

Oxidative stress and the unfulfilled promises of antioxidant agents

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Abstract

It is well known that aging and its associated diseases, including cancer, are triggered by oxidative damage to biological macromolecules. However, antioxidant compounds are still disappointingly distant from any clinical application, so that Jim Watson has declared that antioxidant supplementation may have caused more cancers than it has prevented Watson J ((2013) **Oxidants, antioxidants and the current incurability of metastatic cancers** *Open Biol* **3** DOI: 10.1098/rsob.120144).

To clarify this paradox, here, we describe the mechanisms of oxidative stress focusing in particular on redox balance and physiological oxidative signals.

Keywords: *oxidative stress, reactive oxygen species, mitochondrial respiration, redox signalling, antioxidant, cancer*

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What is oxidative stress?

Oxidative stress indicates a condition occurring when oxidising substances accumulate and accidental oxidative reactions thrive. In mammals, such as in all aerobic eukaryotes, the molecules with substantial oxidising potential contain oxygen. In particular, both reactive oxygen species (ROS), including singlet O_2 , superoxide anion (O_2^-), hydrogen peroxide (H_2O_2), and hydroxyl radical (OH^-), and reactive nitrogen species (RNS), including peroxynitrite ($ONOO^-$), nitrogen dioxide (NO_2), and dinitrogen trioxide (N_2O_3), are potent oxidising agents in living organisms [2].

In cells, the accumulation of ROS and, subsequently, RNS increases the chance of formation of oxidative modifications in proteins, resulting in protein carbonyl content and oxidised- or nitro-modified residues, in lipids, generating hydroperoxide lipid derivatives, and in both purines and pyrimidines, inducing DNA adducts and breaks. Ultimately, the rise of oxidative attacks to biological macromolecules leads to the dysfunctions of proteins, membranes, and nucleic acids.

How do antioxidants work?

Antioxidants are substances that can neutralise ROS and RNS by accepting or donating electrons and, as a consequence, might be converted into a radical form containing unpaired electrons that are less reactive than the neutralised ROS or RNS. Then, another antioxidant molecule may regenerate the paired form of the radical antioxidant. Loosely held hydrogen, large aromatic structures, and unsaturated bonds are all potent electron acceptor sites able to neutralise the free radicals present in antioxidant molecules [3]. Several plant metabolites contain these antioxidant moieties, for example, the large class of polyphenol compounds, including quercetin, myricetin, catechins, anthocyanins, that are present in food sources [4].

Together with these plant derivatives, a variety of different antioxidants are available for clinical purposes. These include endogenous molecules, such as α -tocopherol, β -carotene, glutathione, ascorbic acid, adenosine, lipoic acid, coenzyme Q, lactoferrin, as well as synthetic antioxidants, such as thiols, ebselen (selenium-based peroxide scavenger) [5], idebenone (coenzyme Q-analogue) and the more recently developed mitoQ (mitochondrial-targeted ubiquinone) [6], or α -tocopheryl-succinate nanoparticles [7]. Notably, several cardioprotective drugs, such as probucol and carvediol, or neuroprotective drugs, such as edaravone, are potent and effective antioxidant against the overload of ROS upon ischemia/reperfusion [8].

Other compounds, often included in the category of antioxidants, increase instead the endogenous levels of endogenous antioxidants such as *n*-acetyl cysteine (precursor of glutathione) or inhibit ROS production from cellular oxidases (allopurinol) or from metal ion reactions as in the case of iron chelators (deferrioxamine) [3].

Antioxidant failure in clinical studies

The present day is a golden age for antioxidants, promoted by the popularity of the free-radical theory of aging. Eating food rich in antioxidants protects from cancer and heart disease is common in particular.

Since the last three decades, the establishment of pre-clinical evidence showing that antioxidants protect deoxyribo nucleic acid (DNA) from being damaged by oxygen free radicals, potentially preventing the genetic mutations that cause cancer [9]. Antioxidants have also been consistently shown to reduce oxidative damage to low-density lipoprotein (LDL) cholesterol inside atherosclerotic plaques, thus protecting against atherosclerosis in the walls of arteries [10]. Thus, antioxidant treatments was promising to increase longevity by defeating the putative major cause of aging, that is, oxidative stress, or, more specifically by fighting top killers such as cancer and cardiovascular disease.

Unfortunately, several large randomised clinical trials found that antioxidant supplementation does not reduce the risk of cardiovascular disease or cancer, whereas one antioxidant, β -carotene, actually appears to increase the risk of some types of cancer in smokers. From 1985, the α -tocopherol/ β -carotene cancer prevention trial (ATBC) was the first large study to examine the effect of antioxidants. Aged male smokers who had assumed 20 mg β -carotene for 8 years had an 18% increase in lung cancer incidence, and a less significant increase in prostate cancer, with respect to the placebo group [11]. In the following years, the beneficial effects of supplementation with β -carotene or vitamin A, α -tocopherol, ascorbic acid, and selenium increasingly became the object of debate [12–17].

Presently, after reviewing all the information, the US Preventive Services Task Force recommends against β -carotene or vitamin E to prevent cancer and cardiovascular disease (<http://www.uspreventiveservicestaskforce.org>) [18].

