

Comparative evaluation of the antibacterial and physical properties of conventional glass ionomer cement containing chlorhexidine and antibiotics

Sudhir Mittal, Heena Soni, Devender Kumar Sharma¹, Kavita Mittal²,
Vasundhara Pathania, Samridhi Sharma

Departments of Pedodontics and ¹Microbiology, Himachal Dental College, Sunder Nagar, Himachal Pradesh, ²Department of Pedodontics Guru Nanak Dev Dental College and Research Institute, Sunam, Punjab, India

Corresponding author (email: <care@jalandhardentalcare.com>)

Dr. Sudhir Mittal, Department of Pedodontics, Himachal Dental College, Sunder Nagar - 175 002, Himachal Pradesh, India.

Abstract

Objective: To evaluate the antimicrobial efficacy and compressive strength of conventional glass ionomer cement (GIC) containing chlorhexidine and antibiotics at varying concentrations. **Materials and Methods:** Chlorhexidine diacetate and antibiotics (ciprofloxacin, metronidazole, and minocycline) were incorporated into GIC Fuji IX at 1.5% and 3% w/w ratio to form the experimental groups. The experimental GIC specimens were placed on brain heart infusion agar plates inoculated with *Streptococcus mutans*, and the area of inhibition was measured after 48 h. The 24-h compressive strength of the set specimens was evaluated using a Universal Testing Machine. **Results:** The control group demonstrated no zone of inhibition. All experimental groups showed inhibition against *S. mutans* ($P < 0.05$), with larger zones of inhibition found in the higher concentration groups. Compressive strength at the end of 24 h decreased in the experimental groups as compared to the control group ($P < 0.05$), but no difference was found between the experimental groups ($P > 0.05$). **Conclusion:** The present study demonstrated that experimental GICs containing chlorhexidine diacetate and antibiotics were effective in inhibiting *S. mutans*, and incorporation of 1.5% ABX was optimal to give the appropriate antibacterial and physical properties.

Key words: Antibiotics, chlorhexidine diacetate, glass ionomer cement

INTRODUCTION

Dental caries is a disease that dates back to antiquity and still remains a perennial public health problem.^[1] Scientific research continues to make progress in identifying the best practices for treating and preventing dental caries.^[2] Minimal intervention dentistry is an emerging modern dental practice designed around the principle aim of preservation of as much as natural tooth structure as possible.

Atraumatic restorative treatment is one such paradigm of the Minimal Intervention Dentistry concept. An Alternative Restorative Treatment restoration involves the removal of soft, completely demineralized carious tooth tissue with hand instruments, followed by restoration of the cavity with an adhesive dental material such as glass ionomer cement (GIC) that simultaneously seals any remaining pits and fissures at risk.^[3] However, cavities treated by ART have some residual infected dentin, as

Access this article online	
Quick Response Code:	Website: www.jispcd.org
	DOI: 10.4103/2231-0762.161754

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 3.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms.

For reprints contact: reprints@medknow.com

How to cite this article: Mittal S, Soni H, Sharma DK, Mittal K, Pathania V, Sharma S. Comparative evaluation of the antibacterial and physical properties of conventional glass ionomer cement containing chlorhexidine and antibiotics. J Int Soc Prevent Communit Dent 2015;5:268-75.

manual instruments are not as effective as rotary burs in terms of eliminating bacteria.^[4] Consequently, cariogenic bacteria can survive incarceration under GIC restoration and remain viable for up to 2 years resulting in secondary caries.^[5] Literature has been evidence to the fact that fluoride released from GICs is not sufficiently potent to combat the effects of bacterial destruction over an extensive duration of time.^[6]

