



Review

Cite this article: Bashey F. 2015 Within-host competitive interactions as a mechanism for the maintenance of parasite diversity. *Phil. Trans. R. Soc. B* **370**: 20140301.
<http://dx.doi.org/10.1098/rstb.2014.0301>

Accepted: 14 May 2015

One contribution of 17 to a theme issue 'Within-host dynamics of infection: from ecological insights to evolutionary predictions'.

Subject Areas:

ecology, evolution, health and disease and epidemiology

Keywords:

within-host competition, parasite diversity, bacteriocins, coexistence, public goods, $G \times G$ interactions

Author for correspondence:

Farrah Bashey
e-mail: fbasheyv@indiana.edu

Within-host competitive interactions as a mechanism for the maintenance of parasite diversity

Farrah Bashey

Department of Biology, Indiana University, 1001 East Third Street, Bloomington, IN 47405, USA

Variation among parasite strains can affect the progression of disease or the effectiveness of treatment. What maintains parasite diversity? Here I argue that competition among parasites within the host is a major cause of variation among parasites. The competitive environment within the host can vary depending on the parasite genotypes present. For example, parasite strategies that target specific competitors, such as bacteriocins, are dependent on the presence and susceptibility of those competitors for success. Accordingly, which parasite traits are favoured by within-host selection can vary from host to host. Given the fluctuating fitness landscape across hosts, genotype by genotype ($G \times G$) interactions among parasites should be prevalent. Moreover, selection should vary in a frequency-dependent manner, as attacking genotypes select for resistance and genotypes producing public goods select for cheaters. I review competitive coexistence theory with regard to parasites and highlight a few key examples where within-host competition promotes diversity. Finally, I discuss how within-host competition affects host health and our ability to successfully treat infectious diseases.

1. Introduction

The maintenance of diversity, be it at the species or genotypic level, is a fundamental problem in biology. The root of the issue is that, in a given environment, the fittest type should competitively exclude all others. How diversity is maintained, therefore, has been the focus of a rich body of empirical and theoretical work in ecology and evolutionary biology. Here, I examine how these works inform our understanding of parasite diversity. I start by describing the complex selective environment faced by parasites, focusing on the varied forms of competitive interaction that can occur within a single host individual. Then, I review theoretical treatments of competition and diversity to gain insights into how diversity can be maintained in the face of competition. I then highlight a few key systems where within-host competitive interactions are thought to be crucial in maintaining parasite diversity. Finally, I conclude by discussing the implications of parasite diversity and within-host competitive interactions for host health and the treatment of infectious diseases.

Parasite fitness is dependent on both within- and among-host selection [1]. Within the host, competition with the host's native microbiota or other co-infecting parasites is a key determinant of parasite fitness and successful transmission to new hosts. These competitive interactions can be categorized using classic terms from the fields of ecology and social behaviour [2–4]. Most fundamentally, genotypes or species consuming the same resources compete via exploitative competition (figure 1*a*). Within the host, pathogen traits that enable faster use of host resources can allow a pathogen to outgrow others within the host and to be numerically dominant upon transmission (e.g. [5]). These traits can be exclusive to the pathogen, such as a greater number of nutrient receptors or increased replication machinery (e.g. [6,7]). Alternatively, faster growth rate can be achieved by the release of compounds that enable degradation of host tissues or sequestration of nutrients [8,9]. If these released products can benefit other members of the community, they can be viewed as public goods, and the

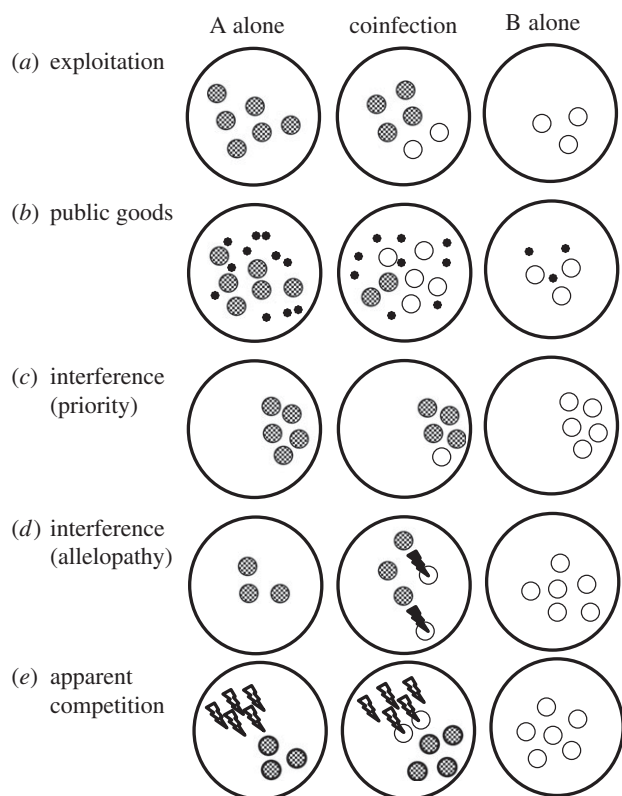


Figure 1. Mechanisms of within-host competition. Each large circle represents a host individual either infected with parasite A (smaller grey circles) alone, parasite B (smaller white circles) alone or simultaneously infected with both parasites. The relative growth rate of each parasite is indicated by the number of parasites present within each host individual. (a) When parasites compete through faster or more efficient exploitation of host resources, within-host selection favours parasites with superior exploitation (here, parasite A). (b) If parasites achieve greater host exploitation through the release of public goods (smallest black circles), then within-host selection favours parasites that produce less of the public good (here, parasite B). (c) When parasites compete over access to host infection sites, the parasite that occupies a site first (here, parasite A) can interfere with the attachment of later arriving parasites. (d) When parasites compete through the production of allelopathic agents (black lightning bolts), within-host selection favours parasites that produce costly competitive toxins (here, parasite A). (e) When parasites compete via the host immune systems, parasites who benefit when infecting alone by being resistant to the host immune response they elicit (white lightning bolts) may also benefit via within-host selection by eliminating immune-sensitive competitors.

interactions between the producers can be viewed as cooperation [4,10]. As these public goods increase access to host resources, producing genotypes will be able to exploit hosts better than genotypes with lower levels of production (figure 1*b*). However, when within the same host, non-cooperating cheaters that avoid cost of production may have a competitive advantage.

