kerbel RS. Angiogenesis as a therapeutic target. *Nature* 2005; **438**: 967-974 [PMID: 16355214 DOI: 10.1038/nature04483]
 - 27 **Gerger A**, LaBonte M, Lenz HJ. Molecular predictors of response to antiangiogenesis therapies. *Cancer J* 2011; **17**: 134-141 [PMID: 21427557 DOI: 10.1097/PPO.0b013e318212db3c]
 - 28 **Hurwitz H**, Fehrenbacher L, Novotny W, Cartwright T, Hainsworth J, Heim W, Berlin J, Baron A, Griffing S, Holmgren E, Ferrara N, Fyfe G, Rogers B, Ross R, Kabbinavar F. Bevacizumab plus irinotecan, fluorouracil, and leucovorin for metastatic colorectal cancer. *N Engl J Med* 2004; **350**: 2335-2342 [PMID: 15175435 DOI: 10.1056/NEJMoa032691]
 - 29 **Scartozzi M**, Galizia E, Chiellini S, Giampieri R, Berardi R, Pierantoni C, Cascinu S. Arterial hypertension correlates with clinical outcome in colorectal cancer patients treated with first-line bevacizumab. *Ann Oncol* 2009; **20**: 227-230 [PMID: 18842611 DOI: 10.1093/annonc/mdn637]
 - 30 **Rudge JS**, Holash J, Hylton D, Russell M, Jiang S, Leidich R, Papadopoulos N, Pyles EA, Torri A, Wiegand SJ, Thurston G, Stahl N, Yancopoulos GD. VEGF Trap complex formation measures production rates of VEGF, providing a biomarker for predicting efficacious angiogenic blockade. *Proc Natl Acad Sci USA* 2007; **104**: 18363-18370 [PMID: 18000042 DOI: 10.1073/pnas.0708865104]
 - 31 **Loupakis F**, Falcone A, Masi G, Fioravanti A, Kerbel RS, Del Tacca M, Bocci G. Vascular endothelial growth factor levels in immunodepleted plasma of cancer patients as a possible pharmacodynamic marker for bevacizumab activity. *J Clin Oncol* 2007; **25**: 1816-1818 [PMID: 17470880 DOI: 10.1200/JCO.2006.1.03051]
 - 32 **Zhu AX**, Duda DG, Sahani DV, Jain RK. HCC and angiogenesis: possible targets and future directions. *Nat Rev Clin Oncol* 2011; **8**: 292-301 [PMID: 21386818 DOI: 10.1038/nrclinonc.2011.30]
 - 33 **Miles DW**, de Haas SL, Dirix LY, Romieu G, Chan A, Pivrot X, Tomczak P, Provencher L, Cortés J, Delmar PR, Scherer SJ. Biomarker results from the AVADO phase 3 trial of first-line bevacizumab plus docetaxel for HER2-negative metastatic breast cancer. *Br J Cancer* 2013; **108**: 1052-1060 [PMID: 23422754 DOI: 10.1038/bjc.2013.69]
 - 34 **Sandler A**, Gray R, Perry MC, Brahmer J, Schiller JH, Dowlati A, Lilenbaum R, Johnson DH. Paclitaxel-carboplatin alone or with bevacizumab for non-small-cell lung cancer. *N Engl J Med* 2006; **355**: 2542-2550 [PMID: 17167137 DOI: 10.1056/NEJMoa061884]
 - 35 **Jubb AM**, Hurwitz HI, Bai W, Holmgren EB, Tobin P, Guerrero AS, Kabbinavar F, Holden SN, Novotny WF, Frantz GD, Hillan KJ, Koeppen H. Impact of vascular endothelial growth factor-A expression, thrombospondin-2 expression, and microvessel density on the treatment effect of bevacizumab in metastatic colorectal cancer. *J Clin Oncol* 2006; **24**: 217-227 [PMID: 16365183 DOI: 10.1200/JCO.2005.01.5388]
 - 36 **Llovet JM**, Peña CE, Lathia CD, Shan M, Meinhardt G, Bruix J. Plasma biomarkers as predictors of outcome in patients with advanced hepatocellular carcinoma. *Clin Cancer Res* 2012; **18**: 2290-2300 [PMID: 22374331 DOI: 10.1158/1078-0432.CCR-11-2175]
 - 37 **Dowlati A**, Gray R, Sandler AB, Schiller JH, Johnson DH. Cell adhesion molecules, vascular endothelial growth factor, and

- basic fibroblast growth factor in patients with non-small cell lung cancer treated with chemotherapy with or without bevacizumab--an Eastern Cooperative Oncology Group Study. *Clin Cancer Res* 2008; **14**: 1407-1412 [PMID: 18316562 DOI: 10.1158/1078-0432.CCR-07-1154]
- 38 **Willett CG**, Boucher Y, Duda DG, di Tomaso E, Munn LL, Tong RT, Kozin SV, Petit L, Jain RK, Chung DC, Sahani DV, Kalva SP, Cohen KS, Scadden DT, Fischman AJ, Clark JW, Ryan DP, Zhu AX, Blaszkowsky LS, Shellito PC, Mino-Kenudson M, Lauwers GY. Surrogate markers for antiangiogenic therapy and dose-limiting toxicities for bevacizumab with radiation and chemotherapy: continued experience of a phase I trial in rectal cancer patients. *J Clin Oncol* 2005; **23**: 8136-8139 [PMID: 16258121 DOI: 10.1200/JCO.2005.02.5635]
