

# Role of Tyrosine Kinase Inhibitors in Indolent and Other Mature B-Cell Neoplasms

Nadine Kutsch<sup>1</sup>, Reinhard Marks<sup>2</sup>, Richard Ratei<sup>3</sup>, Thomas K. Held<sup>3</sup> and Martin Schmidt-Hieber<sup>3</sup>

<sup>1</sup>Department I of Internal Medicine and Center of Integrated Oncology Cologne Bonn, University of Cologne, Cologne, Germany. <sup>2</sup>Clinic for Hematology, Oncology and Stem Cell Transplantation, University Hospital of Freiburg, Freiburg, Germany. <sup>3</sup>Department of Hematology, Oncology and Tumor Immunology, HELIOS Clinic Berlin-Buch, Berlin, Germany.

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**ABSTRACT:** Targeting tyrosine kinases represents a highly specific treatment approach for different malignancies. This also includes non-Hodgkin lymphoma since it is well known that these enzymes are frequently involved in the lymphomagenesis. Hereby, tyrosine kinases might either be dysregulated intrinsically or be activated within signal transduction pathways leading to tumor survival and growth. Among others, Bruton's tyrosine kinase (Btk) is of particular interest as a potential therapeutic target. Btk is stimulated by B-cell receptor signaling and activates different transcription factors such as nuclear factor  $\kappa$ B. The Btk inhibitor ibrutinib has been approved for the treatment of chronic lymphocytic leukemia and mantle-cell lymphoma recently. Numerous clinical trials evaluating this agent in different combinations (eg, with rituximab or classical chemotherapeutic agents) as a treatment option for aggressive and indolent lymphoma are under way. Here, we summarize the role of tyrosine kinase inhibitors in the treatment of indolent and other non-Hodgkin lymphomas (eg, mantle-cell lymphoma).

**KEYWORDS:** indolent lymphoma, treatment, tyrosine kinase inhibitors, Bruton's tyrosine kinase, pathogenesis

**SUPPLEMENT:** Tyrosine kinases in cancer

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**CORRESPONDENCE:** martin.schmidt-hieber@helios-kliniken.de

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## Introduction

Different receptor and nonreceptor tyrosine kinases play a crucial role in normal B-cell development and in the pathogenesis of indolent and aggressive B-cell neoplasms.<sup>1–3</sup> Therefore, targeting these structures is a promising treatment approach for these malignancies.<sup>4</sup> Comprehensive research elucidating the biological function of tyrosine kinases within different signal transduction pathways has been undertaken in recent years. Based on these findings, numerous clinical trials focusing on inhibitors of these enzymes have been conducted or are under way. Inhibition of the Bruton's tyrosine kinase (Btk) – a TEC family nonreceptor (cytoplasmic) tyrosine kinase involved in B-cell receptor (BCR) signaling – is of particular interest.<sup>1,4</sup> Hereby, BCR signaling might either be activated by chronic antigen stimulation or by an intrinsically increased BCR signaling activity (tonic BCR signaling).<sup>1</sup>

The novel Btk inhibitor ibrutinib has recently been approved for the treatment of chronic lymphocytic leukemia (CLL) and mantle-cell lymphoma (MCL). Likewise, receptor tyrosine kinases might be important in the pathogenesis of B-cell neoplasms and potentially be inhibited, such as the receptor tyrosine kinase-like orphan receptor 1 (ROR1),

which is highly overexpressed in CLL and hairy cell leukemia (HCL).<sup>5</sup> Blocking this receptor in a CLL mouse model has shown therapeutic activity.<sup>6</sup> Targeting tyrosine kinases is not restricted to the malignant cell itself but might also affect the microenvironment. For example, osteoclast function was suppressed by the Btk inhibitor CC-292 in multiple myeloma (MM) and showed therapeutic potential when combined with the proteasome inhibitor carfilzomib.<sup>7</sup>

Here, we give an overview on the role of tyrosine kinase inhibitors in the treatment of different mature B-cell neoplasms, including CLL/small lymphocytic lymphoma (SLL), follicular lymphoma (FL), MCL, marginal zone lymphoma (MZL), Waldenstrom macroglobulinemia (WM), HCL, and plasma cell dyscrasias (Table 1). Additionally, we summarize information on the biological function of tyrosine kinases within different signaling pathways, which might be involved in lymphomagenesis and might be targeted in the future.

## Chronic Lymphocytic Leukemia/Small Lymphocytic Lymphoma

The antigen-dependent BCR pathway plays an important role in the survival of CLL cells.<sup>8</sup> Blocking this pathway with the

**Table 1.** Clinical trials investigating tyrosine kinase inhibitors in CLL/SLL, FL, MCL, MZL, WM, HCL, and plasma cell dyscrasias.

TYROSINE KINASE INHIBITOR [TARGETED TYROSINE KINASE*]/ TREATMENT REGIMEN	PHASE OF THE CLINICAL TRIAL	MALIGNANCY/ DISEASE STATUS (N)	OUTCOME	REFERENCE(S)
AZD2171 (cediranib) [VEGFR]	II	CLL/SLL, relapsed/refractory (n = 15)	OR: none (trial closed early due to lack of efficacy)	82
Bafetinib [diverse kinases like Bcr-Abl, Lyn, Fyn]	II	CLL, relapsed/refractory (n = 16)	OR: none [partial nodal response in 7/11 patients (64%)]	83
CC-292 [Btk]	I	CLL/SLL (n = 57), WM (n = 6), other B-NHL (n = 23), updated results for CLL/SLL (n = 83), relapsed/refractory	CLL/SLL: PR: 34%; all 17 efficacy-evaluable B-NHL patients reached SD, updated results for CLL/SLL: OR (PR): 31–67% (depending on dosage)	84,85
Dasatinib [diverse kinases like Bcr-Abl, Src and Btk]	II	MM, relapsed/‘plateau phase’ (n = 21)	OR: 5% (subgroup of 6 patients after dose escalation: 17%)	86
	II	CLL/SLL, relapsed/refractory (n = 15)	OR (PR): 20%	29
Dasatinib [diverse kinases like Bcr-Abl, Src and Btk] + fludarabine	II	CLL, fludarabine-refractory (n = 20)	OR (PR): 17%	30
Dovitinib [FGFR]	II	MM, relapsed/refractory (n = 43)	OR: none, SD: 62% (patients with t(4;14)) vs 35% (patients without t(4;14))	72
Entospletinib [Syk]	II	CLL, relapsed/refractory (n = 41)	OR (PR): 61%	25
	II	FL, relapsed/refractory (n = 41)	55% with reduced tumor bulk (10% decreased in tumor bulk ≥50%), no CR	33
Fostamatinib [Syk]	I/II	phase II part: CLL/SLL (n = 11), FL (n = 21), MCL (n = 9), MZL (n = 3), DLBCL (n = 23), LPL (n = 1), relapsed/refractory	OR (phase II part): CLL/SLL: 55% (PR), FL: 10% (PR), MCL: 11% (PR), DLBCL: 22% (1 patient with CR), none of 3 evaluable patients with MZL or LPL responded	24
Ibrutinib (single agent) [Btk]	I	CLL/SLL (n = 16), FL (n = 16), MCL (n = 9), MZL (n = 4), WM (n = 4), DLBCL (n = 7), relapsed/refractory	OR (evaluable patients): 60% (CR 16%); ITT population: CLL/SLL: 69%, FL: 38%, MCL: 78%, MZL: 25%, WM: 75%, DLBCL: 29%	34
	I/II	CLL/SLL, treatment naïve (n = 31)	OR: 71% (CR: 13%)	87
	I/II	CLL/SLL, relapsed/refractory (n = 85)	OR: 71% (CR: 2%), no difference between the patient groups (420 mg vs 840 mg ibrutinib daily), PFS 75% at 26 months	13
	II	CLL/SLL, presence of TP53 aberration, different disease stages (n = 51)	OR: previously untreated: 97% (PR: 55%, PR with lymphocytosis: 42%), relapsed/refractory: 80% (PR: 40%, PR with lymphocytosis: 40%)	10
	II	MCL, relapsed/refractory (n = 111)	OR: 68% (CR: 21%), estimated median PFS: 13.9 months	42
	II	FL, relapsed/refractory (n = 40)	OR: 30% (CR: 3%)	35
	II	WM, relapsed/refractory (n = 63)	OR: 81%, major response rate (PR or better): 57%	57
	II	HCL, relapsed or ‘unfit’ (n = 8)	No detailed efficacy data available	60
	II**	MM, relapsed/refractory (n = 69)	OR (PR): 5%, up to 25% clinical benefit rate (depending on dosage)	88