Competition over limited resources can also select for strategies that block or attack potential competitors (i.e. interference competition) [11]. Multiple types of interference competition have been recognized among parasites. For example, in a study on acanthocephalan parasites of fish [12], the number of parasites that can establish within an individual host is limited by the amount of space left unoccupied by pre-established parasites (figure 1*c*). Similarly, microbes in a biofilm can pre-empt the attachment of other cells, or alternatively, overgrow established layers, depriving them access to

nutrients [13]. More aggressive forms of interference include contact-dependent killing [14] and allelopathy (i.e. the release of growth-inhibiting compounds). Examples of allelopathy abound in the microbial world, as numerous antibiotics and other secondary metabolites have been found to have inhibitory effects [15]. A key class of allelopathic compounds, bacteriocins, target even closely related strains of the same species and are likely a key mechanism maintaining within-species diversity [16,17]. Critically, allelopathy is only beneficial in the presence of a susceptible competitor (figure 1*d*). In the absence of a suitable target, the costs of production (or even the carriage of production machinery) may lower the fitness of aggressive mechanisms of interference.

Finally, competition between two parasites within a host can be mediated by the host immune system [18,19]. In ecology, this is termed apparent competition, because the negative effect that two species have on each other (i.e. competition) is not due to shared resource use, but due to a shared predator [20]. Under classic apparent competition, increasing density of one prey species promotes population growth of a shared predator, thereby increasing predation on a second prey species. In the case of parasites, the presence of one parasite can stimulate the host immune system. If the immune response is non-specific, it may cross-react with other co-infecting parasite strains or species [2]. This immune-system mediated competition thereby favours pathogens that can escape the immune response by hiding from or being immune to its effects. In fact, the ability to withstand a severe inflammatory host response can favour the emergence of a parasite that provokes the immune response and subsequently benefits from the clearing of less resistant competitors (figure 1*e*) [21].

How do the complex competitive interactions within the host balance with among-host selection? Empirical work has demonstrated that each of the mechanisms described above can alter parasite within-host reproduction and transmission success. However, how different mechanisms of within-host competition affect each other and the full extent to which host epidemiology influences parasite evolution is still an open question [22]. Nevertheless, it is clear that the complex competitive environment within the host can favour different strategies in different host individuals: the competitive strategy that is successful in one host may depend on the presence of a specific competitor, and may be costly in the next host due to absence of that competitor. One result of this complex selective landscape is that parasites can evolve adaptive plastic responses to deal with the varied competitive environments encountered within hosts [23,24]. Host populations can, however, support many parasite species, and considerable genetic diversity within a given parasite species. So how is this diversity maintained given that competition often leads to exclusion of one competitor by another? I next review our understanding of competitive coexistence in relation to parasites.

2. Effects of competition on parasite diversity

Competitive coexistence requires that each species limits its own growth more than it limits the growth of another species [25]. Ecologically, this can be achieved by three main mechanisms: niche partitioning, competition–colonization trade-offs and heterogeneity in the competitive environment. Niche partitioning is the best understood mechanism of coexistence. Niches can be described in numerous dimensions involving

essential resources, abiotic tolerances and susceptibility to natural enemies. If specialization in a given niche dimension is costly, then one species will become competitively dominant in each dimension, and species diversity will be limited by the number of niche dimensions. Thus over evolutionary time, competition is expected to lead to character displacement among species, lessening competition between species and thereby stabilizing the community [26]. Accordingly, one predicted outcome of within-host competition among parasites is specialization of parasites to different host tissues or different host species [27]. Within a host species, niche partitioning can occur due to specific immunity [28]. A successful strain elicits an immune response thereby limiting its own success, while antigenically different strains can still invade. Not surprisingly, fewer strains can be maintained if there are large intrinsic fitness differences among strains or if the immune response is cross-reactive (i.e. resulting in apparent competition). However, relatively simple models of strain competitive ability and immune response can explain the incredible antigenic variation seen within some human pathogens [28,29].

In the absence of niche partitioning, coexistence can occur across homogeneous patches via a competition–colonization trade-off [30,31]. Thus, even if one species is dominant within a patch, the other can persist by reaching more unoccupied patches. For parasites, host individuals form discrete patches, and a strain that is inferior in within-host competition may be better able to disperse to competitor-free hosts. One remarkable example of this trade-off is the mitigation of a competitive asymmetry between two species of pigeon lice by the unique ability of the competitively inferior species to disperse to new hosts by attaching to a fly [32]. Similarly, although the mechanism is not clear, HIV strains favoured by within-host selection are at a disadvantage in transmission to new hosts [33]. Additionally, the competition–colonization trade-off can arise from a trade-off between within-host competition and persistence outside the host [34]. For example, a parasite that is better at free-living survival may be slower to recover or grow within the host, and thus competitively inferior. In support of this idea, a comparative study of the phages of *Escherichia coli* found that slower growing phages were more persistent in the environment, suggesting a fundamental trade-off between these properties [35]. Further supporting a trade-off, experimental evolution of an arbovirus found that higher extracellular survival was associated with reduced fecundity [36], although such trade-offs are not necessarily universal [37] and should be evaluated over both ecological and evolutionary time scales.

A final category for coexistence requires heterogeneity in the competitive environment [25,38]. A corollary to the maxim that species coexistence requires each species to limit its own growth more than it limits the growth of another species, is that species can coexist if each species can increase when rare [39]. Even in the absence of local niche partitioning, negative-frequency dependence can occur if species experience competition on different spatial scales [40] or if species differ in how they respond competitively to spatial and temporal variation in the environment (e.g. through a storage effect [41]). Empirical demonstrations of these mechanisms are less well established than niche partitioning or competition–colonization trade-offs; nevertheless, they suggest that variation in how species experience competition is an important mechanism of coexistence [42,43]. For parasites, transient changes in the environment or among hosts can

result in different pathogen genotypes being successful at different times or in different places. If this variation dampens otherwise successful strains and favours strains that were struggling, then it may promote coexistence. Such equalizing effects are inherent in among-host selection and can be more forceful when epidemiological feedbacks occur. Additionally, within-host selection is expected to vary depending on the composition of the within-host community. For example, parasite traits that increase the speed of host exploitation will be favoured within the host, but parasites with these traits may be susceptible to invasion by parasites that can attack them.

