

## CROSSTALK

**CrossTalk opposing view: 5-HT is not necessary for peristalsis**

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**Contention**

Our contention is that endogenous 5-hydroxytryptamine (5-HT) is not necessary for colonic peristalsis, nor other neurogenic motor patterns in the lower gastrointestinal (GI) tract, such as colonic migrating motor complexes (CMMCs).

**Background**

Large quantities of 5-hydroxytryptamine (5-HT) are synthesized in the gut, in enterochromaffin (EC) cells of the mucosa (via tryptophan hydroxylase-1) and in a small proportion in enteric neurons (via tryptophan hydroxylase-2). It has been well established that 5-HT can be released from the mucosa by mechanical stimulation, including contractile activity of the gut and compression of the gut wall (Bertrand, 2006). Numerous 5-HT receptors are expressed on enteric neurons and extrinsic afferent endings, and exogenous 5-HT has potent effects on neuronal excitability and transmitter release (Gershon, 2000). Despite this, the evidence that 5-HT is necessary for peristalsis or CMMCs in the lower GI-tract is all circumstantial. Extensive evidence now suggests that these motor patterns occur independent of 5-HT.

**Three lines of disputed evidence**

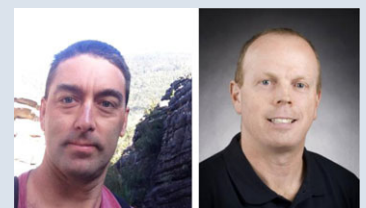
**High quantities of 5-HT are synthesized and released from the mucosa. Release of endogenous 5-HT can occur during peristalsis and CMMCs and exogenous 5-HT can evoke peristalsis or CMMCs.** The observation that mucosal compression causes release of *endogenous* 5-HT at the same time as the onset of peristalsis or CMMCs does not reflect a causal relationship between these events. Mucosal compression distorts multiple cell types in the gut wall, including the underlying myenteric plexus – which is itself exquisitely sensitive to mechanical deformation (Kunze *et al.* 2000). Compression applied to the myenteric plexus still evokes peristaltic reflexes, even after the mucosa has been removed (Spencer *et al.* 2003) and CMMCs also persisted in these preparations (Keating & Spencer, 2010; Zagorodnyuk & Spencer, 2011). Thus, while mucosal stimulation can release large amounts of *endogenous* 5-HT, we contend that this release is not a *necessary* step in initiating peristalsis or CMMCs. The observation that *exogenous* 5-HT (and many other agonists) can initiate peristalsis and CMMCs does not reveal any information about the functional role of *endogenous* 5-HT.

It has been suggested that *endogenous* 5-HT release from the mucosa initiates the colonic peristaltic reflex (Foxy-Orenstein *et al.* 1996; Grider *et al.* 1996) and is ‘critical’ for spontaneous CMMCs (Heredia *et al.* 2009). It was reported that ‘removing the mucosa appeared to abolish spontaneous CMMCs, suggesting that the mucosa is normally critical for their generation’ (Heredia *et al.* 2009) – a conclusion based on four mice. It was further stated that ‘the trigger for the CMMC appears to be spontaneous

or evoked (i.e. a fecal pellet) release of 5-HT from EC cells to stimulate AH neurons’. Although, 5-HT release from the mucosa was not measured, nor were recordings made from neurons to confirm this hypothesis. If 5-HT release from the mucosa was essential for peristalsis or CMMCs, then both these motor patterns would be expected to cease when the mucosa is removed. They don’t. With practice it was demonstrated that the entire mucosa can be reliably removed from an isolated specimen of colon and still record peristalsis (Spencer *et al.* 2011) and *spontaneous* and *evoked* CMMCs (Keating & Spencer, 2010). In these mucosa-free preparations, release of 5-HT that is normally associated with peristaltic or CMMC contractions was abolished. Importantly, when removing the mucosa we observed that if the dissection technique was too vigorous, the delicate neural circuitry required for peristalsis or CMMCs is disrupted. This may explain why spontaneous CMMCs were not recorded after removal of the mucosa in the four mice studied in Heredia *et al.* (2009). We consider it inconceivable that dissection damage in our preparations could have created a whole new motor pattern that mimicked normal peristalsis and CMMCs.

In other studies, electrochemical detection of 5-HT release from the mucosa showed that 5-HT release was only associated with some, *but not all*, contractions underlying peristalsis (Bertrand, 2006) or CMMCs (Keating & Spencer, 2010), again arguing against a causal link between 5-HT release and these colonic motor patterns. If 5-HT release from the mucosa was critical for peristalsis and CMMCs, then deleting the gene responsible for synthesizing mucosal 5-HT (TPH-1) should block CMMCs and affect transit. It doesn’t. The laboratory that concluded 5-HT release from the mucosa

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was critical for CMMCs (Heredia *et al.* 2009) also showed that CMMCs still occurred when mucosal 5-HT synthesis had been prevented (Heredia *et al.* 2013). Furthermore, conscious mice lacking mucosal 5-HT synthesis have *no* inhibitory deficits in GI-transit (Yadav *et al.* 2010; Li *et al.* 2011).