Different classes of GIC have been used for luting, restorative, core build up, lining purposes, as orthodontic cement, and for sealing pits and fissures. Conventional and metal-reinforced glass ionomers have been superseded by highly viscous GICs like Fuji IX GP, Chem-Flex, and Ketac-Molar. The quest to develop an efficient GIC has led to several advances in the material or modifications of the existing one.^[7] The fact that ions can readily travel in and out of the material offers the opportunity to dope the cement with other soluble antimicrobials.^[8] Chlorhexidine, a bisbiguanide, is currently the most potent chemotherapeutic agent used to enhance the antimicrobial properties of GICs.^[9] This cationic antibacterial agent binds to hydroxyapatite and is gradually released at therapeutic levels, a phenomenon named as substantivity.^[10-12] Recently, the addition of antibiotics to glass ionomer has been recommended, with an aim to alleviate the total number of viable bacteria.^[13]

From dental literature it appears that chlorhexidine (diacetate and digluconate) and antibiotics like doxycycline, metronidazole, ciprofloxacin, cefaclor, cetrime, and minocycline have frequently been incorporated into GIC and all studies have demonstrated encouraging results regarding the antimicrobial efficacy of the modified cement. However, the incorporation of these antibacterial agents has resulted in jeopardizing the basic biomechanical characteristics of the material.^[4,7,13] Since the requisites of an ideal restorative material remain unaccomplished without the ability of a material to withstand the traumas of occlusion, this highlights the crucial effect of antibacterial additive concentration on GICs' mechanical performance. Therefore, the present study aims to evaluate the antibacterial activity and physical properties of modified GIC at different concentrations and to determine the optimal concentration of antimicrobials to be incorporated into this emerging biomaterial.

MATERIALS AND METHODS

Preparation of molds

A commercially available plastic tubing (linear low-density polyethylene tubing 1/4" outside diameter

and 0.165" inside diameter) was obtained. The tube was then cut with the help of a microtome to prepare molds of 4 mm diameter and 10 mm height. Care was taken to have a clean cut surface perpendicular to the long axis of the mold. The molds were grouped as groups I, II, and III, where Group I served as the control group and groups II and III as the experimental groups containing chlorhexidine and antibiotics, respectively. The experimental groups were further sub-categorized into two concentrations: 1.5% and 3%. Seven molds each were assigned to the control group as well as the various concentration subgroups, forming a total of 70 molds.

Preparation of antibacterial cement

Preparation of antibacterial cement is presented in Table 1.

A conventional posterior restorative GIC (Fuji IX; GC Corporation, Tokyo, Japan) was used to fill the molds of the control group (Group I). Chlorhexidine diacetate (CHX), which is commercially available as a solid substance, was weighed using a 10⁻⁴ g precision balance (Mettler Toledo Electronic scale) and added to calculated amount of conventional glass ionomer powder, in order to obtain two concentrations of 1.5% and 3% CHX in the GIC formulation.

The antibiotics were obtained in the form of tablets and the surface sugar coating from the tablets was scrapped off. Using a pestle and mortar, the tablets were then ground into fine powder. The antibiotics were proportioned in the ratio 1:1:1 and were added to the GIC powder to form concentrations of 1.5% and 3% w/w antibiotics in the GIC formulation. Following the completion of measurements, all the experimental groups were stored in separate airtight containers containing desiccant (silica gel).

Table 1: Preparation of antibacterial cement

Group	Composition	Concentration of chlorhexidine and antibiotic in GIC
IIA	60 mg of CHX added to 3940 mg of GIC powder	1.5%
IIB	120 mg of CHX added to 3880 mg of GIC powder	3%
IIIA	20 mg of ciprofloxacin, 20 mg of metronidazole, 20 mg of minocycline added to 3940 mg of GIC powder	1.5%
IIIB	40 mg of ciprofloxacin, 40 mg of metronidazole, 40 mg of minocycline added to 3880 mg of GIC powder	3%

Preparation of samples

The powder and liquid (P/L ratio 3.6:1, as per the manufacturer's instruction) for each experimental group were dispensed on a mixing pad and mixed with a plastic spatula for 30 s. The material was transferred into standardized molds with a plastic instrument from one end of the mold till the material extruded from the other end. Excess material was removed from the molds and both ends of the mold were covered with a glass slide held under pressure. This was done to facilitate the setting of material without any surface defects or voids. The specimens were allowed to set for 30 min at room temperature. Following this, the specimens were carefully teased out of the molds by applying pressure at one end with a condenser. [Figure 1].