Furthermore, regardless of the fact that some epidemiological studies have overall shown that vegetable-enriched diets, while increasing antioxidant intake [19], inversely relate to mortality [20] or cancer [21] and stroke [22] risks, and that, in animal models, antioxidant-rich foods appears to inhibit tumorigenesis [23] and to be cardioprotective [24], both the US food and drug administration [25] and the European food safety authority [26] have banned any writing that could imply potential health benefits on the package labels of products with antioxidants.

Actually, the cancer prevention recommendations of eating, mostly, food of plant origin indicated by international health organisations such as the World Health Organisation (www.who.int/dietphysicalactivity/whatworks) or the World Cancer Research Fund International [27] are not related to vegetable antioxidant supply.

Insight into ROS metabolism

The negative effect of oxygen on living organisms has been known for a long time [28]. However, although a negative correlation was observed in vertebrates between the intracellular levels of ROS/oxidative stress and longevity, the levels of endogenous antioxidants were also found to anti-correlate with life span. The supplementation of antioxidants in animal models, including frogs, pigs, rats and mice, did not affect mortality [29]. In actual fact, oxidative stress is determined by the rates of both ROS production and scavenging. Thus, the negative effect of aerobic metabolism could be only partially balanced by antioxidant activities if ROS production is maintained.

ROS are usually considered as a side effect of aerobic metabolism, and mitochondrial respiration is thought to be the main intracellular source of accidental ROS [30]. During mitochondrial respiration, electrons are extracted from nicotinamide adenine dinucleotide (NADH) or succinate and are then transferred to O_2 through a chain of enzymatic complexes. In the final step of this electron-transfer chain (ETC), the cytochrome c oxidase (complex IV) catalyses the full reduction of molecular O_2 to water, without forming O_2 radicals. However, partial reduction of O_2 leading to the formation of O_2^- can occur if O_2 hits sensible reduced sites of the ETC upstream of complex IV [31, 32]. Experimental data indicate that ROS are indeed continuously produced during mitochondrial respiration and that up to 2% of the total O_2 consumption is converted to ROS [30].

Cells are normally able to defend themselves against ROS damage through the use of specific enzymatic (dismutases, catalase, peroxidases) or non-enzymatic (A, C, and E vitamins, uric acid, bilirubin) ROS-reducing mechanisms. The O_2^- dismutase enzyme, for instance, catalyses the conversion of O_2^- into H_2O_2 , which is, in turn, reduced to water by the glutathione peroxidase and the catalase enzymes. In this way, the levels of different ROS are lowered to avoid the excessive oxidation of cellular components [30].

The rate of aging is assumed to be influenced, at least in part, by the rate of ROS production rather than by the rate of ROS scavenging. However, increased ROS production during aging is controversial; in contrast, scavenging activity has been clearly found to decrease over a lifetime and in different degenerative diseases [33–35].

Oxidative stress has a physiological role

Substantial evidence demonstrates that ROS have a physiological role regardless of their toxicity. In fact, ROS have been shown to mediate growth factor/hormone/cytokine signal transduction, to regulate gene expression, and to determine programmed cell death [36, 37]. Among ROS, H_2O_2 is diffusible, less reactive and longer-lived than, for instance, O_2^- and OH^- . H_2O_2 is especially involved in the regulation of intracellular signalling pathways and could be considered as a second messenger [37].

Since the rate of mitochondrial ROS formation depends on the local concentrations of O_2 and energetic substrates, and on ATP cellular demands [38, 39], the emerging picture is that mitochondria generate ROS in a regulated manner to deal with the different metabolic activities of the cell [40] and/or hypoxic conditions [41, 42].

ROS do not only cause irreversible damage to cellular components, they can also lead to fully reversible protein modifications. In particular, H_2O_2 has been demonstrated to directly oxidise cysteinyl thiols inducing formation of disulphide bonds and sulphenic acids. It has also been

shown to induce glutathionylation of cysteine residues or the formation of methionine sulphoxide on methionine residues in a variety of contexts, such as the transcription factors OxyR and Pap1 in bacteria and in the yeast, respectively, the Kinase Sty1 in the yeast, the vacuolar ATPase, Vatp, in plants, the HIV-2 protease in viruses, the arylamine N-acetyl transferase 1, NAT1, the indoleamine 2, 3-dioxygenase, the phospholipase A2, iPLA2Beta, the small ubiquitin-related modifiers SUMO E1 subunit Uba2 and SUMO E2-conjugating enzymes Ubc9, the phosphatases PTP1B (protein tyrosine phosphatase 1B) and PTEN (phosphatase and tensin homologue), the peroxidase enzymes Prx I and II (both only cytosolic) and Prx III (cytosolic and mitochondrial), the annexin A2 protein, the heat shock factor 1 (HSF1) the mitochondrial enzymes aconitase and α -ketoglutarate dehydrogenase, and the subunits of the respiratory complex I, in mammals [43].

Overall, it has been demonstrated that oxidation and reduction of key cellular proteins participate in a redox-dependent regulation of cellular functions, including energy metabolism and response to stress. It has also become clear that intracellular signalling pathways can be activated by changes in intracellular metabolic redox reactions that involve O_2^- and H_2O_2 [44]. Early hypotheses had proposed that exposure to specific environmental factors can induce ROS accumulation, thus triggering abnormal ROS-signalling leading to increase proliferation and malignant transformation. Evidence for this was obtained from studies showing that carcinogen initiators and promoters, including ionising radiation and polycyclic aromatic hydrocarbons, increase ROS formation, which in turn favours tumorigenesis [45]. A clear mechanism of how H_2O_2 in particular, can favour proliferation has emerged from studies on the redox regulation of critical phosphatases involved in signal transduction from plasma membrane receptors, together with the findings that several growth factors, such as EGF or Insulin/IGF, trigger H_2O_2 production directly from their membrane receptors [51, 79].