(Continued)



Table 1. (Continued)

TYROSINE KINASE INHIBITOR [TARGETED TYROSINE KINASE*]/ TREATMENT REGIMEN	PHASE OF THE CLINICAL TRIAL	MALIGNANCY/ DISEASE STATUS (N)	OUTCOME	REFERENCE(S)
Ibrutinib [Btk] (vs ofatumumab)	III	CLL/SLL, relapsed/ refractory (n = 391)	OR (PR): 43% vs 4%, OS (at 12 months): 90% vs 81%	16
Ibrutinib [Btk] + bendamustine + rituximab	I	FL (n = 12), MCL (n = 17), MZL (n = 1), DLBCL (n = 16), transformed (n = 2), different disease stages	OR: 72% (FL: 90%, MCL: 94%, MZL: 100%, DLBCL: 37%, transformed: 50%); CR: 52%	36
	I	CLL/SLL, relapsed/refractory (n = 30)	OR: 93% (CR: 17%)	89
Ibrutinib [Btk] + lenalidomide	I	FL (n = 2), MCL (n = 2), LPL (n = 1), DLBCL (n = 4), transformed (n = 4), relapsed/ refractory	No detailed efficacy data available	38
	I	CLL/SLL, relapsed/refractory (n = 11)	OR (PR): 100%	90
Ibrutinib [Btk] + ofatumumab	I/II	CLL/SLL (n = 66), PLL (n = 2), transformed (n = 3), relapsed/ refractory	OR (CLL/SLL): 83%	91
Ibrutinib [Btk] + rituximab	II	CLL, high risk, different disease stages (n = 40)	OR: 95% (CR: 8%)	92,93
	II	MCL, relapsed/refractory (n = 50)	OR: 87% (CR: 38%)	44
Ibrutinib [Btk] + R-CHOP	I	FL (n = 4), MCL (n = 5), DLBCL (n = 24), treatment-naïve	OR: 91% (PR: 21%, CR: 70%)	37
Imatinib [Bcr-Abl, ckit]	II	MM, relapsed/refractory (n = 23)	OR: none, treatment ended in 18/23 patients (78%) due to PD	75
Imatinib [Bcr-Abl, ckit]*** + chlorambucil	I	CLL, relapsed/refractory (n = 11)	OR: 45%	94
Nintedanib (BIBF 1120) [VEGFR/FGFR/ PDGFR]	I	MM, relapsed/ refractory (n = 17)	OR: none, SD: 13% (evaluable patients)	95
ONO-4059 [Btk]	I	CLL, relapsed/refractory (n = 25)	OR (including modified PR with lymphocytosis): 84%, 89% responses in the 17p-deleted subgroup	18,19
	I	SLL (n = 1), FL (n = 3), MCL (n = 7), WM (n = 1), DLBCL (n = 2), relapsed/ refractory	OR (PR): 42%, OR for MCL: 50%	96
SB1518 [JAK2]	I	SLL (n = 1), FL (n = 10), MCL (n = 5), HL (n = 14), DLBCL (n = 4), relapsed/ refractory	OR at the highest dose level (n = 22): 14%, median PFS (all evaluable patients): 120 days	40
Sorafenib [different kinases such as VEGFR, PDGFR, ckit]	II	CLL (n = 2), FL (n = 4), MCL (n = 2), LPL (n = 1), DLBCL (n = 11), T-cell lymphoma (n = 1), relapsed/refractory	OR: 10%, SD: 42%	97
Sunitinib [different kinases such as VEGFR, PDGFR, ckit]	II	CLL/SLL, relapsed/refractory (n = 18)	OR: none (trial closed early due to lack of efficacy)	82
Vandetanib (ZD6474) [VEGFR/EGFR]	II	MM, relapsed/ refractory (n = 18)	OR: none	80

**Notes:** Tyrosine kinase inhibitors are shown in an alphabetic order. \*Some of the tyrosine kinase inhibitors inhibit further structures than those specified in this table. \*\*Ibrutinib ± dexamethasone. \*\*\*Might also inhibit DNA repair.

**Abbreviations:** Btk, Bruton's tyrosine kinase; CLL, chronic lymphocytic leukemia; CR, complete remission; DLBCL, diffuse large B-cell lymphoma; EGFR, epidermal growth factor receptor; FGFR, fibroblast growth factor receptor; FL, follicular lymphoma; HCL, hairy cell leukemia; HL, Hodgkin lymphoma; ITT, intention to treat; JAK2, Janus kinase 2; LPL, lymphoplasmocytic lymphoma; MCL, mantle-cell lymphoma; MM, multiple myeloma; MZL, marginal zone lymphoma; NHL, non-Hodgkin lymphoma; OR, objective response; OS, overall survival; PD, progressive disease; PDGFR, platelet-derived growth factor receptor; PFS, progression-free survival; PLL, prolymphocytic leukemia; PR, partial response; R-CHOP, rituximab together with cyclophosphamide, doxorubicin, vincristine, and prednisolone; SD, stable disease; SLL, small lymphocytic lymphoma; VEGFR, vascular endothelial growth factor receptor; WM, Waldenström macroglobulinemia.



novel oral inhibitors appears to be highly active in relapsed/refractory CLL and even in high-risk disease defined by the presence of deletion 17p (del(17p)).<sup>9,10</sup>

