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### Research Article

# The Influence of Interleukin 17A and IL17F Polymorphisms on Chronic Periodontitis Disease in Brazilian Patients

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A case-control study was conducted on patients with chronic periodontitis (CP) and healthy controls with the aim of evaluating possible association between interleukin 17A (IL17A) G197A (rs2275913) and IL17F T7488C (rs763780) polymorphisms and periodontitis. Genotypes were determined by PCR-RFLP method. Statistical analyses were conducted using the OpenEpi and SNPStas software to calculate Chi-square with Yates correction or Fisher's exact tests, odds ratios (OR), and 95% confidence intervals (CIs). SNPStas software was used to calculate Hardy-Weinberg equilibrium. IL17A AA genotype was more frequent in patients with chronic periodontitis (CP) in the codominant and recessive models (P = 0.09; OR = 2.53 and P = 0.03; OR = 2.46, resp.), the females with CP (P = 0.01, OR = 4.34), Caucasoid patients with CP (P = 0.01, OR = 3.45), and nonsmoking Caucasian patients with CP (P = 0.04, OR = 3.51). The IL17A A allele was also more frequent in Caucasians with CP (P = 0.04, OR = 1.59). IL17F T7488C polymorphism was not associated with chronic periodontitis. In these patients from Southern Brazil, the IL17A rs2275913 polymorphisms, IL17A AA genotype, and the A allele were associated with a susceptibility to chronic periodontitis.

#### 1. Introduction

Periodontitis is a chronic inflammatory disease that affects the tooth supporting tissue and destroys alveolar bone. It is the most frequent cause of tooth loss in the adult [1]. Epidemiologic studies suggest that up to 60% of the population is affected by the common form of the disease, termed chronic periodontitis (CP) [1, 2]. Periodontitis has been said to have interaction with a number of common human diseases like diabetes mellitus or rheumatoid arthritis. Epidemiological data has confirmed that diabetes has been a major risk factor for the onset of periodontitis and showed a clear relationship between degree of hyperglycemia and severity of periodontitis [3]. Other studies show that patients with rheumatoid arthritis had a higher incidence of periodontitis compared to healthy controls [4, 5], though the mechanism of

periodontitis that interacted with other diseases was not very clear.

Periodontitis is a multifactorial disease and as such, the significant elements include not only the presence of pathogenic bacteria and the immune mechanism, but also the genetic predisposition [6]. From a pathophysiology perspective, periodontitis is the result of host-mediated inflammatory damage of the supporting tissues triggered in response to the microbial infection [7, 8]. More than 700 different bacterial species have been shown to inhabit periodontal biofilms [9] and some species are currently considered to be causally associated with periodontitis; these include Gramnegative species, such as anaerobic *Porphyromonas gingivalis*, a principal pathogen in chronic periodontitis [10].

Specific cytokines are important in the pathogenic process of the periodontitis [8, 11–13]. Interleukin-17 (IL-17) is a

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proinflammatory cytokine secreted by activated T cells [14]. The IL-17 family contains six members, IL-17A, IL-17B, IL-17C, IL-17D, IL-17E (or IL-25), and IL-17F, and five receptors, IL-17RA-RD and SEF. Interleukin-17A is most homologous to IL-17F and the genes encoding them are proximally located on chromosome 6p12 [15]. The IL-17F activity is similar to IL-17A but significantly weaker and is related to inducing the expression of various cytokines, chemokines, matrix metalloproteinases, antimicrobial peptides, and adhesion molecules by human fibroblasts, airway epithelial cells, and vein endothelial cells [16].

Th17 cells were a distinct T lineage that do not share developmental pathways with either Th1 or Th2 cells [17]. Th17 cells have been linked to several autoimmune disorders and are also linked to the development of pathological inflammatory disorders. However Th17 cells are physiologically found in the lamina propria of the intestine [17, 18]. Cytokines related with Th17, such as IL-17 and IL-22, are crucial for host protection against many extracellular pathogens. IL-17 stimulates the production and expression of TNF-alpha and IL-1 beta by human macrophages [19] and induces production of IL-1 beta in osteoblasts [20]. Thus, IL-17 was found to contribute to inflammatory bone pathology as in rheumatoid arthritis and inflammatory bowel diseases and was centrally involved in numerous autoimmune disorders [21–25].

IL-17 cytokine can stimulate fibroblasts, epithelial and endothelial cells, to produce IL-6, CXCL8/IL-8, and prostaglandin E2 (PGE2) [26]. IL-17 also induces the expression of receptor activator of nuclear factor kappa B ligand (RANKL) on osteoblasts and stimulates the differentiation and activation of osteoclasts, which can influence bone resorption mediated by these cells [22]. Many studies have demonstrated the presence of IL-17 in periodontal tissues, crevicular gingival fluid, saliva, and plasma of patients with periodontal disease [27–30].

In order to investigate whether *IL17A* and the *IL17F* polymorphisms are associated with chronic periodontal disease and understand its immunopathogenesis, this study aimed at evaluating the *IL17A* G197A (rs2275913) and *IL17F* T7488C (His161Arg, rs763780) polymorphisms in patients with chronic periodontitis and in a healthy group who had undergone dental care in the North/Northwest of the state of Paraná, Southern Brazil.

#### 2. Materials and Methods

2.1. Sample Selection. A total of 313 individuals were selected from those who sought dental treatment in the dental clinics of the Maringa State University (UEM) and Inga University (UNINGÁ) from January 2012 to December 2014. After taking patient's medical records, clinical periodontal examinations were conducted by two examiners. Clinical parameters of probing depth (PD) and clinical attachment level (CAL) were examined at six sites (mesiovestibular, vestibular, distovestibular, mesiolingual, lingual, and distolingual) of each tooth, as was bleeding on probing (BOP).