### Endogenous 5-HT is synthesized in enteric neurons and numerous 5-HT receptors are expressed in the gut.

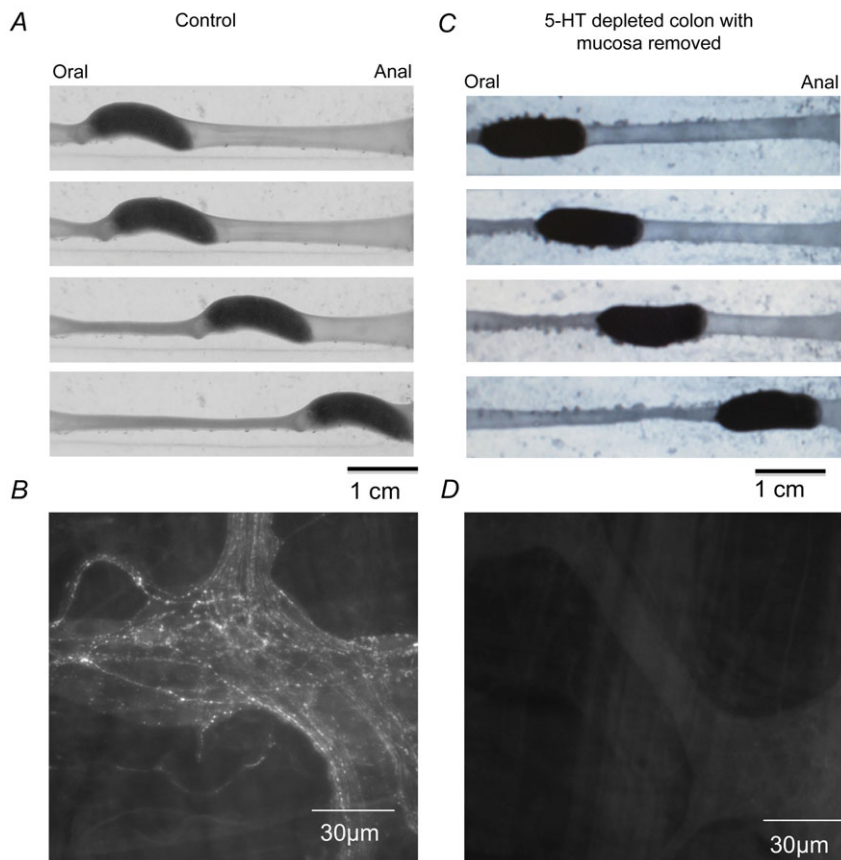
There is some evidence that 5-HT may be a neurotransmitter in a small population of myenteric (Zhou & Galligan, 1999; Galligan *et al.* 2000) and submucosal neurons (Monro *et al.* 2004) of the small intestine, despite only ~1% of enteric neurons expressing 5-HT (Costa *et al.* 1982; Wardell *et al.* 1994; Costa *et al.* 1996). For a substance to be a neurotransmitter, it must be present in the presynaptic terminal and is released in amounts sufficient to exert a defined action on the postsynaptic

neuron or effector organ' (Gardner *et al.* 2000). Based on this criterion, we are not aware of any evidence that *endogenous* 5-HT participates in any synaptic potential in the colon of the mouse (Furukawa *et al.* 1986; Nurgali *et al.* 2004) and rarely (Nurgali *et al.* 2003) or never (Wade & Wood, 1988; Spencer & Smith, 2004) in the colon of the guinea-pig, which are the species we have been studying CMMCs and peristalsis with. In our experience in guinea-pig colon (Spencer & Smith, 2004) and in mouse (Furukawa *et al.* 1986; Nurgali *et al.* 2004), human (Brookes *et al.* 1987) and rat colon (Brookes *et al.* 1988), all fast synaptic potentials are blocked by hexamethonium.