In this context, the function of the p66Shc protein is representative. P66Shc is the largest of the three isoforms encoded by the *ShcA* locus and almost ubiquitously expressed in vertebrates; it functions to regulate intracellular ROS levels and mitochondrial apoptosis. Cytosolic p66Shc mediates activation of the membrane oxidase activity and suppresses catalase and MnSOD expressions [46]. Then, a fraction of p66Shc translocates within the mitochondrial inter-membrane space [47] upon specific stimuli, including pro-apoptotic stresses [48] or growth factor stimulation [49], and oxidises cytochrome c to form H_2O_2 [50], which in turn regulates mitochondrial [51], and cellular functions [49, 52]. Accordingly, cells from p66Shc null mice or p66Shc-depleted by RNAi have reduced ROS levels [49]; however, p66Shc null mice show normal tumour incidence [53] and increased mutation rate [54]. Notably, p66Shc deletion is counter-selected when mice are maintained in harsh settings that mimic conditions in the wild (in an open field in the cold and in competition for food), indicating that the pro-oxidant function of p66Shc is essential for fitness under stressing natural conditions but redundant in a protected environment [55].

The role of oxidative stress in cancer

Tumorigenesis is characterised by major alterations in energetic metabolism, O_2 consumption and ROS accumulation [2] which result in a change in the balance between reduced/oxidised species (redox balance) [56]. Changes in the cellular redox balance affect proliferation, migration, and survival of cancer cells contributing to disease progression [57–59]. Activated oncogenes, such as Myc [60], Bcl-2 [61] or Ras [62], have been reported to affect redox balance. For example, the expression of the oncogenic form of Ras was found to boost [63–65] or to reduce [66] the level of glutathione, depending on the cell line. Furthermore, a reducing environment associates with different types of cancer [67–68]. This “reducing” environment results from the relative concentration of all the oxidant and reducing species (redox species) that exist in the metabolic network [69]. As a consequence, high levels of ROS endogenous scavengers or treatment with antioxidants increase oncogenic transformation [70] or tumour progression [71, 72], whereas increasing oxidation has even been proposed as a therapeutic strategy for cancer [73–76]. Finally, the antioxidant activity of chemicals is considered hazardous for novel classes of anticancer drugs [77, 78].

Conclusion

Antioxidants affecting ROS levels and functions produce different outcomes since ROS have both deleterious and beneficial effects. Although ROS are a by-product of aerobic metabolism, several enzymatic systems have evolved to generate ROS on purpose. In cancer cells, ROS act as secondary messengers of oncogenic signalling pathways and can also induce cellular senescence and apoptosis. As a consequence, oxidative stress during cancer expansion and progression selects for clones with high antioxidant metabolism. Based on this, the failure of antioxidant treatments, as documented in several clinical trials, is not surprising. Antioxidant drugs are not sufficient to inhibit tumorigenesis or, even worst, they may accelerate it.