One of the key kinases in the BCR pathway is Btk, which activates downstream survival signaling, including extracellular-signal regulated kinases 1/2 (ERK 1/2), phosphoinositide-3-kinases (PI3K), and the nuclear factor “kappa-light-chain-enhancer” of activated B-cells (NF-κB) pathway.<sup>11</sup> Ibrutinib binds Btk covalently and thereby inhibits CLL cell proliferation, promotes apoptosis, and unleashes CLL cells from tissues into the blood.<sup>12</sup> In a phase Ib/II multicenter study, 85 patients with refractory/relapsed CLL/SLL were treated with different doses of ibrutinib orally attaining an overall response rate of 71%.<sup>13</sup> At month 26, an estimated progression-free survival (PFS) rate of 75% and overall survival (OS) of 83% were achieved (Table 1). These results were encouraging, especially in the high-risk setting as response was independent of clinical and genomic risk factors. Furthermore, toxicity was mild comprising transient diarrhea, fatigue, and upper respiratory tract infection.<sup>13</sup> Updated data from the 3-year follow-up of this trial showed that longer treatment with ibrutinib yielded an improved response quality. Treatment-related lymphocytosis was mostly asymptomatic even with a duration of >1 year and did not seem to impact PFS and OS. Toxicity including grade 3 or greater cytopenias, fatigue, and infections reduced during longer follow-up.<sup>14</sup> Furthermore, the comparison between ibrutinib and ofatumumab in a phase III trial recruiting pretreated patients with CLL/SLL showed a significantly longer PFS for ibrutinib-treated patients at 16 months (median not reached vs 8.1 months).<sup>15,16</sup> Additionally, OS was better in the ibrutinib arm with 85% vs 78% in the ofatumumab arm at 18 months, although 61% of patients randomized to the ofatumumab arm were censored because they had crossed over to ibrutinib. Patients treated with ibrutinib achieved a best overall response rate of 90% compared to only 25% in the ofatumumab subgroup.<sup>15</sup> Ibrutinib has been approved by the EMA and the FDA on the basis of the phase Ib/II study in 2014.<sup>13</sup> However, treatment fails in some patients. Performing whole exome sequencing in six relapsed patients, the acquired resistance to ibrutinib often involved mutation of a cysteine residue at the ibrutinib binding site (C481) and two additional mutations in phospholipase Cγ2 downstream of Btk.<sup>17</sup> Progression occurs mainly in patients with del(17p) and/or del(11q).<sup>14</sup>

Besides ibrutinib, several other Btk inhibitors are under clinical development. Preliminary data available on a phase I trial with ONO-4059 showed a favorable safety profile along with promising efficacy in 25 heavily pretreated CLL patients with a median treatment duration of 363 days. Best overall response rate was 84% with even 89% responses in the 17p-deleted subgroup.<sup>18,19</sup> Furthermore, ACP-196, a next-generation Btk inhibitor, is currently under investigation as a single agent or within combination therapy in phase I/II clinical trials.

Spleen tyrosine kinase (Syk) is a cytoplasmic tyrosine kinase and a mediator of BCR signaling. Activation of Syk is important for cell survival and proliferation in CLL.<sup>20,21</sup> Syk also influences retention of CLL cells within lymphoid tissues and chemotaxis.<sup>22,23</sup> Disrupting the BCR pathway by Syk inhibitors is another therapeutic approach for CLL. However, it seems to be less efficient than Btk inhibition. Fostamatinib attained partial remission in 55% of eleven patients with CLL/SLL.<sup>24</sup> Entospletinib, another selective Syk inhibitor, was assessed in a phase II study that enrolled 41 CLL patients among patients with other non-Hodgkin lymphoma. At 24 weeks, the PFS rate was 70% with a median PFS of 13.8 months and an objective response rate of 61%. Entospletinib was generally well tolerated, and the most common side effects included dyspnea, pneumonia, febrile neutropenia, dehydration, and pyrexia.<sup>25</sup>

Dasatinib, a tyrosine kinase inhibitor originally developed for the treatment of chronic myelogenous leukemia, inhibits several kinases (eg, Bcr-Abl, Src, Btk), whereas some of them are activated within CLL cells too.<sup>26–28</sup> When used in clinical trials, dasatinib achieved only a low partial remission rate of 20% in a phase II study encompassing 15 patients with CLL/SLL.<sup>29</sup> In combination with fludarabine, even inferior response was achieved in fludarabine-refractory patients (17% of patients with partial remission).<sup>30</sup>

### Follicular Lymphoma

Dysregulation of expression and/or activation in cytoplasmic and receptor tyrosine kinases is observed in FL cells too. Although the impact of tonic BCR signaling in the pathogenesis of FL is still not well understood yet, the BCR-mediated activation of the tyrosine kinase Syk appears to be altered in FL.<sup>31</sup> Newer *in vitro* data showed that Syk is not only overexpressed in FL cells but also involved in the high expression of matrix metalloproteinase 9 and vascular endothelial growth factor (VEGF), thereby putatively promoting invasion and angiogenesis.<sup>32</sup> In spite of these promising data, using Syk inhibitors like fostamatinib in clinical trials for relapsed/refractory FL resulted in an objective response rate of only about 10% of treated patients.<sup>24</sup> In a more recent phase II trial, another selective Syk inhibitor (entospletinib) was evaluated in the same patient population and also a substantial tumor bulk reduction (≥50%) was reported in only 10% of treated patients.<sup>33</sup>

More mature data regarding inhibition of tyrosine kinases involved in BCR signaling of FL were obtained by using ibrutinib in relapsed and de novo FL. Two phase I/II trials with monotherapy in relapsed FL resulted in very similar results with an objective response rate of 38% (6/16 patients, three patients with complete remission) and 30% (including one patient with complete remission) in 40 treated patients, respectively.<sup>34,35</sup> Although active, ibrutinib monotherapy appears to be less effective in FL patients when compared to CLL or MCL, an observation that might reflect differences



in chronic BCR signaling between the separate non-Hodgkin lymphoma entities. Nevertheless, combining this approach with rituximab and bendamustine improves the objective response rate in relapsed/refractory FL to up to 90%.<sup>36</sup> Further phase I trials combining ibrutinib with lenalidomide in relapsed FL or with rituximab together with cyclophosphamide, doxorubicin, vincristine, and prednisolone (R-CHOP) in treatment-naïve FL reported encouraging safety and efficacy results.<sup>37,38</sup>

Since VEGF has been shown to be overexpressed in FL cells, interfering with receptor tyrosine kinase VEGF signaling in the malignant cells or the microenvironment appears to be a logical consequence. In this regard, testing the angiogenesis inhibitor bevacizumab in addition to rituximab in a randomized phase II trial resulted in a significant increase in PFS.<sup>39</sup> As another target for selective inhibition of tyrosine kinases, interfering with cytokine-mediated janus kinase 2 activation led to some objective responses in tumor size in FL patients.<sup>40</sup>

In summary, inhibition of BCR-activated tyrosine kinases shows clinical efficacy in FL patients. Further investigations, especially in combination therapy approaches, will provide additional data on the impact of this therapeutic principle in the treatment of FL patients. In addition, inhibition of tyrosine kinases involved in growth factor or cytokine signaling might be another promising way for further investigations.

### Mantle-Cell Lymphoma

Still being considered an incurable disease, there is no standard upfront treatment regimen for MCL. Typically, three regimens are used, each in combination with rituximab: R-CHOP, bendamustine (R-B), and cyclophosphamide, vincristine, and prednisolone (R-CVP). Despite a usually good initial response, MCL tends to recur early and shows an aggressive course in most patients.<sup>41</sup> Hence, there is a need for treatment options with tolerable side effects.