A 500-grit silicon carbide paper was used to finish any visible irregularities at the ends of the specimen. The height and diameter of each specimen were then measured using digital callipers. The specimens for evaluating antimicrobial efficacy were immediately inoculated, while those for compressive strength evaluation were stored in distilled water at 37°C for 24 h.

Microbial strain and growth media

The antibacterial activity was evaluated against *Streptococcus mutans*. Stock culture of *S. mutans* (MTCC No. 497) procured from MTCC, Institute of Microbial Technology (IMTECH), Chandigarh was used in the present study. A loopful of bacterial inoculum from the lyophilized culture was transferred to Brain Heart Infusion (BHI) broth and incubated for 24 h at 37°C. The bacterial growth was assessed by the appearance of turbidity in the broth after



Figure 1: Specimens allowed to set at room temperature and teased out of molds

incubation [Figure 2]. BHI agar was used as a culture medium for agar diffusion test.

In vitro evaluations

Agar diffusion test

Fresh culture of *S. mutans* from turbid BHI broth was flood inoculated onto the surface of BHI agar plates. Bacterial lawn was prepared by spread plate method with a volume of 500 µl of bacterial inoculum by using a micropipette (100–1000 µl). The specimens were placed in hot air oven for a period of 1 h to achieve suitable sterilization. The specimens were then placed on BHI agar plates having bacterial strain and incubated at 37°C for 24–48 h. Zones of inhibition around the specimens were measured in millimeters using Hi-Veg Media Antibiotic zone scale (Hi Media Laboratories, Mumbai) [Figures 3 and 4].

Evaluation of compressive strength

After preparation, the specimens were stored in distilled water and 24-h compressive strength was evaluated. Prior to testing, the diameter and height of each specimen were determined using a digital calliper. The specimens were placed with the flat ends up between the plates of the Universal Testing Machine (Hounsfield UTM) [Figure 5]. The strength of the specimen was then recorded in MPa by applying a compressive load along the long axis of the cylindrical pellet at a crosshead speed of 0.5 mm/min.

Statistical analysis

The resultant findings for antimicrobial efficacy (48 h) and compressive strength (24 h) were statistically