References

1. Watson J (2013) **Oxidants, antioxidants and the current incurability of metastatic cancers** *Open Biol* **3** DOI: [10.1098/rsob.120144](https://doi.org/10.1098/rsob.120144)
2. Halliwell B and Gutteridge JMC (2007) *Free Radicals in Biology and Medicine* 4th edn Oxford University Press Oxford
3. Cadenas E and Packer L (2002) *Hand Book of Antioxidants* (New York: Marcel Dekker Inc) Introduction, Chapters 1, 6
4. Halliwell B (2002) *Food Derived Antioxidant: How to Evaluate Their Importance in Food and In Vivo* In: Cadenas E, Packer L (ed) *Handbook of antioxidants* (New York: Marcel Inc) Chapter 1.1
5. Fortmann SP, Burda BU, Senger CA, Lin JS and Whitlock EP (2013) **Vitamin and mineral supplements in the primary prevention of cardiovascular disease and cancer: an updated systematic evidence review for the U.S. preventive services task force** *Ann Intern Med* **159** 824–834 PMID: [24217421](https://pubmed.ncbi.nlm.nih.gov/24217421/)
6. Kelso GF, Porteous CM, Coulter CV and Hughes G *et al* (2001) **Selective targeting of a redox-active ubiquinone to mitochondria within cells: antioxidant and antiapoptotic properties** *J Biol Chem* **276** 4588–4596 DOI: [10.1074/jbc.M009093200](https://doi.org/10.1074/jbc.M009093200)
7. Hama S and Kogure K (2014) **Nanoparticles consisting of tocopheryl succinate are a novel drug-delivery system with multifaceted antitumor activity** *Biol Pharm Bull* **37** 196–200 DOI: [10.1248/bpb.b13-00848](https://doi.org/10.1248/bpb.b13-00848) PMID: [24492715](https://pubmed.ncbi.nlm.nih.gov/24492715/)
8. Lefer DJ and Granger DN (2000) **Oxidative stress and cardiac disease** *Am J Med* **109** 315–323 DOI: [10.1016/S0002-9343\(00\)00467-8](https://doi.org/10.1016/S0002-9343(00)00467-8) PMID: [10996583](https://pubmed.ncbi.nlm.nih.gov/10996583/)
9. Ames BN (1983) **Dietary carcinogens and anticarcinogens Oxygen radicals and degenerative diseases** *Science* **221** 1256–1264 DOI: [10.1126/science.6351251](https://doi.org/10.1126/science.6351251) PMID: [6351251](https://pubmed.ncbi.nlm.nih.gov/6351251/)
10. Morel DW, DiCorleto PE and Chisolm GM (1984) **Endothelial and smooth muscle cells alter low density lipoprotein in vitro by free radical oxidation** *Arteriosclerosis* **4** 357–364 DOI: [10.1161/01.ATV.4.4.357](https://doi.org/10.1161/01.ATV.4.4.357) PMID: [6466193](https://pubmed.ncbi.nlm.nih.gov/6466193/)
11. The Alpha-Tocopherol, Beta Carotene Cancer Prevention Study Group (1994) The effect of vitamin E and beta carotene on the incidence of lung cancer and other cancers in male smokers *N Engl J Med* **330** 1029–1235 DOI: [10.1056/NEJM199404143301501](https://doi.org/10.1056/NEJM199404143301501)
12. Lawlor DA, Smith GD, Bruckdorfer KR, Kundu D and Ebrahim S (2004) **Those confounded vitamins: what can we learn from the differences between observational versus randomised trial evidence?** *Lancet* **363** 1724–1727 DOI: [10.1016/S0140-6736\(04\)16260-0](https://doi.org/10.1016/S0140-6736(04)16260-0) PMID: [15158637](https://pubmed.ncbi.nlm.nih.gov/15158637/)
13. Bjelakovic G, Nikolova D, Gluud LL, Simonetti RG and Gluud C (2007) **Mortality in randomized trials of antioxidant supplements for primary and secondary prevention: systematic review and meta-analysis** *JAMA* **297** 842–857 DOI: [10.1001/jama.297.8.842](https://doi.org/10.1001/jama.297.8.842) PMID: [17327526](https://pubmed.ncbi.nlm.nih.gov/17327526/)
14. Greenlee H, Hershman DL and Jacobson JS (2009) **Use of antioxidant supplements during breast cancer treatment: a comprehensive review** *Breast Cancer Res Treat* **115** 437–452 DOI: [10.1007/s10549-008-0193-0](https://doi.org/10.1007/s10549-008-0193-0)
15. Lippman SM *et al* (2009) **Effect of selenium and vitamin E on risk of prostate cancer and other cancers: the selenium and vitamin E cancer prevention trial (SELECT)** *JAMA* **301** 39–51 DOI: [10.1001/jama.2008.864](https://doi.org/10.1001/jama.2008.864)
16. Bjelakovic G, Nikolova D, Gluud LL, Simonetti RG and Gluud C (2012) **Antioxidant supplements for prevention of mortality in healthy participants and patients with various diseases** *Cochrane Database Syst Rev* **3** CD007176 PMID: [22419320](https://pubmed.ncbi.nlm.nih.gov/22419320/)
17. Bjelakovic G, Nikolova D, Gluud LL, Simonetti RG and Gluud C (2013) **Meta-regression analyses, meta-analyses, and trial sequential analyses of the effects of supplementation with beta-carotene, vitamin A, and vitamin E singly or in different combinations on all-cause mortality: do we have evidence for lack of harm?** *PLoS One* **8** e74558 DOI: [10.1371/journal.pone.0074558](https://doi.org/10.1371/journal.pone.0074558) PMID: [24040282](https://pubmed.ncbi.nlm.nih.gov/24040282/) PMCID: [3765487](https://pubmed.ncbi.nlm.nih.gov/3765487/)