Being an attractive target for selective B-cell inhibition, ibrutinib has been studied as a single substance in MCL. In a phase I study, nine patients with MCL were treated with ibrutinib. Seven patients showed a response, of which three had a complete response.<sup>34</sup> On the basis of these results, a phase II study was conducted with 111 patients with MCL who received ibrutinib at a dose of 560 mg daily until disease progression or the occurrence of unacceptable side effects.<sup>42</sup> They had received a median of three therapies before and were stratified according to prior treatment without ( $n = 63$ ) or with bortezomib ( $n = 48$ ). A total of 68% of the patients ( $n = 75$ ) responded, with 21% having a complete and 47% a partial response; there was no significant difference between bortezomib-pretreated and bortezomib-naïve patients. With regard to the secondary endpoints, the estimated median response duration was 17.5 months, the estimated median PFS was 13.9 months, and the median OS was not reached. Moreover, the estimated rate of OS at 18 months was 58%.

Treatment was generally well tolerated; most common adverse events comprised mild-to-moderate fatigue, nausea, diarrhea, and grades 3 and 4 hematologic adverse events, including neutropenia in 16%, thrombocytopenia in 11%, and anemia in 10% of the MCL patients. Ibrutinib showed activity in all analyzed subgroups regardless of the presence or absence of blastoid disease, bulky disease, advanced disease, refractory disease, previous high-intensity therapy, or previous therapy with lenalidomide.<sup>42</sup> Given the fact that most patients had a long-standing, heavily pretreated disease, this study formed the base for the US FDA's Breakthrough Therapy Designation Pathway.<sup>43</sup> The encouraging results spurred the initiation of several trials investigating the combination of ibrutinib with rituximab alone, rituximab and bendamustine, ublituximab or lenalidomide, and comparing ibrutinib against temsirolimus, thus further enhancing the therapeutic possibilities in this disease.<sup>36–38,44</sup> In fact, ibrutinib showed the highest response rate when compared to cladribine, lenalidomide, temsirolimus, rituximab, 90Y-ibrutinib, and everolimus.<sup>45</sup> Several other tyrosine kinase inhibitors are currently studied in MCL besides the Btk inhibitor ibrutinib, such as the Syk inhibitor fostamatinib.

### Marginal Zone Lymphoma

Compared to other indolent B-cell lymphoma (eg, CLL), limited data has been published on the role of tyrosine kinases in the pathogenesis of MZL. Moreover, larger clinical studies investigating kinase inhibitors are lacking for patients with MZL. One possible reason is the fact that the incidence of MZL is lower than that of other indolent B-cell lymphomas.<sup>46</sup> However, it has also to be mentioned that MZL might be cured by anti-infective strategies alone, such as *Helicobacter pylori* eradication for early-stage gastric mucosa-associated lymphoid tissue (MALT) lymphoma.<sup>47</sup> Despite this, targeting tyrosine kinases – in particular in the BCR pathway – might be an important therapeutical approach for MZL.<sup>1</sup> For example, BCR binding of the E2 envelope protein of hepatitis C virus – potentially associated with chronic BCR activation – has been described in extranodal MZL, a fact which might explain their causal relationship.<sup>1,48,49</sup>

Patients with MZL have been included in previous and ongoing clinical trials evaluating inhibition of BCR signaling by ibrutinib.<sup>34</sup> A phase I trial evaluating ibrutinib in relapsed or refractory B-cell lymphoma encompassed three evaluable patients with MZL and reported partial response, stable disease, and progressive disease in each one of them.<sup>34</sup> Besides Btk, the nonreceptor tyrosine kinase Syk is controlled by BCR signaling. A phase I/II trial evaluating fostamatinib was conducted based on the data that Syk inhibition induced apoptosis in various B-cell lines and inhibited lymphoma growth in a xenograft model.<sup>24,50</sup> Three patients with MZL were included, but none of them showed an objective response.<sup>24</sup>

A further tyrosine kinase that might be involved in the pathogenesis of MZL is the orphan receptor tyrosine kinase



ROR1. It is highly overexpressed in CLL and HCL, but can also be detected in a subset of MZL patients.<sup>5,51</sup> Growing evidence suggests that this tyrosine kinase is a survival factor, and thus might be a potential therapeutic target in different malignancies, including MZL.<sup>5,6</sup> Antiangiogenesis treatment using VEGF receptor antibodies might be another interesting approach for gastric MALT lymphoma. It was already demonstrated in a mouse model that these antibodies decreased significantly the size of the tumor accompanied by apoptotic changes of the endothelial cells belonging to the microvascular network.<sup>52,53</sup>

### Waldenstrom Macroglobulinemia

Potential molecular targets for tyrosine kinase inhibitors in WM have only recently been detected. Unlike other B-cell neoplasms such as CLL or diffuse large B-cell lymphoma, the role of the BCR signalosome in the biology of WM remains poorly understood.<sup>49,54</sup> The myeloid differentiation factor 88 (MYD88) L265P somatic mutation occurs in more than 90% of patients with WM or lymphoplasmocytic lymphoma (LPL). Thus, detection of this mutation might help to differentiate WM or non-IgM LPL from other similar B-cell disorders.<sup>55,56</sup>

MYD88 and its two adaptor proteins Btk and Toll/interleukin-1 receptor (TIR) domain containing adaptor protein (TIRAP) are activated after Toll-like receptor 4 binds to its ligand. MYD88 might also directly be activated by ligand binding of interleukin-1 receptor.<sup>55</sup> Further downstream signaling promotes the activation of NF- $\kappa$ B-dependent pro-survival pathways. WM cells with the L265P mutation in the MYD88 gene have enhanced proliferation and survival.<sup>56</sup> Although the exact signaling cascade remains unclear, it has been shown that the abrogation of Btk binding to MYD88 results in inhibition of NF- $\kappa$ B signaling and apoptosis.<sup>56</sup> Therefore, Btk appears to be a potential therapeutic target for tyrosine kinase inhibitors in WM as well. In fact, a recently conducted phase II trial investigating the Btk inhibitor ibrutinib in previously treated patients with WM ( $n = 63$ ) demonstrated a best overall response rate of 81% with a major response rate (partial response or better) of 57% leading to approval by the FDA.<sup>57</sup> Further phase II and III trials investigating different Btk inhibitors in WM are under way.

Taken together, tyrosine kinase inhibitors seem to complement the already existing armory of alkylating agents, nucleoside analogs, monoclonal antibodies, and proteasome inhibitors to treat WM and might be even used in combination therapies.

### Hairy Cell Leukemia

Alterations in the BCR-mediated signaling pathways are also crucial for the pathogenesis of HCL.<sup>1,58</sup> With respect to their BCR, canonical (ie, nonrandom, potentially antigen-driven) somatic hypermutation was observed in rearrangements of classical HCL and to a lesser extent also in HCL variant.<sup>59</sup> In addition, Btk has recently been identified as a promising

target in patients with HCL and clinical trials investigating the Btk inhibitor ibrutinib in these patients are ongoing.<sup>60,61</sup>

As aforementioned, ROR1 is highly overexpressed in HCL compared to healthy controls and other indolent lymphomas such as FL.<sup>5</sup> ROR1 is part of the Wnt pathway with Wnt5a presumably being its ligand.<sup>5,62</sup> It is important for organogenesis but its role in HCL remains largely unknown.<sup>63</sup> Additionally, increased expression of Src – a proto-oncogene with tyrosine-specific protein kinase activity – has been reported in HCL and might be an interesting therapeutic target.<sup>64</sup>

In conclusion, tyrosine kinase inhibitors offer a promising new therapeutic tool for HCL besides BRAF inhibitors such as vemurafenib and dabrafenib after decades of interferon, chemotherapy, and lately monoclonal antibodies, but randomized controlled data are not available yet. Given the low incidence and the prolonged natural course of HCL, it will most likely take decades again to gain evidence on that topic.

