



## Research

**Cite this article:** Genner MJ, Ngatunga BP, Mzighani S, Smith A, Turner GF. 2015 Geographical ancestry of Lake Malawi's cichlid fish diversity. *Biol. Lett.* **11**: 20150232. <http://dx.doi.org/10.1098/rsbl.2015.0232>

Received: 26 March 2015

Accepted: 15 May 2015

### Subject Areas:

evolution, taxonomy and systematics

### Keywords:

adaptive radiation, phylogeny, African fishes

### Author for correspondence:

Martin J. Genner

e-mail: [m.genner@bristol.ac.uk](mailto:m.genner@bristol.ac.uk)

Electronic supplementary material is available at <http://dx.doi.org/10.1098/rsbl.2015.0232> or via <http://rsbl.royalsocietypublishing.org>.

# Geographical ancestry of Lake Malawi's cichlid fish diversity

Martin J. Genner<sup>1</sup>, Benjamin P. Ngatunga<sup>2</sup>, Semvua Mzighani<sup>2</sup>, Alan Smith<sup>3</sup> and George F. Turner<sup>4</sup>

<sup>1</sup>School of Biological Sciences, University of Bristol, Life Sciences Building, Bristol BS81TQ, UK

<sup>2</sup>Tanzania Fisheries Research Institute (TAFIRI), PO Box 9750, Dar-es-Salaam, Tanzania

<sup>3</sup>School of Biological, Biomedical and Environmental Science, University of Hull, Hull HU67RX, UK

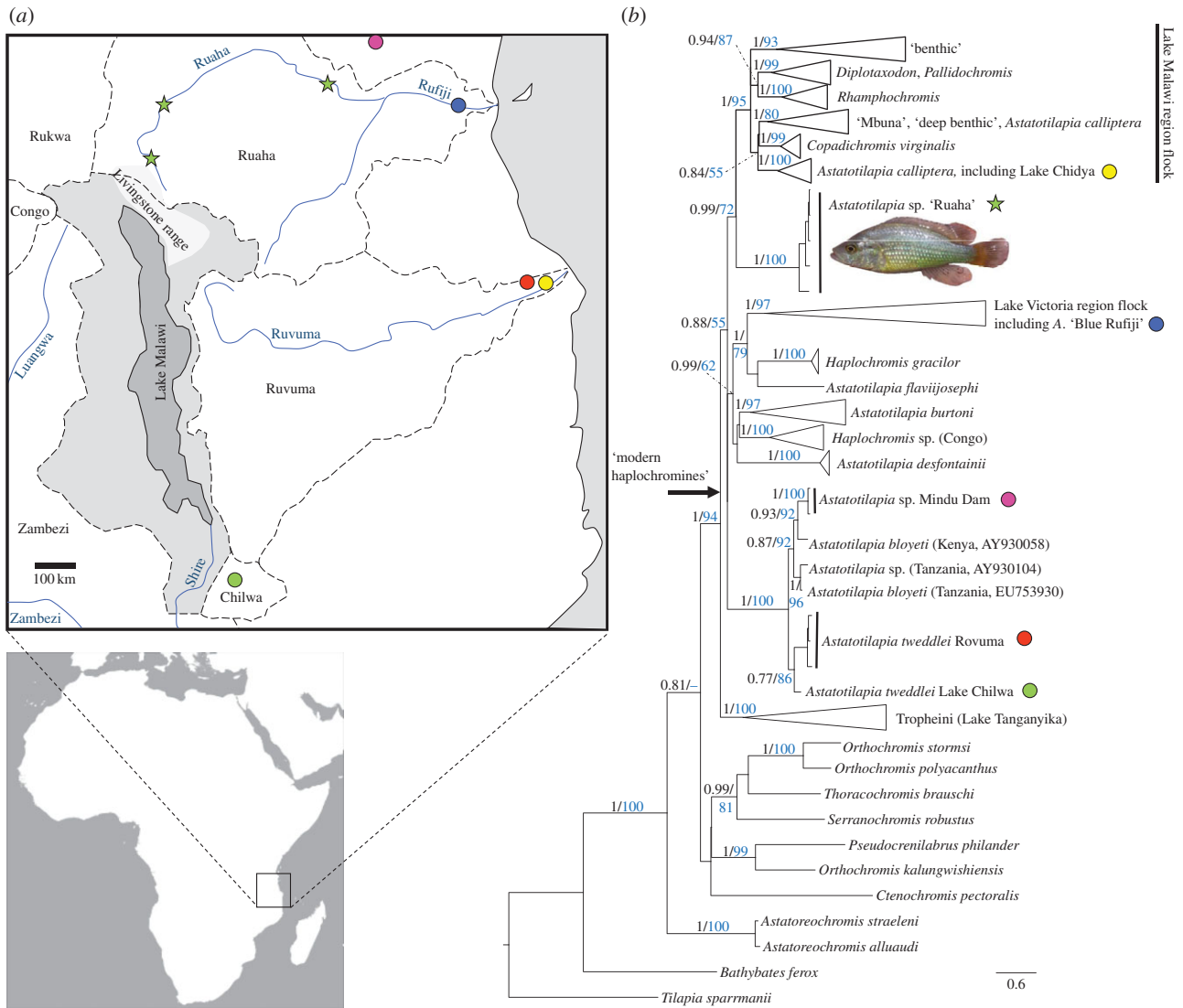
<sup>4</sup>School of Biological Sciences, Bangor University, Bangor, Gwynedd LL572UW, UK

