

SIX1 overexpression in diffuse-type and grade III gastric tumors: Features that are associated with poor prognosis

Modjtaba Emadi-Baygi, Parvaneh Nikpour^{1,2,3}, Elaheh Emadi-Andani¹

Department of Genetics, Research Institute of Biotechnology, Shahrekord University, Shahrekord, ¹Department of Genetics and Molecular Biology, ²Pediatric Inherited Diseases Research Center, ³Child Growth and Development Research Center, Isfahan University of Medical Sciences, Isfahan, Iran

Abstract

Background: Gastric cancer is the second most common cancer worldwide. In Iran, the incidence of gastric cancer is well above the world average, and is the first common cancer in Iranian men and the third one in women. Located at chromosome 14q23, *SIX1* is a homolog of the *Drosophila* 'sine oculis' (*so*) gene and is highly conserved in numerous species. In addition to the role of *SIX1* in the development, its expression is frequently dysregulated in multiple cancers. This study aimed to evaluate the clinicopathological features of the expression of *SIX1* gene in gastric adenocarcinoma.

Materials and Methods: Thirty pairs of gastric tissue samples from patients with gastric adenocarcinoma were evaluated for *SIX1* gene expression using quantitative real-time polymerase chain reaction. A paired *t*-test or one-way ANOVA with *post hoc* multiple comparisons were used to analyze the differences between groups. Statistical significance was defined as $P \leq 0.05$.

Results: *SIX1* expression was decreased in tumoral samples. However, its expression increased significantly in diffuse-type gastric cancer. Furthermore, there was a trend toward statistical significance in increasing *SIX1* gene expression with higher grades. Of note, the difference was significant between grades I and III.

Conclusions: The results suggest that *SIX1* gene expression might be used in the future as a potential biomarker to predict the outcome of the disease as diffuse-type and grade III of gastric tumors are associated with poor prognosis.

Key Words: Diffuse-type gastric cancer, gene expression, poor prognosis *SIX1*, tumor grades

Address for correspondence:

Dr. Parvaneh Nikpour, Department of Genetics and Molecular Biology, Faculty of Medicine, Isfahan University of Medical Sciences, Isfahan, Iran.

E-mail: pnikpour@med.mui.ac.ir

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INTRODUCTION

Gastric cancer is the second most common cancer

worldwide, with an estimated 900,000 new cases and 700,000 gastric cancer-related deaths in the world.^[1] In Iran, the incidence rate of gastric cancer is well above the world average, and is the first common cancer in Iranian men and the third one in women.^[2] Because of the lack of trustworthy early diagnostic methods and effective treatment, more than 80% of patients with advanced gastric cancer die of the disease or recurrent disease within 1 year after diagnosis. The majority of patients with gastric cancer are being diagnosed in advanced stages of the disease such that usual treatment protocols are ineffective in a remarkable number of cases.^[3]

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Therefore, elucidation of the molecular characteristics of gastric tumors is an essential need to develop methods of early cancer detection and reduce its mortality.

Located at chromosome 14q23,^[4] *SIX1* (sineoculis homeobox homolog 1), a member of the Six gene superfamily, is a homolog of the *Drosophila* 'sine oculis' (so) gene and is highly conserved in numerous species from *Drosophila* to human.^[5,6] The *SIX1* gene product functions in concert with *Eya1*,^[6] *Pax* and *Dac* in DNA binding^[7] and regulates the expression of many downstream target genes.^[8] Therefore, the *SIX1* homeoprotein organizes a variety of cellular processes during normal development of some tissues and organs such as promoting progenitor cell population proliferation and their invasion before cell differentiation and specification, survival, migration and apoptosis. However, after development, expression of *SIX1* changes and decreases in most normal adult tissues.^[9-11] Furthermore, *SIX1* indirectly affects cell movement and adhesion between cells and the extracellular matrix (ECM) by regulating the expression of *Ezrin*.^[7]

In addition to the role of *SIX1* in the development, its expression is frequently dysregulated in multiple cancers including breast cancer,^[12] Wilms' tumors,^[13] ovarian cancer,^[14] hepatocellular carcinoma,^[15] alveolar rhabdomyosarcomas,^[16] and cervical cancer.^[17] The misexpression of *SIX1* in cancer can enhance cancer cell proliferation and survival and lead to tumor inception and progression.^[8,18,19] These findings explain that unsuitable *SIX1* expression in adult differentiated tissues results in cell proliferation stimulus and, in turn, leads to initiation and progression of numerous cancers.^[11]

Considering the dysregulation of *SIX1* gene expression in various tumors, in this study, we studied the expression of *SIX1* gene in gastric tumors.

