

Characterization of the role of the photosensitizer, deuterporfin, in the detection of lymphatic metastases in a pancreatic cancer xenograft model

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Abstract. Currently, the use of photosensitizers as tracer agents to detect lymphatic metastases is a developing area of study in the field of pancreatic cancer treatment. In the present study, deuterporfin, a novel photosensitizer, was used as a tracer agent to detect lymphatic metastases in a pancreatic cancer xenograft model. The biodistribution and pharmacokinetics of deuterporfin, following intravenous administration and injection of deuterporfin into the left rear footpad, were investigated in Sprague-Dawley rats. The increased difference in deuterporfin concentration between the cancerous and normal tissues was directly observed through the application of a Wood's lamp. In addition, the highly lymphatic BxPC-3-LN5 human metastatic pancreatic cancer cell line was generated from BxPC-3 cells using a continuous screening and seeding method *in vivo*. A xenograft model of the BxPC-3-LN5 human pancreatic cancer cell line transplanted into the left rear footpad of nude mice, was established. The effects of deuterporfin as a tracer agent in the detection of lymphatic metastases were then characterized in the pancreatic cancer xenograft model. Following intravenous administration, deuterporfin was rapidly enriched in the pancreas and popliteal fossa lymph nodes compared with that of the left rear footpad administration group. In addition, deuterporfin appeared to be selectively enriched in the cancerous pancreatic lymph nodes of the pancreatic cancer xenograft model. These results indicated that deuterporfin may be developed as a novel photosensitizer tracer agent for the detection of lymphatic metastases in pancreatic cancer. The advantages of deuterporfin are that it has a selective tumor-targeting effect due to high tissue uptake, and that it may be administered intravenously and is therefore suitable for surgery.

Introduction

Pancreatic cancer represents the fourth most common cause of cancer-associated mortality in the western world (1,2). Despite significant progress in basic and clinical research, pancreatic cancer remains a significant therapeutic challenge to humans, with 1- and 5-year survival rates of just 20 and 6%, respectively (3,4). The difficult surgical approach, resistance to conventional therapies and subsequent low survival rates all contribute to the high frequency of occurrence of lymphatic metastases (5-7). Currently, the use of tracer agents in the detection of lymphatic metastases is a focus of attention, as it facilitates specification of treatment options for the given diagnosis. However, at present, there is no acceptable and ideal imaging modality or technique for the accurate detection of lymphatic metastases (8,9). For example, the use of isotope tracing is limited due to the potential side-effects of irradiation, as well as the vague surgical resolution provided by the blue dye (10,11). Computed tomography and conventional magnetic resonance imaging are not sufficiently reliable for the accurate detection of lymphatic metastases. The limitations of these tracing methods are due to the following two aspects: i) They possess no tumor targeting effect; and ii) the route of administration is typically through the lymphatic system, which is difficult for the treatment of pancreatic cancer as the pancreas is a retroperitoneal organ (12). Therefore, the development of a novel lymph node tracer and imaging method is required.

Photodynamic therapy (PDT), based on the photogeneration of highly cytotoxic singlet oxygen species and the subsequent induction of oxidative stress that results in the death of stained cells, is currently used in oncology (13). PDT is a minimally invasive treatment that damages target cells by inducing cytotoxicity via the generation of cytotoxic oxygen species. The PDT components, comprising the photosensitizer, light and oxygen, are individually non-toxic. However, exposure of the photosensitizer to light in the presence of oxygen induces the generation of highly reactive singlet oxygen (1O_2) species within the tumor tissue, which results in significant damage of the cells in proximity to the treated area (14-18). PDT is a widely accepted treatment strategy for numerous types of cancerous and precancerous lesion, including those in the bladder, brain, ovary and pancreas (19-21). However, most attention has been directed towards the therapeutic role of the photosensitizer, rather than its potential function as a tracer agent.