The interplay of strategies favouring faster growth and strategies depending on attack has been modelled extensively with regard to bacteriocins. Strains that produce bacteriocins are able to attack sensitive strains, but they pay a cost of production. Thus, producers cannot displace sensitive strains unless they are sufficiently numerous and toxic, or unless the competitive interactions are local [44,45]. In order for both producer and sensitive strains to coexist stably, there must be underlying spatial heterogeneity in resource availability [46] or stochastic environmental disturbances that hit some patches but not others [47]. This spatial heterogeneity allows sensitive strains to gain an advantage in either low resource or empty patches. Notably, when more than two strains are examined, coexistence can occur in the absence of spatial heterogeneity. Resistant strains can evolve either from sensitive strains [48] or represent non-producing cheaters [45]. If resistant strains have higher growth rates than producers (because they are not paying the cost of production) and lower growth rates than sensitives (because they are paying a cost of resistance), then resistant strains can out-compete producers, but are out-competed by sensitives in a rock–paper–scissors-like dynamic (RPS). As each strain can be invaded by another, these non-transitive interactions can maintain diversity through negative-frequency dependence. Theoretical, as well as experimental, evaluations of this dynamic show that stability is contingent upon spatial structure facilitating local interactions [49]. Furthermore, a study using *E. coli* strains in mice found that strains could be transmitted between hosts and replaced each other as predicted by the RPS model, although coexistence was not achieved in the relatively small communities of the experimental set-up [50].

How do non-transitive ecological dynamics play out on an evolutionary timescale? Intriguingly, theory has suggested that spatial structure coupled with non-transitive pairwise interactions favours the evolution of competitive restraint [51,52]. This result can be understood intuitively as more rapacious competitors more effectively eliminate their subordinate partner (be it Resistant > Producer, $P > S$ or $S > R$), placing them more frequently in contact with a partner that can out-compete them. For example, if a more rapacious form of the resistant strain arises, say by mutations that lower the cost of resistance, it will more effectively out-compete the producer strain. However, the resistant strain will then find itself more likely to encounter sensitive strains which limit the spread of the rapacious resistant strain. Thus, restrained behaviour towards ‘the enemy of my enemy’ is beneficial to a strain subject to community-level feedbacks. Such feedbacks favour community coexistence [53]. The importance of spatial structure and non-transitive interactions on the evolution of competitive restraint has been experimentally demonstrated *in vitro* with *E. coli* [54]. Furthermore, ecological feedbacks on community stability extend beyond the three-player game

[53]. Allowing mutation and recombination leads to the prediction that a diverse number of strains can be maintained, with each strain producing one or more allelopathic agents [55,56]. This result is consistent with the diversity of bacteriocins carried among hosts in a natural population of *E. coli* [57].

In summary, although competition often leads to competitive exclusion, there are numerous mechanisms whereby coexistence can occur. Importantly, competition is not a transient effect that quickly becomes minimized due to character displacement. The patchy structure inherent in the parasite lifestyle sets the stage for heterogeneity across hosts to favour coexistence and for competition to act as a continual selective force on parasite traits. The interplay between ecological and evolutionary forces in maintaining parasite diversity is already well recognized with respect to host genetic diversity [58–60]. Analogously, competitive interactions within the host can shape the evolution of parasite phenotypes and can even facilitate the coexistence of multiple parasite types.

3. Key examples linking within-host competitive interactions and parasite diversity

Several parasite systems suggest that competitive interactions within the host are important for the maintenance of parasitic diversity. Here I will focus on three systems where we have a strong understanding of the diversifying effect of within-host competition. *Diplostomum* trematodes have a complex life cycle involving two intermediate hosts (snails and fish) and a definitive bird host. When a snail host is co-infected with multiple genotypes of *D. pseudospathaceum*, transmission success of each genotype is lower relative to a single genotype infection. However, when a fish host is exposed to multiple parasite genotypes simultaneously, the probability of parasite establishment is higher relative to single genotype exposures [61]. Thus, exploitative competition in the snail host reduces diversity, while interactions with the immune system of the fish host can maintain diversity. Exactly how parasite diversity interacts with the host immunity in this system is still under investigation, but there is evidence for cross-reactivity in adaptive immunity as well as specificity in innate immunity, a pattern contrary to the standard paradigm [62]. Furthermore in cross-species infections, some strains of *D. pseudospathaceum* perform better in mixed exposures than when infecting a fish alone, while others do worse [63,64]. Therefore, predicting the outcome of competition among species is dependent on knowing the specific parasite genotypes present within the host: a strain that is disfavoured in one host individual may have a fitness advantage in another due to differences in the parasite community.

Similar genotypic effects are seen in competition among species of entomopathogenic nematodes. These insect-killing nematodes (genus *Steinernema*) are involved in a mutualistic symbiosis with bacteria in the genus *Xenorhabdus*. *Xenorhabdus* bacteria are known to produce a suite of compounds that attack the insect immune system and prevent the insect from being colonized by fungal and other bacterial competitors. Additionally, *Xenorhabdus* produce bacteriocins that are effective at inhibiting the growth of *Xenorhabdus* genotypes distinct from the producing genotype. A diversity of bacteriocin phenotypes have been isolated from a small geographical area [65–67] and distributed in a pattern which suggests that within-insect interactions are important for maintaining this

diversity. Specifically, one bacteriocin phenotype was found to dominate within an insect host, yet on a larger scale, consistent with nematode movement, multiple bacteriocin phenotypes could be detected. Additionally, laboratory-based co-infections have demonstrated a fitness benefit to the inhibiting phenotype that is conditional on the presence of a sensitive competitive partner within the insect host [68]. In the absence of a sensitive competitor, exploitative competition seems paramount: parasites that establish an infection faster are competitively dominant [69]. Thus, the competitive context determines which traits selection favours. Furthermore, transmission in this system requires a free-living stage. Species differ in longevity in this stage, which can influence their relative competitive success (F. Bashey 2011, unpublished data). Conditions within the host can also influence the size of the free-living stage, which influences the probability of successful establishment in a new host [70]. Hence, diverse within-host competitive environments can select for diverse parasite genotypes due to both within- and among-host fitness components.