- 39 **Rini BI**, Michaelson MD, Rosenberg JE, Bukowski RM, Sosman JA, Stadler WM, Hutson TE, Margolin K, Harmon CS, DePrimo SE, Kim ST, Chen I, George DJ. Antitumor activity and biomarker analysis of sunitinib in patients with bevacizumab-refractory metastatic renal cell carcinoma. *J Clin Oncol* 2008; **26**: 3743-3748 [PMID: 18669461 DOI: 10.1200/JCO.2007.15.5416]
- 40 **DePrimo SE**, Bello C. Surrogate biomarkers in evaluating response to anti-angiogenic agents: focus on sunitinib. *Ann Oncol* 2007; **18** Suppl 10: x11-x19 [PMID: 17761718 DOI: 10.1093/annonc/mdm409]
- 41 **Dreys J**, Siegert P, Medinger M, Mross K, Strecker R, Zirrgiebel U, Harder J, Blum H, Robertson J, Jürgensmeier JM, Puchalski TA, Young H, Saunders O, Unger C. Phase I clinical study of AZD2171, an oral vascular endothelial growth factor signaling inhibitor, in patients with advanced solid tumors. *J Clin Oncol* 2007; **25**: 3045-3054 [PMID: 17634482 DOI: 10.1200/JCO.2006.07.2066]
- 42 **Mita MM**, Rowinsky EK, Forero L, Eckhart SG, Izbicke E, Weiss GR, Beeram M, Mita AC, de Bono JS, Tolcher AW, Hammond LA, Simmons P, Berg K, Takimoto C, Patnaik A. A phase II, pharmacokinetic, and biologic study of semaxanib and thalidomide in patients with metastatic melanoma. *Cancer Chemother Pharmacol* 2007; **59**: 165-174 [PMID: 16736151 DOI: 10.1007/s00280-006-0255-0]
- 43 **Motzer RJ**, Michaelson MD, Redman BG, Hudes GR, Wilding G, Figlin RA, Ginsberg MS, Kim ST, Baum CM, DePrimo SE, Li JZ, Bello CL, Theuer CP, George DJ, Rini BI. Activity of SU11248, a multitargeted inhibitor of vascular endothelial growth factor receptor and platelet-derived growth factor receptor, in patients with metastatic renal cell carcinoma. *J Clin Oncol* 2006; **24**: 16-24 [PMID: 16330672 DOI: 10.1200/JCO.2005.02.2574]
- 44 **Saltz LB**, Rosen LS, Marshall JL, Belt RJ, Hurwitz HI, Eckhardt SG, Bergsland EK, Haller DG, Lockhart AC, Rocha Lima CM, Huang X, DePrimo SE, Chow-Maneval E, Chao RC, Lenz HJ. Phase II trial of sunitinib in patients with metastatic colorectal cancer after failure of standard therapy. *J Clin Oncol* 2007; **25**: 4793-4799 [PMID: 17947727 DOI: 10.1200/JCO.2007.12.8637]
- 45 **Schneider BP**, Wang M, Radovich M, Sledge GW, Badve S, Thor A, Flockhart DA, Hancock B, Davidson N, Gralow J, Dickler M, Perez EA, Cobleigh M, Shenkier T, Edgerton S, Miller KD. Association of vascular endothelial growth factor and vascular endothelial growth factor receptor-2 genetic polymorphisms with outcome in a trial of paclitaxel compared with paclitaxel plus bevacizumab in advanced breast cancer: ECOG 2100. *J Clin Oncol* 2008; **26**: 4672-4678 [PMID: 18824714 DOI: 10.1200/JCO.2008.16.1612]
- 46 **Horowitz NS**, Penson RT, Duda DG, di Tomaso E, Boucher Y, Ancukiewicz M, Cohen KS, Berlin S, Krasner CN, Moses MA, Jain RK. Safety, Efficacy, and Biomarker Exploration in a Phase II Study of Bevacizumab, Oxaliplatin, and Gemcitabine in Recurrent Müllerian Carcinoma. *Clin Ovarian Cancer Other Gynecol Malig* 2011; **4**: 26-33 [PMID: 21833345 DOI: 10.1016/j.cloc.2011.04.003]
- 47 **Van Cutsem E**, Tabernero J, Lakomy R, Prenen H, Prausová J, Macarulla T, Ruff P, van Hazel GA, Moiseyenko V, Ferry D, McKendrick J, Polikoff J, Tellier A, Castan R, Allegra C. Addition of aflibercept to fluorouracil, leucovorin, and irinotecan improves survival in a phase III randomized trial in patients with metastatic colorectal cancer previously treated with an oxaliplatin-based regimen. *J Clin Oncol* 2012; **30**: 3499-3506 [PMID: 22949147 DOI: 10.1200/JCO.2012.42.8201]
- 48 **Duda DG**, Willett CG, Ancukiewicz M, di Tomaso E, Shah M, Czito BG, Bentley R, Poleski M, Lauwers GY, Carroll M, Tyler D, Mantyh C, Shellito P, Clark JW, Jain RK. Plasma soluble VEGFR-1 is a potential dual biomarker of response and toxicity for bevacizumab with chemoradiation in locally advanced rectal cancer. *Oncologist* 2010; **15**: 577-583 [PMID: 20484123 DOI: 10.1634/theoncologist.2010-0029]