The Lake Malawi haplochromine cichlid flock is one of the largest vertebrate adaptive radiations. The geographical source of the radiation has been assumed to be rivers to the south and east of Lake Malawi, where extant representatives of the flock are now present. Here, we provide mitochondrial DNA evidence suggesting the sister taxon to the Lake Malawi radiation is within the Great Ruaha river in Tanzania, north of Lake Malawi. Estimates of the time of divergence between the Lake Malawi flock and this riverine sister taxon range from 2.13 to 6.76 Ma, prior to origins of the current radiation 1.20–4.06 Ma. These results are congruent with evaluations of 2–3.75 Ma fossil material that suggest past faunal connections between Lake Malawi and the Ruaha. We propose that ancestors of the Malawi radiation became isolated within the catchment during Pliocene rifting that formed both Lake Malawi and the Kipengere/Livingstone mountain range, before colonizing rivers to the south and east of the lake region and radiating within the lake basin. Identification of this sister taxon allows tests of whether standing genetic diversity has predisposed Lake Malawi cichlids to rapid speciation and adaptive radiation.

## 1. Introduction

Adaptive radiations make up a high proportion of biodiversity. In many cases, ancestors or sister species of these flocks have been identified, as with Galapagos finches [1], Hawaiian silverswords [2] and Canadian three-spined sticklebacks [3]. Identification of their origins has enabled discussion of events that initiated adaptive radiation, and allowed tests of whether diversification has been promoted by novel mutations that have arisen since colonization, or instead whether adaptation is based primarily on pre-existing genetic variation [4]. This is an important issue to resolve, because it can explain why only some colonizing lineages radiate when provided with ecological opportunity, and how parallel adaptive radiation can take place rapidly in geographically separated habitats.

The evolutionary origins of cichlid fishes radiations in East African lakes are largely elusive or speculative [5–8]. This is partly because of incomplete geographical and genomic sampling of riverine species within and surrounding lake basins. However, it is also due to intrinsic complexity of cichlid evolutionary relationships, as radiations may have been seeded by multiple riverine ancestors [8,9], and rivers can be recolonized by species with lacustrine ancestry [10]. A greater understanding of geographical and phylogenetic ancestry of cichlids is required to test whether functional genetic variation under divergent selection within lake radiations is present within riverine ancestors, and whether this variation has been shared among riverine cichlids through intraspecific gene flow and interspecific hybridization [9].