MATERIALS AND METHODS

Tumor and non-tumor tissues

Thirty pairs of gastric tissue samples (tumor and their adjacent non-tumor tissues) from patients with gastric adenocarcinoma were provided from the Iran Tumoural Bank (Tehran, Iran) as described previously.^[20] The clinicopathological characteristics of the specimens are shown in Table 1. Written informed consent from all subjects was obtained by the Iran Tumoural Bank. The experimental procedures were approved by the Ethics Committee of the Isfahan University of Medical Sciences. The samples were frozen in liquid nitrogen and kept at -80°C until analysis.

RNA isolation and reverse transcription

Total RNAs from frozen gastric cancer tissue samples were extracted using Qiazol reagent and RNeasy columns (Qiagen, Hilden, Germany) following the manufacturer's protocol. RNA integrity was examined by running on a 1% agarose gel and total RNA concentrations determined spectrophotometrically. Two micrograms of total RNA were reverse transcribed using random hexamer primers (TAG Copenhagen) and MMLV Reverse Transcriptase (Fermentas, Vilnius, Lithuania) according to the manufacturer's instructions.

Quantitative real-time reverse transcriptase-polymerase chain reaction (RT-PCR)

Synthesized cDNAs were subjected to quantitative real-time PCR using the Maxima SYBR Green/ROX qPCR Master Mix (Fermentas, Vilnius, Lithuania) and specific primers for *SIX1* and *TBP*^[21] as an endogenous control in a total volume of 20 µL reaction mixture and were run on the Rotor-gene 6000 system. The following primers were used to amplify *SIX1*:

SIX1 forward primer:

5'- TAAGAACCGGAGGCAAAGAG -3'

SIX1 reverse primer:

5'- AGTTTGAGCTCCTGGCGTG -3'

The amplification conditions for *SIX1* were as follows: An initial denaturation step at 95°C for 10 min followed by 45 amplification cycles consisting of denaturation at 95°C for 20 s, annealing for 20 s at 55°C and an extension at 72°C for 20 s. For each sample, measurements were performed at least in triplicate. The standard curve method was used to calculate relative gene expression. For further verification of the identity of the PCR products, agarose gel electrophoresis was performed.

Statistical analysis

Data are represented as means ± standard error of mean (SEM) from at least three separate experiments.

Table 1: Clinicopathological parameters of gastric cancer samples

Characteristics	Numbers (%)
Sex	
Male	18 (60)
Female	12 (40)
Age (years)	
≥70	15 (50)
<70	15 (50)
Tumor grades	
Grade I	10 (33.3)
Grade II	8 (26.6)
Grade III	12 (40)
Tumor types	
Diffuse	15 (50)
Intestinal	15 (50)

To compare the gene expression levels between the tumor and non-tumor tissues and associated clinicopathological characteristics with gene expression, Student's *t* test and ANOVA statistical tests were performed. The SPSS program, version 20.0, was utilized for statistical analyses, and differences were considered significant if $P < 0.05$.

RESULTS

SIX1 is underexpressed in human gastric adenocarcinoma tissues

Relative quantitation of the expression levels of *SIX1* in gastric adenocarcinomas showed that the relative levels of *SIX1* transcripts were significantly decreased (around 1.5-fold, P value = 0.018) in cancerous tissues compared with adjacent non-cancerous tissues: 0.48 ± 0.03 versus 0.64 ± 0.06 , respectively, as shown in Figure 1.