- 49 **Batchelor TT**, Duda DG, di Tomaso E, Ancukiewicz M, Plotkin SR, Gerstner E, Eichler AF, Drappatz J, Hochberg FH, Benner T, Louis DN, Cohen KS, Chea H, Exarhopoulos A, Loeffler JS, Moses MA, Ivy P, Sorensen AG, Wen PY, Jain RK. Phase II study of cediranib, an oral pan-vascular endothelial growth factor receptor tyrosine kinase inhibitor, in patients with recurrent glioblastoma. *J Clin Oncol* 2010; **28**: 2817-2823 [PMID: 20458050 DOI: 10.1200/JCO.2009.26.3988]
- 50 **Meyerhardt JA**, Ancukiewicz M, Abrams TA, Schrag D, Enzinger PC, Chan JA, Kulke MH, Wolpin BM, Goldstein M, Blaszkowsky L, Zhu AX, Elliott M, Regan E, Jain RK, Duda DG. Phase I study of cetuximab, irinotecan, and vandetanib (ZD6474) as therapy for patients with previously treated metastatic colorectal cancer. *PLoS One* 2012; **7**: e38231 [PMID: 22701615 DOI: 10.1371/journal.pone.0038231]
- 51 **Zhu AX**, Ancukiewicz M, Supko JG, Sahani DV, Blaszkowsky LS, Meyerhardt JA, Abrams TA, McCleary NJ, Bhargava P, Muzikansky A, Sheehan S, Regan E, Vasudev E, Knowles M, Fuchs CS, Ryan DP, Jain RK, Duda DG. Efficacy, safety, pharmacokinetics, and biomarkers of cediranib monotherapy in advanced hepatocellular carcinoma: a phase II study. *Clin Cancer Res* 2013; **19**: 1557-1566 [PMID: 23362324 DOI: 10.1158/1078-0432.CCR-12-3041]
- 52 **Lambrechts D**, Claes B, Delmar P, Reumers J, Mazzone M, Yesilyurt BT, Devlieger R, Verslype C, Tejpar S, Wildiers H, de Haas S, Carmeliet P, Scherer SJ, Van Cutsem E. VEGF pathway genetic variants as biomarkers of treatment outcome with bevacizumab: an analysis of data from the AVITA and AVOREN randomised trials. *Lancet Oncol* 2012; **13**: 724-733 [PMID: 22608783 DOI: 10.1016/S1470-2045(12)70231-0]
- 53 **Sorensen AG**, Batchelor TT, Zhang WT, Chen PJ, Yeo P, Wang M, Jennings D, Wen PY, Lahdenranta J, Ancukiewicz M, di Tomaso E, Duda DG, Jain RK. A "vascular normalization index" as potential mechanistic biomarker to predict survival after a single dose of cediranib in recurrent glioblastoma patients. *Cancer Res* 2009; **69**: 5296-5300 [PMID: 19549889 DOI: 10.1158/0008-5472.CAN-09-0814]
- 54 **Wilson PM**, Yang D, Azuma M, Shi MM, Danenberg KD, Lebowitz D, Sherrod A, Ladner RD, Zhang W, Danenberg PV, Trarbach T, Folprecht G, Meinhardt G, Lenz HJ. Intratumoral expression profiling of genes involved in angiogenesis in colorectal cancer patients treated with chemotherapy plus the VEGFR inhibitor PTK787/ZK 222584 (vatalanib). *Pharmacogenomics J* 2013; **13**: 410-416 [PMID: 22664478 DOI: 10.1038/tpj.2012.23]
- 55 **Koukourakis MI**, Giatromanolaki A, Sivridis E, Gatter KC, Trarbach T, Folprecht G, Shi MM, Lebowitz D, Jalava T, Laurent D, Meinhardt G, Harris AL. Prognostic and predictive role of lactate dehydrogenase 5 expression in colorectal cancer patients treated with PTK787/ZK 222584 (vatalanib) antiangiogenic therapy. *Clin Cancer Res* 2011; **17**: 4892-4900 [PMID: 21632858 DOI: 10.1158/1078-0432.CCR-10-2918]
- 56 **Hanrahan EO**, Lin HY, Kim ES, Yan S, Du DZ, McKee KS, Tran HT, Lee JJ, Ryan AJ, Langmuir P, Johnson BE, Heymach JV. Distinct patterns of cytokine and angiogenic factor modulation and markers of benefit for vandetanib and/or chemotherapy in patients with non-small-cell lung cancer. *J Clin Oncol* 2010; **28**: 193-201 [PMID: 19949019 DOI: 10.1200/JCO.2009.22.4279]
- 57 **Schultheis AM**, Lurje G, Rhodes KE, Zhang W, Yang D, Garcia AA, Morgan R, Gandara D, Scudder S, Oza A, Hirte H, Fleming

- G, Roman L, Lenz HJ. Polymorphisms and clinical outcome in recurrent ovarian cancer treated with cyclophosphamide and bevacizumab. *Clin Cancer Res* 2008; **14**: 7554-7563 [PMID: 19010874 DOI: 10.1158/1078-0432.CCR-08-0351]
- 58 **Duda DG**, Kozin SV, Kirkpatrick ND, Xu L, Fukumura D, Jain RK. CXCL12 (SDF1alpha)-CXCR4/CXCR7 pathway inhibition: an emerging sensitizer for anticancer therapies? *Clin Cancer Res* 2011; **17**: 2074-2080 [PMID: 21349998 DOI: 10.1158/1078-0432.CCR-10-2636]