### 5-HT antagonists can block peristalsis and CMMCs and retard transit.

Perhaps the strongest evidence that endogenous 5-HT is necessary for peristalsis is based on the finding that 5-HT

antagonists can substantially inhibit or block colonic peristalsis (Foxx-Orenstein *et al.* 1996; Grider *et al.* 1996; Kadowaki *et al.* 1996). However, when these experiments were repeated with the same antagonists (SDZ-205-557 and ondansetron) they were only found to cause a temporary blockade of peristalsis. In the continued presence of both antagonists peristalsis reappeared (Sia *et al.* 2013*a,b*). Importantly, these antagonists also blocked peristalsis in colonic preparations from mucosa-free, reserpine-treated animals, where endogenous 5-HT had been depleted. This showed that endogenous 5-HT may not be necessary for activation of 5-HT<sub>3</sub> and 5-HT<sub>4</sub> receptors. We suggest that 5-HT<sub>3</sub> and 5-HT<sub>4</sub> antagonists bind to 5-HT<sub>3</sub> and 5-HT<sub>4</sub> receptors which are constitutively active even in the absence of any 5-HT. Indeed, ligand-gated 5-HT<sub>3</sub> receptors (Hu & Peoples, 2008) and G-protein coupled 5-HT<sub>4</sub> receptors (Berthouze *et al.* 2005) both show constitutive activity. These



**Figure 1. Colonic Peristalsis occurs without endogenous 5-HT**

A, peristalsis in the distal colon. B, 5-HT immunoreactivity. C, in mucosa-free colon also depleted of neuronal 5-HT, peristalsis still occurs. D, confirmation of depletion of neuronal 5-HT after reserpine.

antagonists could work as inverse agonists, reducing the constitutive activity of the receptors, rather than blocking the effects of endogenous 5-HT.

At present, we are unsure of the role of 5-HT in the lower GI tract. We do know that 5-HT can have many effects and it *may* modulate enteric circuits and motility. However, recent evidence from a number of laboratories, using a number of techniques, shows that neither mucosal nor neuronal 5-HT is necessary for colonic peristalsis or CMMCs.

## References

- Berthouze M, Ayoub M, Russo O, Rivail L, Sicsic S, Fischmeister R, Berque-Bestel I, Jockers R & Lezoualc'h F (2005). Constitutive dimerization of human serotonin 5-HT<sub>4</sub> receptors in living cells. *FEBS Lett* **579**, 2973–2980.
- Bertrand PP (2006). Real-time measurement of serotonin release and motility in guinea pig ileum. *J Physiol* **577**, 689–704.
- Brookes SJ, Ewart WR & Wingate DL (1987). Intracellular recordings from myenteric neurones in the human colon. *J Physiol* **390**, 305–318.
- Brookes SJ, Ewart WR & Wingate DL (1988). Intracellular recordings from cells in the myenteric plexus of the rat duodenum. *Neuroscience* **24**, 297–307.
- Costa M, Brookes SJ, Steele PA, Gibbins I, Burcher E & Kandiah CJ (1996). Neurochemical classification of myenteric neurons in the guinea-pig ileum. *Neuroscience* **75**, 949–967.
- Costa M, Furness JB, Cuello AC, Verhofstad AA, Steinbusch HW & Elde RP (1982). Neurons with 5-hydroxytryptamine-like immunoreactivity in the enteric nervous system: their visualization and reactions to drug treatment. *Neuroscience* **7**, 351–363.
- Foxx-Orenstein AE, Kuemmerle JF & Grider JR (1996). Distinct 5-HT receptors mediate the peristaltic reflex induced by mucosal stimuli in human and guinea pig intestine. *Gastroenterology* **111**, 1281–1290.
- Furukawa K, Taylor GS & Bywater RA (1986). An intracellular study of myenteric neurons in the mouse colon. *J Neurophysiol* **55**, 1395–1406.
- Galligan JJ, LePard KJ, Schneider DA & Zhou X (2000). Multiple mechanisms of fast excitatory synaptic transmission in the enteric nervous system. *J Auton Nerv Syst* **81**, 97–103.
- Gardner EP, Martin JH & Jessell TM (2000). The bodily senses. In *Principles of Neural Science*, 4th edn, ed. Kandel ER, Schwartz JH & Jessell TM, pp. 430–450. McGraw-Hill.
- Gershon MD (2000). 5-HT (serotonin) physiology and related drugs. *Curr Opin Gastroenterol* **16**, 113–120.
- Grider JR, Kuemmerle JF & Jin JG (1996). 5-HT released by mucosal stimuli initiates peristalsis by activating 5-HT<sub>4</sub>/5-HT<sub>1p</sub> receptors on sensory CGRP neurons. *Am J PhysiolGastrointestLiver Physiol* **270**, G778–782.
- Heredia DJ, Dickson EJ, Bayguinov PO, Hennig GW & Smith TK (2009). Localized release of serotonin (5-hydroxytryptamine) by a fecal pellet regulates migrating motor complexes in murine colon. *Gastroenterology* **136**, 1328–1338.
- Heredia DJ, Gershon MD, Koh SD, Corrigan RD, