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The immunopathogenesis of Crohn's disease: a three-stage model

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Abstract

The pathogenesis of Crohn's disease (CD) has remained an enigma for at least a century. There was considerable optimism that genetic linkage and genome-wide association (GWA) studies had identified genes causally responsible. However, the realisation that these genes make a relatively minor contribution to the development of CD has led to the acceptance of a 'missing heritability'. In contrast to the weak genetic effects, patients with CD almost without exception exhibit a gross phenotype, namely a profound systemic failure of the acute inflammatory response. This results in markedly delayed clearance of bacteria from the tissues, leading to local chronic granulomatous inflammation and compensatory adaptive immunological changes, as well as constitutional symptoms.

Introduction

'Inflammatory bowel disease' encompasses several distinct clinical entities, the most common being Crohn's disease (CD) and ulcerative colitis (UC). CD is a chronic, relapsing–remitting inflammatory condition predominantly affecting the terminal ileum and colon, associated with distinctive pathological features [1]. Unsuccessful attempts have been made to identify its cause since it first entered the literature at the turn of the 20th century.

Putative 'causes' have included infections with a variety of organisms such as mycobacteria, L-form bacteria, *E. coli* and measles virus. A host of other mechanisms have also been postulated, including auto-immunity and disordered T cell function [2].

The advent of molecular biological and gene sequencing technologies spawned studies relating disease phenotypes to particular regions of the genome and then to specific genes. This search appeared particularly fruitful in relation to CD where a number of statistically significant associations were identified.

Furthermore, gene targeting technologies produced mouse models of relevance to CD, either because the targeted gene corresponded to one of the associated genes described above, or its disruption predisposed to bowel inflammation interpreted as having some similarity to human 'inflammatory bowel disease'.

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Conflict of interest statement

None of the authors have any conflicts of interest to declare.

This review discusses the concept that the development of CD occurs in three stages. The first of these is ingress of bacteria and antigenic material into the bowel wall. In stage 2, a weak acute inflammatory response results in impaired clearance of this material. Chronic granulomatous inflammation and adaptive immune responses are subsequently provoked in stage 3, culminating in the development of CD. The potential relevance of recently identified susceptibility genes and the relationship of animal models to this pathogenic scheme will be evaluated.

Genetic studies and Crohn's disease

Twin and family studies confirmed a strong genetic influence on the acquisition of CD. For example, approximately 50% of monozygotic twins and 30% of offspring of two affected parents develop disease [3]. Linkage analysis and positional cloning strategies, together with subsequent genome-wide association (GWA) studies, have identified over 30 distinct genetic loci that confer susceptibility. Some of the most strongly associated genes included *CARD15*, the *IBD5* locus, the autophagy genes *ATG16L1* and *IRGM* and the IL-23 receptor [4[•]].

Whilst such studies have provided several important clues into genetic susceptibility and pathogenesis of CD, it must be stressed that all the polymorphisms identified to date cannot be regarded as 'causal'. The variants have very low penetrance, even in the case of *CARD15*. In a random sample of 100 000 people, an estimated 15 000 would be heterozygous for one *CARD15* variant, with around 500 homozygous or compound heterozygous [5]. Approximately 100 in the whole sample will develop CD, but only 28 of these patients will be simple homozygous or compound heterozygous for *CARD15* polymorphisms [6]. A similar situation exists for *ATG16L1*, but with even weaker effects.

It is increasingly apparent that GWA studies do not provide a comprehensive description of the entire genetic risk for any one condition. This is illustrated by three recent studies that sought to identify genetic factors determining human height (a classic polygenic trait with a strong genetic component), using data from approximately 63 000 individuals [7–9]. Whilst more than 40 important genes were identified, they accounted for just over 5% of normal height variation. A similar situation exists in CD; it is estimated that all the genes identified so far account for less than 20% of the total genetic risk [4[•]]. This has led to the recent suggestion of 'missing heritability' — that both the individual and cumulative effects of susceptibility genes identified to date are very small, and far less than the total heritability [10^{••}].

The unifying findings in all CD patients are therefore phenotypic abnormalities, rather than defects in single genes. Polymorphisms in the identified genes, and many more that are as yet undiscovered, may confer susceptibility by contributing to mucosal barrier dysfunction, the innate immunodeficiency state or by influencing the propagation of chronic inflammation in the tertiary phase of the disease.

Animal models and Crohn's disease

Numerous animal models of inflammatory bowel disease have been described, including chemically induced colitides, inbred animal strains, knockout or transgenic animals and adoptive transfer models [11,12]. These have been useful tools to investigate the physiology of bowel inflammation, highlighting roles for immune–microflora interactions. However, the direct applicability of most to human CD is unclear. Firstly, there are striking differences in pathological features, with few models demonstrating granuloma formation, discontinuous transmural inflammation and extraintestinal manifestations. Another limitation is the short lifespan of the mouse, which typically does not exceed four years. Given that onset of CD in humans peaks in the second and third decade, these models may well not adequately take into account the time dependence for the development of lesions. Furthermore, GWA studies have failed to demonstrate significant associations between many of the manipulated genes in animal models and human CD. Indeed, even between different animal strains, the effects of certain genetic alterations and exogenous manipulations are variable [11].