### Plasma Cell Dyscrasias

Various tyrosine kinases are dysregulated in clonal vs normal plasma cells and might play an important role in the pathogenesis of plasma cell dyscrasias. The translocation t(4;14) – usually a primary event – is detected in 10%–20% of all patients with MM and associated with an unfavorable outcome.<sup>65–67</sup> Presence of t(4;14) frequently results in dysregulated expression of fibroblast growth factor receptor 3 (FGFR3), a receptor tyrosine kinase.<sup>68</sup> However, FGFR3 might also be activated constitutively after somatic mutation at a later disease stage. It acts through the mitogen-activated protein kinase pathway and promotes tumor progression in MM.<sup>68,69</sup> Dovitinib (CHIR-258) is a receptor tyrosine kinase inhibitor, which showed inhibitory activity against various receptors, including FGFR *in vitro* and induced tumor growth reduction in a MM xenograft model with activating FGFR3 mutations.<sup>70,71</sup> Based on these promising preclinical data, a phase II study evaluating this agent in patients with relapsed or refractory MM was recently conducted.<sup>72</sup> Objective response was not observed in any patient, although 62% of patients carrying t(4;14) (vs 35% of patients lacking this translocation) showed disease stabilization. Despite this, dovitinib might have a role as part of a combination regimen in MM treatment.

CD117 (ckit), a receptor tyrosine kinase activated by its ligand stem cell factor is expressed by clonal plasma cells in about 30% of MM patients, 45% of patients with smoldering MM (SMM), and 70% of patients with monoclonal gammopathy of undetermined significance (MGUS).<sup>73</sup> Conversely, normal plasma cells are uniquely CD117.<sup>74</sup> Despite the fact that the biological function of ckit is largely unknown in plasma cell dyscrasias, it was shown that its expression is associated with an altered maturation of the myeloid and lymphoid hematopoietic cells in the bone marrow.<sup>73</sup> Imatinib, which inhibits ckit besides Bcr-Abl, has been evaluated as a



treatment option for refractory/relapsed MM but did not have significant efficacy.<sup>75</sup>

Furthermore, the proline-rich tyrosine kinase 2 (Pyk2), a nonreceptor protein kinase belonging to the focal adhesion kinase family, is overexpressed in MM, SMM, and MGUS compared to normal bone marrow.<sup>76</sup> Interestingly, it has also been demonstrated that Pyk2 might be an effector of FGFR3 activation, which regulates cell survival as aforementioned.<sup>77</sup> Pyk2 silencing suppressed Wnt/ $\beta$ -catenin signaling and reduced MM cell proliferation, adhesion, and cell cycle progression.<sup>76</sup> In addition, Pyk2 silencing prolonged survival in an MM xenograft model.<sup>76</sup> Taken together, these data indicate that Pyk2 might be involved in the pathogenesis of plasma cell dyscrasias and be a potential therapeutic target in the future.

Although clonal plasma cells, like other B-cell neoplasms, express the cytoplasmic nonreceptor tyrosine kinase Btk, its inhibition by the novel Btk inhibitor CC-292 had no potent anti-MM activity *in vitro*.<sup>7,78</sup> This observation might be explained by the specific *in vitro* conditions (including a low CC-292 dose) used in this study.<sup>7</sup> However, Btk is also expressed by osteoclasts that contribute to bone destruction in MM.<sup>78</sup> It was shown that CC-292 suppresses osteoclast function, in particular if combined with the proteasome inhibitor carfilzomib.<sup>7</sup> A multicenter phase I/II trial evaluating the combination of carfilzomib together with ibrutinib in patients with refractory or relapsed MM is currently ongoing.

In summary, tyrosine kinase inhibitors do not play a role in the clinical routine treatment of plasma cell dyscrasias yet. However, several tyrosine kinase inhibitors (eg, ibrutinib, PTK787/ZK222584) are further explored *in vitro* and *in vivo* in MM and might play a role in the treatment of this disease in the future.<sup>79–81</sup>

## Conclusion

Recently published phase I and II trials demonstrated promising objective response rates of 70%–90% by using ibrutinib as a single agent for relapsed or refractory CLL and MCL. Ibrutinib is characterized by a favorable toxicity profile, and preliminary data suggest that ibrutinib-induced lymphocytosis has no adverse impact on outcome. It is noteworthy that ibrutinib showed high rates of remission even in CLL patients having adverse cytogenetic changes like del(17p). First data demonstrate that ibrutinib, given as a single agent, has a similar efficacy in WM than it has in CLL or MCL. In contrast, the substance, used as monotherapy, has only moderate activity in FL exhibiting an objective response rate of about 30%. Combined with classical chemotherapy or monoclonal antibodies, ibrutinib is currently under investigation in different subtypes of indolent lymphomas. Other tyrosine kinase inhibitors are evaluated as well, of which the Syk inhibitors, fostamatinib and entospletinib, might be of particular interest in the near future. But also elucidating resistance mechanisms is important as possible combination

therapies might help to overcome these in the future. Research does not stop here, however, and next-generation Btk inhibitors are already in sight.

## Author Contributions

Contributed to the writing of the manuscript: NK, RM, RR, TKH, MSH. Agree with manuscript results and conclusions: NK, RM, RR, TKH, MSH. Jointly developed the structure and arguments for the paper: NK, RM, RR, TKH, MSH. Made critical revisions and approved final version: NK, RM, RR, TKH, MSH. All authors reviewed and approved of the final manuscript.