After the periodontal examination, participants were assigned to two different groups: the chronic periodontitis

group (N = 140) composed of individuals who had at least 5 sites in different teeth with PD  $\geq$ 5 mm, CAL  $\geq$ 3 mm, and more than 25% of BOP and the control group (N = 173), formed by individuals who did not have sites with reduced CAL, displayed a PD of less than 4 mm, and exhibited less than 25% of BOP. Both patients and controls were from the North and Northwest regions of the state of Paraná (between 22°29′30″-26°42′59″S and 48°02′24″-54°37′38″W), Southern Brazil, over 30 years of age, from all ethnic groups, and with at least 20 teeth in the oral cavity. Information on the patient's ethnic background and smoking history was obtained by interviewing the individual (anamnesis). Exclusion criterions were patients and control subjects with diabetes mellitus and acute infections and patients with aggressive periodontitis and who had periodontal treatment in the last 6 months.

All individuals who agreed to participate in this research were informed about the nature of the study and signed an informed consent form. This study was approved by the Human Research Ethics Committee of the Maringa State University (UEM: number 719/2011, 02/12/2011).

2.2. DNA Extraction. To extract the DNA, the buffy coat was obtained from 4 mL of peripheral blood collected in EDTA by centrifugation (210 g for 15 min). The DNA was extracted using the salting-out method [31]. The concentration and quality of the DNA were analyzed by optical density in a Thermo Scientific Nanodrop 2000 apparatus (Wilmington, USA).

2.3. Genotyping Analysis. Single nucleotide polymorphisms (SNP) to IL17A G197A (rs2275913) and IL17F T7488C (rs763780, His161Arg) genotyping were performed by Polymerase Chain Reaction-Restriction Fragment Length Polymorphism (PCR-RFLP) [32] with some modifications. Primer sequences for IL17A G197A were sense 5'-AACAAG-TAAĜAATGAAAAGAGGACATGGT-3' and antisense 5'-CCCCCAATGAGGTCATAGAAGAATC-3' and for IL17F T7488C were sense 5'-ACCAAGGCTGCTCTGTTTCT-3' and antisense 5'-GGTAAGGAGTGGCATTTCTA-3'. PCR amplification was performed in a total volume of 30  $\mu$ L mixture containing 100 ng genomic DNA, 1.0  $\mu$ M of each primer, 200  $\mu$ M of each dNTP, 2.0 mM of MgCl<sub>2</sub>, 3  $\mu$ L 10x PCR buffer, and 1.5 U Taq DNA polymerase (Invitrogen Life Technologies, Grand Island, NY, USA). PCR products were digested for one hour at 37°C with XagI (Fermentas, Canada) for IL17A G197A and NlaIII (New England Biolabs) for IL17F T7488C and then separated by electrophoresis on 3% agarose with SYBR Green (Invitrogen Life Technologies, Grand Island, NY, USA).

2.4. Statistical Analysis. Allele and genotype frequencies of IL17A G197A and IL17F T7488C were obtained by direct counts. The association between genetic polymorphisms and chronic periodontitis was evaluated using the Chi-square test with Yates correction or the Fisher's exact test and the correlation was deemed present by an odds ratio with 95% confidence intervals only for significant P values. Adjusting the

TABLE 1: Characteristics of patients with chronic periodontitis (CP) and controls.

	CP patients	Controls	P	OR (95% CI)	
	n (%)	n (%)	Γ	OR (95% CI)	
	N = 140	N = 173			
Gender					
Male	66 (47)	56 (32)			
Female	74 (53)	117 (68)			
Age					
Mean $\pm$ SD (year)	$47.03 \pm 9.21$	$45.61 \pm 9.18$			
Ethnic origin					
Caucasian	84 (60)	118 (68)			
Mixed	36 (26)	40 (23)			
Black	18 (13)	15 (9)			
Not declared	2 (1)	0			
Smoking					
Nonsmokers	56 (40)	127 (73)	< 0.001	0.24 (0.149-0.39)	
Smokers	33 (24)	20 (12)	0.008	2.78 (1.33–5.78)	
Ex-smokers	51 (36)	26 (15)	< 0.001	3.24 (1.89-5.56)	
Nonsmoker	<i>N</i> = 56	N = 127			
Gender					
Male	20 (35.7)	33 (26.0)			
Female	36 (64.28)	94 (74.0)			
Age					
Mean $\pm$ SD (year)	$46.5 \pm 9.3$	$45.5 \pm 9.9$			
Ethnic origin					
Caucasian	36 (64.28)	86 (67.7)			
Mixed	16 (28.6)	28 (22.0)			
Black	3 (5.4)	13 (10.3)			
Not declared	1 (1.8)	0			

n: number; P = P value; OR: odds ratio; CI: confidence interval.

genotypic differences for the effect of age, gender, and smoking status was applied. All tests were carried out using a significance level of 5%. For these analyses and calculating Hardy-Weinberg equilibrium OpenEpi program Version 2.3.1 and SNPStas software (http://bioinfo.iconcologia.net/index.php) were used.

#### 3. Results

Polymorphisms in the *IL17A* G197A (rs2275913) and *IL17F* T7488C (rs763780) were analyzed in 140 CP patients and 173 control subjects. Most participants were female (61.0%), Caucasian (64.5%), and nonsmokers (58.5%). The characteristics of patient and control subjects are described in Table 1.