Conversely, animals with targeted manipulations of many of the recently identified 'CD susceptibility genes' do not develop a clear CD phenotype, in spite of immunological abnormalities. Macrophages isolated from *CARD15* knockout mice secrete reduced IL-12 in response to MDP stimulation, but do not develop spontaneous bowel inflammation [13]. Spontaneous colitis is likewise not observed in mice with a targeted deletion of *ATG16L1* in haematopoietic cells, nor in animals hypomorphic for *ATG16L1* expression, despite demonstrating increased susceptibility to acute DSS colitis, enhanced macrophage IL-1 β secretion after LPS stimulation [14] and abnormalities in Paneth cells [15].

Therefore, in the absence of an obvious pathogenesis of CD, we propose a three-stage mechanism for the development of CD lesions (illustrated in Figure 1). This takes into account and integrates much of the existing knowledge of CD, including genetic factors and data from animal models.

Stage 1: penetration of luminal contents into the bowel wall

The development of CD is dependent on bowel contents. Diversion of the faecal stream in such patients is associated with the induction of remission [16,17], and experimental reintroduction of ileostomy effluent into the excluded ileum results in recurrence [18]. To produce CD lesions, components of the faecal stream must penetrate the intestinal barrier and gain access to the underlying bowel tissues.

The gastrointestinal epithelial cells normally form a relatively impermeable physical barrier to luminal contents, facilitated by tight junctions that impede paracellular transport [19]. A thick layer of mucus, largely composed of mucin glycoproteins, overlies the epithelial cell layer. Defensins, immunoglobulins and various other molecules are also contained within this mucus layer, which form an additional defensive component. These substances are secreted by enterocytes, lymphocytes, goblet cells and Paneth cells (the last being generally restricted to the crypts of Lieberkuhn in the small bowel) [20].

Several studies have reported increased intestinal permeability in CD patients [21–24]. The underlying mechanism remains debated, as does whether it is a primary phenomenon or secondary to a chronic inflammatory state in the bowel. Although many studies investigated patients with active CD, increased permeability has also been reported in macroscopically normal small bowel of patients [25], which may predict recurrence [26]. These observations are compatible with the hypothesis that increased permeability is a primary initiating factor in CD.

Subsequent studies demonstrated increased permeability in a proportion of both healthy first-degree relatives [27] and spouses [28] of CD patients, arguing in favour of both environmental and genetic determinants. The underlying bowel permeability would make the mucosa more susceptible to damage. Whereas in most cases penetration of foreign material probably follows viral or bacterial infection, clearly a less robust mucosa will have reduced resistance. Other environmental factors such as trauma, high intra-luminal pressure, non-steroidal anti-inflammatory drugs and hypoxia could also play roles in causing mucosal damage.

Polymorphisms in mucin genes such as *MUC19* are associated with increased risk of CD [4[•]]; these variants could foreseeably increase the susceptibility of the mucosal barrier to injury. Alterations in expression and post-translational modifications of mucins have also been reported in CD [29]. Another CD susceptibility locus is located in a gene desert region on chromosome 5p13.1, which may be associated with altered expression of the prostaglandin E4 receptor (*PTGE4R*) [30]. *PTGE4R* may have roles in maintenance of mucosal barrier integrity, which is reflected in the increased susceptibility of *PTGE4R* knockout mice to dextran sodium sulphate (DSS) induced colitis [31].

Several mouse models underscore the importance of mucosal barrier dysfunction in the induction of bowel inflammation. Transgenic mice manipulated to express a dominant negative N-cadherin, a junctional adhesion protein, develop spontaneous inflammatory bowel disease [32]. Furthermore, introduction of mutations in the mucin encoding gene *MUC2* also results in spontaneous colitis, although the histopathological features are more reminiscent of UC [33]. Interestingly, the inbred SAMP1/Yit (Samp) mouse strain displays increased ileal permeability, which precedes the onset of a spontaneous ileitis [34]. In this model, the inflammation described is not dissimilar to human CD, with discontinuous, transmural leukocytic infiltrates and coalescence of macrophages into aggregates.

Stage 2: impaired clearance of foreign material from the bowel wall

It was realised over three decades ago that the consequences of the penetration of faecal contents into the underlying tissues would depend upon the adequacy of the acute inflammatory response. In CD this response is defective as the result of a primary failure of acute inflammation [35^{••}]. Neutrophil accumulation to 'skin windows', as well as traumatised bowel mucosa, is impaired in CD patients, as a consequence of diminished concentrations of acute inflammatory mediators such as IL-8 and IL-1β at these sites [36^{••}]. The neutrophils themselves function normally *in vitro* [37], and the addition of exogenous IL-8 to skin windows of CD patients corrected neutrophil influx to normal levels [36^{••}].

Pro-inflammatory cytokines are primarily secreted by resident tissue macrophages [38], which in the bowel are derived and continually replenished from peripheral blood monocytes [39]. Dramatically impaired secretion of pro-inflammatory cytokines by macrophages from CD patients was observed in response to stimulation with heat-killed *E. coli* [67^{••}]. Subsequent experiments, including microarray analysis of the transcriptome in these cells, indicated defects in macrophage secretory systems as the underlying cause of this phenomenon, which results in mistargeting of cytokines to lysosomal compartments.