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Deuteporfin, one of the novel generation of photosensitizers, is particularly photosensitive to light with wavelengths of 400-700 nm. Chemically, the structure of deuteporfin is composed of four distinct sub-porphyrin derivatives: Methoxyethyl-hydroxyethyl-dipropionic, di-methoxyethyl-dipropionic, hydroxyethyl-vinyl-dipropionic and methoxyethyl-vinyl-dipropionic (Fig. 1). Compared with traditional lymph node tracers, deuteporfin has a significant advantage in terms of two aspects: i) Selective targeting of tumors due to high tissue uptake and ii) that it may be administered intravenously. The present study aimed to exploit these features of deuteporfin for use as a tracer agent, in order to detect lymphatic metastases in a pancreatic cancer xenograft model. The study was approved by the ethics committee of Fudan University, Shanghai, China.

Materials and methods

Reagents. Deuteporfin was obtained from the R&D Center of Fudan-Zhangjiang Bio-Pharmaceutical Co. Ltd. (Shanghai, China) and was used without further purification. Normal saline was purchased from Shanghai Baxter Healthcare Co., Ltd., Shanghai, China. Absolute ethanol, high glucose-Dulbecco's modified Eagle's medium (H-DMEM), fetal bovine serum (FBS), trypsin, EDTA, formalin, paraffin, and hematoxylin and eosin (H&E) were purchased from Sinopharm Chemical Reagent Co., Ltd. (Shanghai, China). In addition, deionized water was from Fudan University (Shanghai, China), and Tris-buffered saline and phosphate-buffered saline were prepared in our laboratory. All solvents and chemicals were of analytical grade.

Experimental animals. Sprague-Dawley (SD) rats and BALB/C-nu/nu nude mice were purchased from the Shanghai Laboratory Animal Co. Ltd. (Shanghai, China). The SD rats and BALB/c-nu/nu male mice were housed in a pathogen-free animal facility. The temperature was maintained at 23°C with a humidity of 50-60%, and the mice were subjected to a 10/14-h light/dark cycle. All mice were provided with food and water 3 times a week by professional technicians. All animal studies were in compliance with the approved animal protocols and the guidelines of the Institutional Animal Care and Use Committee of Fudan University. The BxPC-3 human pancreatic cancer cell line was obtained from the Shanghai Branch of the Chinese Academy of Science, Shanghai, China.

Cell culture. The highly lymphatic metastatic pancreatic cancer cell line was generated from BxPC-3 cells by a continuous screening and seeding method *in vivo*. Briefly, 1×10^7 BxPC-3 cells in 100 μ l normal saline were subcutaneously injected into the left rear footpad of male BALB/C-nu/nu nude mice (n=5; aged 6-8 weeks). Six weeks later, the mice were anesthetized by intravenous injection with chloral hydrate and sacrificed by cervical dislocation. The lymph nodes located in the left knee, groin, side of the iliac artery and porta renis (depression in the kidney) were then collected. All the conjunctive tissues were removed and the lymph nodes were gently crushed on a 200-mesh sieve (Sinopharm Chemical Reagent Co., Ltd.). The cells routinely cultured in H-DMEM containing 10% FBS in an incubator at a temperature of 37°C for 2-3 days. Subsequently, cells were digested by incubation

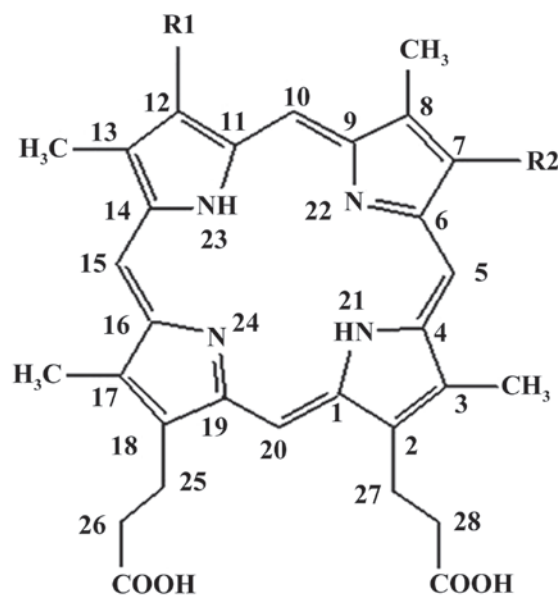


Figure 1. Chemical structure of deuteporfin. R1=CH₃CH (OCH₃) and R2=CH₃CHOH or R1=CH₃CHOH and R2=CH₃CH (OCH₃) or R1=CH₃CH (OCH₃) and R2=CH₃CH (OCH₃).