Differences were noted in smoking history when the groups were compared, and to eliminate the smoking as a confounding factor, all analyses were also done in the non-smoker patients versus nonsmoker controls. There was no significant difference with respect to gender, age, and ethnic background distributions.

Genotype distribution of *IL17A* G197A and *IL17F* T7488C in CP patient and control groups was consistent with the

Hardy-Weinberg equilibrium (P > 0.05). The *IL17A* and *IL17F* were not in linkage disequilibrium (P = 0.51).

The genotype and allele frequencies distributions are summarized in Table 2. There were significant differences in the codominant and recessive models for *IL17A* AA genotype between all CP patients and controls. No significant difference was observed for *IL17F* in the recessive, dominant, and codominant models.

We analyzed *IL17A* and *IL17F* genotype and allele frequencies in CP and controls after stratifying according to gender and ethnic background. No significant difference was observed for *IL17F* polymorphism after stratification. However *IL17A* AA genotype was more frequent in female CP patients (16.2% *versus* 4.3%, P=0.01, OR = 4.34, and 95% CI:1.46–12.87) (Table 3). *IL17A* AA genotype and A allele were more frequent in Caucasoid CP patients than in controls (17.8% *versus* 5.9%, P=0.01, OR = 3.45, and 95% CI: 1.34–8.88, and 38.7% *versus* 28.4%, P=0.04, OR = 1.59, and 95% CI: 1.05–2.42, resp.). Difference was also found for *IL17A* AA genotype that was more frequent in the Caucasian and nonsmoking CP patients than in the Caucasian nonsmoking controls (22.2% *versus* 8.1%, P=0.048, OR = 3.51, and 95% CI: 1.17–10.55) (Table 4).

TABLE 2: Genotypes and allele distribution of *IL17A* rs2275913 and *IL17F* rs763780 in the patients with chronic periodontitis and controls from Southern Brazil.

	All subjects			Nonsmokers		
Genotype	Patients	Controls		Patients	Controls	
	N = 140	N = 173		N = 56	N = 127	
	n (%)	n (%)		n (%)	n (%)	
rs2275913						
GG	67 (47.9)	87 (50.3)	Ref.	25 (44.6)	61 (48.0)	Ref.
AA	18 (12.9)	12 (7.0)	*	9 (16.1)	10 (7.9)	
AG	55 (39.2)	74 (42.7)		22 (39.3)	56 (44.1)	
AA/AG	73 (52.1)	86 (49.7)		31 (55.4)	66 (52.0)	
GG/AG	122 (87.1)	99 (57.2)		47 (83.9)	117 (92.1)	
Allele						
A	91 (32.5)	98 (28.3)		40 (35.7)	76 (30.0)	
G	189 (67.5)	248 (71.7)		72 (64.3)	178 (70.0)	
rs763780						
TT	125 (89.3)	158 (91.3)	Ref.	49 (87.5)	116 (91.3)	Ref.
CC	1 (0.7)	0		1 (1.8)	0	
TC	14 (10.0)	15 (8.7)		6 (10.7)	11 (8.7)	
CC/TC	15 (10.7)	15 (8.7)		7 (92.9)	11 (8.7)	
TT/TC	139 (99.3)	173 (100)		55 (98.2)	127 (100)	
Allele						
T	264 (94.3)	331 (95.7)		104 (92.9)	243 (95.7)	
С	16 (5.7)	15 (4.3)		8 (7.1)	11 (4.3)	

<sup>\*</sup>Codominant model: P = 0.09; OR = 2.53; 95% CI = 1.07–5.99; recessive model: P = 0.03; OR = 2.46; 95% CI = 1.08–5.59.

Table 3:  $\it IL17A$  genotype and allele frequencies in chronic periodontitis Brazilian patients and controls stratified according to gender\* in the total samples and nonsmoking individuals.

	<i>IL17A</i> rs2275913	CP patients n (%)	Controls n (%)	
	Genotype	N = 74	N = 117	
	GG	36 (48.7)	63 (53.8)	
	$AA^{**}$	12 (16.2)	5 (4.3)	
Female	GA	26 (35.1)	49 (41.9)	
	Allele			
	A	50 (33.8)	59 (25.2)	
	G	98 (66.2)	175 (74.8)	
	Genotype	N = 14	N = 33	
	GG	6 (42.9)	11 (33.3)	
Nonsmoker	AA	2 (14.2)	5 (15.2)	
female	GA	6 (42.9)	17 (51.5)	
	Allele			
	A	10 (35.7)	27 (40.9)	
	G	18 (64.3)	39 (59.1)	

CP: chronic periodontitis; \* only significant differences were showed; \*\*P = 0.01, OR = 4.34, and 95% CI: 1.46–12.87.

#### 4. Discussion

In this study we investigated a possible role of *IL17A* and *IL17F* polymorphisms in immunopathogenic mechanism for CP in a Southern Brazilian population. We observed that the *IL17A* 197AA genotype was more frequent in patients with chronic periodontitis (CP), females with CP, and the Caucasian and nonsmoking Caucasian patients with CP than in respective controls, and this could be correlated to the risk of disease. The *IL17F* T7488C polymorphism was not associated with CP risk in this population.