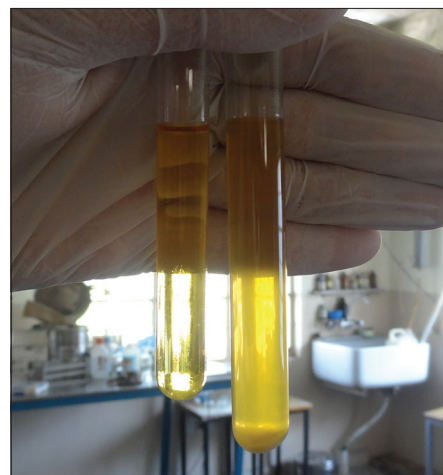


Figure 2: Bacterial growth assessed by appearance of turbidity in the broth

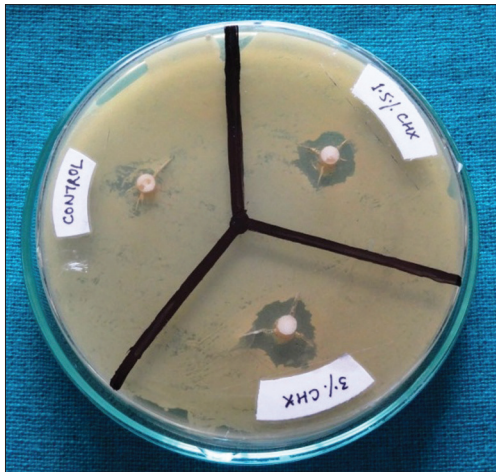


Figure 3: Zones of inhibition around specimens containing chlorhexidine

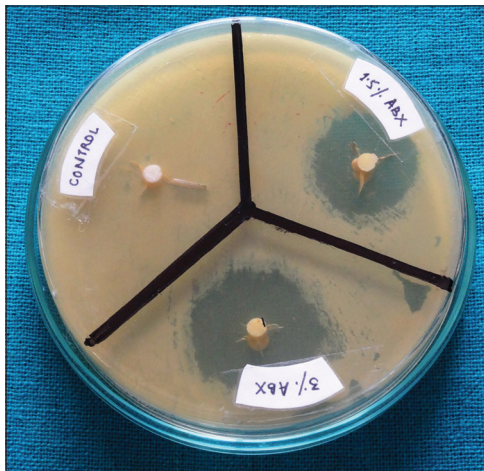


Figure 4: Zones of inhibition around specimens containing antibiotics



Figure 5: Specimen being tested for compressive strength using Universal Testing Machine

analyzed using one-way analysis of variance (ANOVA) [Table 2] and Tukey's *post hoc* test.

RESULTS

Antimicrobial activity screening test

At the end of 48 h, Group I (control) exhibited no zone of inhibition against the test organism. However, large zones of inhibition, as determined by the Hi-Veg Media Antibiotic zone scale, were observed around specimens of groups II and III [Figure 6]. Also, the size of inhibition zones was dependant on the amount of antimicrobial agent added.

Significant differences existed in the size of the inhibition zones produced among the control and experimental groups ($P < 0.05$). Zones of inhibition recorded for Group III were significantly higher ($P < 0.05$) as compared to Group II.

Evaluation of compressive strength

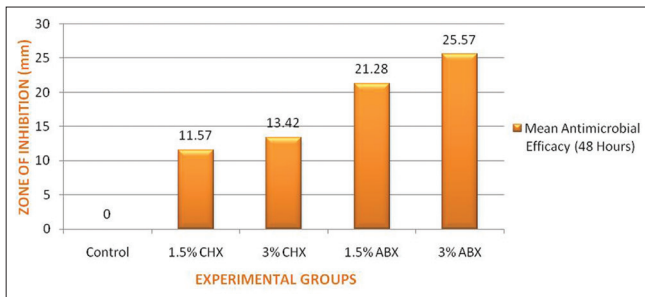
At the end of 24 h, the experimental groups showed lower compressive strength when compared to the control group, with a statistically significant difference ($P < 0.05$). However, there was no significant difference ($P > 0.05$) in the mean compressive strength values of groups II and III for both concentrations. The compressive strength decreased in a concentration-dependant manner [Figure 7].

DISCUSSION

The quest to search an ideal restorative material has been a challenge for the researchers and academicians in the fraternity of restorative dentistry. Glass ionomers are a class of biomaterials in widespread use in modern dentistry.^[14] GICs are capable of releasing fluoride, which contributes to some reduction in the number of residual bacteria in cavities as well as remineralization of the softened dentin.^[15-18] However, even after the removal of infected dentin and adequate sealing, viable bacteria have been found in the remaining affected dentine after different periods of evaluation.^[19] Literature is a testimony to the fact that therapeutic benefits have been gained when antimicrobial substances like chlorhexidine and antibiotics are used in association with GIC; however, a compromise of the strength characteristics has always emerged unconcealed.^[4,7,13,16] Therefore, a comparison of the influence of incorporating CHX and triple antibiotic mixture (ABX) consisting of ciprofloxacin, metronidazole, and minocycline on the antibacterial efficacy and compressive strength of GIC was the prime objective of our study.

Table 2: Results of analysis of variance (ANOVA) comparing different groups for compressive strength and antimicrobial efficacy