with 0.025% trypsin/0.002% EDTA and then reinoculated into another group of mice. This procedure was repeated 5 times (total mice, n=25), until the BxPC-3-LN5 subline was obtained, comprising BxPC-3 cells with an increased capacity to metastasize.

Pharmacokinetics and tissue distribution of deuteporfin *in vivo*. Seventy-two SD rats were randomly divided into two equal groups, comprising the caudal vein administration group (n=36) and the left rear footpad administration group (n=36). The rats were injected with 2.5 mg/ml deuteporfin either intravenously or in the left rear footpad. At various time-points (1, 3, 6, 12, 24 and 48 h) post-injection, the rats were sacrificed by cervical dislocation and blood samples were obtained from the aorta abdominalis (n=6 rats per time-point). The tissues of interest (including the heart, liver, spleen, lung, kidney, pancreas, popliteal lymph nodes and dorsal skin) were collected immediately, weighed and homogenized with an adequate quantity of physiological saline. Following centrifugation at 644 x g at 25-26°C for 5 min, the homogenate was filtered and the filtrate was boiled, cooled and centrifuged again (644 x g at 25-26°C for 5 min). The deuteporfin content of each tissue of interest was determined by the fluorometric method, using a fluorescence spectrometer (F-4500; Hitachi High-Technologies Corp., Tokyo, Japan) at excitation and emission wavelengths of 295 and 610 nm, respectively. The standard curve of deuteporfin was obtained by plotting the concentration vs. fluorescence intensity.

Generation of popliteal fossa lymphatic metastasis model. The popliteal fossa (depression behind the knee joint) lymphatic metastases model was generated as a xenograft of the BxPC-3-LN5 human cancer cell line. Briefly, $\sim 1 \times 10^7$ BxPC-3-LN5 cells in 100 μ l normal saline were injected subcutaneously into the left rear footpad of thymus-deficient 8-week-old BALB/C nude mice (n=5; weighing 18-20 g).

Five weeks later, a lymph node intumescence or swelling that was 3-5 mm in diameter was able to be observed in the left knee. This procedure was repeated five times (total mice, n=25). The mice were sacrificed by cervical dislocation and the intumescent lymph nodes were removed and fixed in 10% neutral buffered formalin. The specimens were dehydrated in a graded series of alcohol, and embedded in paraffin. Serial 5- μ m sections were deparaffinized, hydrated and stained with H&E in order to detect pancreatic cancer lymphatic metastases.

Localization of deuterporfin in tumors at inoculation site and sites of lymphatic metastases. The BALB/C-nu/nu nude mice with pancreatic cancer metastases were intravenously injected with deuterporfin at a dose of 2.5 mg/ml. At given time intervals (1, 2, 4, 6, 12, 24 and 48 h) following the injection, the mice were sacrificed by cervical dislocation. The lymph nodes in the left (metastatic side) and right (control side) popliteal fossa, respectively, were collected immediately. The deuterporfin content was measured according to the aforementioned fluorescence method.

Wood's lamp observations. The difference in deuterporfin concentration between the cancerous and normal tissues was directly observed using a Wood's lamp (model, BS-WL1) purchased from the Sincery International Ltd. (Shanghai, China). Briefly, the light was turned off and the cover shade was secured prior to usage. The Wood's lamp was placed parallel to the mice at a distance of 15-20 cm and then turned on. Characteristics of the mice were identified by the appearance of different colors under the ultraviolet ray.

Statistical analysis. The number of independent replicates is listed individually for each experiment. All data are expressed as the mean \pm standard deviation. Statistical analyses were performed with analysis of variance using SPSS 20.0 software (IBM SPSS, Armonk, NY, USA) and P<0.05 was considered to indicate a statistically significant difference.