Perhaps the best understood example of within-host competition from a mechanistic basis comes from work on the opportunistic pathogen *Pseudomonas aeruginosa*. In response to low levels of iron in the within-host environment, this species releases pyoverdinin, its primary siderophore or iron scavenging molecule [71]. Pyoverdinin constitutes a public good: non-producing cells can bind to this molecule and thereby gain iron. In the absence of competition, increased pyoverdinin production increases population growth; however, in the presence of a competitor that produces less pyoverdinin, fitness is reduced due to the higher cost of pyoverdinin production [72]. Thus, selection on this trait depends upon the social context. Strains also compete through interference competition. *Pseudomonas* produces several types of bacteriocins [73] as well as possessing the ability to attack neighbouring cells in a contact-dependent manner [74]. The fitness benefit of bacteriocin production has also been shown to be contingent upon the nature of the competitive environment, depending not just on the presence of susceptible competitors, but also on the local density of kin [75]. Moreover, selection on bacteriocin production shifts depending on the level of pyoverdinin production of the susceptible strain [76], demonstrating further the context dependence of within-host fitness. Similarly, contact-dependent killing has been found to be responsive to the behaviour of competitors in a tit-for-tat fashion [77]. Although most of this work has been elucidated in an *in vitro* setting, multiple competitive phenotypes can be isolated from a host population [78] or a host individual [79]. Moreover, receptors for these molecules demonstrate high levels of diversifying selection [80].

4. Implications of within-host competition to host health

Parasite growth within the host is a major cause of the negative effects that parasites have on host health [81,82]. As outlined above (figure 1), different mechanisms of within-host competition select for different parasite traits, some of which may increase or decrease within-host growth. Accordingly, a high prevalence of multiple infections is predicted to have varied effects on host health [22,83]. The earliest theoretical treatment of the effect of multiple infections on the evolution of virulence assumed that pathogens compete within the host

via exploitative competition. Thus, they predicted that due to within-host selection for increased growth, pathogen virulence should evolve to be higher in mixed infections relative to single genotype infections [84,85]. While this prediction has been upheld in some systems [5], other systems are dominated by different forms of within-host interactions. When public goods allow greater host exploitation [8,86] or virulent immuno-provoking forms of apparent competition [87], mixed infections are predicted to result in lower virulence due to within-host selection favouring cheats. However, incorporating epidemiological feedbacks can change these predictions [88,89]. Similarly, spiteful forms of interference competition have been shown to lower virulence [90], but they can raise virulence depending on the scale of competition and the kin structure within the host [91]. As models more explicitly incorporate multiple forms of interactions within the host into an epidemiological framework [92], we will further improve our ability to connect knowledge of the competitive environment to host health outcomes.

Already, understanding within-host competition has helped to explain strains with puzzling phenotypes. For example, highly virulent forms of *Streptococcus pneumoniae* are not transmitted from the host once they invade the body, yet they persist because they can withstand apparent competition that occurs in the nasal cavity [93]. Additionally, coupling selection in the within-host environment with selection occurring outside the host can provide a more accurate understanding of how variation in virulence is maintained in nature [94,95]. Within-host competition has also explained why some forms of methicillin-resistant *Staphylococcus aureus* (MRSA) do not respond to treatment with vancomycin [96]. Unlike other cases where antibiotic resistance occurs after antibiotic treatment or due to high levels of resistance circulating in the community, some forms of vancomycin resistance in MRSA appear to occur due to within-host competition. Specifically, resistant variants were found to arise spontaneously within hosts due to competition favouring a bacteriocin-producing form that could attack the wild-type, followed by the evolution of a bacteriocin-resistant strain. Traits conferring bacteriocin resistance also provided partial resistance to vancomycin. These examples illustrate that within-host selection pressures are profound and of significant clinical relevance.

While clinicians have mainly noted when the presence of one pathogen species complicates treatment of another [97], the presence of a competitor can improve the outcome for the host when interference or apparent competition slows disease progression [90,98,99]. In fact, the health benefits of many probiotics are thought to be due in part to their

effectiveness in interference competition [100]. Interest in using bacteriocins as an alternative to traditional, broad-spectrum antibiotics is growing [101–103]. Historically, the narrow killing range of bacteriocins limited their utility; however, faster diagnostic tests, the ability to engineer bacteriocins to target novel sites [104], and our awareness of the consequences of using broad-spectrum antibiotics has made the therapeutic use of bacteriocins more viable.

As within-host competition alters selection on parasite traits and increases parasite diversity, it is further important to realize that variation in disease outcomes could be due as much to the within-host parasite community as due to host genotypic effects. Indeed, some cases that we have viewed as parasite interactions with the host genotype (i.e. $G_{\text{parasite}} \times G_{\text{host}}$) may in fact be due to differences in the within-host competitive environment [105]. Critically, variation in disease that is due to $G_{\text{parasite}} \times G_{\text{parasite}}$ may be more amenable to intervention than variation caused by host genotypic differences. Capitalizing on within-host selection is a major tenet of Hamiltonian medicine [106] and understanding how parasite traits influence parasite fitness across spatial scales is crucial to predicting when novel therapies, such as targeting virulence factors, will be successful [107,108].

In summary, the lifestyle of pathogens where selection occurs both within the host and in transmission between hosts creates opportunities for selection to maintain diverse pathogen phenotypes. This occurs in two main ways. First, the rich diversity of competitive strategies that parasites employ within the host can engender reciprocal selection between parasites. Parasites more successful at exploiting the host than competitors can be invaded by attacking or freeloading strategies, which then select for resistance and a restarting of this cycle. Second, heterogeneity among hosts in the composition of the within-host competitive environment itself can favour different parasite traits across hosts. As the relative costs and benefits of different forms of competition depend on the identity of the competitors, selection due to competition can be continual and within-host competition should be viewed as a potentially major driver of parasite diversity. Better understanding this driving force on parasite traits will enable us to more effectively treat infectious diseases.

Competing interests. I declare I have no competing interests.

Funding. This study was supported by NSF DEB 0919015.

Acknowledgements. I thank C. Lively and the various iterations of his laboratory group for thought-provoking discussion on the maintenance of diversity. Additionally, I thank A. Gibson, A. Bhattacharya, O. Restif and two anonymous reviewers for their insightful comments on previous versions of this manuscript.