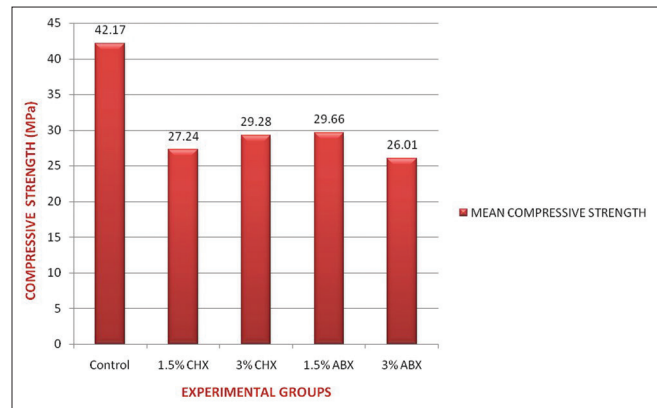
Variable	Intercomparison	Sum of squares	df	Mean square	F	Sig.
Compressive strength (24 h)	Between groups	1512.092	6	252.015	4.980	0.001
	Within groups	2125.232	42	50.601		
	Total	3637.324	48			
Antimicrobial efficacy (48 h)	Between groups	3590.490	6	598.415	2094.452	0.000
	Within groups	12.000	42	0.286		
	Total	3602.490	48			

**Figure 6:** Mean values of zone of inhibition for control and experimental groups in millimeters

Fuji IX GIC was used as the control in the present study, since this is the most frequently reported material in *in vivo* and *in vitro* studies in the past.^[4,13,20] Considering the material as a gold standard in high-strength posterior restoratives, it could be efficiently used for the assessment of compressive strength of the modified cement. CHX and triple antibiotic mixture were the preferred choice of antimicrobials. The efficacy of chlorhexidine has been proven against oral pathogens, primarily *S. mutans*.^[21] Different salts of chlorhexidine, mainly digluconate and diacetate, are commercially available as pre-weighed packages. CHX was preferred to other CHX derivatives (chlorhexidine gluconate) in the present study, as it is a more stable material, not prone to decomposition, and can be easily blended with GIC powder.^[22]

The antibacterial efficacy of the modified cements was illustrated against *S. mutans*. *S. mutans* bacteria are the most cariogenic pathogens as they induce an acid tolerance response that enables this pathogen to survive and grow in low-pH environments. Considering the impact of *S. mutans* as an initiator of the pathological process of dental caries, it was selected as the test organism.

Considering that the development of dental caries is attributed to a diverse, abundant, and complex microbial community, the use of a mixture of antibiotics is a better alternative than the use of a single antibiotic.^[13,23] Pinherio *et al.*^[24] suggested that a GIC containing triple antibiotic mixture may be used for the treatment

**Figure 7:** Mean values of compressive strength for control and experimental groups in megapascals

of carious lesions, as it reduces total viable bacteria. These studies were contemplated and ciprofloxacin, metronidazole, and minocycline were selected for the mixture of antibiotics that was tested in the present research.

The concentration of antibiotics and material preparation were determined based on the study conducted by Yesilyurt *et al.*^[13] CHX was proportioned into GIC in similar ratios to ensure adequate intercomparison between the two antimicrobial agents. The antibacterial efficacy of the modified cements was illustrated against *S. mutans*. Agar plate diffusion was the method of choice to evaluate the antimicrobial efficacy as the process is relatively inexpensive and can be performed rapidly and easily with a large number of specimens.^[16] The diameter of the inhibition zone is related to the susceptibility of the isolate and to the diffusion rate of the drug through the agar medium.^[25-27]

The agar diffusion test in our study demonstrated that Fuji IX GIC showed no antibacterial effect against *S. mutans*. These results were consistent with the findings of Botelho *et al.*,^[28] Yap *et al.*,^[29] and Yesilyurt *et al.*^[13] On the contrary, Shashibhushan *et al.*^[30] demonstrated that some degree of growth inhibition of mutans streptococci is exhibited by

GICs due to the release of fluoride and zinc ions into an aqueous medium, which may inhibit the growth of mutans streptococci. However, the release of these ions from GICs is governed by intrinsic and extrinsic factors such as preparation of the material, its P/L ratio, manipulation time, temperature, specimen geometry, surface protection, storage/dissolution of the medium, and the analytic method used. A combination of these factors could have possibly contributed to the absence of inhibition zones around the control specimens.