Association of *SIX1* expression with clinicopathological parameters in gastric adenocarcinoma tissues

Next, we analyzed the association of *SIX1* relative gene expression with the reported clinicopathological characteristics of the tumors (histological classifications and grade). As shown in Figure 2, the expression level of *SIX1* was different in both diffuse- and intestinal-type tumors. We found that *SIX1* overexpressed in diffuse-type gastric tumors (mean: 0.56) compared with intestinal-type tumors (mean: 0.40) (P value: 0.025). Furthermore, there was no significant association between the expression levels of *SIX1* and different grades of the tumors (P value: 0.09). However, *SIX1* was overexpressed in grade III of gastric tumors compared with grade I (P value: 0.03) [Figure 3].

DISCUSSION

To the best of our knowledge, this is the first study that evaluates the expression of *SIX1* gene in gastric adenocarcinoma using quantitative real-time RT-PCR. Our study showed that the relative expression of *SIX1* is significantly downregulated in tumoral tissues compared with the adjacent non-tumoral tissues ($P = 0.018$). However, our results showed that expression of *SIX1* increased significantly in diffuse-type gastric tumors in comparison with the intestinal-type gastric tumors ($P = 0.025$). Furthermore, *SIX1* expression significantly increased in grade III gastric tumors in comparison with grade I gastric tumors ($P = 0.03$).

SIX1 is an important developmental regulator in several diverse tissues/organs,^[10,22-25] and induces the expression of diverse genes (e.g. *Cyclin D1*, *Cyclin A1* and *c-myc*) in various cell types.^[8] It has

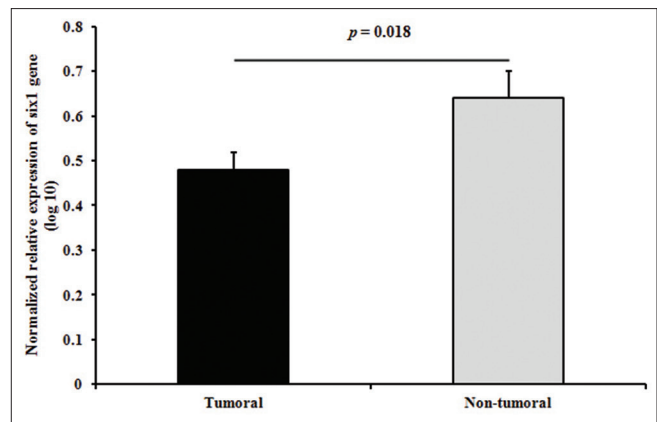


Figure 1: The relative expression levels of *SIX1* in tumoral versus non-tumoral gastric samples. Error bars represent standard error of mean (SEM)

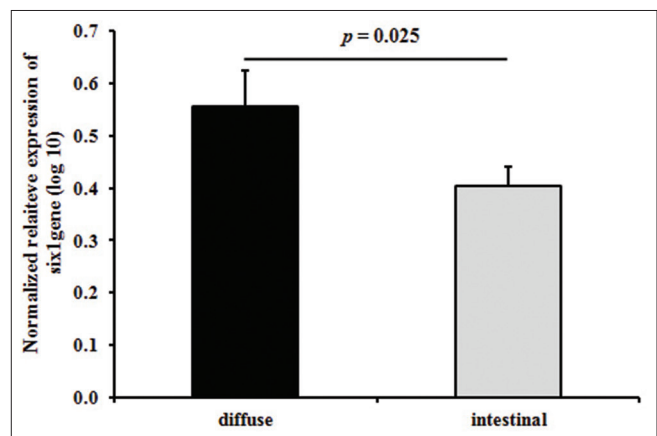


Figure 2: Relationship of the relative expression levels of *SIX1* with the histological classifications of the gastric tumors (diffuse vs. intestinal types)