In contrast to previous studies, many of which describe elevated cytokine levels in biopsies or peripheral blood mononuclear cells (PBMCs) from CD patients, in our studies samples were obtained from patients with quiescent disease. Pure cultures of a single cell type (macrophages) were obtained, and their responses to defined stimuli examined. It is therefore improbable that the results obtained were secondary to a chronic inflammatory state in the bowel, further bolstered by the distinct cytokine profiles observed from macrophages cultured from UC patients.

It was hypothesised that delayed neutrophil recruitment would impair clearance of intestinal contents gaining access to the bowel tissues. We have recently been able to test this experimentally by injecting heat-killed *E. coli*, a gram negative coliform, into the subcutaneous tissues of the forearm. The accumulation of ¹¹¹indium labelled neutrophils to these sites was markedly delayed in CD compared to healthy controls. To determine whether this affected clearance of bacteria, the *E. coli* were labelled with ³²P, and the rate of disappearance of radioactivity from injection sites determined. Clearance was dramatically lower in CD, requiring, on average, 40 days compared to 10 days in healthy controls [67^{••}].

In certain scenarios, the ability of neutrophils to degrade and remove the bacteria and other bowel contents might be compromised as a result of an inherited defect. Such a situation exists in patients with congenital, monogenic disorders of phagocyte function. Patients with chronic granulomatous disease (CGD), an immunodeficiency disorder caused by mutations in NADPH oxidase, frequently suffer from bowel inflammation that is indistinguishable from CD [40]. The cellular defect in this condition is not limited to bacterial killing — digestion is also severely impaired as a consequence of abnormal pH and charge compensation [41]. This therefore supports the hypothesis that clearance of foreign material is a critical factor in the development of CD.

Strong associations between other monogenic disorders of neutrophil function and CD have also been highlighted in recent reviews [42,43]. Mutations in single genes result in failure of neutrophil production and accumulation (congenital neutropenias and leukocyte adhesion deficiency), impaired digestion (CGD and Glycogen Storage Disease-1b) and defective phagolysosomal fusion and vesicle trafficking (Chediak–Higashi and Hermansky–Pudlak syndrome). As a consequence of these defects, these patients also have a highly impaired ability to clear foreign material and, as would be expected, have an increased susceptibility to CD.

The immunodeficiency phenotype is also consistent with findings from genetic studies. *CARD15* was the first CD susceptibility locus identified. It encodes NOD2, which is

expressed in mononuclear phagocytes, epithelial cells and Paneth cells [44,45]. It is thought to interact with MDP, a constituent of bacterial cell walls, leading to activation of the transcription factor NF-kB and induction of pro-inflammatory cytokines. Polymorphisms in the leucine rich repeat domain predispose to CD, and are associated with reduced proinflammatory cytokine responses to MDP [36^{••},46]. *In vivo*, addition of exogenous MDP was shown to correct the defect in neutrophil accumulation in CD patients wild-type for *CARD15*, an ability that was abrogated by the presence of CD-associated polymorphisms. This suggests a possible 'compensatory' role for NOD2 in boosting weak acute inflammatory responses to bacteria [36^{••}]. *CARD15* sequence variants may also be associated with reduced alpha defensin expression by Paneth cells [47], and altered antiinflammatory cytokine production by PBMCs [48], suggesting that NOD2 could exert effects in multiple stages of CD pathogenesis.

GWA studies have also identified variants in the *ATG16L1* and *IRGM* genes that are associated with CD [49°,50°,51]. CD-associated mutations in *ATG16L1*, as well as functional knockdown of *ATG16L1* or *IRGM*, results in impaired autophagy and elimination of intracellular pathogens [52,53], in keeping with the concept that clearance of foreign material is important in the development of bowel inflammation.

There are few animal models that convincingly describe bowel inflammation occurring in conjunction with innate immunodeficiency. Mice with targeted deletions of *STAT3* in the bone marrow are a possible exception, where impaired innate immune function, including reduced NADPH oxidase activity, was demonstrated. Interestingly, these mice had histopathological features reminiscent of CD, with transmural inflammation and granuloma formation [54].

If CD arises from a systemic defect in innate immunity, one might expect patients to manifest increased susceptibility to bacterial infection. This may indeed be the case, and several studies have reported increased incidences of acute gastroenteritis [55] and urinary tract infection [56]. Larger scale studies are required to confirm this finding, which must take into account important confounding factors such as surgery, malnutrition and use of immunosuppressant therapy. Another consideration is bacterial load: most acute infections arise from multiplication of a small number of initial inoculating organisms, which even the partially attenuated innate immune response in CD might be sufficient to control. In contrast, the terminal ileum and colon, the sites most commonly affected in CD, contain large number of bacteria (approximately $10^8/g$ and $10^{11}/g$, respectively [57]) which could foreseeably 'overwhelm' the impaired clearance mechanisms.