## REFERENCES

1. Niemann CU, Wiestner A. B-cell receptor signaling as a driver of lymphoma development and evolution. *Semin Cancer Biol.* 2013;23(6):410–21.
2. Sakata-Yanagimoto M, Enami T, Yokoyama Y, Chiba S. Disease-specific mutations in mature lymphoid neoplasms: recent advances. *Cancer Sci.* 2014;105(6):623–9.
3. Toffalini F, Demoulin J. New insights into the mechanisms of hematopoietic cell transformation by activated receptor tyrosine kinases. *Blood.* 2010;116(14):2429–37.
4. Robak T, Robak E. Tyrosine kinase inhibitors as potential drugs for B-cell lymphoid malignancies and autoimmune disorders. *Expert Opin Investig Drugs.* 2012;21(7):921–47.
5. Daneshmanesh AH, Porwit A, Hojjat-Farsangi M, et al. Orphan receptor tyrosine kinases ROR1 and ROR2 in hematological malignancies. *Leuk Lymphoma.* 2013;54(4):843–50.
6. Mani R, Mao Y, Frizzera FW, et al. Tumor antigen ROR1 targeted drug delivery mediated selective leukemic but not normal B-cell cytotoxicity in chronic lymphocytic leukemia. *Leukemia.* 2015;29(2):346–55.
7. Eda H, Santo L, Cirstea DD, et al. A novel Bruton's tyrosine kinase inhibitor CC-292 in combination with the proteasome inhibitor carfilzomib impacts the bone microenvironment in a multiple myeloma model with resultant antimyeloma activity. *Leukemia.* 2014;28(9):1892–901.
8. Chiorazzi N, Ferrarini M. B cell chronic lymphocytic leukemia: lessons learned from studies of the B cell antigen receptor. *Annu Rev Immunol.* 2003;21:841–94.
9. Davids MS, Brown JR. Targeting the B cell receptor pathway in chronic lymphocytic leukemia. *Leuk Lymphoma.* 2012;53(12):2362–70.
10. Farooqui MZ, Valdez J, Martyr S, et al. Ibrutinib for previously untreated and relapsed or refractory chronic lymphocytic leukaemia with TP53 aberrations: a phase 2, single-arm trial. *Lancet Oncol.* 2015;16(2):169–76.
11. Herman SE, Gordon AL, Hertlein E, et al. Bruton tyrosine kinase represents a promising therapeutic target for treatment of chronic lymphocytic leukemia and is effectively targeted by PCI-32765. *Blood.* 2011;117(23):6287–96.
12. Burger JA, Li K, Keating M, et al. Functional evidence from deuterated water labeling that the bruton tyrosine kinase inhibitor ibrutinib blocks leukemia cell proliferation and trafficking and promotes leukemia cell death in patients with chronic lymphocytic leukemia and small lymphocytic lymphoma. *Blood.* 2014;124(21):326.
13. Byrd JC, Furman RR, Coutre SE, et al. Targeting BTK with ibrutinib in relapsed chronic lymphocytic leukemia. *N Engl J Med.* 2013;369(1):32–42.
14. Byrd JC, Furman RR, Coutre SE, et al. Three-year follow-up of treatment-naïve and previously treated patients with CLL and SLL receiving single-agent ibrutinib. *Blood.* 2015;125(16):2497–506.
15. Brown JR, Hillmen P, O'Brien S, et al. RESONATE Investigators. Updated efficacy including genetic and clinical subgroup analysis and overall safety in the phase 3 RESONATE™ trial of ibrutinib versus ofatumumab in previously treated chronic lymphocytic leukemia/small lymphocytic lymphoma. *Blood.* 2014;124(21):3331.
16. Byrd JC, Brown JR, O'Brien S, et al. Ibrutinib versus ofatumumab in previously treated chronic lymphoid leukemia. *N Engl J Med.* 2014;371(3):213–23.
17. Woyach JA, Furman RR, Liu TM, et al. Resistance mechanisms for the Bruton's tyrosine kinase inhibitor ibrutinib. *N Engl J Med.* 2014;370(24):2286–94.
18. Karlin L, Rule S, Shah N, et al. A phase I study of the oral btk inhibitor ONO-4059 in patients with relapsed/refractory and high risk chronic lymphocytic leukaemia (CLL). *Blood.* 2013;122(21):676.
19. Fegan C, Bagshawe J, Salles G, et al. The Bruton's tyrosine kinase (BTK) inhibitor ONO-4059: promising single agent activity and well tolerated in patients with high risk chronic lymphocytic leukaemia (CLL). *Blood.* 2014;124(21):3328.