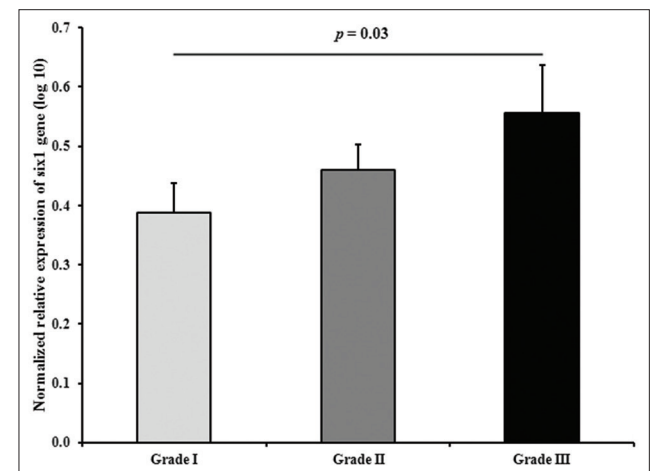


Figure 3: The *SIX1* relative expression stratified according to different tumor grades. The difference between grades I and III was statistically significant ($P = 0.03$)

been postulated that dysregulation of *SIX1* leads to cancer.^[7,26]

Overexpression of *SIX1* has been documented in several types of cancer, including breast cancer,^[12] Wilms' tumors,^[13] ovarian cancer,^[14] hepatocellular carcinoma,^[15] alveolar rhabdomyosarcomas^[16] and cervical cancer,^[17] where it facilitates proliferation and metastasis of the cancerous cells.^[26] In the same vein, we observed that *SIX1* expression increased significantly in diffuse type and grade III gastric cancer. Of note, microarray analyses have also shown that *SIX1* overexpresses in diffuse-type gastric tumors versus intestinal-type gastric tumors.^[27,28] However, Matsusaka *et al.* recently reported that EYA1, a Six 1 coactivator, is often methylated in both EBV+ and EBV/high methylation gastric cancers.^[29] As EYA1 interacts with and functions upstream of the homeobox gene *Six 1* in the development of some organs including ear and kidney,^[24,30] it is plausible that *SIX1* was co-underexpressed with *EYA1* in gastric cancer. Furthermore, microarray analysis performed by Cui *et al.* showed that *SIX1* expression decreases in gastric tumors versus normal gastric tissues.^[31]

Located within a critical interval on chromosome 14q23, Ruf *et al.* identified a 3-bp deletion in the *SIX1* gene in branchio-otic syndrome.^[32] Furthermore, a loss of 14q23 has been reported in breast cancer,^[33] gastrointestinal stromal tumors^[34] and neuroblastomas.^[35] In the same vein, deletions have been observed in 14q in gastric cancer.^[36] Moreover, Gümüs-Akay *et al.* recently reported that the most common losses in gastric adenocarcinomas were found on arms 18q (26%), 5q (21%) and 14q (21%).^[37] Taken together, the overall underexpression of *SIX1* in gastric cancer may be attributed to the loss of 14q.

In conclusion, this is the first report that evaluates the expression of *SIX1* in gastric cancer. Our results showed that *SIX1* is significantly downregulated in gastric tumors. However, our results showed that expression of *SIX1* increased significantly in diffuse-type and grade III gastric tumors. Taken together, this gene might be used in the future as a potential biomarker to predict the outcome of the disease as diffuse-type and grade III gastric tumors are associated with poor prognosis.^[38] Further studies should be carried out to elucidate the mechanisms which cause *SIX1* underexpression in gastric tumors and to find out how *SIX1* and *EYA1* function in gastric cancer.