Stage 3: compensatory adaptive immune responses

In the absence of adequate neutrophil recruitment, the remaining uncleared debris will be phagocytosed by macrophages. These cells subsequently form granulomata, in an attempt to contain this material. Macrophage activation will then result in a 'second wave' of secretion of pro-inflammatory cytokines and chemokines that will drive recruitment of T cells to the site, as well as their polarisation to the characteristic Th1 phenotype. This phase of chronic inflammation is temporally distinct (occurring days to weeks after penetration) from the

initial acute inflammatory response described in stage 2 (which occurs within several hours). At this stage, even if net production of cytokines by each cell were lower than normal, the overall number of cells will be so great that damaging concentrations of cytokines will be produced. Local tissue damage, as well as systemic responses then ensue, giving rise to the symptoms of CD.

In this model, granuloma formation and lymphocytic infiltration represent compensatory mechanisms for the initial failure of acute inflammation, rather than primary pathogenic defects. In support of this, CD does not fulfil Witebsky's postulates for defining an autoimmune condition [58]. Furthermore, whilst there have been reports of autoreactive T cells and autoantibodies in CD patients, their mechanistic relevance has not been demonstrated. These may therefore be generated secondary to a chronic inflammatory state in the bowel, characterised by a high degree of cell turnover and presentation of antigens to the adaptive immune system.

The development and perpetuation of the chronic inflammatory response will involve products of variety of different genes, which could further contribute to CD susceptibility. For example, polymorphisms in genes encoding the IL-23 receptor, and the IL-12B subunit are associated with CD, with the minor alleles conferring protection [4•,59]. The proteins encoded by these genes may modify the chronic inflammatory response to bacteria through the induction of Th17 cells [60], effects on immunosuppressive T-reg cells [61], driving granuloma formation [62] or by other mechanisms. The association of these genes with other chronic inflammatory conditions, including UC [63], is noteworthy.

Despite having a secondary role in disease pathogenesis, the chronic inflammatory response is nevertheless important clinically, being responsible for the local lesions such as ulceration, fistulation and stricturing, as well as systemic symptoms. Immunosuppressive therapies and drugs directed against TNFa target this stage of disease pathogenesis by direct cytokine blockade, induction of leukocyte apoptosis [64] and stimulation of T-reg cells [65]. Whilst suppressing chronic inflammation in the final stage of CD pathogenesis, thereby inducing remission, therapies such as corticosteroids are not efficacious at preventing relapse [66], and may indeed exacerbate the underlying innate immunodeficiency.

Conclusion

In conclusion, we propose a model for the pathogenesis of CD that involves three temporally distinct stages: penetration of faecal contents into the underlying tissues (stage 1), which is followed by a weak acute inflammatory response to this material that arises from a generalised defect in pro-inflammatory cytokine secretion by macrophages. The diminished neutrophil recruitment that occurs as a consequence of this defect results in impaired clearance of antigenic material (stage 2). Compensatory responses will then come into play (stage 3), which includes the recruitment of T cells and their polarisation to a Th1 phenotype. The model is consistent with findings from recent genetic studies, as well as animal models of inflammatory bowel disease.

This scheme of CD pathogenesis has significant implications, both in terms of future research and therapies. It identifies an immediate need to understand the mechanisms of cytokine trafficking within immune cells, and locating variants in these processes that may contribute to the development of CD. More fundamentally, the phenotype that appears to predispose to CD is one of an impotent acute inflammatory response. The magnitude of the latter will be determined by the collective effects of a very large number of different gene products and environmental factors. A bioinformatics approach will be required to understand how these inflammatory networks interact, the individual factors summate, and the combinations that predispose to CD.

This model highlights the value of a 'phenotype to gene' approach to CD research. Further functional characterisation of the macrophage phenotype and mucosal barrier function, as well as discovering the underlying cellular and molecular mechanisms, will help elucidate the identity of the genes that make up the 'missing heritability', which are likely to be of low effect size and penetrance. Prerequisites to any further studies will be precise clinical phenotyping of patient groups (not using a generic 'inflammatory bowel disease' cohort), and careful distinction between primary phenomena and those occurring secondary to chronic inflammation or its treatment. The model will also be useful in evaluation of future candidate susceptibility genes identified in GWA and familial studies. Given the complex, polygenic nature of CD, the precise molecular lesions will differ between patients and may well be impractical to correct clinically. However, therapies directed at protection of the mucosal barrier, and stimulating the acute inflammatory response may prove efficacious for the prevention of relapse in patients with quiescent disease. In a wider context, this model of CD pathogenesis may be equally applicable to many other chronic inflammatory conditions.

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References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- •• of outstanding interest
- 1. Podolsky DK. Inflammatory bowel disease. N Engl J Med. 2002; 347:417–429. [PubMed: 12167685]
- Korzenik JR. Past and current theories of etiology of IBD: toothpaste, worms, and refrigerators. J Clin Gastroenterol. 2005; 39:S59–S65. [PubMed: 15758661]
- Halme L, Paavola-Sakki P, Turunen U, Lappalainen M, Farkkila M, Kontula K. Family and twin studies in inflammatory bowel disease. World J Gastroenterol. 2006; 12:3668–3672. [PubMed: 16773682]
- 4 •. Barrett JC, Hansoul S, Nicolae DL, Cho JH, Duerr RH, Rioux JD, Brant SR, Silverberg MS, Taylor KD, Barmada MM, et al. Genome-wide association defines more than 30 distinct susceptibility loci for Crohn's disease. Nat Genet. 2008; 40:955–962. [PubMed: 18587394] A

meta-analysis of three GWA studies for CD, which resulted in the identification of over 20 new susceptibility loci, and verification of genes previously identified as associated with CD.