References

- Ewald PW. 1983 Host-parasite relations, vectors, and the evolution of disease severity. *Ann. Rev. Ecol. Syst.* **14**, 465–485. (doi:10.1146/annurev.es.14.110183.002341)
- Mideo N. 2009 Parasite adaptations to within-host competition. *Trends Parasitol.* **25**, 261–268. (doi:10.1016/j.pt.2009.03.001)
- Brown SP, Inglis RF, Taddei F. 2009 Evolutionary ecology of microbial wars: within-host competition and (incidental) virulence. *Evol. Appl.* **2**, 32–39. (doi:10.1111/j.1752-4571.2008.00059.x)
- West SA, Griffin AS, Gardner A, Diggle SP. 2006 Social evolution theory for microorganisms. *Nat. Rev. Microbiol.* **4**, 597–607. (doi:10.1038/nrmicro1461)
- de Roode JC *et al.* 2005 Virulence and competitive ability in genetically diverse malaria infections. *Proc. Natl Acad. Sci. USA* **102**, 7624–7628. (doi:10.1073/pnas.0500078102)
- Dalton JP, Skelly P, Halton DW. 2004 Role of the tegument and gut in nutrient uptake by parasitic platyhelminths. *Can. J. Zool.* **82**, 211–232. (doi:10.1139/z03-213)
- de Wit E, Munster VJ, van Riel D, Beyer WE, Rimmelzwaan GF, Kuiken T, Osterhaus AD, Fouchier RA. 2010 Molecular determinants of adaptation of highly pathogenic avian influenza H7N7 viruses to efficient replication in the