In a case-control study, pairing between study subjects is necessary in order to avoid bias in the final results. CP can be related to individual risk factors such as stress, diabetes, osteoporosis, and arthritis [3-5, 33-37] and, in this studied population, they were considered confounding factors and were an exclusion criterion. On the other hand, smoking habits, also a predisposition factor with oral diseases and especially in the chronic periodontitis [38-41], were more frequent in CP patients (smoker + ex-smoker: 60% versus 40%; P < 0.001, OR = 4.12, and 95% CI: 2.56-6.69); thus, to exclude smoking as a predisposing factor, statistical analyses were performed in all the individuals as well as in the nonsmokers CP versus nonsmokers in control group, and stratifying by smoking habits was not considered. Some studies alert that gender can be a confounding factor to periodontitis [42] although others showed the higher frequency in male was principally related to personal hygiene habits [43]; furthermore, it was revealed that there was no meaningful relation to the role of IL-17 polymorphism in men and in women when compared to each other [44]. With regard to ethnic background, the distributions of patients and controls were similar: this is important because IL17A and IL17F alleles and genotypes differ in different populations (http://www.snpedia.com/index.php/Rs2275913 and /Rs763780) and Brazilians are an admixed population. Although ethnicity was not a confounding variable, the analyses were done after stratification by the Caucasian, Black, and mixed population.

In the present report IL17F was not associated with periodontal disease in all the patients nor in nonsmoking patients and controls who were investigated in the recessive, dominant, and codominant models. However, IL17A 197AA genotype was more frequent in CP patients, in female patients, and in the Caucasian CP patients as well as in the nonsmoking Caucasian patients with CP. Furthermore, IL17A 197A allele was also more frequent in Caucasians with CP. These results suggest that IL17A 197AA genotype and A allele could be related to higher risk for the development of chronic periodontitis. In our Caucasian group with CP the IL17A 197G allele was less frequent and could be a resistant factor. The polymorphism in the promoter region of cytokines may be related to higher expression of the specific cytokine. According to Espinoza et al. [45] the IL17A 197A allele correlates to more efficient IL-17 secretion and higher affinity for the nuclear factor of activated T cells (NFAT), which is a critical regulator of the *IL17* promoter gene [46].

The *IL17A* 197AA genotype was associated with tumorigenesis susceptibility as in gastric cancer [47, 48], breast

Table 4: IL17A genotype and allele frequencies in chronic periodontitis Brazilian patients and controls stratified according to ethnic group\*.

	<i>IL17A</i> rs2275913	CP patients n (%)	Controls n (%)	P	OR (95% CI)
Caucasian	Genotype	N = 84	N = 118		
	GG	34 (40.5)	58 (49.2)		
	AA	15 (17.8)	7 (5.9)	0.01	3.45 (1.34-8.88)
	GA	35 (41.7)	53 (44.9)		
	Allele				
	A	65 (38.7)	67 (28.4)	0.04	1.59 (1.05-2.42)
	G	103 (61.3)	169 (71.6)	0.04	0.63 (0.41-0.96)
Nonsmoker Caucasian	Genotype	N = 36	N = 86		
	GG	15 (41.7)	41 (47.7)		
	AA	8 (22.2)	7 (8.1)	0.048	3.51 (1.17–10.55)
	GA	13 (36.1)	38 (44.2)		
	Allele				
	A	29 (40.3)	52 (30.2)		
	G	43 (59.7)	120 (69.8)		

CP: chronic periodontitis; \*only significant differences were showed.

cancer [49], and cervical cancer [50], as well as autoimmunity diseases such as ulcerative colitis [51, 52] and rheumatoid arthritis [53]. As for periodontitis diseases, only two previous studies in the Brazilian patients were reported. Corrêa et al. [54] showed similar results to our study: *IL17A* 197AA genotype and A allele were associated with worse clinical and inflammatory periodontal parameters. Saraiva et al. [55] conducted another study on chronic periodontitis in Brazil and severe periodontitis patients with the aim of investigating the phenotypic expression of *IL17A* and the polymorphisms of *IL17A* and *IL17F* within different clinical forms and severity of the disease. However, differently to our and Corrêa et al.'s results [54], their data suggested that the IL-17 and *IL17A* A allele were associated with the absence of periodontal disease, and the *IL17A* GG genotype and G allele were associated with risk factors.

According to the distribution of *IL17A* genotypes frequencies in our control group, *IL17A* AA was 7.0%, *IL17A* AG was 42.7%, and *IL17A* GG was 50.3%, similar to those in the study conducted by Saraiva et al. [55] which were 10% for AA, 44% for AG, and 46% for GG but very different from those of Corrêa et al. [54] which were 25.9% for AA, 14.81% for AG, and 59.26% for GG.

The *IL17F* T7488C polymorphism was not associated with chronic periodontitis in this study. This result was similar to Corrêa et al. [54] and Saraiva et al. [55] in CP Brazilian patients. *IL17F* has been associated with several diseases like asthma, Crohn's disease, multiple sclerosis, inflammatory bowel disease, autoimmune thyroid diseases, tuberculosis, and dilated cardiomyopathy as well as a high risk of recurrent pregnancy loss [51, 56–61]. According to the literature, the *IL-17F* activity is similar to *IL-17A* but significantly weaker, and the variant form of *IL-17* protein (His121Arg) suppresses the expression and the activity of wild type [16, 62].

According to the *IL17F* genotype frequency distribution in our control group, we had 8.7% for *IL17F* TC and 91.3% for *IL17F* TT; the *IL17F* CC genotype was absent in our control

population. Again, it was similar to the study conducted by Saraiva et al. [55] which found 6.4% for TC and 93.6% for TT and did not find the *IL17F* CC genotype. However, the distribution of genotype frequencies found by Corrêa et al. [54] was also very different: they found 16.66% for *IL17F* TC, 56.66% for *IL17F* TT, and 23.33% for *IL17F* CC in their Brazilian control group. The distribution of genotype frequency in our populations was consistent with a low frequency of polymorphic genotype of *IL17F* T7488C found in other populations (http://www.snpedia.com/index.php/Rs763780).