- Hugot JP, Zaccaria I, Cavanaugh J, Yang H, Vermeire S, Lappalainen M, Schreiber S, Annese V, Jewell DP, Fowler EV, et al. Prevalence of CARD15/NOD2 mutations in Caucasian healthy people. Am J Gastroenterol. 2007; 102:1259–1267. [PubMed: 17319929]
- Cuthbert AP, Fisher SA, Mirza MM, King K, Hampe J, Croucher PJ, Mascheretti S, Sanderson J, Forbes A, Mansfield J, et al. The contribution of NOD2 gene mutations to the risk and site of disease in inflammatory bowel disease. Gastroenterology. 2002; 122:867–874. [PubMed: 11910337]
- Weedon MN, Lango H, Lindgren CM, Wallace C, Evans DM, Mangino M, Freathy RM, Perry JR, Stevens S, Hall AS, et al. Genome-wide association analysis identifies 20 loci that influence adult height. Nat Genet. 2008; 40:575–583. [PubMed: 18391952]
- Lettre G, Jackson AU, Gieger C, Schumacher FR, Berndt SI, Sanna S, Eyheramendy S, Voight BF, Butler JL, Guiducci C, et al. Identification of ten loci associated with height highlights new biological pathways in human growth. Nat Genet. 2008; 40:584–591. [PubMed: 18391950]
- Gudbjartsson DF, Walters GB, Thorleifsson G, Stefansson H, Halldorsson BV, Zusmanovich P, Sulem P, Thorlacius S, Gylfason A, Steinberg S, et al. Many sequence variants affecting diversity of adult human height. Nat Genet. 2008; 40:609–615. [PubMed: 18391951]
- 10 ••. Maher B. Personal genomes: the case of the missing heritability. Nature. 2008; 456:18–21.
 [PubMed: 18987709] This *Nature* news feature discusses the concept of 'missing heritability' in the light of recent 'disease susceptibility genes' identified by GWA studies.
- Wirtz S, Neurath MF. Mouse models of inflammatory bowel disease. Adv Drug Deliv Rev. 2007; 59:1073–1083. [PubMed: 17825455]
- 12. Hoffmann JC, Pawlowski NN, Kuhl AA, Hohne W, Zeitz M. Animal models of inflammatory bowel disease: an overview. Pathobiology. 2002; 70:121–130. [PubMed: 12571415]
- Pauleau AL, Murray PJ. Role of NOD2 in the response of macrophages to toll-like receptor agonists. Mol Cell Biol. 2003; 23:7531–7539. [PubMed: 14560001]
- Saitoh T, Fujita N, Jang MH, Uematsu S, Yang BG, Satoh T, Omori H, Noda T, Yamamoto N, Komatsu M, et al. Loss of the autophagy protein Atg16L1 enhances endotoxin-induced IL-1beta production. Nature. 2008; 456:264–268. [PubMed: 18849965]
- Cadwell K, Liu JY, Brown SL, Miyoshi H, Loh J, Lennerz JK, Kishi C, Kc W, Carrero JA, Hunt S, et al. A key role for autophagy and the autophagy gene Atg16l1 in mouse and human intestinal Paneth cells. Nature. 2008; 456:259–263. [PubMed: 18849966]
- Edwards CM, George BD, Jewell DP, Warren BF, Mortensen NJ, Kettlewell MG. Role of a defunctioning stoma in the management of large bowel Crohn's disease. Br J Surg. 2000; 87:1063–1066. [PubMed: 10931051]
- Winslet MC, Allan A, Poxon V, Youngs D, Keighley MR. Faecal diversion for Crohn's colitis: a model to study the role of the faecal stream in the inflammatory process. Gut. 1994; 35:236–242. [PubMed: 8307475]
- D'Haens GR, Geboes K, Peeters M, Baert F, Penninckx F, Rutgeerts P. Early lesions of recurrent Crohn's disease caused by infusion of intestinal contents in excluded ileum. Gastroenterology. 1998; 114:262–267. [PubMed: 9453485]
- Berkes J, Viswanathan VK, Savkovic SD, Hecht G. Intestinal epithelial responses to enteric pathogens: effects on the tight junction barrier, ion transport, and inflammation. Gut. 2003; 52:439–451. [PubMed: 12584232]
- McGuckin MA, Eri R, Simms LA, Florin TH, Radford-Smith G. Intestinal barrier dysfunction in inflammatory bowel diseases. Inflamm Bowel Dis. 2009; 15:100–113. [PubMed: 18623167]
- Olaison G, Leandersson P, Sjodahl R, Tagesson C. Intestinal permeability to polyethyleneglycol 600 in Crohn's disease. Peroperative determination in a defined segment of the small intestine. Gut. 1988; 29:196–199. [PubMed: 3345930]
- Ukabam SO, Clamp JR, Cooper BT. Abnormal small intestinal permeability to sugars in patients with Crohn's disease of the terminal ileum and colon. Digestion. 1983; 27:70–74. [PubMed: 6414866]