**Figure 1.** (a) Lake Malawi and surrounding major river systems; (b) Bayesian phylogeny based on 544 mtDNA NADH2 sequences. Numbers above branches indicate posterior probabilities (black, values greater than 0.7 shown) and maximum-likelihood bootstrap support (blue, values greater than 70% shown). (Online version in colour.)

Lake Malawi contains a radiating flock of at least 450 haplochromine species [11]. Early phylogenetic reconstructions suggested that the lake radiation was monophyletic [12,13]. More recent phylogenies show two species outside the Lake Malawi catchment also fall within the flock, namely *Astatotilapia calliptera* and *Astatotilapia swynnertoni* [5,8]. There have been indications that these are sister lineages to the radiating flock [5,8], but the hypothesis has not been well supported by either nuclear or mitochondrial DNA [5,8,14,15]. There is evidence that riverine representatives of the flock outside the catchment have seeded some lacustrine diversity in the radiation [8,16], but preceding this they may have escaped from Lake Malawi into neighbouring drainages. Given such uncertainty, and evidence of recent gene flow across catchment boundaries in *A. calliptera* [16], there is a need to further resolve relationships of Malawi endemics to cichlids in neighbouring drainages.

To date, phylogenetic reconstructions have included haplochromines from many of the surrounding catchments (figure 1), including the Zambezi, Lake Chilwa, Ruvuma, Congo and Lake Rukwa catchments [5,8,13,17]. However, no published phylogenies have included haplochromines from the Great Ruaha river [18]. Here, we show phylogenetic reconstructions including an undescribed taxon *Astatotilapia*

sp. 'Ruaha' from this region which place it as a sister taxon to the Lake Malawi flock.

## 2. Material and methods

Genetic samples (fin clips) were collected from riverine haplochromines (electronic supplementary material, table S1; figure 1) and preserved in 95% ethanol. DNA was isolated using the Promega Wizard kit. Sequences of the mitochondrial gene NADH2 [7] were generated and aligned with sequences of other haplochromines and outgroup taxa, using CLUSTALW in DAMBE [19]. This resulted in an alignment of 1047 bp with 544 sequences (electronic supplementary material, table S2). Bayesian phylogenies were generated in MRBAYES v. 3.2.4 [20], using Partitionfinder [21] models, and two runs of 10 million generations. Resultant trees were combined after removal of 50% as burn-in. Maximum-likelihood phylogenetic analysis were conducted in RAXML [22], Partitionfinder models and 100 bootstrap replicates. We dated divergence times using a subset of 40 NADH2 sequences (electronic supplementary material, table S2). Time-calibrated trees were generated with BEAST v. 1.8.0 [23] using Partitionfinder models and two sets of calibrations [24,25] employed independently (electronic supplementary material, table S3). Random local clocks were used in runs of 50 million generations, with 20% of trees removed as burn-in. Consensus trees from

### 3. Results and discussion

An as yet undescribed representative of the 'modern haplochromine' group, *Astatotilapia* sp. 'Ruaha', was present at three Great Ruaha sites (figure 1). On the basis of mitochondrial NADH2 DNA sequences, the species was resolved as an immediate sister taxon to the radiating flock (figure 1; electronic supplementary material, figure S1). The results of the analyses suggest they diverged between 2.13 Ma (95% highest posterior density (HPD) 1.52–2.84 Ma; using non-cichlid fossil derived calibrations from Friedman *et al.* [24]) and 6.76 Ma (95% HPD 3.76–10.12 Ma; using non-cichlid fossil derived calibrations from Schwarzer *et al.* [25]). This divergence took place before initial divergence of extant representatives of the Lake Malawi flock estimated at 1.2 Ma (95% HPD 1.52–2.84 Ma) or 4.06 Ma (95% HPD 2.02–6.59 Ma), from Friedman *et al.* [24] and Schwarzer *et al.* [25] calibrations, respectively. The *Astatotilapia* sp. 'Ruaha' lineage is geographically separated from the Malawi catchment by the Livingstone/Kipengere mountain range. This comprises steep mountainous areas and high altitude plateau, and it is plausible that both geography and low temperatures impose barriers to habitat occupancy and dispersal across the boundary [16]. The range was formed during Pliocene rifting that initiated formation of Lake Malawi [26], perhaps driving simultaneous population division and ecological opportunity for species flock formation.