CP progression was related to a host inflammatory response that mediates tissue damage, and several previous studies had been relating immune genetic factors to CP disease, especially cytokines genes polymorphism [63-69]. The IL-17, a proinflammatory cytokine, was detected in periodontal tissues, crevicular gingival fluid, saliva, and plasma of patients with periodontal disease [27-30, 70]. IL-17, especially when combined with IFN-gamma, may have a role in immune modulation through stimulation of human gingival fibroblasts in periodontal disease [71]. This occurs by triggering the release of other proinflammatory, metalloproteinase, and neutrophil-mobilizing cytokines [47, 49, 51, 72] and having effects on osteoclasts maturity as a stimulating factor [73, 74]. IL-17A and IL-17F have a very similar amino acid sequence and both play similar functions and have the ability to induce chemokines which is crucial to the neutrophil recruitment and activation [75], the first wall in the periodontal diseases. Thus, understanding *IL17* polymorphism is necessary in order to infer the role of IL-17 in the CP immunopathogenesis.

In these Southern Brazilian patients, the *IL17A* rs2275913 polymorphisms, AA genotype, and A allele were associated with a susceptibility to chronic periodontitis disease, in females with CP, and in the Caucasian and nonsmoking Caucasian patients. Furthermore, the possible immunopathogenic mechanism would be investigated in the future through histological studies.

#### 5. Conclusion

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In conclusion we can infer that *IL17A* G197A rs2275913 polymorphism, *IL17A* AA genotype, and A allele could be associated with a susceptibility to chronic periodontitis but no evidence showed for risk or protection associations for *IL17F* T7488C rs763780. Additional studies are necessary for understanding the functional role of rs2275913 polymorphisms in chronic periodontitis.

#### **Conflict of Interests**

The authors declare that they have no conflict of interests.

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#### References

- [1] R. C. Page, "Milestones in periodontal research and the remaining critical issues," *Journal of Periodontal Research*, vol. 34, no. 7, pp. 331–339, 1999.
- [2] R. T. Demmer and P. N. Papapanou, "Epidemiologic patterns of chronic and aggressive periodontitis," *Periodontology 2000*, vol. 53, no. 1, pp. 28–44, 2010.
- [3] P. M. Preshaw, A. L. Alba, D. Herrera et al., "Periodontitis and diabetes: a two-way relationship," *Diabetologia*, vol. 55, no. 1, pp. 21–31, 2012.
- [4] D. Potikuri, K. C. Dannana, S. Kanchinadam et al., "Periodontal disease is significantly higher in non-smoking treatment-naive rheumatoid arthritis patients: results from a case-control study," *Annals of the Rheumatic Diseases*, vol. 71, no. 9, pp. 1541–1544, 2012.
- [5] J. Detert, N. Pischon, G. R. Burmester, and F. Buttgereit, "The association between rheumatoid arthritis and periodontal disease," *Arthritis Research and Therapy*, vol. 12, no. 5, article 218, 2010
- [6] L. N. Borrell and P. N. Papapanou, "Analytical epidemiology of periodontitis," *Journal of Clinical Periodontology*, vol. 32, no. 6, pp. 132–158, 2005.
- [7] Y.-C. G. Liu, U. H. Lerner, and Y.-T. A. Teng, "Cytokine responses against periodontal infection: protective and destructive roles," *Periodontology* 2000, vol. 52, no. 1, pp. 163–206, 2010.
- [8] D. Graves, "Cytokines that promote periodontal tissue destruction," *Journal of Periodontology*, vol. 79, no. 8, pp. 1585–1591, 2008.
- [9] P. S. Kumar, A. L. Griffen, M. L. Moeschberger, and E. J. Leys, "Identification of candidate periodontal pathogens and beneficial species by quantitative 16S clonal analysis," *Journal of Clinical Microbiology*, vol. 43, no. 8, pp. 3944–3955, 2005.
- [10] R. P. Darveau, "Periodontitis: a polymicrobial disruption of host homeostasis," *Nature Reviews Microbiology*, vol. 8, no. 7, pp. 481– 490, 2010.
- [11] P. C. Trevilatto, A. P. de Souza Pardo, R. M. Scarel-Caminaga et al., "Association of IL1 gene polymorphisms with chronic periodontitis in Brazilians," *Archives of Oral Biology*, vol. 56, no. 1, pp. 54–62, 2011.

[12] K. S. Kornman, A. Crane, H. Y. Wang et al., "The interleukin-1 genotype as a severity factor in adult periodontal disease," *Journal of Clinical Periodontology*, vol. 24, no. 1, pp. 72–77, 1997.