18. Coulter I, Hardy M and Shekelle P *et al* (2003) Effect of the Supplemental Use of Antioxidants Vitamin C, Vitamin E, and Coenzyme Q10 for the Prevention and Treatment of Cancer Evidence Report/Technology Assessment Number 75. (Prepared by Southern California Evidence-based Practice Center under Contract No. 290-97-0001) AHRQ Publication No. 04-E003 (Rockville, MD: Agency for Healthcare Research and Quality)
19. Park SY, Ollberding NJ and Woolcott CG *et al* (2013) **Fruit and vegetable intakes are associated with lower risk of bladder cancer among women in the multiethnic cohort study** *J Nutr* **143** 1283–1292 DOI: [10.3945/jn.113.174920](https://doi.org/10.3945/jn.113.174920) PMID: [23739308](https://pubmed.ncbi.nlm.nih.gov/23739308/) PMCID: [3709993](https://pubmed.ncbi.nlm.nih.gov/3709993/)
20. Masala G, Assedi M and Bendinelli B *et al* **Fruit and vegetables consumption and breast cancer risk: the EPIC Italy study** *Breast Cancer Res Treat* **132** 1127–1136 PMID: [22215387](https://pubmed.ncbi.nlm.nih.gov/22215387/)
21. Genkinger JM, Platz EA, Hoffman SC, Comstock GW and Helzlsouer KJ (2004) **Fruit, vegetable, and antioxidant intake and all-cause, cancer, and cardiovascular disease mortality in a community-dwelling population in Washington County, Maryland** *Am J Epidemiol* **160** 1223–1233 DOI: [10.1093/aje/kwh339](https://doi.org/10.1093/aje/kwh339) PMID: [15583375](https://pubmed.ncbi.nlm.nih.gov/15583375/)
22. Wang X, Ouyang Y and Liu J *et al* (2014) **Fruit and vegetable consumption and mortality from all causes, cardiovascular disease, and cancer: systematic review and dose-response meta-analysis of prospective cohort studies** *BMJ* **349** g4490 DOI: [10.1136/bmj.g4490](https://doi.org/10.1136/bmj.g4490) PMID: [25073782](https://pubmed.ncbi.nlm.nih.gov/25073782/) PMCID: [4115152](https://pubmed.ncbi.nlm.nih.gov/4115152/)
23. Butelli E, Titta L and Giorgio M *et al* (2008) **Enrichment of tomato fruit with health-promoting anthocyanins by expression of select transcription factors** *Nat Biotechnol* **26** 1301–1308 DOI: [10.1038/nbt.1506](https://doi.org/10.1038/nbt.1506) PMID: [18953354](https://pubmed.ncbi.nlm.nih.gov/18953354/)
24. Testai L, Martelli A and Cristofaro M *et al* (2013) **Cardioprotective effects of different flavonoids against myocardial ischaemia/reperfusion injury in Langendorff-perfused rat hearts** *J Pharm Pharmacol* **65** 750–756 DOI: [10.1111/jphp.12032](https://doi.org/10.1111/jphp.12032) PMID: [23600393](https://pubmed.ncbi.nlm.nih.gov/23600393/)
25. **Guidance for industry, food labeling; nutrient content claims; definition for “high potency” and definition for “antioxidant” for use in nutrient content claims for dietary supplements and conventional foods** U.S. department of health and human services, food and drug administration Center for food safety and applied nutrition June 2008
26. **Scientific opinion on the substantiation of health claims related to various food(s)/food constituent(s) and protection of cells from premature aging, antioxidant activity, antioxidant content and antioxidant properties, and protection of DNA, proteins and lipids from oxidative damage pursuant to Article 13(1) of Regulation (EC) No 1924/2006** EFSA panel on dietetic products, nutrition and allergies (NDA)2, 3 European food safety authority (EFSA), Parma, Italy, *EFSA J* 2010 **8** 1489
27. World Cancer Research Fund/American Institute for Cancer Research (2007) *Food, Nutrition, Physical Activity, and the Prevention of Cancer: A Global Perspective* Washington DC: AICR
28. Lorrain SJ (1899) **The pathological effects due to increase of oxygen tension in the air breathed** *J Physiol* **24** 19 DOI: [10.1113/jphysiol.1899.sp000746](https://doi.org/10.1113/jphysiol.1899.sp000746)
29. Barja G (2004) **Aging in vertebrates, and the effect of caloric restriction: a mitochondrial free radical production-DNA damage mechanism?** *Biol Rev Camb Philos Soc* **79** 235–251 DOI: [10.1017/S1464793103006213](https://doi.org/10.1017/S1464793103006213) PMID: [15191224](https://pubmed.ncbi.nlm.nih.gov/15191224/)
30. Chance B, Sies H and Boveris A (1979) **Hydroperoxide metabolism in mammalian organs** *Physiol Rev* **59** 527–605 PMID: [37532](https://pubmed.ncbi.nlm.nih.gov/37532/)
31. Turrens JF (2003) **Mitochondrial formation of reactive oxygen species** *J Physiol* **552** 335–344 DOI: [10.1113/jphysiol.2003.049478](https://doi.org/10.1113/jphysiol.2003.049478) PMID: [14561818](https://pubmed.ncbi.nlm.nih.gov/14561818/) PMCID: [2343396](https://pubmed.ncbi.nlm.nih.gov/2343396/)
32. Brand MD (2010) **The sites and topology of mitochondrial superoxide production** *Exp Gerontol* **45** 466–472 DOI: [10.1016/j.exger.2010.01.003](https://doi.org/10.1016/j.exger.2010.01.003) PMID: [20064600](https://pubmed.ncbi.nlm.nih.gov/20064600/) PMCID: [2879443](https://pubmed.ncbi.nlm.nih.gov/2879443/)
33. Frisard M and Ravussinm E (2006) **Energy metabolism and oxidative stress: impact on the metabolic syndrome and the aging process** *Endocrine* **29** 27–32 DOI: [10.1385/ENDO:29:1:27](https://doi.org/10.1385/ENDO:29:1:27) PMID: [16622290](https://pubmed.ncbi.nlm.nih.gov/16622290/)