The addition of triple antibiotic mixture and CHX to GIC enhanced its antimicrobial efficacy in a concentration-dependant manner. The results of our study were complementary to the findings of Ribeiro *et al.*^[31] and Prabhakar *et al.*,^[7] who demonstrated a similar dose response effect. The antimicrobial efficacy of both concentrations of the antibiotic group was significantly higher as compared to the chlorhexidine group. This could possibly be accredited to the difference in mechanism of action of the two antimicrobial agents. At low concentrations, the bacteriostatic effect of CHX is based on disturbance of bacterial cell functions, enzymes, and cell receptors and at high concentrations, CHX causes cytoplasmic precipitation or coagulation.^[32,33] In contrast, the triple antibiotic mixture has a broader spectrum of antibacterial activity and a versatile antibacterial action. Though metronidazole is a narrow-spectrum antibiotic effective only against obligate anaerobes, ciprofloxacin and minocycline have proven efficacy against *S. mutans*.^[34] Moreover, the agar diffusion assay is highly affected by material diffusibility through the agar. The powdered antibiotic particles easily absorb water and disseminate through the agar medium, as compared to CHX.

The clinical utility of a material is defined by its ability to endure the stresses and strains induced during mastication and function. The most commonly used strength value to characterize dental cements is compressive strength.^[4] In the present study, compressive strength of the experimental groups was considerably lower as compared to the control group. The recorded values of compressive strength (MPa) for both control and experimental groups were less than that reported in literature.^[4,16,35] This could be due to the difference in the method of casting the specimens and the mechanical testing procedure. Since the detection of internal defects was beyond the confines of our study, their effect on the compressive strength of the material cannot be ignored.

The compressive strength of experimental groups containing CHX and antibiotics decreased in a concentration-dependant manner. The cross-linking in GIC is because of the coordination of Al^{3+} and Ca^{2+} with the COOH groups on the acidic polymers. Due to vitrification of GIC with antimicrobials, many of these COOH groups are prevented from participating in these coordination complexes.^[36] In addition, variation in the P/L ratio by addition of antimicrobials may also have attributed to the decrease observed in compressive strength.^[37-41] Moreover, the powdered antibiotic particles which are added to GIC easily absorb water.^[13] The absorption of water can decrease the compressive strength of the GIC.

Since the addition of antimicrobial agents to GIC can affect the mechanical properties of the cement,^[42] the particular antimicrobial agent and its quantity are important aspects to be determined. The antimicrobial efficacy of the 1.5% ABX group was significantly higher than that in the 1.5% CHX and 3% CHX groups. The mean compressive strength of 1.5% ABX was also better as compared to both 1.5% CHX and 3% CHX. Although the compressive strength of the 1.5% ABX group was significantly compromised as compared to the control group, therapeutic properties obtained from these materials might outdo the disadvantages of altered mechanical properties. Nevertheless, the amount of antimicrobial agent must be kept as low as possible, as high amounts of additives would weaken the scaffold and the glass ionomer network, thereby compromising the physical properties.

CONCLUSION

The present *in vitro* study demonstrated that the addition of chlorhexidine and antibiotics to GIC decreased the compressive strength in all experimental groups. However, within the limitations of the present study, incorporation of 1.5% ABX into glass ionomer appears to provide an acceptable combination of properties. Further *in vivo* studies are required to test the clinical efficacy of this concentration before advocating the use of antibiotic-modified GIC in ART procedures. Endeavors to compare lower concentrations of chlorhexidine with antibiotics for other properties like fluoride release, bond strength, diametral tensile strength, and biaxial flexure strength must be attempted in future studies. Therefore, until the long-term effects of the antibiotic-modified GIC are investigated, this modified cement can be used as a base material under the conventional glass ionomer restorations.

Financial support and sponsorship

Nil.

Conflicts of interest

There are no conflicts of interest.