Close evolutionary relationships between Malawi and upper Ruaha haplochromines are mirrored by recent observations from fish fossils of fluvial deposits of the Chiwondo beds dated to between 2 and 3.75 Ma [18]. The Chiwondo fauna includes claroteid catfishes and tigerfish (*Hydrocynus*) [18], but geographically the nearest system containing extant representatives of these non-cichlid families is the Ruaha. It has been proposed on the basis of these fossils that rivers currently in Lake Malawi catchment were once extensions of the Great Ruaha system in pre-rift times [18]. Our results are compatible with this concept and imply further molecular studies may identify this region as a source of genetic diversity of other elements of the Malawi fauna. Notably, although the Chiwondo fauna includes representatives of Cichlidae, it has not been possible to identify remains to a lower taxonomic level [18].

It has been proposed that the ancestor of the Lake Malawi haplochromine flock is a riverine haplochromine similar to *Astatotilapia bloyeti* or *A. calliptera* [27]. Our study places specimens assigned to *A. bloyeti* in a sister clade to *Astatotilapia tveddlei*, consistent with previous analyses of both nuclear and mitochondrial markers [8], and our results suggest both taxa are more distantly related to Malawi cichlids than

*Astatotilapia* sp. 'Ruaha'. Our results also show that *A. calliptera* outside the Lake Malawi catchment are part of a geographically broader 'Lake Malawi region' flock. It remains equivocal whether the species secondarily colonized external rivers from Lake Malawi, or instead whether there have been multiple colonizations of *A. calliptera* from outside the catchment along with maintenance of the ancestral riverine phenotype [5,8,14]. In either case, given mitochondrial DNA evidence suggesting that *Astatotilapia* sp. 'Ruaha' is a sister species to the flock, and fossil evidence of historic connectivity of the Ruaha and Lake Malawi, it seems plausible that extant representatives of the Malawi flock are biogeographically derived from a species with a former distribution that encompassed both the Ruaha and Lake Malawi catchments. Further phylogenetic analyses based on nuclear genome data will help to provide further resolution of the relationship between *Astatotilapia* sp. 'Ruaha' and Malawi cichlids. Genome-wide data will also help to resolve whether *A. calliptera* occupy a basal, sister or derived position in the flock, which may force reconsideration of the biogeographic scenario suggested here.

Recent results show a high proportion of genomic diversity present within Lake Malawi cichlids is also present in riverine cichlids [9]. It has been proposed that riverine species may be active transporters of genomic material enabling rapid adaptation within lacustrine flocks. However, such situations require introgression among riverine taxa at contact zones, and gene flow across catchment boundaries. There is support for the concept of intraspecific gene flow across watersheds within Africa [16], but currently only indirect evidence of interspecific hybridization among river cichlids [27], and there is no evidence of interspecific hybridization among riverine haplochromines in the region surrounding Lake Malawi. A greater understanding of taxonomic and spatial patterns of genetic diversity within and among potentially ancestral riverine cichlids is required, including *Astatotilapia* sp. 'Ruaha'. This would enable tests of the importance of active transport of genes through riverine species and hybridization events for explaining shared genomic diversity among lacustrine radiations [9].

**Ethics.** Tanzania Commission for Research and Technology (COSTECH) issued a permit for this study (no. 2011-205-NA-2011-103).

**Data accessibility.** DNA sequences can be accessed from Genbank (KR010448–KR010461).

**Authors' contributions.** M.J.G., B.P.N. and G.F.T. conceived the study and wrote the manuscript. M.J.G., B.P.N., S.M., A.S. and M.J.G. collected field samples and data. M.J.G. generated and analysed sequence data. All authors critically revised and approved the manuscript.