- [13] D. T. Graves, "The potential role of chemokines and inflammatory cytokines in periodontal disease progression," *Clinical Infectious Diseases*, vol. 28, no. 3, pp. 482–490, 1999.
- [14] H. E. Broxmeyer, "Is interleukin 17, an inducible cytokine that stimulates production of other cytokines, merely a redundant player in a sea of other biomolecules?" *The Journal of Experimental Medicine*, vol. 183, no. 6, pp. 2411–2415, 1996.
- [15] C. T. Weaver, L. E. Harrington, P. R. Mangan, M. Gavrieli, and K. M. Murphy, "Th17: an effector CD4 T Cell lineage with regulatory T Cell ties," *Immunity*, vol. 24, no. 6, pp. 677–688, 2006.
- [16] N. Hizawa, M. Kawaguchi, S.-K. Huang, and M. Nishimura, "Role of interleukin-17F in chronic inflammatory and allergic lung disease," *Clinical & Experimental Allergy*, vol. 36, no. 9, pp. 1109–1114, 2006.
- [17] R. A. Kastelein, C. A. Hunter, and D. J. Cua, "Discovery and biology of IL-23 and IL-27: related but functionally distinct regulators of inflammation," *Annual Review of Immunology*, vol. 25, pp. 221–242, 2007.
- [18] C. Dong, "TH17 cells in development: an updated view of their molecular identity and genetic programming," *Nature Reviews Immunology*, vol. 8, no. 5, pp. 337–348, 2008.
- [19] D. V. Jovanovic, J. A. Di Battista, J. Martel-Pelletier et al., "IL-17 stimulates the production and expression of proinflammatory cytokines, IL- $\beta$  and TNF- $\alpha$ , by human macrophages," *The Journal of Immunology*, vol. 160, no. 7, pp. 3513–3521, 1998.
- [20] L. Rifas and L. V. Avioli, "A novel T cell cytokine stimulates interleukin-6 in human osteoblastic cells," *Journal of Bone and Mineral Research*, vol. 14, no. 7, pp. 1096–1103, 1999.
- [21] S. L. Gaffen, "Biology of recently discovered cytokines: interleukin-17—a unique inflammatory cytokine with roles in bone biology and arthritis," *Arthritis Research and Therapy*, vol. 6, no. 6, pp. 240–247, 2004.
- [22] S. Kotake, N. Udagawa, N. Takahashi et al., "IL-17 in synovial fluids from patients with rheumatoid arthritis is a potent stimulator of osteoclastogenesis," *Journal of Clinical Investigation*, vol. 103, no. 9, pp. 1345–1352, 1999.
- [23] T. A. Moseley, D. R. Haudenschild, L. Rose, and A. H. Reddi, "Interleukin-17 family and IL-17 receptors," *Cytokine & Growth Factor Reviews*, vol. 14, no. 2, pp. 155–174, 2003.
- [24] W. Ouyang, J. K. Kolls, and Y. Zheng, "The biological functions of T helper 17 cell effector cytokines in inflammation," *Immunity*, vol. 28, no. 4, pp. 454–467, 2008.
- [25] N. J. Wilson, K. Boniface, J. R. Chan et al., "Development, cytokine profile and function of human interleukin 17–producing helper T cells," *Nature Immunology*, vol. 8, no. 9, pp. 950– 957, 2007.
- [26] F. Fossiez, O. Djossou, P. Chomarat et al., "T cell interleukin-17 induces stromal cells to produce proinflammatory and hematopoietic cytokines," *The Journal of Experimental Medicine*, vol. 183, no. 6, pp. 2593–2603, 1996.
- [27] R. B. Johnson, N. Wood, and F. G. Serio, "Interleukin-11 and IL-17 and the pathogenesis of periodontal disease," *Journal of Periodontology*, vol. 75, no. 1, pp. 37–43, 2004.
- [28] T. Honda, Y. Aoki, N. Takahashi et al., "Elevated expression of IL-17 and IL-12 genes in chronic inflammatory periodontal disease," *Clinica Chimica Acta*, vol. 395, no. 1-2, pp. 137–141, 2008.

- [29] N. Dutzan, J. Gamonal, A. Silva, M. Sanz, and R. Vernal, "Over-expression of forkhead box P3 and its association with receptor activator of nuclear factor-κ B ligand, interleukin (IL) -17, IL-10 and transforming growth factor-β during the progression of chronic periodontitis," *Journal of Clinical Periodontology*, vol. 36, no. 5, pp. 396–403, 2009.
- [30] L. Zhao, Y. Zhou, Y. Xu, Y. Sun, L. Li, and W. Chen, "Effect of non-surgical periodontal therapy on the levels of Th17/Th1/Th2 cytokines and their transcription factors in Chinese chronic periodontitis patients," *Journal of Clinical Periodontology*, vol. 38, no. 6, pp. 509–516, 2011.
- [31] D. M. Cardozo, G. A. Guelsin, S. L. Clementino et al., "DNA extraction from coagulated human blood for application in genotyping techniques for human leukocyte antigen and immunoglobulin-like receptors," *Revista da Sociedade Brasileira* de Medicina Tropical, vol. 42, no. 6, pp. 651–656, 2009.
- [32] X. Wu, Z. Zeng, B. Chen et al., "Association between polymorphisms in interleukin–17A and interleukin–17F genes and risks of gastric cancer," *International Journal of Cancer*, vol. 127, no. 1, pp. 86–92, 2010.
- [33] I. Bakri, C. W. I. Douglas, and A. Rawlinson, "The effects of stress on periodontal treatment: a longitudinal investigation using clinical and biological markers," *Journal of Clinical Periodontology*, vol. 40, no. 10, pp. 955–961, 2013.
- [34] S. Ayilavarapu, A. Kantarci, H. Hasturk, and T. E. van Dyke, "IPLA2 mRNA expression by human neutrophils in type 2 diabetes and chronic periodontitis," *Journal of the International Academy of Periodontology*, vol. 16, no. 4, pp. 121–126, 2014.
- [35] F. F. Lopes, F. H. Loureiro, A. d. Pereira, A. L. Pereira, and C. M. Alves, "Associação entre osteoporose e doença periodontal em mulheres na pós-menopausa," *Revista Brasileira de Ginecologia e Obstetrícia*, vol. 30, no. 8, pp. 379–383, 2008.
- [36] A. Pejčić, D. Kojović, I. Grigorov, and B. Stamenković, "Periodontitis and osteoporosis," *Facta Universitatis, Series: Medicine and Biology*, vol. 12, no. 2, pp. 100–103, 2005.
- [37] A. Stabholz, W. A. Soskolne, and L. Shapira, "Genetic and environmental risk factors for chronic periodontitis and aggressive periodontitis," *Periodontology 2000*, vol. 53, no. 1, pp. 138–153, 2010.
- [38] K. S. Kornman, "Diagnostic and prognostic tests for oral diseases: practical applications," *Journal of Dental Education*, vol. 69, no. 5, pp. 498–508, 2005.
- [39] J. Bergström, "Tobacco smoking and chronic destructive periodontal disease," *Odontology*, vol. 92, no. 1, pp. 1–8, 2004.
- [40] H. Preber, T. Kant, and J. Bergstrom, "Cigarette smoking, oral hygiene and periodontal health in Swedish army conscripts," *Journal of Clinical Periodontology*, vol. 7, no. 2, pp. 106–113, 1980.
- [41] J. Haber, J. Wattles, M. Crowley, R. Mandell, K. Joshipura, and R. L. Kent, "Evidence for cigarette smoking as a major risk factor for periodontitis," *Journal of Periodontology*, vol. 64, no. 1, pp. 16–23, 1993.
- [42] S. Reichert, J. Stein, A. Gautsch, H.-G. Schaller, and H. K. G. Machulla, "Gender differences in HLA phenotype frequencies found in German patients with generalized aggressive periodontitis and chronic periodontitis," *Oral Microbiology and Immunology*, vol. 17, no. 6, pp. 360–368, 2002.
- [43] L. Machion, P. M. de Freitas, J. B. C. Neto, G. R. N. Filho, and F. H. Nociti Jr., "The influence of gender and age on the prevalence of periodontal pockets," *Pesquisa Odontológica Brasileira*, vol. 14, no. 1, pp. 33–37, 2000.
- [44] M. Kadkhodazadeh, A. R. Ebadian, R. Amid, N. Youssefi, and A. R. Mehdizadeh, "Interleukin 17 receptor gene polymorphism in