REFERENCES

- Ahluwalia P, Chopra S, Thomas AM. Strength characteristics and marginal sealing ability of chlorhexidine-modified glass ionomer cement: An *in vitro* study. J Indian Soc Pedod Prev Dent 2012;30:41-6.
- Moses J, Rangeeth BN, Gurunathan D. Prevalence of dental caries, socio-economic status and treatment needs among 5 to 15 year old school going children of Chidambaram. J Clin Diagn Res 2011;5:146-51.
- Frencken JE, van Amerongen WE. The atraumatic restorative treatment approach. In: Fejerskov O, Kidd E, Bente N, editors. Dental Caries: The Disease and its Clinical Management. 2nd ed. Oxford, UK: Blackwell Munksgaard; 2008. p. 427-42.
- Deepalakshmi M, Poorni S, Miglani R, Rajamani I, Ramachandran S. Evaluation of the antibacterial and physical properties of glass ionomer cements containing Chlorhexidine and Cetrimide: An *in-vitro* study. Indian J Dent Res 2010;21:552-6.
- Weerheijm KL, Kreulen CM, de Soet JJ, Groen HJ, van Amerongen WE. Bacterial counts in carious dentine under restorations: 2-year *in vivo* effects. Caries Res 1999;33:130-4.
- Weng Y, Guo X, Gregory R, Xie D. A novel antibacterial dental glass-ionomer cement. Eur J Oral Sci 2010;118:531-4.
- Prabhakar AR, Prahlad D, Kumar SR. Antibacterial activity, fluoride release, and physical properties of an antibiotic-modified glass ionomer cement. Pediatr Dent 2013;35:411-5.
- Hook ER, Owen OJ, Bellis CA, Holder JA, O'Sullivan DJ, Barbour ME. Development of a novel antimicrobial-releasing glass ionomer cement functionalized with chlorhexidine hexametaphosphate nanoparticles. J Nanobiotechnology 2014;12:3.
- Emilson CG. Potential efficacy of chlorhexidine against mutans streptococci and human dental caries. J Dent Res 1994;73:682-91.
- Rölla G, Löe H, Schiott CR. The affinity of chlorhexidine for hydroxyapatite and salivary mucins. J Periodontol Res 1970;5:90-5.
- Rosenthal S, Spångberg L, Safavi K. Chlorhexidine substantivity in root canal dentine. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2004;98:488-92.
- Khadeimi AA, Mohammadi Z, Havaee A. Evaluation of the antibacterial substantivity of several intra-canal agents. Aust Endod J 2006;32:112-5.
- Yesilyurt C, Er K, Tasdemir T, Buruk K, Celik D. Antibacterial activity and physical properties of glass-ionomer containing antibiotics. Oper Dent 2009;34:18-23.
- Sidhu SK. Glass ionomer cement restorative materials: A sticky subject? Aust Dent J 2011;56(Suppl 1):23-30.
- Herrera M, Castillo A, Baca P, Carrión P. Antibacterial activity of glass-ionomer restorative cements exposed to cavity-producing microorganisms. Oper Dent 1999;24:286-91.
- Türkün LS, Türkün M, Ertuğrul F, Ateş M, Brugger S. Long-term antibacterial effects and physical properties of a chlorhexidine-containing glass ionomer cement. J Esthet Restor Dent 2008;20:29-45.
- Herrera M, Castillo A, Bravo M, Liébana J, Carrión P. Antibacterial activity of resin adhesives, glass ionomer and resin-modified glass ionomer cements and a compomer in contact with dentin caries samples. Oper Dent 2000;25:265-9.
- Massara ML, Alves JB, Brandão PR. Atraumatic restorative treatment: Clinical, ultrastructural and chemical analysis. Caries Res 2002;36:430-6.
- Björndal L, Larsen T, Thylstrup A. A clinical and microbiological study of deep carious lesion during stepwise excavation using long treatment intervals. Caries Res 1997;31:411-7.
- Tuzuner T, Kuşgöz A, Kursat ER, Taşdemir T, Buruk K, Kemer B. Antibacterial activity and physical properties of conventional glass-ionomer cements containing chlorhexidine diacetate/cetrimide mixtures. J Esthet Restor Dent 2011;23:46-55.
- Emilson CG. Susceptibility of various microorganisms to chlorhexidine. Scand J Dent Res 1977;85:255-65.
- Block SS. Disinfection, Sterilization and Preservation. 4th ed. Malvern: Lea and Febiger; 1991. p. 274-5.
- van Houte J. Role of micro-organisms in caries etiology. J Dent Res 1994;73:672-81.
- Pinheiro SL, Simionato MR, Imparato JC, Oda M. Antibacterial activity of glass-ionomer cement containing antibiotics on caries lesion microorganisms. Am J Dent 2005;18:261-6.
- Al-Khatib ZZ, Baum RH, Morse DR, Yesilsoy C, Bhambhani S, Furst ML. The antimicrobial effect of various endodontic sealers. Oral Surg Oral Med Oral Pathol 1990;70:784-90.
- Abdulkader A, Deguid R, Saunders EM. The antimicrobial activity of endodontic sealers to anaerobic bacteria. Int Endod J 1996;29:280-3.
- Siqueira JF Jr, Favieri A, Gahyva SM, Moraes SR, Lima KC, Lopes HP. Antimicrobial activity and flow rate of newer and established root canal sealers. J Endod 2000;26:274-7.
- Botelho MG. Inhibitory effects on selected oral bacteria of antibacterial agents incorporated in a glass ionomer cement. Caries Res 2003;37:108-14.
- Yap AU, Khor E, Foo SH. Fluoride release and antibacterial properties of new generation tooth-colored restoratives. Oper Dent 1999;24:297-305.
- Shashibhushan KK, Basappa N, Subba Reddy VV. Comparison of antibacterial active ity of three fluorides- and zinc-releasing commercial glass ionomer cements on strains of mutans streptococci: An *in vitro* study. J Indian Soc Pedod Prev Dent 2008;26(Suppl 2):S56-61.
- Ribeiro J, Ericson D. *In vitro* antibacterial effect of chlorhexidine added to glass-ionomer cements. Scand J Dent Res 1991;99:533-40.
- Marsh PD, Keevil CW, McDermid AS, Williamson MI, Ellwood DC. Inhibition by the antimicrobial agent chlorhexidine of acid production and sugar transport in oral streptococcal bacteria. Arch Oral Biol 1983;28:233-40.
- Hennessey TD. Antibacterial properties of Hibitane. J Clin Periodontol 1977;4:36-48.
- Choudhary S, Singh V, Chauhan PK, Tyagi A, Kumar M. *In vitro* antibacterial activity of *Engenia Jambolana* against *Streptococcus mutans* causing dental plaque formation. Int J Inst Pharm Life Sci 2011;1:91-9.