**Competing interests.** We declare we have no competing interests.

**Funding.** This study was financially supported by the Royal Society-Leverhulme Trust Africa Award AA100023.

**Acknowledgements.** We thank TAFIRI staff and J. Swanstrom for field assistance.

### References

1. Sato A, Tichy H, O'hUigin C, Grant PR, Grant BR, Klein J. 2001 On the origin of Darwin's finches. *Mol. Biol. Evol.* **18**, 299–311. (doi:10.1093/oxfordjournals.molbev.a003806)
2. Barrier M, Baldwin BG, Robichaux RH, Purugganan MD. 1999 Interspecific hybrid ancestry of a plant adaptive radiation: allopolyploidy of the Hawaiian silversword alliance (Asteraceae) inferred from floral homeotic gene duplications. *Mol. Biol. Evol.* **16**, 1105–1113. (doi:10.1093/oxfordjournals.molbev.a026200)
3. Taylor EB, McPhail JD. 1999 Evolutionary history of an adaptive radiation in species pairs of threespine sticklebacks (*Gasterosteus*): insights from mitochondrial DNA. *Biol. J. Linn. Soc.* **3**, 271–291. (doi:10.1111/j.1095-8312.1999.tb01891.x)
4. Feulner PG *et al.* 2013 Genome-wide patterns of standing genetic variation in a marine population of three-spined sticklebacks. *Mol. Ecol.* **22**, 635–649. (doi:10.1111/j.1365-294X.2012.05680.x)