- Bjarnason I, O'Morain C, Levi AJ, Peters TJ. Absorption of 51chromium-labeled ethylenediaminetetraacetate in inflammatory bowel disease. Gastroenterology. 1983; 85:318–322. [PubMed: 6407889]
- Casellas F, Aguade S, Soriano B, Accarino A, Molero J, Guarner L. Intestinal permeability to 99mTc-diethylenetriaminopentaacetic acid in inflammatory bowel disease. Am J Gastroenterol. 1986; 81:767–770. [PubMed: 3529937]
- Peeters M, Ghoos Y, Maes B, Hiele M, Geboes K, Vantrappen G, Rutgeerts P. Increased permeability of macroscopically normal small bowel in Crohn's disease. Dig Dis Sci. 1994; 39:2170–2176. [PubMed: 7924738]
- Wyatt J, Vogelsang H, Hubl W, Waldhoer T, Lochs H. Intestinal permeability and the prediction of relapse in Crohn's disease. Lancet. 1993; 341:1437–1439. [PubMed: 8099141]
- Hollander D, Vadheim CM, Brettholz E, Petersen GM, Delahunty T, Rotter JI. Increased intestinal permeability in patients with Crohn's disease and their relatives. A possible etiologic factor. Ann Intern Med. 1986; 105:883–885. [PubMed: 3777713]
- Breslin NP, Nash C, Hilsden RJ, Hershfield NB, Price LM, Meddings JB, Sutherland LR. Intestinal permeability is increased in a proportion of spouses of patients with Crohn's disease. Am J Gastroenterol. 2001; 96:2934–2938. [PubMed: 11693329]
- Moehle C, Ackermann N, Langmann T, Aslanidis C, Kel A, Kel-Margoulis O, Schmitz-Madry A, Zahn A, Stremmel W, Schmitz G. Aberrant intestinal expression and allelic variants of mucin genes associated with inflammatory bowel disease. J Mol Med. 2006; 84:1055–1066. [PubMed: 17058067]
- Libioulle C, Louis E, Hansoul S, Sandor C, Farnir F, Franchimont D, Vermeire S, Dewit O, de Vos M, Dixon A, et al. Novel Crohn disease locus identified by genome-wide association maps to a gene desert on 5p13.1 and modulates expression of PTGER4. PLoS Genet. 2007; 3:e58. [PubMed: 17447842]
- 31. Kabashima K, Saji T, Murata T, Nagamachi M, Matsuoka T, Segi E, Tsuboi K, Sugimoto Y, Kobayashi T, Miyachi Y, et al. The prostaglandin receptor EP4 suppresses colitis, mucosal damage and CD4 cell activation in the gut. J Clin Invest. 2002; 109:883–893. [PubMed: 11927615]
- Hermiston ML, Gordon JI. Inflammatory bowel disease and adenomas in mice expressing a dominant negative N-cadherin. Science. 1995; 270:1203–1207. [PubMed: 7502046]
- 33. Heazlewood CK, Cook MC, Eri R, Price GR, Tauro SB, Taupin D, Thornton DJ, Png CW, Crockford TL, Cornall RJ, et al. Aberrant mucin assembly in mice causes endoplasmic reticulum stress and spontaneous inflammation resembling ulcerative colitis. PLoS Med. 2008; 5:e54. [PubMed: 18318598]
- Olson TS, Reuter BK, Scott KG, Morris MA, Wang XM, Hancock LN, Burcin TL, Cohn SM, Ernst PB, Cominelli F, et al. The primary defect in experimental ileitis originates from a nonhematopoietic source. J Exp Med. 2006; 203:541–552. [PubMed: 16505137]
- 35 ••. Segal AW, Loewi G. Neutrophil dysfunction in Crohn's disease. Lancet. 1976; 2:219–221.
 [PubMed: 59239] The first demonstration of impaired acute inflammation in patients with CD, using a skin window technique.
- 36 ••. Marks DJ, Harbord MW, MacAllister R, Rahman FZ, Young J, Al Lazikani B, Lees W, Novelli M, Bloom S, Segal AW. Defective acute inflammation in Crohn's disease: a clinical investigation. Lancet. 2006; 367:668–678. [PubMed: 16503465] In this study, impaired recruitment of neutrophils to both skin windows and sites of trauma in the bowel, was demonstrated in CD patients. This was related to reduced concentrations of pro-inflammatory mediators at these sites, including IL-8 and IL-1β, with associated cellular defects identified at the level of the macrophage. The pathophysiological relevance of these findings was illustrated by demonstrating grossly attenuated acute inflammatory responses to bacteria subcutaneously inoculated into CD patients.
- Morain CO, Segal AA, Walker D, Levi AJ. Abnormalities of neutrophil function do not cause the migration defect in Crohn's disease. Gut. 1981; 22:817–822. [PubMed: 7028577]
- Medzhitov R. Origin and physiological roles of inflammation. Nature. 2008; 454:428–435. [PubMed: 18650913]