- periimplantitis and chronic periodontitis," *Acta Medica Iranica*, vol. 51, no. 6, pp. 353–358, 2013.
- [45] J. L. Espinoza, A. Takami, K. Nakata et al., "A genetic variant in the IL-17 promoter is functionally associated with acute graft-versus-host disease after unrelated bone marrow transplantation," *PLoS ONE*, vol. 6, no. 10, Article ID e26229, 2011.
- [46] X. K. Liu, X. Lin, and S. L. Gaffen, "Crucial role for nuclear factor of activated T cells in T cell receptor-mediated regulation of human interleukin-17," *The Journal of Biological Chemistry*, vol. 279, no. 50, pp. 52762–52771, 2004.
- [47] T. Shibata, T. Tahara, I. Hirata, and T. Arisawa, "Genetic polymorphism of interleukin-17A and -17F genes in gastric carcinogenesis," *Human Immunology*, vol. 70, no. 7, pp. 547–551, 2009.
- [48] H. Yu, S. Sun, F. Liu, and Q.-H. Xu, "Meta-analysis of associations between interleukin-17 gene polymorphisms and risk of gastric cancer," *Asian Pacific Journal of Cancer Prevention*, vol. 15, no. 20, pp. 8709–8713, 2014.
- [49] L. Wang, Y. Jiang, Y. Zhang et al., "Association analysis of IL-17A and IL-17F polymorphisms in Chinese han women with breast cancer," *PLoS ONE*, vol. 7, no. 3, Article ID e34400, 2012.
- [50] Y. Quan, B. Zhou, Y. Wang et al., "Association between IL17 polymorphisms and risk of cervical cancer in Chinese women," *Clinical and Developmental Immunology*, vol. 2012, Article ID 258293, 6 pages, 2012.
- [51] T. Arisawa, T. Tahara, T. Shibata et al., "The influence of polymorphisms of interleukin-17A and interleukin-17F genes on the susceptibility to ulcerative colitis," *Journal of Clinical Immunology*, vol. 28, no. 1, pp. 44–49, 2008.
- [52] R. Hayashi, T. Tahara, H. Shiroeda et al., "Influence of IL17A polymorphisms (rs2275913 and rs3748067) on the susceptibility to ulcerative colitis," *Clinical and Experimental Medicine*, vol. 13, no. 4, pp. 239–244, 2013.
- [53] G. B. N. Nordang, M. K. Viken, J. E. Hollis-Moffatt et al., "Association analysis of the interleukin 17A gene in Caucasian rheumatoid arthritis patients from Norway and New Zealand," *Rheumatology*, vol. 48, no. 4, pp. 367–370, 2009.
- [54] J. D. Corrêa, M. F. M. Madeira, R. G. Resende et al., "Association between polymorphisms in interleukin-17A and -17F genes and chronic periodontal disease," *Mediators of Inflammation*, vol. 2012, Article ID 846052, 9 pages, 2012.
- [55] A. M. Saraiva, M. R. M. Alves e Silva, J. D. F. Correia Silva et al., "Evaluation of IL17A expression and of IL17A, IL17F and IL23R gene polymorphisms in Brazilian individuals with periodontitis," *Human Immunology*, vol. 74, no. 2, pp. 207–214, 2013.
- [56] C. D. Ramsey, R. Lazarus, C. A. Camargo Jr., S. T. Weiss, and J. C. Celedón, "Polymorphisms in the interleukin 17F gene (IL17F) and asthma," Genes and Immunity, vol. 6, no. 3, pp. 236–241, 2005.
- [57] J. Seiderer, I. Elben, J. Diegelmann et al., "Role of the novel Th17 cytokine IL–17F in inflammatory bowel disease (IBD): upregulated colonic IL–17F expression in active Crohn's disease and analysis of the IL17F p.His161Arg polymorphism in IBD," *Inflammatory Bowel Diseases*, vol. 14, no. 4, pp. 437–445, 2008.
- [58] S. Najafi, H. Hadinedoushan, G. Eslami, and A. Aflatoonian, "Association of IL-17A and IL-17 F gene polymorphisms with recurrent pregnancy loss in Iranian women," *Journal of Assisted Reproduction and Genetics*, vol. 31, no. 11, pp. 1491–1496, 2014.
- [59] S. Wang, H. Zhai, Y. Su, and Y. Wang, "IL-17F but not IL-17A gene polymorphism confers risk to multiple sclerosis in a Chinese Han population," *Journal of the Neurological Sciences*, vol. 342, no. 1, pp. 133–136, 2014.