5. Seehausen O *et al.* 2003 Nuclear markers reveal unexpected genetic variation and a Congolese–Nilotic origin of the Lake Victoria cichlid species flock. *Proc. R. Soc. Lond. B* **270**, 129–137. (doi:10.1098/rspb.2002.2153)
6. Verheyen E, Salzburger W, Snoeks J, Meyer A. 2003 Origin of the superflock of cichlid fishes from Lake Victoria, East Africa. *Science* **300**, 325–329. (doi:10.1126/science.1080699)
7. Salzburger W, Mack T, Verheyen E, Meyer A. 2005 Out of Tanganyika: genesis, explosive speciation, key-innovations and phylogeography of the haplochromine cichlid fishes. *BMC Evol. Biol.* **5**, 17. (doi:10.1186/1471-2148-5-17)
8. Joyce DA, Lunt DH, Genner MJ, Turner GF, Bills R, Seehausen O. 2011 Repeated colonization and hybridization characterize the Lake Malawi cichlid fish species flock. *Curr. Biol.* **21**, R108–R109. (doi:10.1016/j.cub.2010.11.029)
9. Loh YH *et al.* 2013 Origins of shared genetic variation in African cichlids. *Mol. Biol. Evol.* **30**, 906–917. (doi:10.1093/molbev/mss326)
10. Sturmbauer C, Salzburger W, Duftner N, Schelly R, Koblmüller S. 2010 Evolutionary history of the Lake Tanganyika cichlid tribe Lamprologini (Teleostei: Perciformes) derived from mitochondrial and nuclear DNA data. *Mol. Phylogenet. Evol.* **57**, 266–284. (doi:10.1016/j.ympev.2010.06.018)
11. Genner MJ, Seehausen O, Cleary DFR, Knight ME, Michel E, Turner GF. 2004 How does the taxonomic status of allopatric populations influence species richness within African cichlid fish assemblages? *J. Biogeogr.* **31**, 93–102. (doi:10.1046/j.0305-0270.2003.00986.x)
12. Meyer A, Kocher TD, Basasibwaki P, Wilson AC. 1990 Monophyletic origin of Lake Victoria cichlid fishes suggested by mitochondrial DNA sequences. *Nature* **347**, 550–553. (doi:10.1038/347550a0)
13. Nagl S, Tichy H, Mayer WE, Takezaki N, Takahata N, Klein J. 2000 The origin and age of haplochromine fishes in Lake Victoria, east Africa. *Proc. R. Soc. Lond. B* **267**, 1049–1061. (doi:10.1098/rspb.2000.1109)
14. Genner MJ, Turner GF. 2012 Ancient hybridization and phenotypic novelty within Lake Malawi's cichlid fish radiation. *Mol. Biol. Evol.* **29**, 195–206. (doi:10.1093/molbev/msr183)
15. Hulsey CD, Keck BP, Alamillo H, O'Meara BC. 2013 Mitochondrial genome primers for Lake Malawi cichlids. *Mol. Ecol. Res.* **13**, 347–353. (doi:10.1111/1755-0998.12066)
16. Nichols P, Genner MJ, Van Oosterhout C, Smith A, Swanstrom J, Parsons P, Sungani H, Joyce DA. 2015 Secondary contact seeds phenotypic novelty in cichlid fishes. *Proc. R. Soc. B* **282**, 20142272. (doi:10.1098/rspb.2014.2272)
17. Joyce DA, Lunt DH, Bills R, Turner GF, Katongo C, Duftner N, Sturmbauer C, Seehausen O. 2005 An extant cichlid fish radiation emerged in an extinct Pleistocene lake. *Nature* **435**, 90–95. (doi:10.1038/nature03489)
18. Stewart KM, Murray AM. 2013 Earliest fish remains from the Lake Malawi Basin, Malawi, and biogeographical implications. *J. Vert. Paleontol.* **33**, 532–539. (doi:10.1080/02724634.2013.741086)
19. Xia X. 2013 DAMBES: a comprehensive software package for data analysis in molecular biology and evolution. *Mol. Biol. Evol.* **30**, 1720–1728. (doi:10.1093/molbev/mst064)
20. Ronquist F, Huelsenbeck JP. 2003 MrBAYES 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* **19**, 1572–1574. (doi:10.1093/bioinformatics/btg180)
21. Lanfear R, Calcott B, Ho SYH, Guindon S. 2012 PartitionFinder: combined selection of partitioning schemes and substitution models for phylogenetic analyses. *Mol. Biol. Evol.* **29**, 1695–1701. (doi:10.1093/molbev/mss020)
22. Stamatakis A. 2006 RAxML-VI-HPC: maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. *Bioinformatics* **22**, 2688–2690. (doi:10.1093/bioinformatics/btl446)
23. Drummond AJ, Suchard MA, Xie D, Rambaut A. 2012 Bayesian phylogenetics with BEAUti and the BEAST 1.7. *Mol. Biol. Evol.* **29**, 1969–1973. (doi:10.1093/molbev/mss075)
24. Friedman M, Keck BP, Dornburg A, Eytan RI, Martin CH, Hulsey CD, Wainwright PC, Near TJ. 2013 Molecular and fossil evidence place the origin of cichlid fishes long after Gondwanan rifting. *Proc. R. Soc. B* **280**, 20131733. (doi:10.1098/rspb.2013.1733)
25. Schwarzer J, Misof B, Tautz D, Schlieven UK. 2009 The root of the East African cichlid radiations. *BMC Evol. Biol.* **9**, 186. (doi:10.1186/1471-2148-9-186)
26. Danley PD, Husemann M, Ding B, Dipietro LM, Beverly EJ, Peppe DJ. 2012 The impact of the geologic history and paleoclimate on the diversification of East African cichlids. *Int. J. Evol. Biol.* **2012**, 574851. (doi:10.1155/2012/574851)
27. Schwarzer J, Swartz ER, Vreven E, Snoeks J, Cotterill FP, Misof B, Schlieven UK. 2012 Repeated trans-watershed hybridization among haplochromine cichlids (Cichlidae) was triggered by Neogene landscape evolution. *Proc. R. Soc. B* **279**, 4389–4398. (doi:10.1098/rspb.2012.1667)