- Smythies LE, Maheshwari A, Clements R, Eckhoff D, Novak L, Vu HL, Mosteller-Barnum LM, Sellers M, Smith PD. Mucosal IL-8 and TGF-beta recruit blood monocytes: evidence for crosstalk between the lamina propria stroma and myeloid cells. J Leukoc Biol. 2006; 80:492–499. [PubMed: 16793909]
- Marks DJ, Miyagi K, Rahman FZ, Novelli M, Bloom SL, Segal AW. Inflammatory bowel disease in CGD reproduces the clinicopathological features of Crohn's disease. Am J Gastroenterol. 2009; 104:117–124. [PubMed: 19098859]
- Reeves EP, Lu H, Jacobs HL, Messina CG, Bolsover S, Gabella G, Potma EO, Warley A, Roes J, Segal AW. Killing activity of neutrophils is mediated through activation of proteases by K⁺ flux. Nature. 2002; 416:291–297. [PubMed: 11907569]
- 42. Rahman FZ, Marks DJ, Hayee BH, Smith AM, Bloom SL, Segal AW. Phagocyte dysfunction and inflammatory bowel disease. Inflamm Bowel Dis. 2008; 14:1443–1452. [PubMed: 18421761]
- 43. Korzenik JR, Dieckgraefe BK. Is Crohn's disease an immunodeficiency? A hypothesis suggesting possible early events in the pathogenesis of Crohn's disease. Dig Dis Sci. 2000; 45:1121–1129. [PubMed: 10877227]
- 44. Ogura Y, Bonen DK, Inohara N, Nicolae DL, Chen FF, Ramos R, Britton H, Moran T, Karaliuskas R, Duerr RH, et al. A frameshift mutation in NOD2 associated with susceptibility to Crohn's disease. Nature. 2001; 411:603–606. [PubMed: 11385577]
- 45. Hugot JP, Chamaillard M, Zouali H, Lesage S, Cezard JP, Belaiche J, Almer S, Tysk C, O'Morain CA, Gassull M, et al. Association of NOD2 leucine-rich repeat variants with susceptibility to Crohn's disease. Nature. 2001; 411:599–603. [PubMed: 11385576]
- 46. Li J, Moran T, Swanson E, Julian C, Harris J, Bonen DK, Hedl M, Nicolae DL, Abraham C, Cho JH. Regulation of IL-8 and IL-1beta expression in Crohn's disease associated NOD2/CARD15 mutations. Hum Mol Genet. 2004; 13:1715–1725. [PubMed: 15198989]
- 47. Wehkamp J, Harder J, Weichenthal M, Schwab M, Schaffeler E, Schlee M, Herrlinger KR, Stallmach A, Noack F, Fritz P, et al. NOD2 (CARD15) mutations in Crohn's disease are associated with diminished mucosal alpha-defensin expression. Gut. 2004; 53:1658–1664. [PubMed: 15479689]
- Noguchi E, Homma Y, Kang X, Netea MG, Ma X. A Crohn's disease-associated NOD2 mutation suppresses transcription of human IL10 by inhibiting activity of the nuclear ribonucleoprotein hnRNP-A1. Nat Immunol. 2009; 10:471–479. [PubMed: 19349988]
- 49 •. Rioux JD, Xavier RJ, Taylor KD, Silverberg MS, Goyette P, Huett A, Green T, Kuballa P, Barmada MM, Datta LW, et al. Genome-wide association study identifies new susceptibility loci for Crohn disease and implicates autophagy in disease pathogenesis. Nat Genet. 2007; 39:596–604. [PubMed: 17435756] This GWA study confirmed ATG16L1 as a CD susceptibility gene, and identified a functional role for the ATG16L1 protein in autophagy of intracellular bacteria.
- 50 •. Hampe J, Franke A, Rosenstiel P, Till A, Teuber M, Huse K, Albrecht M, Mayr G, De LV, Briggs J, et al. A genome-wide association scan of nonsynonymous SNPs identifies a susceptibility variant for Crohn disease in ATG16L1. Nat Genet. 2007; 39:207–211. [PubMed: 17200669] A GWA study of over 1000 individuals that led to the discovery of ATG16L1 as a CD susceptibility gene.
- Genome-wide association study of 14,000 cases of seven common diseases and 3,000 shared, controls. Nature. 2007; 447:661–678. [PubMed: 17554300]
- Singh SB, Davis AS, Taylor GA, Deretic V. Human IRGM induces autophagy to eliminate intracellular mycobacteria. Science. 2006; 313:1438–1441. [PubMed: 16888103]
- Kuballa P, Huett A, Rioux JD, Daly MJ, Xavier RJ. Impaired autophagy of an intracellular pathogen induced by a Crohn's disease associated ATG16L1 variant. PLoS ONE. 2008; 3:e3391. [PubMed: 18852889]
- 54. Welte T, Zhang SS, Wang T, Zhang Z, Hesslein DG, Yin Z, Kano A, Iwamoto Y, Li E, Craft JE, et al. STAT3 deletion during hematopoiesis causes Crohn's disease-like pathogenesis and lethality: a critical role of STAT3 in innate immunity. Proc Natl Acad Sci U S A. 2003; 100:1879–1884. [PubMed: 12571365]