Results

Intravenous administration of deuterporfin is advantageous over footpad administration. The efficiency of intravenous administration of deuterporfin was compared with that of administration via the left rear footpad. In general, the injected SD rats appeared to be healthy throughout the experimental period and no symptoms of a toxic reaction to light, for example partially inflamed skin or abscesses, were observed. Furthermore, deuterporfin was observed to be distributed in all tissues of the body following intravenous or left rear footpad administration. However, the distribution of deuterporfin was uneven due to differences in blood perfusion and the tissue microenvironment.

The mean deuterporfin levels in the blood and tissues following intravenous administration are exhibited in Table IA. It was observed that deuterporfin levels following intravenous treatment decreased as follows: Blood>liver>kidney>lymph node>pancreas>skin>heart>lungs>spleen. The time-course changes of deuterporfin levels in the blood, liver, kidney, heart, lungs and spleen were similar, with the highest concentration

at 1 h post-injection, followed by a time-dependent decrease. The highest expression values attained were 108.31 \pm 1.02, 75.18 \pm 3.57, 58.52 \pm 5.19, 13.01 \pm 2.10, 1.89 \pm 2.48 and 0.36 \pm 0.57 μ g/l, respectively. Notably, the normal liver exhibited a higher level of accumulation of deuterporfin. Previously, evidence of the poor sensitivity of cancerous liver tissue to photosensitization was observed when compared with that of the normal liver tissue (22). Therefore, caution must be taken when using PDT in the treatment of hepatic disease.

The mean levels of deuterporfin in the blood and tissues following left rear footpad administration are displayed in Table IB. In comparison to the intravenous administration group, the entry rates of deuterporfin into the tissues were markedly slower. For example, deuterporfin levels in the liver, kidney, lymph node and blood were demonstrated to reach a peak level at 3 h post-injection, and reached levels of 66.81 \pm 3.01, 82.25 \pm 15.60, 71.75 \pm 15.68 and 73.26 \pm 2.87 μ g/l, respectively. However, the deuterporfin levels in the heart, spleen, lungs and pancreas peaked at 1 h post-injection and were subsequently eliminated with time. Additionally, the spleen and lungs exhibited their removal or resisting effects to deuterporfin, as the highest deuterporfin concentrations observed were only 1.11 \pm 1.42 and 2.93 \pm 0.88 μ g/l, respectively. As for the skin, a higher level of deuterporfin (30.57 μ g/l) was observed at 1 h post-injection and the peak concentration was detected at 12 h post-injection. It is notable that, following footpad administration, deuterporfin may directly enter the popliteal lymph nodes along the lymphatic vessels.

Deuterporfin may be used as a tracer to detect lymphatic metastases in a pancreatic cancer xenograft model. Regional lymphatic metastases from pancreatic cancer is considered to be relevant to tumor spread and progress (23). Therefore, a reliable lymphatic metastases model of pancreatic cancer that effectively mimics human pancreatic cancer and may be used to test the efficacy of therapeutic strategies, including resection and tracing, is urgently required (24-26). Xenografts of the BxPC-3-LN5 pancreatic cancer cell line with lymphatic metastases were generated in athymic nude mice. Tumors developed at the site of direct implantation of BxPC-3-LN5 cells in all the animals (Fig. 2A). The incidence of lymphatic metastases was 100% and the metastatic growth of tumor cells was observed in all cases. Swelling of the popliteal fossa lymph node was also observed 5 weeks following implantation on the metastatic side, but not on the control side (Fig. 2B). Immunohistochemical evaluation revealed an aggressive invasion of cancer cells in the lymph nodes. Initially, cancer cells were able to enter the marginal sinus and form isolated tumor cells or small groups of metastatic niduses (Fig. 2C). Subsequently, tumor cells accumulated to form large metastatic niduses. Certain tumor cells were also able to infiltrate the intermediate sinuses and proliferate, diffuse toward medullary sinuses and invade the whole lymph node (Fig. 2D).