- human host. *J. Virol.* **84**, 1597–1606. (doi:10.1128/JVI.01783-09)
8. West SA, Buckling A. 2003 Cooperation, virulence and siderophore production in bacterial parasites. *Proc. R. Soc. Lond. B* **270**, 37–44. (doi:10.1098/rspb.2002.2209)
 9. Brown SP, Hochberg ME, Grenfell BT. 2002 Does multiple infection select for raised virulence? *Trends Microbiol.* **10**, 401–405. (doi:10.1016/S0966-842X(02)02413-7)
 10. Sachs JL, Mueller UG, Wilcox TP, Bull JJ. 2004 The evolution of cooperation. *Q. Rev. Biol.* **79**, 135–160. (doi:10.1086/383541)
 11. Schoener TW. 1983 Field experiments on interspecific competition. *Am. Nat.* **122**, 240–285. (doi:10.1086/284133)
 12. Brown AF. 1986 Evidence for density-dependent establishment and survival of *Pomphorhynchus laevis* (Müller, 1776) (Acanthocephala) in laboratory-infected *Salmo gairdneri* Richardson and its bearing on wild populations in *Leuciscus cephalus* (L.). *J. Fish Biol.* **28**, 659–669. (doi:10.1111/j.1095-8649.1986.tb05201.x)
 13. An DD, Danhorn T, Fuqua C, Parsek MR. 2006 Quorum sensing and motility mediate interactions between *Pseudomonas aeruginosa* and *Agrobacterium tumefaciens* in biofilm cocultures. *Proc. Natl Acad. Sci. USA* **103**, 3828–3833. (doi:10.1073/pnas.0511323103)
 14. Ruhe ZC, Low DA, Hayes CS. 2013 Bacterial contact-dependent growth inhibition. *Trends Microbiol.* **21**, 230–237. (doi:10.1016/j.tim.2013.02.003)
 15. Martínez JL. 2008 Antibiotics and antibiotic resistance genes in natural environments. *Science* **321**, 365–367. (doi:10.1126/science.1159483)
 16. Riley MA, Gordon DM. 1999 The ecological role of bacteriocins in bacterial competition. *Trends Microbiol.* **7**, 129–133. (doi:10.1016/S0966-842X(99)01459-6)
 17. Kerr B. 2007 The ecological and evolutionary dynamics of model bacteriocin communities. In *Bacteriocins: ecology and evolution* (eds MA Riley, MA Chavan), pp. 112–134. Berlin: Springer.
 18. Read AF, Taylor LH. 2001 The ecology of genetically diverse infections. *Science* **292**, 1099–1102. (doi:10.1126/science.1059410)
 19. Roode LR, Jacobus CD, Bell AS, Stamou P, Gray D, Read AF. 2006 The role of immune-mediated apparent competition in genetically diverse malaria infections. *Am. Nat.* **168**, 41–53. (doi:10.1086/505160)
 20. Holt R. 1977 Predation, apparent competition and the structure of prey communities. *Theor. Pop. Ecol.* **12**, 197–229. (doi:10.1016/0040-5809(77)90042-9)
 21. Brown SP, Le Chat L, Taddei F. 2008 Evolution of virulence: triggering host inflammation allows invading pathogens to exclude competitors. *Ecol. Lett.* **11**, 44–51. (doi:10.1111/j.1461-0248.2007.01125.x)
 22. Alizon S, de Roode JC, Michalakis Y. 2013 Multiple infections and the evolution of virulence. *Ecol. Lett.* **16**, 556–567. (doi:10.1111/ele.12076)
 23. Cornforth DM, Foster KR. 2013 Competition sensing: the social side of bacterial stress responses. *Nat. Rev Microbiol.* **11**, 285–293. (doi:10.1038/nrmicro2977)
 24. Leggett HC, Brown SP, Reece SE. 2014 War and peace: social interactions in infections. *Phil. Trans. R. Soc. B* **369**, 20130365. (doi:10.1098/rstb.2013.0365)
 25. Chesson P. 2000 Mechanisms of maintenance of species diversity. *Ann. Rev. Ecol. Syst.* **31**, 343–366. (doi:10.1146/annurev.ecolsys.31.1.343)
 26. Lawlor LR, Smith JM. 1976 The coevolution and stability of competing species. *Am. Nat.* **110**, 79–99. (doi:10.2307/2459878)
 27. Karvonen A, Terho P, Seppälä O, Jokela J, Valtonen ET. 2006 Ecological divergence of closely related *Diplostomum* (Trematoda) parasites. *Parasitology* **133**, 229–235. (doi:10.1017/S0031182006000242)
 28. Cobey S, Lipsitch M. 2012 Niche and neutral effects of acquired immunity permit coexistence of pneumococcal serotypes. *Science* **335**, 1376–1380. (doi:10.1126/science.1215947)
 29. Watkins ER, Grad YH, Gupta S, Buckee CO. 2014 Contrasting within- and between-host immune selection shapes *Neisseria* Opa repertoires. *Sci. Rep.* **4**, 6554. (doi:10.1038/srep06554)
 30. Kneitel JM, Chase JM. 2004 Trade-offs in community ecology: linking spatial scales and species coexistence. *Ecol. Lett.* **7**, 69–80. (doi:10.1046/j.1461-0248.2003.00551.x)
 31. Amarasekare P. 2000 Coexistence of competing parasitoids on a patchily distributed host: local versus spatial mechanisms. *Ecology* **81**, 1286–1296. (doi:10.1890/0012-9658(2000)0811286:COCP0A]2.0.CO;2)
 32. Harbison CW, Bush SE, Malenke JR, Clayton DH. 2008 Comparative transmission dynamics of competing parasite species. *Ecology* **89**, 3186–3194. (doi:10.1890/07-1745.1)
 33. Alizon S, Fraser C. 2013 Within and between host evolutionary rates across the HIV genome. *Retrovirology* **10**, 49–59. (doi:10.1186/1742-4690-10-49)
 34. Caraco T, Wang IN. 2008 Free-living pathogens: life-history constraints and strain competition. *J. Theor. Biol.* **250**, 569–579. (doi:10.1016/j.jtbi.2007.10.029)
 35. De Paepe M, Taddei F. 2006 Viruses' life history: towards a mechanistic basis of a trade-off between survival and reproduction among phages. *PLoS Biol.* **4**, e193. (doi:10.1371/journal.pbio.0040193)
 36. Wasik BR, Bhushan A, Ogbunugafor CB, Turner PE. 2015 Delayed transmission selects for increased survival of vesicular stomatitis virus. *Evolution* **69**, 117–125. (doi:10.1111/evo.12544)
 37. Gallet R, Lenormand T, Wang IN. 2012 Phenotypic stochasticity protects lytic bacteriophage populations from extinction during the bacterial stationary phase. *Evolution* **66**, 3485–3494. (doi:10.1111/j.1558-5646.2012.01690.x)
 38. Amarasekare P. 2003 Competitive coexistence in spatially structured environments: a synthesis. *Ecol. Lett.* **6**, 1109–1122. (doi:10.1046/j.1461-0248.2003.00530.x)
 39. Snyder R, Chesson P. 2004 How the spatial scales of dispersal, competition, and environmental heterogeneity interact to affect coexistence. *Am. Nat.* **164**, 645–650. (doi:10.1086/424969)
 40. Murrell DJ, Law R. 2003 Heteromyopia and the spatial coexistence of similar competitors. *Ecol. Lett.* **6**, 48–59. (doi:10.1046/j.1461-0248.2003.00397.x)
 41. Chesson P. 2000 General theory of competitive coexistence in spatially-varying environments. *Theor. Popul. Biol.* **58**, 211–237. (doi:10.1006/tpbi.2000.1486)
 42. Sears ALW, Chesson P. 2007 New methods for quantifying the spatial storage effect: an illustration with desert annuals. *Ecology* **88**, 2240–2247. (doi:10.1890/06-0645.1)
 43. Angert AL, Huxman TE, Chesson P, Venable DL. 2009 Functional tradeoffs determine species coexistence via the storage effect. *Proc. Natl Acad. Sci. USA* **106**, 11 641–11 645. (doi:10.1073/pnas.0904512106)
 44. Chao L, Levin BR. 1981 Structured habitats and the evolution of anticompetitor toxins in bacteria. *Proc. Natl Acad. Sci. USA* **78**, 6324–6328. (doi:10.1073/pnas.78.10.6324)
 45. Durrett R, Levin S. 1997 Allelopathy in spatially distributed populations. *J. Theor. Biol.* **185**, 165–171. (doi:10.1006/jtbi.1996.0292)
 46. Frank SA. 1994 Spatial polymorphism of bacteriocins and other allelopathic traits. *Evol. Ecol.* **8**, 369–386. (doi:10.1007/BF01238189)
 47. Czarán TL, Hoekstra RF. 2003 Killer-sensitive coexistence in metapopulations of micro-organisms. *Proc. R. Soc. Lond. B* **270**, 1373–1378. (doi:10.1098/rspb.2003.2338)
 48. Feldgarden M, Riley MA. 1999 The phenotypic and fitness effects of colicin resistance in *Escherichia coli* K-12. *Evolution* **53**, 1019–1027. (doi:10.2307/2640807)
 49. Kerr B, Riley MA, Feldman MW, Bohannan BJM. 2002 Local dispersal promotes biodiversity in a real-life game of rock–paper–scissors. *Nature* **418**, 171–174. (doi:10.1038/nature00823)
 50. Kirkup BC, Riley MA. 2004 Antibiotic-mediated antagonism leads to a bacterial game of rock–paper–scissors *in vivo*. *Nature* **428**, 412–414. (doi:10.1038/nature02429)
 51. Prado F, Kerr B. 2008 The evolution of restraint in bacterial biofilms under nontransitive competition. *Evolution* **62**, 538–548.
 52. Frean M, Abraham ER. 2001 Rock–scissors–paper and the survival of the weakest. *Proc. R. Soc. Lond. B* **268**, 1323–1327. (doi:10.1098/rspb.2001.1670)
 53. Johnson CR, Seinen I. 2002 Selection for restraint in competitive ability in spatial competition systems. *Proc. R. Soc. Lond. B* **269**, 655–663. (doi:10.1098/rspb.2001.1948)
 54. Nahum JR, Harding BN, Kerr B. 2011 Evolution of restraint in a structured rock–paper–scissors community. *Proc. Natl Acad. Sci. USA* **108**, 10 831–10 838. (doi:10.1073/pnas.1100296108)
 55. Czarán TL, Hoekstra RF, Pagie L. 2002 Chemical warfare between microbes promotes biodiversity.