[60] R. Peng, J. Yue, M. Han, Y. Zhao, L. Liu, and L. Liang, "The IL-17F sequence variant is associated with susceptibility to tuberculosis," *Gene*, vol. 515, no. 1, pp. 229–232, 2013.

- [61] Y. Peng, B. Zhou, Y. Y. Wang et al., "Analysis of IL-17 gene poly-morphisms in Chinese patients with dilated cardiomyopathy," Human Immunology, vol. 74, no. 5, pp. 635–639, 2013.
- [62] M. Kawaguchi, D. Takahashi, N. Hizawa et al., "IL-17F sequence variant (His161Arg) is associated with protection against asthma and antagonizes wild-type IL-17F activity," *Journal of Allergy* and Clinical Immunology, vol. 117, no. 4, pp. 795–801, 2006.
- [63] Y. Yan, H. Weng, Z. Shen, L. Wu, and X. Zeng, "Association between interleukin-4 gene –590 C/T, –33 C/T, and 70-base-pair polymorphisms and periodontitis susceptibility: a meta-analysis," *Journal of Periodontology*, vol. 85, no. 11, pp. e354–e362, 2014.
- [64] Z.-G. Li, J.-J. Li, C.-A. Sun, Y. Jin, and W.-W. Wu, "Interleukin-18 promoter polymorphisms and plasma levels are associated with increased risk of periodontitis: a meta-analysis," *Inflammation Research*, vol. 63, no. 1, pp. 45–52, 2014.
- [65] J. Huang, C. Ding, X. Chen, R. He, and N. Chen, "Association of TGF- $\beta$ 1- 509C/T,+ 869T/C, and+ 915G/C polymorphisms with periodontitis susceptibility," *Oral Diseases*, vol. 21, no. 4, pp. 443-450, 2015.
- [66] X. Chen, J. Huang, L. Zhong, and C. Ding, "Quantitative assessment of the associations between interleukin-8 polymorphisms and periodontitis susceptibility," *Journal of Periodontology*, vol. 86, no. 2, pp. 292–300, 2015.
- [67] C. Ding, X. Ji, X. Chen, Y. Xu, and L. Zhong, "TNF-α gene promoter polymorphisms contribute to periodontitis susceptibility: evidence from 46 studies," *Journal of Clinical Periodontology*, vol. 41, no. 8, pp. 748–759, 2014.
- [68] J.-S. Deng, P. Qin, X.-X. Li, and Y.-H. Du, "Association between interleukin-1β C (3953/4)T polymorphism and chronic periodontitis: evidence from a meta-analysis," *Human Immunology*, vol. 74, no. 3, pp. 371–378, 2013.
- [69] C. M. Albuquerque, A. J. Cortinhas, F. J. Morinha, J. C. Leitão, C. A. Viegas, and E. M. Bastos, "Association of the IL-10 polymorphisms and periodontitis: a meta-analysis," *Molecular Biology Reports*, vol. 39, no. 10, pp. 9319–9329, 2012.
- [70] P. Behfarnia, R. Birang, A. R. Andalib, and S. Asadi, "Comparative evaluation of IFNγ, IL4 and IL17 cytokines in healthy gingiva and moderate to advanced chronic periodontitis," *Dental Research Journal*, vol. 7, no. 2, pp. 45–50, 2010.
- [71] R. Mahononda, P. Jitprasertwong, N. Sa-Ard-Iam et al., "Effects of IL-17 on human gingival fibroblasts," *Journal of Dental Research*, vol. 87, no. 3, pp. 267–272, 2008.
- [72] M. S. Maddur, P. Miossec, S. V. Kaveri, and J. Bayry, "Th17 cells: biology, pathogenesis of autoimmune and inflammatory diseases, and therapeutic strategies," *The American Journal of Pathology*, vol. 181, no. 1, pp. 8–18, 2012.
- [73] K. Sato, A. Suematsu, K. Okamoto et al., "Th17 functions as an osteoclastogenic helper T cell subset that links T cell activation and bone destruction," *The Journal of Experimental Medicine*, vol. 203, no. 12, pp. 2673–2682, 2006.
- [74] S. Takahashi, M. Fukuda, A. Mitani et al., "Follicular dendritic cell-secreted protein is decreased in experimental periodontitis concurrently with the increase of interleukin-17 expression and the Rankl/Opg mRNA ratio," *Journal of Periodontal Research*, vol. 49, no. 3, pp. 390–397, 2014.
- [75] S. Xu and X. Cao, "Interleukin-17 and its expanding biological functions," *Cellular & Molecular Immunology*, vol. 7, no. 3, pp. 164–174, 2010.