- 59 **Raut CP**, Boucher Y, Duda DG, Morgan JA, Quek R, Ancukiewicz M, Lahdenranta J, Eder JP, Demetri GD, Jain RK. Effects of sorafenib on intra-tumoral interstitial fluid pressure and circulating biomarkers in patients with refractory sarcomas (NCI protocol 6948). *PLoS One* 2012; **7**: e26331 [PMID: 22347360 DOI: 10.1371/journal.pone.0026331]
- 60 **Willett CG**, Duda DG, Ancukiewicz M, Shah M, Czito BG, Bentley R, Poleski M, Fujita H, Lauwers GY, Carroll M, Tyler D, Mantyh C, Shellito P, Chung DC, Clark JW, Jain RK. A safety and survival analysis of neoadjuvant bevacizumab with standard chemoradiation in a phase I/II study compared with standard chemoradiation in locally advanced rectal cancer. *Oncologist* 2010; **15**: 845-851 [PMID: 20667969 DOI: 10.1634/theoncologist.2010-0030]
- 61 **Loges S**, Mazzone M, Hohensinner P, Carmeliet P. Silencing or fueling metastasis with VEGF inhibitors: antiangiogenesis revisited. *Cancer Cell* 2009; **15**: 167-170 [PMID: 19249675 DOI: 10.1016/j.ccr.2009.02.007]
- 62 **Páez-Ribes M**, Allen E, Hudock J, Takeda T, Okuyama H, Viñals F, Inoue M, Bergers G, Hanahan D, Casanovas O. Antiangiogenic therapy elicits malignant progression of tumors to increased local invasion and distant metastasis. *Cancer Cell* 2009; **15**: 220-231 [PMID: 19249680 DOI: 10.1016/j.ccr.2009.01.027]
- 63 **Lieu C**, Heymach J, Overman M, Tran H, Kopetz S. Beyond VEGF: inhibition of the fibroblast growth factor pathway and antiangiogenesis. *Clin Cancer Res* 2011; **17**: 6130-6139 [PMID: 21953501 DOI: 10.1158/1078-0432.CCR-11-0659]
- 64 **Kopetz S**, Hoff PM, Morris JS, Wolff RA, Eng C, Glover KY, Adinin R, Overman MJ, Valero V, Wen S, Lieu C, Yan S, Tran HT, Ellis LM, Abbruzzese JL, Heymach JV. Phase II trial of infusional fluorouracil, irinotecan, and bevacizumab for metastatic colorectal cancer: efficacy and circulating angiogenic biomarkers associated with therapeutic resistance. *J Clin Oncol* 2010; **28**: 453-459 [PMID: 20008624 DOI: 10.1200/JCO.2009.24.8252]
- 65 **Tolaney SM**, Duda DG, Boucher Y. A phase II study of preoperative (preop) bevacizumab (bev) followed by dose-dense (dd) doxorubicin (A)/cyclophosphamide (C)/paclitaxel (T) in combination with bev in HER2-negative operable breast cancer (BC). *J Clin Oncol* 2012; **30** Suppl: abstr 1026
- 66 **Batchelor TT**, Gerstner ER, Emblem KE, Duda DG, Kalpathy-Cramer J, Snuderl M, Ancukiewicz M, Polaskova P, Pinho MC, Jennings D, Plotkin SR, Chi AS, Eichler AF, Dietrich J, Hochberg FH, Lu-Emerson C, Iafrate AJ, Ivy SP, Rosen BR, Loeffler JS, Wen PY, Sorensen AG, Jain RK. Improved tumor oxygenation and survival in glioblastoma patients who show increased blood perfusion after cediranib and chemoradiation. *Proc Natl Acad Sci USA* 2013; **110**: 19059-19064 [PMID: 24190997 DOI: 10.1073/pnas.1318022110]
- 67 **Hegde PS**, Jubb AM, Chen D, Li NF, Meng YG, Bernaards C, Elliott R, Scherer SJ, Chen DS. Predictive impact of circulating vascular endothelial growth factor in four phase III trials evaluating bevacizumab. *Clin Cancer Res* 2013; **19**: 929-937 [PMID: 23169435 DOI: 10.1158/1078-0432.CCR-12-2535]
- 68 **Yang SX**, Steinberg SM, Nguyen D, Wu TD, Modrusan Z, Swain SM. Gene expression profile and angiogenic marker correlates with response to neoadjuvant bevacizumab followed by bevacizumab plus chemotherapy in breast cancer. *Clin Cancer Res* 2008; **14**: 5893-5899 [PMID: 18794102 DOI: 10.1158/1078-0432.CCR-07-4762]
- 69 **Xu L**, Duda DG, di Tomaso E, Ancukiewicz M, Chung DC, Lauwers GY, Samuel R, Shellito P, Czito BG, Lin PC, Poleski M, Bentley R, Clark JW, Willett CG, Jain RK. Direct evidence that bevacizumab, an anti-VEGF antibody, up-regulates SDF1alpha, CXCR4, CXCL6, and neuropilin 1 in tumors from patients with rectal cancer. *Cancer Res* 2009; **69**: 7905-7910 [PMID: 19826039 DOI: 10.1158/0008-5472.CAN-09-2099]