34. Harman D (1998) **Aging: phenomena and theories** *Ann N Y Acad Sci* **854** 1–7 DOI: [10.1111/j.1749-6632.1998.tb09886.x](https://doi.org/10.1111/j.1749-6632.1998.tb09886.x)
35. Balaban RS, Nemoto S and Finkel T (2005) **Mitochondria, oxidants, and aging** *Cell* **120** 483–495 DOI: [10.1016/j.cell.2005.02.001](https://doi.org/10.1016/j.cell.2005.02.001) PMID: [15734681](https://pubmed.ncbi.nlm.nih.gov/15734681/)
36. Orrenius S, Gogvadze V and Zhivotovsky B **Mitochondrial oxidative stress: implications for cell death** *Annu Rev Pharmacol Toxicol* **47**(2007) 143–183 PMID: [17029566](https://pubmed.ncbi.nlm.nih.gov/17029566/)
37. Stone JR and Yang S (2006) **Hydrogen peroxide: a signaling messenger** *Antioxid Redox Signal* **8** 243–270 DOI: [10.1089/ars.2006.8.243](https://doi.org/10.1089/ars.2006.8.243) PMID: [16677071](https://pubmed.ncbi.nlm.nih.gov/16677071/)
38. Hoffman DL and Brookes PSJ (2009) **Oxygen sensitivity of mitochondrial reactive oxygen species generation depends on metabolic conditions** *Biol Chem* **284** 16236–16245 DOI: [10.1074/jbc.M809512200](https://doi.org/10.1074/jbc.M809512200)
39. Murphy MP (2009) **How mitochondria produce reactive oxygen species** *Biochem J* **417** 1–13 DOI: [10.1042/BJ20081386](https://doi.org/10.1042/BJ20081386)
40. Marcu R, Rapino S and Trinei M *et al* (2012) **Electrochemical study of hydrogen peroxide formation in isolated mitochondria** *Bioelectrochemistry* **85** 21–28 DOI: [10.1016/j.bioelechem.2011.11.005](https://doi.org/10.1016/j.bioelechem.2011.11.005)
41. Hoffman DL, Salter JD and Brookes PS (2007) **Response of mitochondrial reactive oxygen species generation to steady-state oxygen tension: implications for hypoxic cell signaling** *Am J Physiol Heart Circ Physiol* **292** H101–H108 DOI: [10.1152/ajp-heart.00699.2006](https://doi.org/10.1152/ajp-heart.00699.2006)
42. Bell EL, Klimova TA and Eisenbart J *et al* (2007) **The Qo site of the mitochondrial complex III is required for the transduction of hypoxic signaling via reactive oxygen species production** *Cell Biol* **177** 1029–1036 DOI: [10.1083/jcb.200609074](https://doi.org/10.1083/jcb.200609074)
43. Giorgio M, Trinei M and Migliaccio E *et al* (2007) **Hydrogen peroxide: a metabolic by-product or a common mediator of ageing signals?** *Nat Rev Mol Cell Biol* **8** 722–728 DOI: [10.1038/nrm2240](https://doi.org/10.1038/nrm2240) PMID: [17700625](https://pubmed.ncbi.nlm.nih.gov/17700625/)
44. Valko M, Leibfritz D and Moncol J *et al* (2007) **Free radicals and antioxidants in normal physiological functions and human disease** *Int J Biochem Cell Biol* **39** 44–84 DOI: [10.1016/j.biocel.2006.07.001](https://doi.org/10.1016/j.biocel.2006.07.001)
45. Gius D and Spitz DR (2006) **Redox signaling in cancer biology** *Antioxid Redox Signal* **8** 1249–1252 DOI: [10.1089/ars.2006.8.1249](https://doi.org/10.1089/ars.2006.8.1249) PMID: [16910772](https://pubmed.ncbi.nlm.nih.gov/16910772/)
46. Trinei M, Berniakovich I and Beltrami E *et al* (2009) **P66Shc signals to age** *Giorgio Aging* **1** 503–585
47. Orsini F, Moroni M and Contursi C *et al* (2006) **Regulatory effects of the mitochondrial energetic status on mitochondrial p66Shc** *Biol Chem* **387** 1405–1410 DOI: [10.1515/BC.2006.176](https://doi.org/10.1515/BC.2006.176) PMID: [17081113](https://pubmed.ncbi.nlm.nih.gov/17081113/)
48. Pinton P, Rimessi A and Marchi S *et al* (2007) **Protein kinase C beta and prolyl isomerase 1 regulate mitochondrial effects of the life-span determinant p66Shc** *Science* **315** 659–663 DOI: [10.1126/science.1135380](https://doi.org/10.1126/science.1135380) PMID: [17272725](https://pubmed.ncbi.nlm.nih.gov/17272725/)
49. Berniakovich I, Trinei M and Stendardo M *et al* (2008) **P66Shc-generated oxidative signal promotes fat accumulation** *J Biol Chem* **283** 34283–34293 DOI: [10.1074/jbc.M804362200](https://doi.org/10.1074/jbc.M804362200) PMID: [18838380](https://pubmed.ncbi.nlm.nih.gov/18838380/) PMID: [2590696](https://pubmed.ncbi.nlm.nih.gov/2590696/)
50. Trinei M, Migliaccio E and Bernardi P (2013) **P66Shc, mitochondria, and the generation of reactive oxygen species** *Methods Enzymol* **528** 99–110 DOI: [10.1016/B978-0-12-405881-1.00006-9](https://doi.org/10.1016/B978-0-12-405881-1.00006-9) PMID: [23849861](https://pubmed.ncbi.nlm.nih.gov/23849861/)
51. Trinei M, Berniakovich I and Pelicci PG *et al* (2006) **Mitochondrial DNA copy number is regulated by cellular proliferation: a role for Ras and p66(Shc)** *Biochim Biophys Acta* **1757** 624–630 DOI: [10.1016/j.bbabi.2006.05.029](https://doi.org/10.1016/j.bbabi.2006.05.029) PMID: [16829231](https://pubmed.ncbi.nlm.nih.gov/16829231/)
52. Frijhoff J, Dagnell M and Augsten M *et al* (2014) **The mitochondrial reactive oxygen species regulator p66Shc controls PDGF-induced signaling and migration through protein tyrosine phosphatase oxidation** *Free Radic Biol Med* **68** 268–277 DOI: [10.1016/j.freeradbiomed.2013.12.022](https://doi.org/10.1016/j.freeradbiomed.2013.12.022) PMID: [24378437](https://pubmed.ncbi.nlm.nih.gov/24378437/)