Based on this lymphatic metastases model of pancreatic cancer, the extent of accumulation of deuterporfin in the bilateral popliteal fossa lymph nodes, following intravenous administration, was determined. As shown in Fig. 3, the two popliteal fossa lymph nodes exhibited similar metabolic kinetic trends, and deuterporfin accumulated with the highest concentration at 6 h following administration. However, the

Table I. Deuteporfin levels in blood and tissues of Sprague-Dawley rats following intravenous administration.

A, Following intravenous administration of deuteporfin									
Concentration of deuteporfin, $\mu\text{g/l}$									
Time, h	Blood	Heart	Liver	Spleen	Lung	Kidney	Pancreas	Lymph nodes	Skin
1	108.31 \pm 1.02	13.01 \pm 2.10	75.18 \pm 3.57	0.36 \pm 0.57	1.89 \pm 2.48	58.52 \pm 5.19	30.14 \pm 2.56	28.95 \pm 7.18	26.87 \pm 0.51
3	103.00 \pm 1.14	6.76 \pm 4.11	60.47 \pm 6.21	0.33 \pm 0.52	1.18 \pm 0.46	35.47 \pm 11.83	25.89 \pm 2.29	22.87 \pm 4.42	28.08 \pm 6.37
6	37.67 \pm 0.55	6.15 \pm 1.96	60.93 \pm 7.01	0.06 \pm 0.11	1.19 \pm 1.55	31.34 \pm 13.15	30.38 \pm 4.05	25.89 \pm 5.82	32.24 \pm 4.95
12	30.00 \pm 0.27	4.95 \pm 0.83	50.11 \pm 4.5	0.02 \pm 0.05	0.86 \pm 0.23	44.43 \pm 9.34	31.60 \pm 2.85	24.78 \pm 7.26	23.74 \pm 1.50
24	26.5 \pm 0.51	3.16 \pm 0.33	47.19 \pm 10.89	0.03 \pm 0.07	0.85 \pm 1.29	52.80 \pm 11.28	30.97 \pm 0.86	16.57 \pm 2.19	25.76 \pm 5.04
48	24.18 \pm 0.07	3.03 \pm 0.25	35.29 \pm 7.05	0.02 \pm 0.06	0.81 \pm 0.69	42.51 \pm 7.09	31.95 \pm 7.93	22.47 \pm 1.72	30.86 \pm 8.29

B, Following left rear footpad administration of deuteporfin									
Concentration of deuteporfin, $\mu\text{g/l}$									
Time, h	Blood	Heart	Liver	Spleen	Lung	Kidney	Pancreas	Lymph nodes	Skin
1	38.29 \pm 1.86	8.37 \pm 2.83	64.62 \pm 13.96	1.11 \pm 1.42	2.93 \pm 0.88	39.64 \pm 16.26	36.45 \pm 4.29	16.50 \pm 1.17	30.57 \pm 2.64
3	73.26 \pm 2.87	6.46 \pm 1.44	66.81 \pm 3.01	0.60 \pm 0.17	2.52 \pm 1.63	82.25 \pm 15.60	31.88 \pm 1.29	71.75 \pm 15.68	29.75 \pm 1.09
6	35.42 \pm 5.78	5.61 \pm 4.35	48.52 \pm 3.58	0.60 \pm 0.11	1.10 \pm 0.84	46.47 \pm 13.39	30.08 \pm 1.95	32.11 \pm 3.60	30.88 \pm 1.85
12	30.21 \pm 4.23	4.28 \pm 3.54	44.90 \pm 10.44	0.49 \pm 0.58	1.36 \pm 1.01	42.44 \pm 8.73	31.18 \pm 4.67	24.39 \pm 16.11	33.87 \pm 6.23
24	20.28 \pm 3.65	4.63 \pm 3.50	37.07 \pm 5.64	0.28 \pm 0.31	0.99 \pm 1.08	36.06 \pm 6.84	21.00 \pm 9.09	18.05 \pm 3.59	24.96 \pm 2.50
48	19.43 \pm 2.51	4.25 \pm 0.95	39.20 \pm 6.55	0.27 \pm 0.42	0.03 \pm 0.05	27.95 \pm 9.82	30.00 \pm 8.73	21.86 \pm 3.08	27.69 \pm 2.23

Dose of deuteporfin administered was 2.5 mg/ml. Data are expressed mean \pm standard deviation, n=6.