20. Kipps TJ. The B-cell receptor and ZAP-70 in chronic lymphocytic leukemia. *Best Pract Res Clin Haematol.* 2007;20(3):415–24.
21. Stevenson FK, Caligaris-Cappio F. Chronic lymphocytic leukemia: revelations from the B-cell receptor. *Blood.* 2004;103(12):4389–95.
22. Buchner M, Baer C, Prinz G, et al. Spleen tyrosine kinase inhibition prevents chemokine- and integrin-mediated stromal protective effects in chronic lymphocytic leukemia. *Blood.* 2010;115(22):4497–506.
23. Quiroga MP, Balakrishnan K, Kurtova AV, et al. B-cell antigen receptor signaling enhances chronic lymphocytic leukemia cell migration and survival: specific targeting with a novel spleen tyrosine kinase inhibitor, R406. *Blood.* 2009;114(5):1029–37.
24. Friedberg JW, Sharman J, Sweetenham J, et al. Inhibition of Syk with fostamatinib disodium has significant clinical activity in non-Hodgkin lymphoma and chronic lymphocytic leukemia. *Blood.* 2010;115(13):2578–85.
25. Sharman J, Hawkins M, Kolibaba K, et al. An open-label phase 2 trial of entospletinib (GS-9973), a selective Syk inhibitor, in chronic lymphocytic leukemia. *Blood.* 2015;125(15):2336–43.
26. Bantscheff M, Eberhard D, Abraham Y, et al. Quantitative chemical proteomics reveals mechanisms of action of clinical ABL kinase inhibitors. *Nat Biotechnol.* 2007;25(9):1035–44.
27. Hantschel O, Rix U, Schmidt U, et al. The Btk tyrosine kinase is a major target of the Bcr-Abl inhibitor dasatinib. *Proc Natl Acad Sci USA.* 2007;104(33):13283–8.
28. Rix U, Hantschel O, Dürnberger G, et al. Chemical proteomic profiles of the BCR-ABL inhibitors imatinib, nilotinib, and dasatinib reveal novel kinase and nonkinase targets. *Blood.* 2007;110(12):4055–63.
29. Amrein PC, Attar EC, Takvorian T, et al. Phase II study of dasatinib in relapsed or refractory chronic lymphocytic leukemia. *Clin Cancer Res.* 2011;17(9):2977–86.
30. Kater AP, Spiering M, Liu RD, et al. Dasatinib in combination with fludarabine in patients with refractory chronic lymphocytic leukemia: a multicenter phase 2 study. *Leuk Res.* 2014;38(1):34–41.
31. Irish JM, Czerwinski DK, Nolan GP, Levy R. Altered B-cell receptor signaling kinetics distinguish human follicular lymphoma B cells from tumor-infiltrating nonmalignant B cells. *Blood.* 2006;108(9):3135–42.
32. Fruchon S, Kheirallah S, Al Saati T, et al. Involvement of the Syk-mTOR pathway in follicular lymphoma cell invasion and angiogenesis. *Leukemia.* 2012;26(4):795–805.
33. Sharman JP, Klein LM, Boxer M, et al. Phase 2 trial of entospletinib (GS-9973), a selective SYK inhibitor, in follicular lymphoma (FL). *Blood.* 2014;124(21):4419.
34. Advani RH, Buggy JJ, Sharman JP, et al. Bruton tyrosine kinase inhibitor ibrutinib (PCI-32765) has significant activity in patients with relapsed/refractory B-cell malignancies. *J Clin Oncol.* 2013;31(1):88–94.
35. Bartlett NL, LaPlant BR, Qi J, et al. Ibrutinib monotherapy in relapsed/refractory follicular lymphoma (FL): preliminary results of a phase 2 consortium (P2C) trial. *Blood.* 2014;124(21):800.
36. Maddocks K, Christian B, Jaglowski S, et al. A phase 1/1b study of rituximab, bendamustine, and ibrutinib in patients with untreated and relapsed/refractory non-Hodgkin lymphoma. *Blood.* 2015;125(2):242–8.
37. Younes A, Thieblemont C, Morschhauser F, et al. Combination of ibrutinib with rituximab, cyclophosphamide, doxorubicin, vincristine, and prednisone (R-CHOP) for treatment-naïve patients with CD20-positive B-cell non-Hodgkin lymphoma: a non-randomised, phase 1b study. *Lancet Oncol.* 2014;15(9):1019–26.
38. Christian B, Kuruvilla J, Smith S, et al. A phase I study of ibrutinib and lenalidomide in patients with relapsed and refractory B-cell non-Hodgkin's lymphoma. *Blood.* 2014;124(21):4476.
39. Hainsworth JD, Greco FA, Raefsky EL, et al. Rituximab with or without bevacizumab for the treatment of patients with relapsed follicular lymphoma. *Clin Lymphoma Myeloma Leuk.* 2014;14(4):277–83.
40. Younes A, Romaguera J, Fanale M, et al. Phase I study of a novel oral Janus kinase 2 inhibitor, SB1518, in patients with relapsed lymphoma: evidence of clinical and biologic activity in multiple lymphoma subtypes. *J Clin Oncol.* 2012;30(33):4161–7.
41. Ghilmini M, Zucca E. How I treat mantle cell lymphoma. *Blood.* 2009;114(8):1469–76.
42. Wang ML, Rule S, Martin P, et al. Targeting BTK with ibrutinib in relapsed or refractory mantle-cell lymphoma. *N Engl J Med.* 2013;369(6):507–16.
43. Cameron F, Sanford M. Ibrutinib: first global approval. *Drugs.* 2014;74(2):263–71.
44. Wang ML, Hagemeister F, Westin J, et al. Ibrutinib and rituximab are an efficacious and safe combination in relapsed mantle cell lymphoma: preliminary results from a phase II clinical trial. *Blood.* 2014;124(21):627.
45. Fowler NH. Novel treatment approaches to mantle cell lymphoma. *Clin Adv Hematol Oncol.* 2013;11(suppl 18):14–7.
46. Morton LM, Wang SS, Devesa SS, Hartge P, Weisenburger DD, Linet MS. Lymphoma incidence patterns by WHO subtype in the United States, 1992–2001. *Blood.* 2006;107(1):265–76.
47. Parsonnet J, Hansen S, Rodriguez L, et al. *Helicobacter pylori* infection and gastric lymphoma. *N Engl J Med.* 1994;330(18):1267–71.
48. Quinn ER, Chan CH, Hadlock KG, Fountzakis SK, Flint M, Levy S. The B-cell receptor of a hepatitis C virus (HCV)-associated non-Hodgkin lymphoma binds the viral E2 envelope protein, implicating HCV in lymphomagenesis. *Blood.* 2001;98(13):3745–9.
49. Young RM, Staudt LM. Targeting pathological B cell receptor signalling in lymphoid malignancies. *Nat Rev Drug Discov.* 2013;12(3):229–43.
50. Spurgeon SE, Coffey G, Fletcher LB, et al. The selective SYK inhibitor P505-15 (PRT062607) inhibits B cell signaling and function in vitro and in vivo and augments the activity of fludarabine in chronic lymphocytic leukemia. *J Pharmacol Exp Ther.* 2013;344(2):378–87.
51. Barna G, Mihalik R, Timár B, et al. ROR1 expression is not a unique marker of CLL. *Hematol Oncol.* 2011;29(1):17–21.
52. Nakamura M, Matsui H, Takahashi T, et al. Suppression of lymphangiogenesis induced by Flt-4 antibody in gastric low-grade mucosa-associated lymphoid tissue lymphoma by *Helicobacter heilmannii* infection. *J Gastroenterol Hepatol.* 2010;25(suppl 1):S1–6.
53. Nakamura M, Takahashi T, Matsui H, et al. New pharmaceutical treatment of gastric MALT lymphoma: anti-angiogenesis treatment using VEGF receptor antibodies and celecoxib. *Curr Pharm Des.* 2014;20(7):1097–103.
54. Argyropoulos KV, Ziegler CGK, Altan-Bonnet G, et al. Multiparameter phosphoproteomic profiling of the B-cell receptor pathway in Waldenström's macroglobulinemia. *Blood.* 2014;124(21):141.
55. Treon SP, Xu L, Yang G, et al. MYD88 L265P somatic mutation in Waldenström's macroglobulinemia. *N Engl J Med.* 2012;367(9):826–33.
56. Yang G, Zhou Y, Liu X, et al. A mutation in MYD88 (L265P) supports the survival of lymphoplasmacytic cells by activation of Bruton tyrosine kinase in Waldenström macroglobulinemia. *Blood.* 2013;122(7):1222–32.
57. Tripsas CK, Yang G, Cao Y, et al. A prospective multicenter study of the Bruton's tyrosine kinase inhibitor ibrutinib in patients with relapsed or refractory Waldenström's macroglobulinemia. *Blood.* 2013;122(21):251.
58. Avalos AM, Meyer-Wentrup F, Ploegh HL. B-cell receptor signaling in lymphoid malignancies and autoimmunity. *Adv Immunol.* 2014;123:1–49.
59. Arons E, Roth L, Sapolsky J, Suntum T, Stetler-Stevenson M, Kreitman RJ. Evidence of canonical somatic hypermutation in hairy cell leukemia. *Blood.* 2011;117(18):4844–51.
60. Jones JA, Andritsos LA, Lucas DM, et al. Preliminary safety and efficacy of the Bruton's tyrosine kinase (BTK) inhibitor ibrutinib (IBR) in patients (pts) with hairy cell leukemia (HCL). *J Clin Oncol.* 2014;32(suppl):7063.
61. Sivina M, Kreitman RJ, Arons E, Ravandi F, Burger JA. The Bruton tyrosine kinase inhibitor ibrutinib (PCI-32765) blocks hairy cell leukemia survival, proliferation and B cell receptor signalling: a new therapeutic approach. *Br J Haematol.* 2014;166(2):177–88.
62. Minami Y, Oishi I, Endo M, Nishita M. Ror-family receptor tyrosine kinases in noncanonical Wnt signaling: their implications in developmental morphogenesis and human diseases. *Dev Dyn.* 2010;239(1):1–15.
63. Al-Shawi R, Ashton SV, Underwood C, Simons JP. Expression of the Ror1 and Ror2 receptor tyrosine kinase genes during mouse development. *Dev Genes Evol.* 2001;211(4):161–71.
64. Lynch SA, Brugge JS, Fromowitz F, et al. Increased expression of the src proto-oncogene in hairy cell leukemia and a subgroup of B-cell lymphomas. *Leukemia.* 1993;7(9):1416–22.
65. Avet-Loiseau H, Soulier J, Feraud JP, et al. IFM and MAG Groups. Impact of high-risk cytogenetics and prior therapy on outcomes in patients with advanced relapsed or refractory multiple myeloma treated with lenalidomide plus dexamethasone. *Leukemia.* 2010;24(3):623–8.
66. Gutiérrez NC, Castellanos MV, Martín ML, et al. GEM/PETHEMA Spanish Group. Prognostic and biological implications of genetic abnormalities in multiple myeloma undergoing autologous stem cell transplantation: t(4;14) is the most relevant adverse prognostic factor, whereas RB deletion as a unique abnormality is not associated with adverse prognosis. *Leukemia.* 2007;21(1):143–50.
67. Schmidt-Hieber M, Gutiérrez ML, Pérez-Andrés M, et al. Cytogenetic profiles in multiple myeloma and monoclonal gammopathy of undetermined significance: a study in highly purified aberrant plasma cells. *Haematologica.* 2013;98(2):279–87.
68. Chesi M, Brents LA, Ely SA, et al. Activated fibroblast growth factor receptor 3 is an oncogene that contributes to tumor progression in multiple myeloma. *Blood.* 2001;97(3):729–36.
69. Kanai M, Gōke M, Tsunekawa S, Podolsky DK. Signal transduction pathway of human fibroblast growth factor receptor 3. Identification of a novel 66-kDa phosphoprotein. *J Biol Chem.* 1997;272(10):6621–8.
70. Trudel S, Li ZH, Wei E, et al. CHIR-258, a novel, multitargeted tyrosine kinase inhibitor for the potential treatment of t(4;14) multiple myeloma. *Blood.* 2005;105(7):2941–2948.