- 70 **Choueiri TK**, Regan MM, Rosenberg JE, Oh WK, Clement J, Amato AM, McDermott D, Cho DC, Atkins MB, Signoretti S. Carbonic anhydrase IX and pathological features as predictors of outcome in patients with metastatic clear-cell renal cell carcinoma receiving vascular endothelial growth factor-targeted therapy. *BJU Int* 2010; **106**: 772-778 [PMID: 20230385 DOI: 10.1111/j.1464-410X.2010.09218.x]
- 71 **Ince WL**, Jubb AM, Holden SN, Holmgren EB, Tobin P, Sridhar M, Hurwitz HI, Kabbinavar F, Novotny WF, Hillan KJ, Koeppen H. Association of k-ras, b-raf, and p53 status with the treatment effect of bevacizumab. *J Natl Cancer Inst* 2005; **97**: 981-989 [PMID: 15998951 DOI: 10.1093/jnci/dji174]
- 72 **Loupakis F**, Ruzzo A, Salvatore L, Cremolini C, Masi G, Frumento P, Schirripa M, Catalano V, Galluccio N, Canestrari E, Vincenzi B, Santini D, Bencardino K, Ricci V, Manzoni M, Danova M, Tonini G, Magnani M, Falcone A, Graziano F. Retrospective exploratory analysis of VEGF polymorphisms in the prediction of benefit from first-line FOLFIRI plus bevacizumab in metastatic colorectal cancer. *BMC Cancer* 2011; **11**: 247 [PMID: 21669012 DOI: 10.1186/1471-2407-11-247]
- 73 **Gerger A**, El-Khoueiry A, Zhang W, Yang D, Singh H, Bohanes P, Ning Y, Winder T, Labonte MJ, Wilson PM, Benhaim L, Paez D, El-Khoueiry R, Absenger G, Lenz HJ. Pharmacogenetic angiogenesis profiling for first-line Bevacizumab plus oxaliplatin-based chemotherapy in patients with metastatic colorectal cancer. *Clin Cancer Res* 2011; **17**: 5783-5792 [PMID: 21791631 DOI: 10.1158/1078-0432.CCR-11-1115]
- 74 **Zhang WW**, Cortes JE, Yao H, Zhang L, Reddy NG, Jabbour E, Kantarjian HM, Jones D. Predictors of primary imatinib resistance in chronic myelogenous leukemia are distinct from those in secondary imatinib resistance. *J Clin Oncol* 2009; **27**: 3642-3649 [PMID: 19506164 DOI: 10.1200/JCO.2008.19.4076]
- 75 **Jiang T**, Zhu AX, Sahani DV. Established and novel imaging biomarkers for assessing response to therapy in hepatocellular carcinoma. *J Hepatol* 2013; **58**: 169-177 [PMID: 22944253 DOI: 10.1016/j.jhep.2012.08.022]
- 76 **Zhu AX**, Holalkere NS, Muzikansky A, Horgan K, Sahani DV. Early antiangiogenic activity of bevacizumab evaluated by computed tomography perfusion scan in patients with advanced hepatocellular carcinoma. *Oncologist* 2008; **13**: 120-125 [PMID: 18305056 DOI: 10.1634/theoncologist.2007-0174]
- 77 **Hsu CY**, Shen YC, Yu CW, Hsu C, Hu FC, Hsu CH, Chen BB, Wei SY, Cheng AL, Shih TT. Dynamic contrast-enhanced magnetic resonance imaging biomarkers predict survival and response in hepatocellular carcinoma patients treated with sorafenib and metronomic tegafur/uracil. *J Hepatol* 2011; **55**: 858-865 [PMID: 21338641 DOI: 10.1016/j.jhep.2011.01.032]
- 78 **García-Figueiras R**, Goh VJ, Padhani AR, Baleato-González S, Garrido M, León L, Gómez-Caamaño A. CT perfusion in oncologic imaging: a useful tool? *AJR Am J Roentgenol* 2013; **200**: 8-19 [PMID: 23255736 DOI: 10.2214/AJR.11.8476]
- 79 **Ng CS**, Chandler AG, Wei W, Herron DH, Anderson EF, Kurzrock R, Charnsangavej C. Reproducibility of CT perfusion parameters in liver tumors and normal liver. *Radiology* 2011; **260**: 762-770 [PMID: 21788525 DOI: 10.1148/radiol.11110331]
- 80 **Carmeliet P**. VEGF as a key mediator of angiogenesis in cancer. *Oncology* 2005; **69** Suppl 3: 4-10 [PMID: 16301830 DOI: 10.1159/000088478]
- 81 **Jain RK**, Duda DG, Clark JW, Loeffler JS. Lessons from phase III clinical trials on anti-VEGF therapy for cancer. *Nat Clin Pract Oncol* 2006; **3**: 24-40 [PMID: 16407877 DOI: 10.1038/ncon0403]
- 82 **Seong J**, Park HC, Han KH, Lee DY, Lee JT, Chon CY, Moon YM, Suh CO. Local radiotherapy for unresectable hepatocellular carcinoma patients who failed with transcatheter arterial chemoembolization. *Int J Radiat Oncol Biol Phys* 2000; **47**: 1331-1335 [PMID: 10889387]