REFERENCES

- Gomceli I, Demiriz B, Tez M. Gastric carcinogenesis. *World J Gastroenterol* 2012;18:5164-70.
- Kolahdoozan S, Sadjadi A, Radmard AR, Khademi H. Five common cancers in Iran. *Arch Iran Med* 2010;13:143-6.
- Malekzadeh R, Derakhshan MH, Malekzadeh Z. Gastric cancer in Iran: Epidemiology and risk factors. *Arch Iran Med* 2009;12:576-83.
- Boucher CA, Carey N, Edwards YH, Siciliano MJ, Johnson KJ. Cloning of the human *SIX1* gene and its assignment to chromosome 14. *Genomics* 1996;33:140-2.
- Seo HC, Curtiss J, Mlodzik M, Fjose A. Six class homeobox genes in drosophila belong to three distinct families and are involved in head development. *Mech Dev* 1999;83:127-39.
- Buller C, Xu X, Marquis V, Schwanke R, Xu PX. Molecular effects of *Eya1* domain mutations causing organ defects in BOR syndrome. *Hum Mol Genet* 2001;10:2775-81.
- Yu Y, Davicioni E, Triche TJ, Merlino G. The homeoprotein *six 1* transcriptionally activates multiple protumorigenic genes but requires *e2f1* to promote metastasis. *Cancer Res* 2006;66:1982-9.
- Coletta RD, Christensen KL, Micalizzi DS, Jedlicka P, Varella-Garcia M, Ford HL. *Six 1* overexpression in mammary cells induces genomic instability and is sufficient for malignant transformation. *Cancer Res* 2008;68:2204-13.
- McCoy EL, Iwanaga R, Jedlicka P, Abbey NS, Chodosh LA, Heichman KA, *et al.* *Six 1* expands the mouse mammary epithelial stem/progenitor cell pool and induces mammary tumors that undergo epithelial-mesenchymal transition. *J Clin Invest* 2009;119:2663-77.
- Christensen KL, Patrick AN, McCoy EL, Ford HL. The *six* family of homeobox genes in development and cancer. *Adv Cancer Res* 2008;101:93-126.
- Kumar JP. The *sine oculis* homeobox (*SIX*) family of transcription factors as regulators of development and disease. *Cell Mol Life Sci* 2009;66:565-83.
- Ford HL, Kabingu EN, Bump EA, Mutter GL, Pardee AB. Abrogation of the G2 cell cycle checkpoint associated with overexpression of *HSIX 1*: A possible mechanism of breast carcinogenesis. *Proc Natl Acad Sci U S A* 1998;95:12608-13.
- Li CM, Guo M, Borczuk A, Powell CA, Wei M, Thaker HM, *et al.* Gene expression in Wilms' tumor mimics the earliest committed stage in the metanephric mesenchymal-epithelial transition. *Am J Pathol* 2002;160:2181-90.
- Behbakht K, Qamar L, Aldridge CS, Coletta RD, Davidson SA, Thorburn A, *et al.* *Six 1* overexpression in ovarian carcinoma causes resistance to TRAIL-mediated apoptosis and is associated with poor survival. *Cancer Res* 2007;67:3036-42.
- Ng KT, Man K, Sun CK, Lee TK, Poon RT, Lo CM, *et al.* Clinicopathological significance of homeoprotein *Six 1* in hepatocellular carcinoma. *Br J Cancer* 2006;95:1050-5.
- Yu Y, Khan J, Khanna C, Helman L, Meltzer PS, Merlino G. Expression profiling identifies the cytoskeletal organizer *e2f1* and the developmental homeoprotein *Six-1* as key metastatic regulators. *Nat Med* 2004;10:175-81.
- Wan F, Miao X, Quraishi I, Kennedy V, Creek KE, Pirisi L. Gene expression changes during HPV-mediated carcinogenesis: A comparison between an *in vitro* cell model and cervical cancer. *Int J Cancer* 2008;123:32-40.
- Coletta RD, Christensen K, Reichenberger KJ, Lamb J, Micomonaco D, Huang L, *et al.* The *Six 1* homeoprotein stimulates tumorigenesis by reactivation of cyclin A1. *Proc Natl Acad Sci U S A*. 2004;101:6478-83.
- Li Z, Tian T, Lv F, Chang Y, Wang X, Zhang L, *et al.* *Six 1* promotes proliferation of pancreatic cancer cells via upregulation of cyclin D1 expression. *PLoS One* 2013;8:e59203.
- Nikpour P, Emadi-Baygi M, Mohammad-Hashem F, Maracy MR, Haghjooy-Javanmard S. Differential expression of *ZFX* gene in gastric cancer. *J Biosci* 2012;37:85-90.
- Nikpour P, Baygi ME, Steinhoff C, Hader C, Luca AC, Mowla SJ, *et al.* The RNA binding protein *Musashi1* regulates apoptosis, gene expression and stress granule formation in urothelial carcinoma cells. *J Cell Mol Med* 2011;15:1210-24.
- Li Z, Deng D, Huang H, Tian L, Chen Z, Zou Y, *et al.* Overexpression of *Six 1* leads to retardation of myogenic differentiation in C2C12 myoblasts. *Mol Biol Rep* 2013;40:217-23.
- Zheng W, Huang L, Wei ZB, Silvius D, Tang B, Xu Px. The role of *Six 1* in mammalian auditory system development. *Development* 2003;130:3989-4000.
- Xu PX, Zheng W, Huang L, Maire P, Laclef C, Silvius D. *Six 1* is required