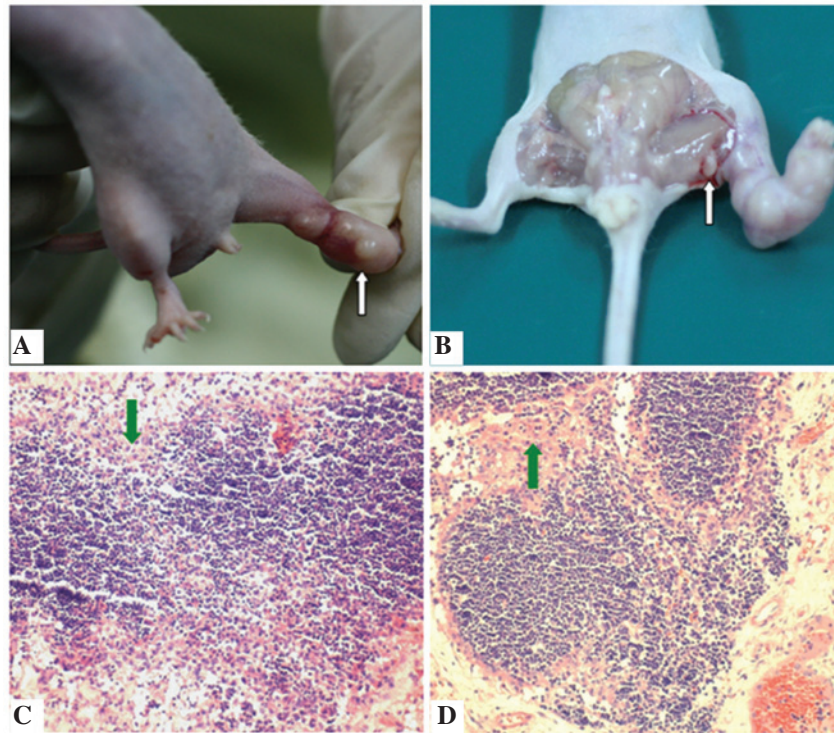


Figure 2. Pathological changes observed in a xenograft model of lymphatic metastatic pancreatic cancer cell line BxPC-3-LN5. (A) Primary tumors at the site of direct implantation of BxPC-3-LN5 cells in nude mice (white arrow). (B) Lymph node metastases (white arrow). (C) Immunohistochemical evaluation of cancer cell invasion to lymph nodes at the marginal sinus (green arrows). (D) Immunohistochemical evaluation indicated that metastatic cancer cells were able to grow in regional lymph nodes (green arrows). Hematoxylin and eosin staining; magnification, x200.

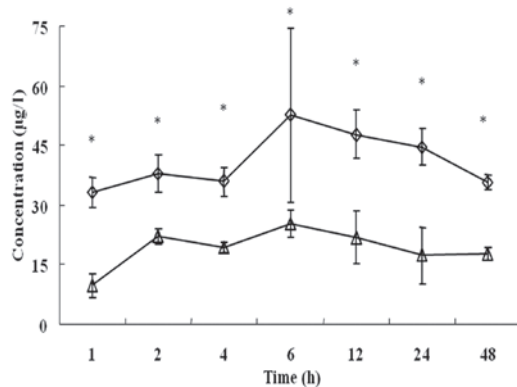


Figure 3. Time-course changes in deuteporfin levels in the popliteal fossa lymph nodes of the metastatic side (◊) and the untreated side (Δ) of the human pancreatic cancer BxPC-3-LN5 cell xenograft mouse model. Values are expressed as the mean ± standard deviation. *P<0.05.