- 83 **Cheng JC**, Chuang VP, Cheng SH, Huang AT, Lin YM, Cheng TI, Yang PS, You DL, Jian JJ, Tsai SY, Sung JL, Horng CF. Local radiotherapy with or without transcatheter arterial chemoembolization for patients with unresectable hepatocellular carcinoma. *Int J Radiat Oncol Biol Phys* 2000; **47**: 435-442 [PMID: 10802371]
- 84 **Herfarth KK**, Debus J, Lohr F, Bahner ML, Rhein B, Fritz P, Höss A, Schlegel W, Wannenmacher MF. Stereotactic single-dose radiation therapy of liver tumors: results of a phase I/II trial. *J Clin Oncol* 2001; **19**: 164-170 [PMID: 11134209]
- 85 **Liang PC**, Ch'ang HJ, Hsu C, Tseng SS, Shih TT, Wu Liu T. Dynamic MRI signals in the second week of radiotherapy relate to treatment outcomes of hepatocellular carcinoma: a preliminary result. *Liver Int* 2007; **27**: 516-528 [PMID: 17403192 DOI: 10.1111/j.1478-3231.2007.01456.x]
- 86 **Camphausen K**, Moses MA, Beecken WD, Khan MK, Folkman J, O'Reilly MS. Radiation therapy to a primary tumor accelerates metastatic growth in mice. *Cancer Res* 2001; **61**: 2207-2211 [PMID: 11280788]
- 87 **Liang PC**, Ch'ang HJ, Hsu C, Chen LT, Shih TT, Liu TW. Perfusion parameters of dynamic contrast-enhanced magnetic resonance imaging predict outcomes of hepatocellular carcinoma receiving radiotherapy with or without thalidomide. *Hepatol Int* 2015; **9**: 258-268 [PMID: 25788178 DOI: 10.1007/s12072-014-9557-1]
- 88 **Seong J**, Lee IJ, Shim SJ, Lim do H, Kim TH, Kim JH, Jang HS, Kim MS, Chie EK, Kim JH, Nam TK, Lee HS, Han CJ. A multicenter retrospective cohort study of practice patterns and clinical outcome on radiotherapy for hepatocellular carcinoma in Korea. *Liver Int* 2009; **29**: 147-152 [PMID: 18795897 DOI: 10.1111/j.1478-3231.2008.01873.x]
- 89 **Ch'ang HJ**, Hsu C, Chen CH, Chang YH, Chang JS, Chen LT. Phase II study of concomitant thalidomide during radiotherapy for hepatocellular carcinoma. *Int J Radiat Oncol Biol Phys* 2012; **82**: 817-825 [PMID: 21277098 DOI: 10.1016/j.ijrobp.2010.10.067]
- 90 **Ansiaux R**, Baudelet C, Jordan BF, Beghein N, Sonveaux P, De Wever J, Martinive P, Grégoire V, Feron O, Gallez B. Thalidomide radiosensitizes tumors through early changes in the tumor microenvironment. *Clin Cancer Res* 2005; **11**: 743-750 [PMID: 15701864]
- 91 **Crookart N**, Radermacher K, Jordan BF, Baudelet C, Cron GO, Grégoire V, Beghein N, Bouzin C, Feron O, Gallez B. Tumor radiosensitization by antiinflammatory drugs: evidence for a new mechanism involving the oxygen effect. *Cancer Res* 2005; **65**: 7911-7916 [PMID: 16140962 DOI: 10.1158/0008-5472.CAN-05-1288]
- 92 **Hsu C**, Chen CN, Chen LT, Wu CY, Yang PM, Lai MY, Lee PH, Cheng AL. Low-dose thalidomide treatment for advanced hepatocellular carcinoma. *Oncology* 2003; **65**: 242-249 [PMID: 14657598]
- 93 **Chiou HE**, Wang TE, Wang YY, Liu HW. Efficacy and safety of thalidomide in patients with hepatocellular carcinoma. *World J Gastroenterol* 2006; **12**: 6955-6960 [PMID: 17109516]
- 94 **Shiah HS**, Chao Y, Chen LT, Yao TJ, Huang JD, Chang JY, Chen PJ, Chuang TR, Chin YH, Whang-Peng J, Liu TW. Phase I and pharmacokinetic study of oral thalidomide in patients with advanced hepatocellular carcinoma. *Cancer Chemother Pharmacol* 2006; **58**: 654-664 [PMID: 16520988]
- 95 **Martin LK**, Grecula J, Jia G, Wei L, Yang X, Otterson GA, Wu X, Harper E, Kefauver C, Zhou BS, Yen Y, Bloomston M, Knopp M, Ivy SP, Grever M, Bekaii-Saab T. A dose escalation and pharmacodynamic study of triapine and radiation in patients with locally advanced pancreas cancer. *Int J Radiat Oncol Biol Phys* 2012; **84**: e475-e481 [PMID: 22818416 DOI: 10.1016/j.ijrobp.2012.06.003]
- 96 **Filipovic M**, Vuissoz PA, Codreanu A, Claudon M, Felblinger J. Motion compensated generalized reconstruction for free-breathing dynamic contrast-enhanced MRI. *Magn Reson Med* 2011; **65**: 812-822 [PMID: 20882640]
- 97 **Zhang XM**, Yu D, Zhang HL, Dai Y, Bi D, Liu Z, Prince MR, Li C. 3D dynamic contrast-enhanced MRI of rectal carcinoma at 3T: correlation with microvascular density and vascular endothelial growth factor markers of tumor angiogenesis. *J Magn Reson Imaging* 2008; **27**: 1309-1316 [PMID: 18504761]