- for the early organogenesis of mammalian kidney. *Development* 2003;130:3085-94.
25. Zou D, Silviu D, Fritsch B, Xu Px. *Eya1* and *Six 1* are essential for early steps of sensory neurogenesis in mammalian cranial placodes. *Development* 2004;131:5561-72.
 26. Abate-Shen C. Deregulated homeobox gene expression in cancer: Cause or consequence? *Nat Rev Cancer* 2002;2:777-85.
 27. Ooi CH, Ivanova T, Wu J, Lee M, Tan IB, Tao J, *et al.* Oncogenic pathway combinations predict clinical prognosis in gastric cancer. *PLoS Genet* 2009;5:e1000676.
 28. Förster S, Gretschel S, Jons T, Yashiro M, Kemmner W. THBS4, a novel stromal molecule of diffuse-type gastric adenocarcinomas, identified by transcriptome-wide expression profiling. *Mod Pathol* 2011;24:1390-403.
 29. Matsusaka K, Kaneda A, Nagae G, Ushiku T, Kikuchi Y, Hino R, *et al.* Classification of Epstein-Barr virus-positive gastric cancers by definition of DNA methylation epigenotypes. *Cancer Res* 2011;71:7187-97.
 30. Ahmed M, Wong EY, Sun J, Xu J, Wang F, Xu PX. *Eya1-Six 1* interaction is sufficient to induce hair cell fate in the cochlea by activating *Atoh1* expression in cooperation with *Sox 2*. *Dev Cell* 2012;22:377-90.
 31. Cui J, Chen Y, Chou WC, Sun L, Chen L, Suo J, *et al.* An integrated transcriptomic and computational analysis for biomarker identification in gastric cancer. *Nucleic Acids Res* 2011;39:1197-207.
 32. Ruf RG, Xu PX, Silviu D, Otto EA, Beekmann F, Muerb UT, *et al.* *SIX1* mutations cause branchio-oto-renal syndrome by disruption of *EYA1-SIX1*-DNA complexes. *Proc Natl Acad Sci U S A* 2004;101:8090-5.
 33. Tanner MM, Karhu RA, Nupponen NN, Borg A, Baldetorp B, Pejovic T, *et al.* Genetic aberrations in hypodiploid breast cancer: Frequent loss of chromosome 4 and amplification of cyclin D1 oncogene. *Am J Pathol* 1998;153:191-9.
 34. El-Rifai W, Sarlomo-Rikala M, Andersson LC, Miettinen M, Knuutila S. High-resolution deletion mapping of chromosome 14 in stromal tumors of the gastrointestinal tract suggests two distinct tumor suppressor loci. *Genes Chromosomes Cancer* 2000;27:387-91.
 35. Thompson PM, Seifried BA, Kyemba SK, Jensen SJ, Guo C, Maris JM, *et al.* Loss of heterozygosity for chromosome 14q in neuroblastoma. *Med Pediatr Oncol* 2001;36:28-31.
 36. Koo SH, Kwon KC, Shin SY, Jeon YM, Park JW, Kim SH, *et al.* Genetic alterations of gastric cancer: Comparative genomic hybridization and fluorescence *In situ* hybridization studies. *Cancer Genet Cytogenet* 2000;117:97-103.
 37. Gümüs -Akay G, Unal AE, Elhan AH, Bayar S, Karadayt K, Sunguroglu A, *et al.* DNA copy number changes in gastric adenocarcinomas: High resolution-comparative genomic hybridization study in Turkey. *Arch Med Res* 2009;40:551-60.
 38. Dicken BJ, Bigam DL, Cass C, Mackey JR, Joy AA, Hamilton SM. Gastric adenocarcinoma: Review and considerations for future directions. *Ann Surg* 2005;241:27-39.

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