- Porter CK, Tribble DR, Aliaga PA, Halvorson HA, Riddle MS. Infectious gastroenteritis and risk of developing inflammatory bowel disease. Gastroenterology. 2008; 135:781–786. [PubMed: 18640117]
- 56. Kyle J. Urinary complications of Crohn's disease. World J Surg. 1980; 4:153–160. [PubMed: 7405253]
- 57. Neish AS. Microbes in gastrointestinal health and disease. Gastroenterology. 2009; 136:65–80. [PubMed: 19026645]
- Marks DJ, Mitchison NA, Segal AW, Sieper J. Can unresolved infection precipitate autoimmune disease? Curr Top Microbiol Immunol. 2006; 305:105–125. [PubMed: 16724803]
- Duerr RH, Taylor KD, Brant SR, Rioux JD, Silverberg MS, Daly MJ, Steinhart AH, Abraham C, Regueiro M, Griffiths A, et al. A genome-wide association study identifies IL23R as an inflammatory bowel disease gene. Science. 2006; 314:1461–1463. [PubMed: 17068223]
- Iwakura Y, Ishigame H. The IL-23/IL-17 axis in inflammation. J Clin Invest. 2006; 116:1218– 1222. [PubMed: 16670765]
- Izcue A, Hue S, Buonocore S, Arancibia-Carcamo CV, Ahern PP, Iwakura Y, Maloy KJ, Powrie F. Interleukin-23 restrains regulatory T cell activity to drive T cell-dependent colitis. Immunity. 2008; 28:559–570. [PubMed: 18400195]
- 62. Mizoguchi A, Ogawa A, Takedatsu H, Sugimoto K, Shimomura Y, Shirane K, Nagahama K, Nagaishi T, Mizoguchi E, Blumberg RS, et al. Dependence of intestinal granuloma formation on unique myeloid DC-like cells. J Clin Invest. 2007; 117:605–615. [PubMed: 17318261]
- 63. Fisher SA, Tremelling M, Anderson CA, Gwilliam R, Bumpstead S, Prescott NJ, Nimmo ER, Massey D, Berzuini C, Johnson C, et al. Genetic determinants of ulcerative colitis include the ECM1 locus and five loci implicated in Crohn's disease. Nat Genet. 2008; 40:710–712. [PubMed: 18438406]
- 64. ten Hove T, van Montfrans C, Peppelenbosch MP, van Deventer SJ. Infliximab treatment induces apoptosis of lamina propria T lymphocytes in Crohn's disease. Gut. 2002; 50:206–211. [PubMed: 11788561]
- Ricciardelli I, Lindley KJ, Londei M, Quaratino S. Anti tumour necrosis-alpha therapy increases the number of FOXP3 regulatory T cells in children affected by Crohn's disease. Immunology. 2008; 125:178–183. [PubMed: 18422560]
- 66. Gonvers JJ, Juillerat P, Mottet C, Felley C, Burnand B, Vader JP, Michetti P, Froehlich F. Maintenance of remission in Crohn's disease. Digestion. 2005; 71:41–48. [PubMed: 15711049]
- 67 ••. Smith AM, Rahman FZ, Hayee BH, Graham SJ, Marks DJ, Sewell GW, Palmer CD, Wilde J, Foxwell BM, Gloger IS, et al. Disordered macrophage cytokine secretion underlies impaired acute inflammation and bacterial clearance in Crohn's disease. J Exp Med. in press. In this study, impaired clearance of bacteria was demonstrated in CD patients, which was related to defects in the innate immune system. Macrophages cultured from CD patients were found to secrete deficient levels of pro-inflammatory cytokines in response to stimulation with heat killed *E. coli*. This was shown to arise from abnormal routing of cytokines to the lysosomal compartments in these cells, which results in degradation rather than release through the normal secretory pathway.



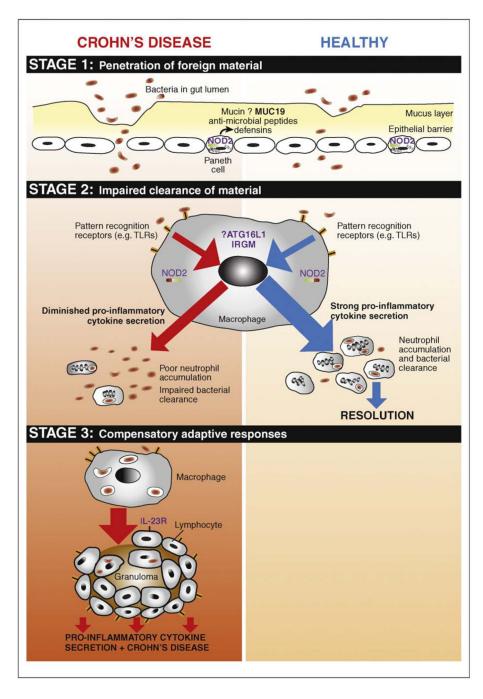


Figure 1.

The immunopathogenesis of CD occurs in three temporally distinct stages. Penetration of luminal contents into underlying tissues occurs in stage 1, which may be facilitated by environmental factors such as infection, or inherent defects in the mucosal barrier. In healthy individuals, resident macrophages secrete pro-inflammatory cytokines in response to this material, resulting in neutrophil accumulation, clearance of the material, and thereby resolution. In CD patients, defective secretion of pro-inflammatory cytokines by macrophages results in impaired neutrophil influx and clearance of foreign material (stage

2). Subsequently, chronic inflammatory responses (stage 3) will be triggered, giving rise to the characteristic features of the CD lesion.

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