- Proc. Natl Acad. Sci. USA* **99**, 786–790. (doi:10.1073/pnas.012399899)
56. Pagie L, Hogeweg P. 1999 Colicin diversity: a result of eco-evolutionary dynamics. *J. Theor. Biol.* **196**, 251. (doi:10.1006/jtbi.1998.0838)
 57. Gordon DM, O'Brien CL. 2006 Bacteriocin diversity and the frequency of multiple bacteriocin production in *Escherichia coli*. *Microbiology* **152**, 3239–3244. (doi:10.1099/mic.0.28690-0)
 58. Kirchner JW, Roy BD. 2002 Evolutionary implications of host–pathogen specificity: fitness consequences of pathogen virulence traits. *Evol. Ecol. Res.* **4**, 27–48.
 59. Marston MF, Pierciey FJ, Shepard A, Gearin G, Qi J, Yandava C, Schuster SC, Henn MR, Martiny JBH. 2012 Rapid diversification of coevolving marine *Synechococcus* and a virus. *Proc. Natl Acad. Sci. USA* **109**, 4544–4549. (doi:10.1073/pnas.1120310109)
 60. Paterson S *et al.* 2010 Antagonistic coevolution accelerates molecular evolution. *Nature* **464**, 275–278. (doi:10.1038/nature08798)
 61. Karvonen A, Rellstab C, Louhi KR, Jokela J. 2012 Synchronous attack is advantageous: mixed genotype infections lead to higher infection success in trematode parasites. *Proc. R. Soc. B* **279**, 171–176. (doi:10.1098/rspb.2011.0879)
 62. Rellstab C, Karvonen A, Louhi KR, Jokela J. 2013 Genotype-specific versus cross-reactive host immunity against a macroparasite. *PLoS ONE* **8**, e78427. (doi:10.1371/journal.pone.0078427)
 63. Seppala O, Karvonen A, Valtonen ET, Jokela J. 2009 Interactions among co-infecting parasite species: a mechanism maintaining genetic variation in parasites? *Proc. R. Soc. B* **276**, 691–697. (doi:10.1098/rspb.2008.1229)
 64. Seppälä O, Karvonen A, Rellstab C, Louhi K-R, Jokela J. 2012 Reciprocal interaction matrix reveals complex genetic and dose-dependent specificity among coinfecting parasites. *Am. Nat.* **180**, 306–315. (doi:10.1086/666985)
 65. Hawlena H, Bashey F, Lively CM. 2010 The evolution of spite: population structure and bacteriocin-mediated antagonism in two natural populations of *Xenorhabdus* bacteria. *Evolution* **64**, 3198–3204. (doi:10.1111/j.1558-5646.2010.01070.x)
 66. Hawlena H, Bashey F, Mendes Soares H, Lively CM. 2010 Spiteful interactions in a natural population of the bacterium *Xenorhabdus bovienii*. *Am. Nat.* **175**, 374–381. (doi:10.1086/650375)
 67. Hawlena H, Bashey F, Lively CM. 2012 Bacteriocin-mediated interactions within and between coexisting species. *Ecol. Evol.* **2**, 2521–2526. (doi:10.1002/ece3.354)
 68. Bashey F, Young SK, Hawlena H, Lively CM. 2012 Spiteful interactions between sympatric natural isolates of *Xenorhabdus bovienii* benefit kin and reduce virulence. *J. Evol. Biol.* **25**, 431–437. (doi:10.1111/j.1420-9101.2011.02441.x)
 69. Bashey F, Hawlena H, Lively CM. 2013 Alternative paths to success in a parasite community: within-host competition can favor higher virulence or direct interference. *Evolution* **67**, 900–907. (doi:10.1111/j.1558-5646.2012.01825.x)
 70. Therese MO, Bashey F. 2012 Natal-host environmental effects on juvenile size, transmission success, and operational sex ratio in the entomopathogenic nematode *Steinernema carpocapsae*. *J. Parasitol.* **98**, 1095–1100. (doi:10.1645/GE-3069.1)
 71. Kummerli R, Jiricny N, Clarke LS, West SA, Griffin AS. 2009 Phenotypic plasticity of a cooperative behaviour in bacteria. *J. Evol. Biol.* **22**, 589–598. (doi:10.1111/j.1420-9101.2008.01666.x)
 72. Ghouil M, West SA, Diggle SP, Griffin AS. 2014 An experimental test of whether cheating is context dependent. *J. Evol. Biol.* **27**, 551–556. (doi:10.1111/jeb.12319)
 73. Michel-Briand Y, Baysse C. 2002 The pyocins of *Pseudomonas aeruginosa*. *Biochimie* **84**, 499–510. (doi:10.1016/S0300-9084(02)01422-0)
 74. Russell AB, Hood RD, Bui NK, LeRoux M, Vollmer W, Mougous JD. 2011 Type VI secretion delivers bacteriolytic effectors to target cells. *Nature* **475**, 343–347. (doi:10.1038/nature10244)
 75. Inglis RF, Roberts PG, Gardner A, Buckling A. 2011 Spite and the scale of competition in *Pseudomonas aeruginosa*. *Am. Nat.* **178**, 276–285. (doi:10.1086/660827)
 76. Inglis RF, Brown SP, Buckling A. 2012 Spite versus cheats: competition among social strategies shapes virulence in *Pseudomonas aeruginosa*. *Evolution* **66**, 3472–3484. (doi:10.1111/j.1558-5646.2012.01706.x)
 77. Basler M, Ho BT, Mekalanos JJ. 2013 Tit-for-tat: type VI secretion system counterattack during bacterial cell-cell interactions. *Cell* **152**, 884–894. (doi:10.1016/j.cell.2013.01.042)
 78. Bakkal S, Robinson SM, Ordonez CL, Waltz DA, Riley MA. 2010 Role of bacteriocins in mediating interactions of bacterial isolates taken from cystic fibrosis patients. *Microbiology* **156**, 2058–2067. (doi:10.1099/mic.0.036848-0)
 79. Köhler T, Buckling A, van Delden C. 2009 Cooperation and virulence of clinical *Pseudomonas aeruginosa* populations. *Proc. Natl Acad. Sci. USA* **106**, 6339–6344. (doi:10.1073/pnas.0811741106)
 80. Smith EE, Sims EH, Spencer DH, Kaul R, Olson MV. 2005 Evidence for diversifying selection at the pyoverdine locus of *Pseudomonas aeruginosa*. *J. Bacteriol.* **187**, 2138–2147. (doi:10.1128/JB.187.6.2138-2147.2005)
 81. de Roode JC, Yates AJ, Altizer S. 2008 Virulence-transmission trade-offs and population divergence in virulence in a naturally occurring butterfly parasite. *Proc. Natl Acad. Sci. USA* **105**, 7489–7494. (doi:10.1073/pnas.0710909105)
 82. Ebert D. 1994 Virulence and local adaptation of a horizontally transmitted parasite. *Science* **265**, 1084–1086. (doi:10.1126/science.265.5175.1084)
 83. Buckling A, Brockhurst M. 2008 Kin selection and the evolution of virulence. *Heredity* **100**, 484–488. (doi:10.1038/sj.hdy.6801093)
 84. Bremermann HJ, Pickering J. 1983 A game-theoretical model of parasite virulence. *J. Theor. Biol.* **100**, 411–426. (doi:10.1016/0022-5193(83)90438-1)
 85. Frank SA. 1996 Models of parasite virulence. *Q. Rev. Biol.* **71**, 37–78. (doi:10.1086/419267)
 86. Chao L, Hanley KA, Burch CL, Dahlberg C, Turner PE. 2000 Kin selection and parasite evolution: higher and lower virulence with hard and soft selection. *Q. Rev. Biol.* **75**, 261–275. (doi:10.1086/393499)
 87. Ewald PW. 1994 *Evolution of infectious disease*, p. 298. Oxford, UK: Oxford University Press.
 88. Alizon S, van Baalen M. 2008 Multiple infections, immune dynamics, and the evolution of virulence. *Am. Nat.* **172**, E150–E168. (doi:10.1086/590958)
 89. Alizon S, Lion S. 2011 Within-host parasite cooperation and the evolution of virulence. *Proc. R. Soc. B* **278**, 3738–3747. (doi:10.1098/rspb.2011.0471)
 90. Vigneux F, Bashey F, Sicard M, Lively CM. 2008 Low migration decreases interference competition among parasites and increases virulence. *J. Evol. Biol.* **21**, 1245–1251. (doi:10.1111/j.1420-9101.2008.01576.x)
 91. Gardner A, West SA, Buckling A. 2004 Bacteriocins, spite and virulence. *Proc. R. Soc. Lond. B* **271**, 1529–1535. (doi:10.1098/rspb.2004.2756)
 92. Sofonea MT, Alizon S, Michalakakis Y. 2015 From within-host interactions to epidemiological competition: a general model for multiple infections. *Phil. Trans. R. Soc. B* **370**, 20140303. (doi:10.1098/rstb.2014.0303)
 93. Lysenko ES, Lijek RS, Brown SP, Weiser JN. 2010 Within-host competition drives selection for the capsule virulence determinant of *Streptococcus pneumoniae*. *Curr. Biol.* **20**, 1222–1226. (doi:10.1016/j.cub.2010.05.051)
 94. Barrett LG, Bell T, Dwyer G, Bergelson J. 2011 Cheating, trade-offs and the evolution of aggressiveness in a natural pathogen population. *Ecol. Lett.* **14**, 1149–1157. (doi:10.1111/j.1461-0248.2011.01687.x)
 95. Mikonranta L, Friman VP, Laakso J. 2012 Life history trade-offs and relaxed selection can decrease bacterial virulence in environmental reservoirs. *PLoS ONE* **7**, e43801. (doi:10.1371/journal.pone.0043801)
 96. Koch G, Yepes A, Forstner KU, Wermser C, Stengel ST, Modamio J, Ohlson K, Foster KR, Lopez D. 2014 Evolution of resistance to a last-resort antibiotic in *Staphylococcus aureus* via bacterial competition. *Cell* **158**, 1060–1071. (doi:10.1016/j.cell.2014.06.046)
 97. Singer M. 2010 Pathogen-pathogen interaction: a syndemic model of complex biosocial processes in disease. *Virulence* **1**, 10–18. (doi:10.4161/viru.1.1.9933)
 98. Balmer O, Stearns SC, Schötzau A, Brun R. 2009 Intraspecific competition between co-infecting parasite strains enhances host survival in African trypanosomes. *Ecology* **90**, 3367–3378. (doi:10.1890/08-2291.1)
 99. Massey R, Buckling A, French-Constant R. 2004 Interference competition and parasite virulence. *Proc. R. Soc. Lond. B* **271**, 785–788. (doi:10.1098/rspb.2004.2676)
 100. Gillor O, Etzion A, Riley MA. 2008 The dual role of bacteriocins as anti- and probiotics. *Appl. Microbiol. Biotechnol.* **81**, 591–606. (doi:10.1007/s00253-008-1726-5)