deuteporfin concentration in the lymph nodes was ~2-3 times higher on the metastatic side than that of the control side (P<0.05). Furthermore, the concentration of deuteporfin reached $33.10 \pm 3.81 \mu\text{g/l}$ at 1 h and $52.59 \pm 21.91 \mu\text{g/l}$ at 6 h, respectively, on the metastatic side; while the concentration of deuteporfin on the control side was $9.72 \pm 2.95 \mu\text{g/l}$ at 1 h and $25.13 \pm 3.43 \mu\text{g/l}$ at 6 h, respectively. The deuteporfin concentration peaked at 24 h on the metastatic and control sides. Subsequently, the concentration of deuteporfin in the bilateral popliteal fossa lymph nodes gradually decreased and reached 17.64 ± 1.56 and $35.58 \pm 1.95 \mu\text{g/l}$ at 48 h post-injection, respectively. A previous study reported that pancreatic cancer cells

are selective to the uptake of photosensitizers, and that the ratio of uptake of deuteporfin between pancreatic cancer cells and normal tissues is 3:1 (27). Furthermore, the selectivity of cancer tissues for the photosensitizer is likely explained by their characteristic abnormal physiology, including poor lymphatic drainage, leaky vasculature, lower pH environment, higher level of receptors for low-density lipoproteins and abnormal stromal composition (19,28,29). In the present study, the concentration of deuteporfin in the bilateral popliteal fossa lymph nodes of normal nude mice was also evaluated, and reached 28.19 ± 1.71 and $26.16 \pm 1.5 \mu\text{g/l}$, respectively, 24 h post-injection. This result indicated that the selectivity of cancer tissue to deuteporfin led to a significant decrease in the deuteporfin concentration in the corresponding normal tissue (P=0.002). The increased difference in drug concentration between the cancerous and normal tissues was directly observed through the application of the Wood's lamp (Fig. 4).

Discussion

PDT, through the use of an intravenously administered photosensitizer, has been used as an anti-vascular and anti-tumor therapeutic strategy for several decades (30). Furthermore, the development of lymph node-specific PDT has generated notable interest. Kilarski *et al* (31) reported that anti-lymph node PDT allows the control of lymphatic ablation and regeneration by alteration of the light fluence and photosensitizer dose. Other potential therapeutic uses of lymph node-specific PDT include the inhibition of the spread of lymph node-trafficking parasites or pathogens, as well