71. Xin X, Abrams TJ, Hollenbach PW, et al. CHIR-258 is efficacious in a newly developed fibroblast growth factor receptor 3-expressing orthotopic multiple myeloma model in mice. *Clin Cancer Res*. 2006;12(16):4908–15.
72. Scheid C, Reece D, Beksac M, et al. Phase 2 study of dovitinib in patients with relapsed or refractory multiple myeloma with or without t(4;14) translocation. *Eur J Haematol*. 2014;doi: 10.1111/ejh.12491.
73. Schmidt-Hieber M, Pérez-Andrés M, Paiva B, et al. CD117 expression in gammopathies is associated with an altered maturation of the myeloid and lymphoid hematopoietic cell compartments and favorable disease features. *Haematologica*. 2011;96(2):328–32.
74. Liu D, Lin P, Hu Y, et al. Immunophenotypic heterogeneity of normal plasma cells: comparison with minimal residual plasma cell myeloma. *J Clin Pathol*. 2012;65(9):823–9.
75. Dispenzieri A, Gertz MA, Lacy MQ, et al. A phase II trial of imatinib in patients with refractory/relapsed myeloma. *Leuk Lymphoma*. 2006;47(1):39–42.
76. Zhang Y, Moschetta M, Huynh D, et al. Pyk2 promotes tumor progression in multiple myeloma. *Blood*. 2014;124(17):2675–86.
77. Meyer AN, Gastwirt RF, Schlaepfer DD, Donoghue DJ. The cytoplasmic tyrosine kinase Pyk2 as a novel effector of fibroblast growth factor receptor 3 activation. *J Biol Chem*. 2004;279(27):28450–7.
78. Bam R, Ling W, Khan S, et al. Role of Bruton's tyrosine kinase in myeloma cell migration and induction of bone disease. *Am J Hematol*. 2013;88(6):463–71.
79. de Brito LR, Batey MA, Zhao Y, et al. Comparative pre-clinical evaluation of receptor tyrosine kinase inhibitors for the treatment of multiple myeloma. *Leuk Res*. 2011;35(9):1233–40.
80. Kovacs MJ, Reece DE, Marcellus D, et al. A phase II study of ZD6474 (Zactima), a selective inhibitor of VEGFR and EGFR tyrosine kinase in patients with relapsed multiple myeloma – NCIC CTG IND.145. *Invest New Drugs*. 2006;24(6):529–35.
81. Lin B, Podar K, Gupta D, et al. The vascular endothelial growth factor receptor tyrosine kinase inhibitor PTK787/ZK222584 inhibits growth and migration of multiple myeloma cells in the bone marrow microenvironment. *Cancer Res*. 2002;62(17):5019–26.
82. Shanafelt T, Zent C, Byrd J, et al. Phase II trials of single-agent anti-VEGF therapy for patients with chronic lymphocytic leukemia. *Leuk Lymphoma*. 2010;51(12):2222–9.
83. Kadia T, Delioukina ML, Kantarjian HM, et al. A pilot phase II study of the Lyn kinase inhibitor bafetinib in patients with relapsed or refractory B cell chronic lymphocytic leukemia. *Blood*. 2011;118:2858.
84. Brown J, Harb W, Sharman J, et al. Phase I study of single agent CC-292, a highly selective Bruton's tyrosine kinase (BTK) inhibitor, in relapsed/refractory chronic lymphocytic leukemia (CLL) and B-cell non-Hodgkin lymphoma (BNHL). *European Hematology Association*. 2013:3793.
85. Harb WA, Hill BT, Gabrilove J, et al. Phase 1 study of single agent CC-292, a highly selective bruton's tyrosine kinase (BTK) inhibitor, in relapsed/refractory chronic lymphocytic leukemia (CLL). *Blood*. 2013;122(21):1630.
86. Wildes TM, Procknow E, Gao F, Dipersio JF, Vij R. Dasatinib in relapsed or plateau-phase multiple myeloma. *Leuk Lymphoma*. 2009;50(1):137–40.
87. O'Brien S, Furman RR, Coutre SE, et al. Ibrutinib as initial therapy for elderly patients with chronic lymphocytic leukaemia or small lymphocytic lymphoma: an open-label, multicentre, phase 1b/2 trial. *Lancet Oncol*. 2014;15(1):48–58.
88. Vij R, Huff CA, Bensinger WI, et al. Ibrutinib, single agent or in combination with dexamethasone, in patients with relapsed or relapsed/refractory multiple myeloma (MM): preliminary phase 2 results. *Blood*. 2014;124(21):31.
89. Barrientos JC, Barr PM, Flinn I, et al. Ibrutinib in combination with bendamustine and rituximab is active and tolerable in patients with relapsed/refractory CLL/SLL: final results of a phase 1b study. *Blood*. 2013;122(21):525.
90. Pollyea DA, Coutre S, Gore L, et al. A dose escalation study of ibrutinib with lenalidomide for relapsed and refractory chronic lymphocytic leukemia/small lymphocytic lymphoma. *Blood*. 2014;124(21):1987.
91. Jaglowski SM, Jones JA, Flynn JM. A phase 1b/2 study evaluating activity and tolerability of the BTK inhibitor ibrutinib in combination with ofatumumab in patients with chronic lymphocytic leukemia/small lymphocytic lymphoma (CLL/SLL) and related diseases. *J Clin Oncol*. 2014;32:7009.
92. Kim E, Werner L, Keating MJ, et al. Addition of rituximab abrogates ibrutinib-induced lymphocytosis and promotes more rapid decrease in absolute lymphocyte counts in patients with relapsed chronic lymphocytic leukemia. *Blood*. 2014;124(21):1998.
93. Keating MJ, Wierda WG, Hoellenriegel J, et al. Ibrutinib in combination with rituximab (iR) is well tolerated and induces a high rate of durable remissions in patients with high-risk chronic lymphocytic leukemia (CLL): new, updated results of a phase II trial in 40 patients. *Blood*. 2013;122(21):675.
94. Hebb J, Assouline S, Rousseau C, et al. A phase I study of imatinib mesylate in combination with chlorambucil in previously treated chronic lymphocytic leukemia patients. *Cancer Chemother Pharmacol*. 2011;68(3):643–51.
95. Kropff M, Kienast J, Bisping G, et al. An open-label dose-escalation study of BIBF 1120 in patients with relapsed or refractory multiple myeloma. *Anticancer Res*. 2009;29(10):4233–8.
96. Rule S, Shah N, Salles GA, et al. A phase I study of the oral Btk inhibitor ONO-4059 in patients with relapsed/refractory B-cell lymphoma. *Blood*. 2013;122:4397.
97. Guidetti A, Carlo-Stella C, Devizzi L, et al. Preliminary results of a phase II trial with the multikinase inhibitor sorafenib in heavily pretreated patients with relapsed/refractory non-hodgkin lymphoma (NHL). *Blood*. 2009;114:1658.