53. Beltrami E, Valtorta S and Moresco R *et al* (2013) **The p53-p66Shc apoptotic pathway is dispensable for tumor suppression whereas the p66Shc-generated oxidative stress initiates tumorigenesis** *Curr Pharm Des* **19** 2708–2714 DOI: [10.2174/1381612811319150005](https://doi.org/10.2174/1381612811319150005)
54. Beltrami E, Ruggiero A and Busuttill R *et al* (2013) **Deletion of p66Shc in mice increases the frequency of size-change mutations in the lacZ transgene** *Aging Cell* **12** 177–183 DOI: [10.1111/accel.12036](https://doi.org/10.1111/accel.12036)
55. Giorgio M, Berry A and Berniakovich I *et al* (2012) **The p66(Shc) knocked out mice are short lived under natural condition** *Aging Cell* **11** 162–168 DOI: [10.1111/j.1474-9726.2011.00770.x](https://doi.org/10.1111/j.1474-9726.2011.00770.x)
56. Gupta SC, Hevia D and Patchva S *et al* (2012) **Upsides and downsides of reactive oxygen species for cancer: the roles of reactive oxygen species in tumorigenesis, prevention, and therapy** *Antioxid Redox Signal* **16** 1295–1322 DOI: [10.1089/ars.2011.4414](https://doi.org/10.1089/ars.2011.4414) PMID: [3324815](https://pubmed.ncbi.nlm.nih.gov/3324815/)
57. Burhans WC and Heintz NH (2009) **The cell cycle is a redox cycle: linking phase-specific targets to cell fate** *Free Radic Biol Med* **47** 1282–1293 DOI: [10.1016/j.freeradbiomed.2009.05.026](https://doi.org/10.1016/j.freeradbiomed.2009.05.026) PMID: [19486941](https://pubmed.ncbi.nlm.nih.gov/19486941/)
58. Hitchler MJ and Domann FE (2012) **Redox regulation of the epigenetic landscape in cancer: a role for metabolic reprogramming in remodeling the epigenome** *Free Radic Biol Med* **53** 2178–2187 DOI: [10.1016/j.freeradbiomed.2012.09.028](https://doi.org/10.1016/j.freeradbiomed.2012.09.028) PMID: [23022407](https://pubmed.ncbi.nlm.nih.gov/23022407/) PMID: [3508253](https://pubmed.ncbi.nlm.nih.gov/3508253/)
59. Ward PS and Thompson CB (2012) **Metabolic reprogramming: a cancer hallmark even warburg did not anticipate** *Cancer Cell* **21** 297–308 DOI: [10.1016/j.ccr.2012.02.014](https://doi.org/10.1016/j.ccr.2012.02.014) PMID: [22439925](https://pubmed.ncbi.nlm.nih.gov/22439925/) PMID: [3311998](https://pubmed.ncbi.nlm.nih.gov/3311998/)
60. Egler RA, Fernandes E and Rothermund K *et al* (2005) **Regulation of reactive oxygen species, DNA damage, and c-Myc function by peroxiredoxin 1** *Oncogene* **24** 8038–8050 DOI: [10.1038/sj.onc.1208821](https://doi.org/10.1038/sj.onc.1208821) PMID: [16170382](https://pubmed.ncbi.nlm.nih.gov/16170382/)
61. Low IC, Kang J and Pervaiz S (2011) **Bcl-2: a prime regulator of mitochondrial redox metabolism in cancer cells** *Antioxid Redox Signal* **15** 2975–2987 DOI: [10.1089/ars.2010.3851](https://doi.org/10.1089/ars.2010.3851) PMID: [21574773](https://pubmed.ncbi.nlm.nih.gov/21574773/)
62. Armeni T, Ercolani L and Urbanelli L (2012) **Cellular redox imbalance and changes of protein S-glutathionylation patterns are associated with senescence induced by oncogenic H-ras** *PLoS One* **7** e52151 DOI: [10.1371/journal.pone.0052151](https://doi.org/10.1371/journal.pone.0052151)
63. Chuang JIM, Chang TY and Liu HS (2003) **Glutathione depletion-induced apoptosis of Ha-ras-transformed NIH3T3 cells can be prevented by melatonin** *Oncogene* **22** 1349–1357 DOI: [10.1038/sj.onc.1206289](https://doi.org/10.1038/sj.onc.1206289) PMID: [12618760](https://pubmed.ncbi.nlm.nih.gov/12618760/)
64. Recktenwald CV, Kellner R and Lichtenfels R (2008) **Altered detoxification status and increased resistance to oxidative stress by K-ras transformation** *Cancer Res* **68** 10086–10093 DOI: [10.1158/0008-5472.CAN-08-0360](https://doi.org/10.1158/0008-5472.CAN-08-0360) PMID: [19074874](https://pubmed.ncbi.nlm.nih.gov/19074874/)
65. Vincenzini MT, Marraccini P and Iantomasi T *et al* (1993) **Altered metabolism of glutathione in cells transformed by oncogenes which cause resistance to ionizing radiations** *FEBS Lett* **320** 219–223 DOI: [10.1016/0014-5793\(93\)80590-Q](https://doi.org/10.1016/0014-5793(93)80590-Q) PMID: [8096467](https://pubmed.ncbi.nlm.nih.gov/8096467/)
66. Choudhary S, Rathore K and Wang HC (2011) **Differential induction of reactive oxygen species through Erk1/2 and Nox-1 by FK228 for selective apoptosis of oncogenic H-Ras-expressing human urinary bladder cancer J82 cells** *J Cancer Res Clin Oncol* **137** 471–480 DOI: [10.1007/s00432-010-0910-z](https://doi.org/10.1007/s00432-010-0910-z)
67. Schumacker PT (2006) **Reactive oxygen species in cancer cells: live by the sword, die by the sword** *Cancer Cell* **10** 175–176 DOI: [10.1016/j.ccr.2006.08.015](https://doi.org/10.1016/j.ccr.2006.08.015) PMID: [16959608](https://pubmed.ncbi.nlm.nih.gov/16959608/)
68. Smart DK, Ortiz KL and Mattson D *et al* (2004) **Thioredoxin reductase as a potential molecular target for anticancer agents that induce oxidative stress** *Cancer Res* **64** 6716–6724 DOI: [10.1158/0008-5472.CAN-03-3990](https://doi.org/10.1158/0008-5472.CAN-03-3990) PMID: [15374989](https://pubmed.ncbi.nlm.nih.gov/15374989/)
69. Schafer FQ and Buettner GR (2001) **Redox environment of the cell as viewed through the redox state of the glutathione disulfide/glutathione couple** *Free Radic Biol Med* **30** 1191–1212 DOI: [10.1016/S0891-5849\(01\)00480-4](https://doi.org/10.1016/S0891-5849(01)00480-4) PMID: [11368918](https://pubmed.ncbi.nlm.nih.gov/11368918/)