- 98 **Tuncbilek N**, Kaplan M, Altaner S, Atakan IH, Süt N, Inci O, Demir MK. Value of dynamic contrast-enhanced MRI and correlation with tumor angiogenesis in bladder cancer. *AJR Am J Roentgenol* 2009; **192**: 949-955 [PMID: 19304699 DOI: 10.2214/AJR.08.1332]
- 99 **Thomassin-Naggara I**, Bazot M, Daraï E, Callard P, Thomassin J, Cuenod CA. Epithelial ovarian tumors: value of dynamic contrast-enhanced MR imaging and correlation with tumor angiogenesis. *Radiology* 2008; **248**: 148-159 [PMID: 18458244 DOI: 10.1148/radiol.2481071120]
- 100 **Liu G**, Rugo HS, Wilding G, McShane TM, Evelhoch JL, Ng C, Jackson E, Kelcz F, Yeh BM, Lee FT, Charnsangavej C, Park JW, Ashton EA, Steinfeldt HM, Pithavala YK, Reich SD, Herbst RS. Dynamic contrast-enhanced magnetic resonance imaging as a pharmacodynamic measure of response after acute dosing of AG-013736, an oral angiogenesis inhibitor, in patients with advanced solid tumors: results from a phase I study. *J Clin Oncol* 2005; **23**: 5464-5473 [PMID: 16027440 DOI: 10.1200/JCO.2005.04.143]
- 101 **Atkin G**, Taylor NJ, Daley FM, Stirling JJ, Richman P, Glynn-Jones R, d'Arcy JA, Collins DJ, Padhani AR. Dynamic contrast-enhanced magnetic resonance imaging is a poor measure of rectal cancer angiogenesis. *Br J Surg* 2006; **93**: 992-1000 [PMID: 16673354]
- 102 **Su MY**, Cheung YC, Fruehauf JP, Yu H, Nalcioglu O, Mechetner E, Kyshtobayeva A, Chen SC, Hsueh S, McLaren CE, Wan YL. Correlation of dynamic contrast enhancement MRI parameters with microvessel density and VEGF for assessment of angiogenesis in breast cancer. *J Magn Reson Imaging* 2003; **18**: 467-477 [PMID: 14508784]
- 103 **Shao YY**, Lin ZZ, Chen TJ, Hsu C, Shen YC, Hsu CH, Cheng AL. High circulating endothelial progenitor levels associated with poor survival of advanced hepatocellular carcinoma patients receiving sorafenib combined with metronomic chemotherapy. *Oncology* 2011; **81**: 98-103 [PMID: 21986371 DOI: 10.1159/000331684]
- 104 **Miyahara K**, Nouse K, Tomoda T, Kobayashi S, Hagihara H, Kuwaki K, Toshimori J, Onishi H, Ikeda F, Miyake Y, Nakamura S, Shiraha H, Takaki A, Yamamoto K. Predicting the treatment effect of sorafenib using serum angiogenesis markers in patients with hepatocellular carcinoma. *J Gastroenterol Hepatol* 2011; **26**: 1604-1611 [PMID: 22011296]
- 105 **Xiang Q**, Chen W, Ren M, Wang J, Zhang H, Deng DY, Zhang L, Shang C, Chen Y. Cabozantinib suppresses tumor growth and metastasis in hepatocellular carcinoma by a dual blockade of VEGFR2 and MET. *Clin Cancer Res* 2014; **20**: 2959-2970 [PMID: 24700742 DOI: 10.1158/1078-0432.CCR-13-2620]
- 106 **Harmon CS**, DePrimo SE, Raymond E, Cheng AL, Boucher E, Douillard JY, Lim HY, Kim JS, Lechuga MJ, Lanzalone S, Lin X, Faivre S. Mechanism-related circulating proteins as biomarkers for clinical outcome in patients with unresectable hepatocellular carcinoma receiving sunitinib. *J Transl Med* 2011; **9**: 120 [PMID: 21787417 DOI: 10.1186/1479-5876-9-120]
- 107 **Sahani DV**, Jiang T, Hayano K, Duda DG, Catalano OA, Ancukiewicz M, Jain RK, Zhu AX. Magnetic resonance imaging biomarkers in hepatocellular carcinoma: association with response and circulating biomarkers after sunitinib therapy. *J Hematol Oncol* 2013; **6**: 51 [PMID: 23842041 DOI: 10.1186/1756-8722-6-51]
- 108 **Boige V**, Malka D, Bourredjem A, Dromain C, Baey C, Jacques N, Pignon JP, Vimond N, Bouvet-Forteau N, De Baere T, Ducreux M, Farace F. Efficacy, safety, and biomarkers of single-agent bevacizumab therapy in patients with advanced hepatocellular carcinoma. *Oncologist* 2012; **17**: 1063-1072 [PMID: 22707516 DOI: 10.1634/theoncologist.2011-0465]
- 109 **Kaseb AO**, Garrett-Mayer E, Morris JS, Xiao L, Lin E, Onicescu G, Hassan MM, Hassabo HM, Iwasaki M, Deaton FL, Abbruzzese JL, Thomas MB. Efficacy of bevacizumab plus erlotinib for advanced hepatocellular carcinoma and predictors of outcome: final results