101. Cotter PD, Ross RP, Hill C. 2013 Bacteriocins—a viable alternative to antibiotics? *Nat. Rev. Microbiol.* **11**, 95–105. (doi:10.1038/nrmicro2937)
102. Riley MA, Robinson SM, Roy CM, Dennis M, Liu V, Dorit RL. 2012 Resistance is futile: the bacteriocin model for addressing the antibiotic resistance challenge. *Biochem. Soc. Trans.* **40**, 1438–1442. (doi:10.1042/BST20120179)
103. Scholl D, Martin DW. 2008 Antibacterial efficacy of R-type pyocins towards *Pseudomonas aeruginosa* in a murine peritonitis model. *Antimicrob Agents Chemother* **52**, 1647–1652. (doi:10.1128/AAC.01479-07)
104. Williams SR, Gebhart D, Martin DW, Scholl D. 2008 Retargeting R-type pyocins to generate novel bactericidal protein complexes. *Appl. Environ. Microbiol.* **74**, 3868–3876. (doi:10.1128/aem.00141-08)
105. Koch H, Schmid-Hempel P. 2012 Gut microbiota instead of host genotype drive the specificity in the interaction of a natural host-parasite system. *Ecol. Lett.* **15**, 1095–1103. (doi:10.1111/j.1461-0248.2012.01831.x)
106. Crespi B, Foster K, Ubeda F. 2014 First principles of Hamiltonian medicine. *Phil. Trans. R. Soc. B* **369**, 20130366. (doi:10.1098/rstb.2013.0366)
107. Köhler T, Perron GG, Buckling A, van Delden C. 2010 Quorum sensing inhibition selects for virulence and cooperation in *Pseudomonas aeruginosa*. *PLoS Pathog.* **6**, e1000883. (doi:10.1371/journal.ppat.1000883)
108. Allen RC, Popat R, Diggle SP, Brown SP. 2014 Targeting virulence: can we make evolution-proof drugs? *Nat. Rev. Microbiol.* **12**, 300–308. (doi:10.1038/nrmicro3232)