70. DeNicola GM *et al* (2011) **Oncogene-induced Nrf2 transcription promotes ROS detoxification and tumorigenesis** *Nature* **475** 106–109 DOI: [10.1038/nature10189](https://doi.org/10.1038/nature10189) PMID: [21734707](https://pubmed.ncbi.nlm.nih.gov/21734707/) PMCID: [3404470](https://pubmed.ncbi.nlm.nih.gov/3404470/)
71. Schafer ST *et al* (2009) **Antioxidant and oncogene rescue of metabolic defects caused by loss of matrix attachment** *Nature* **461** 109–113 DOI: [10.1038/nature08268](https://doi.org/10.1038/nature08268) PMID: [19693011](https://pubmed.ncbi.nlm.nih.gov/19693011/) PMCID: [2931797](https://pubmed.ncbi.nlm.nih.gov/2931797/)
72. Sayin VI, Ibrahim MX and Larsson E *et al* (2014) **Antioxidants accelerate lung cancer progression in mice** *Sci Transl Med* **6** 221ra15 DOI: [10.1126/scitranslmed.3007653](https://doi.org/10.1126/scitranslmed.3007653) PMID: [24477002](https://pubmed.ncbi.nlm.nih.gov/24477002/)
73. Montero AJ and Jassem J (2011) **Cellular redox pathways as a therapeutic target in the treatment of cancer** *Drugs* **71** 1385–1396 DOI: [10.2165/11592590-000000000-00000](https://doi.org/10.2165/11592590-000000000-00000) PMID: [21812504](https://pubmed.ncbi.nlm.nih.gov/21812504/)
74. Gorrini C, Harris IS and Mak TW (2013) **Modulation of oxidative stress as an anticancer strategy** *Nat Rev Drug Discov* **12** 931–947 DOI: [10.1038/nrd4002](https://doi.org/10.1038/nrd4002) PMID: [24287781](https://pubmed.ncbi.nlm.nih.gov/24287781/)
75. Glasauer A, Sena LA and Diebold LP *et al* (2014) **Targeting SOD1 reduces experimental non-small-cell lung cancer** *J Clin Invest* **124** 117–128 DOI: [10.1172/JCI71714](https://doi.org/10.1172/JCI71714) PMCID: [3871252](https://pubmed.ncbi.nlm.nih.gov/3871252/)
76. Sullivan LB and Chandel NS (2014) **Mitochondrial reactive oxygen species and cancer** *Cancer Metab* **2** 17 DOI: [10.1186/2049-3002-2-17](https://doi.org/10.1186/2049-3002-2-17)
77. Neuzil J, Tomasetti M and Zhao Y *et al* (2007) **Vitamin E analogs, a novel group of “mitocans,” as anticancer agents: the importance of being redox-silent** *Mol Pharmacol* **71** 1185–1199 DOI: [10.1124/mol.106.030122](https://doi.org/10.1124/mol.106.030122) PMID: [17220355](https://pubmed.ncbi.nlm.nih.gov/17220355/)
78. Lawenda BD, Kelly KM and Ladas EJ *et al* (2008) **Should supplemental antioxidant administration be avoided during chemotherapy and radiation therapy?** *J Natl Cancer Inst* **100** 773–783 DOI: [10.1093/jnci/djn148](https://doi.org/10.1093/jnci/djn148) PMID: [18505970](https://pubmed.ncbi.nlm.nih.gov/18505970/)
79. Heneberg P (2014) **Reactive nitrogen species and hydrogen sulfide as regulators of protein tyrosine phosphatase activity** *Antioxid Redox Signal* **29** 2191–209 DOI: [10.1089/ars.2013.5493](https://doi.org/10.1089/ars.2013.5493) PMID: [24328688](https://pubmed.ncbi.nlm.nih.gov/24328688/)