- of a phase II trial. *Oncology* 2012; **82**: 67-74 [PMID: 22327795 DOI: 10.1159/000335963]
- 110 **Hsu C**, Chen CN, Chen LT, Wu CY, Hsieh FJ, Cheng AL. Effect of thalidomide in hepatocellular carcinoma: assessment with power doppler US and analysis of circulating angiogenic factors. *Radiology* 2005; **235**: 509-516 [PMID: 15858091 DOI: 10.1148/radiol.2352040271]
- 111 **Shao YY**, Lin ZZ, Hsu C, Lee KD, Hsiao CH, Lu YS, Huang CC, Shen YC, Hsu CH, Cheng AL. Efficacy, safety, and potential biomarkers of thalidomide plus metronomic chemotherapy for advanced hepatocellular carcinoma. *Oncology* 2012; **82**: 59-66 [PMID: 22310088 DOI: 10.1159/000336126]
- 112 **Kanai F**, Yoshida H, Tateishi R, Sato S, Kawabe T, Obi S, Kondo Y, Taniguchi M, Tagawa K, Ikeda M, Morizane C, Okusaka T, Arioka H, Shiina S, Omata M. A phase I/II trial of the oral antiangiogenic agent TSU-68 in patients with advanced hepatocellular carcinoma. *Cancer Chemother Pharmacol* 2011; **67**: 315-324 [PMID: 20390419]
- 113 **Frampas E**, Lassau N, Zappa M, Vullierme MP, Koscielny S, Vilgrain V. Advanced Hepatocellular Carcinoma: early evaluation of response to targeted therapy and prognostic value of Perfusion CT and Dynamic Contrast Enhanced-Ultrasound. Preliminary results. *Eur J Radiol* 2013; **82**: e205-e211 [PMID: 23273822 DOI: 10.1016/j.ejrad.2012.12.004]
- 114 **Moschouris H**, Malagari K, Gkoutzios P, Kalokairinou M, Stamatiou K, Chatzimichail K, Kornezos I, Karagiannis E, Kiltenis M, Papadaki MG. Intermediate and advanced hepatocellular carcinoma treated with the antiangiogenic agent sorafenib. Evaluation with unenhanced and contrast-enhanced ultrasonography. *Med Ultrason* 2012; **14**: 87-94 [PMID: 22675707]
- 115 **Sacco R**, Faggioni L, Bargellini I, Ginanni B, Battaglia V, Romano A, Bertini M, Bresci G, Bartolozzi C. Assessment of response to sorafenib in advanced hepatocellular carcinoma using perfusion computed tomography: results of a pilot study. *Dig Liver Dis* 2013; **45**: 776-781 [PMID: 23578581 DOI: 10.1016/j.dld.2013.03.004]
- 116 **Lassau N**, Koscielny S, Chami L, Chebil M, Benatsou B, Roche A, Ducreux M, Malka D, Boige V. Advanced hepatocellular carcinoma: early evaluation of response to bevacizumab therapy at dynamic contrast-enhanced US with quantification--preliminary results. *Radiology* 2011; **258**: 291-300 [PMID: 20980447 DOI: 10.1148/radiol.10091870]
- 117 **Jiang T**, Kambadakone A, Kulkarni NM, Zhu AX, Sahani DV. Monitoring response to antiangiogenic treatment and predicting outcomes in advanced hepatocellular carcinoma using image biomarkers, CT perfusion, tumor density, and tumor size (RECIST). *Invest Radiol* 2012; **47**: 11-17 [PMID: 21512396]
- 118 **Hayano K**, Lee SH, Yoshida H, Zhu AX, Sahani DV. Fractal analysis of CT perfusion images for evaluation of antiangiogenic treatment and survival in hepatocellular carcinoma. *Acad Radiol* 2014; **21**: 654-660 [PMID: 24703479 DOI: 10.1016/j.acra.2014.01.020]
- 119 **Petralia G**, Fazio N, Bonello L, D'Andrea G, Radice D, Bellomi M. Perfusion computed tomography in patients with hepatocellular carcinoma treated with thalidomide: initial experience. *J Comput Assist Tomogr* 2011; **35**: 195-201 [PMID: 21412089]
- 120 **Yau T**, Chen PJ, Chan P, Curtis CM, Murphy PS, Suttle AB, Gauvin J, Hodge JP, Dar MM, Poon RT. Phase I dose-finding study of pazopanib in hepatocellular carcinoma: evaluation of early efficacy, pharmacokinetics, and pharmacodynamics. *Clin Cancer Res* 2011; **17**: 6914-6923 [PMID: 21831954 DOI: 10.1158/1078-0432.CCR-11-0793]

P- Reviewer: Elshimali YI S- Editor: Gong XM
L- Editor: A E- Editor: Liu SQ





Published by **Baishideng Publishing Group Inc**

8226 Regency Drive, Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: bpgoffice@wjgnet.com

Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>

<http://www.wjgnet.com>