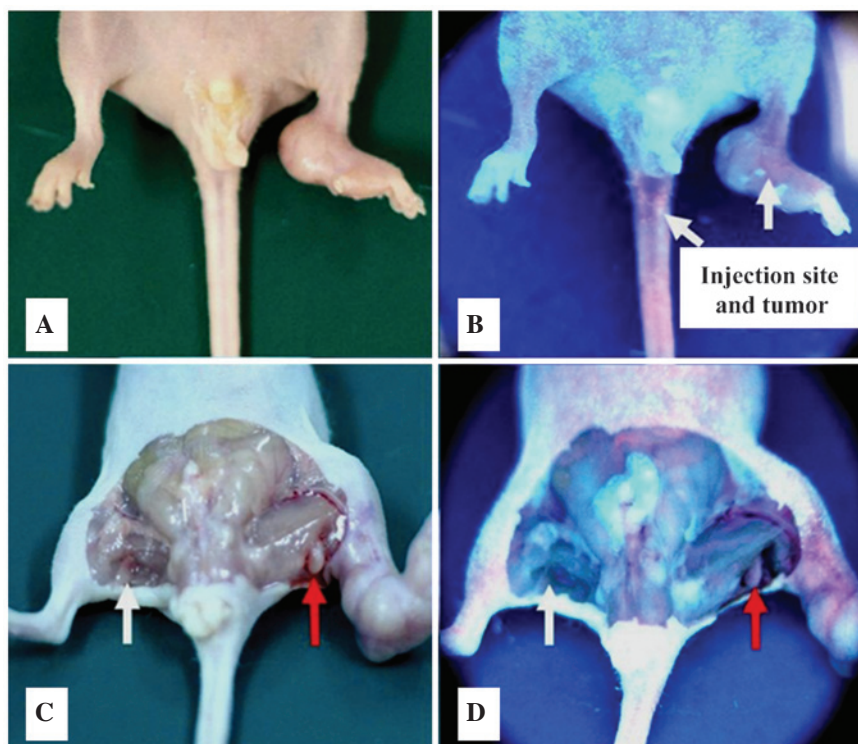


Figure 4. Tracking of deuterporfin to the tumor sites of the lymph node metastatic model of a pancreatic cancer cell line under irradiation of a Wood's lamp. Mice 1 h after intravenous administration of deuterporfin (A) without and (B) with the Wood's lamp. Excised mice 1 h after intravenous administration of deuterporfin (C) without and (D) with the Wood's lamp. White arrows indicate the normal popliteal fossa lymph nodes and the red arrows indicate the metastatic lymph nodes. Blue fluorescence indicates the effect of the Wood's Lamp.

as slowing the clearance of locally delivered drugs (32,33). However, the application of a photosensitizer as a tracing agent for the detection of lymphatic metastases of pancreatic cancer has not previously been reported or fully explored, to the best of our knowledge.

Deuterporfin, one of the latest generation of photosensitizers, is characterized by clear and relatively pure chemical constituents, and has been demonstrated to possess good photodynamic anti-tumor activity (34); however, its role as a photosensitizer has not previously been studied. The present study aimed to elucidate the preclinical pharmacokinetic characteristics of deuterporfin, as well as using it in the detection of lymphatic metastases in a pancreatic cancer xenograft model. Similarly to previous reports (34), deuterporfin was rapidly eliminated throughout the mouse model, which suggested that it may be a promising drug with few side effects.

The current study was divided into two sections. The first section aimed to identify an appropriate route of administration for deuterporfin. The biodistribution and pharmacokinetics of deuterporfin in SD rats, following administration via separate routes, were therefore investigated. In the second section, a lymphatic metastatic pancreatic cancer xenograft model was established, and the biodistribution of deuterporfin in cancerous and normal lymph nodes was subsequently compared. It was demonstrated that the concentration of deuterporfin in the metastatic lymph nodes was highest 6 h following administration, and was ~2-3 times higher than that in the normal lymph nodes.

Following intravenous administration, only 1.89 ± 2.48 and 0.36 ± 0.57 $\mu\text{g}/\text{l}$ of deuterporfin were detected in the lungs and spleen, respectively, 1 h post-injection. These low values may

be a result of specific removal or resistance mechanisms of these organs to xenentesis (the introduction of a novel drug in the organ). Notably, the metabolic and retention effects of deuterporfin in the lymph nodes, skin and pancreas stabilized, and deuterporfin reached relatively stable levels in these organs at 48 h post-injection. These characteristics of deuterporfin distribution indicate that it may be helpful in the detection of lymphatic metastases and in the treatment of pancreatic cancer.

By contrast, the lymph node deuterporfin concentration in the footpad administration group was significantly higher than that in the intravenous administration group ($P < 0.05$). However, the administration of deuterporfin in the left rear footpad is limited in a clinical setting due to the inconvenience of the required operation, particularly for intraperitoneal organs, including the pancreas. In addition, the longer distribution-equilibrium time of deuterporfin in tissues associated with footpad administration may prolong the lucifugal or light-repelling time of the patients. Taken together, these results indicated that intravenous administration was the most appropriate method for deuterporfin, compared with administration via the left rear footpad.

Deuterporfin appeared to be selectively enriched in cancerous pancreatic lymph nodes, and it was therefore hypothesized that deuterporfin may be a potential tracer agent for the detection of lymphatic metastases.

To the best of our knowledge, the present study was the first to explore the potential use of deuterporfin as a tracer for identifying the lymphatic metastases of pancreatic cancer. Two administration routes were compared, and intravenous administration was found to be more practical than the local lymphatic route for human pancreatic cancer treatment, with

the advantages of ease of administration and more rapid metabolism. The application of deuterporfin in the BxPC-3-LN5 human pancreatic cancer xenograft model, clearly exhibited the potential of deuterporfin as a tracer agent in the lymphatic metastases model of a pancreatic cancer cell line.

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