35. Chandana PS, Munaga S, Reddy MN, Devabhaktuni D, Swathi CL. Evaluation of compressive strength for a combination of glass ionomer cement and antibiotics. *J Orofac Res* 2013;3:245-8.
36. Moshaverinia A, Brantley WA, Chee WW, Rohpour N, Ansari S, Zheng F, *et al.* Measure of microhardness, fracture toughness and flexure strength of N-vinylcaprolactam (NVC)- containing glass-ionomer cement. *Dent Mater* 2010;26:1137-43.
37. Billington RW, Williams JA, Pearson GJ. Variation in powder/liquid ratio of restorative glass-ionomer cement used in dental practice. *Br Dent J* 1990;169:164-7.
38. Crisp S, Lewis BG, Wilson AD. Characterization of glass-ionomer cements. 2. Effect of the powder: Liquid ratio on the physical properties. *J Dent* 1976;4:287-90.
39. Ewoldsen N, Covey D, Lavin M. The physical and adhesive properties of dental cements used for atraumatic restorative treatment. *Spec Care Dentist* 1997;17:19-24.
40. Kerby RE, Knobloch L. Strength characteristics of glass-ionomer cements. *Oper Dent* 1992;17:170-4.
41. Wilder AD, Boghsian AA, Bayne SC, Heymann HO, Sturdevant JR, Roberson TM. Effect of powder/liquid ratio on the clinical and laboratory performance of resin-modified glass-ionomers. *J Dent* 1998;26:369-77.
42. Sanders BJ, Gregory RL, Moore K, Avery DR. Antibacterial and physical properties of resin modified glass-ionomers combined with chlorhexidine. *J Oral Rehabil* 2002;29:553-8.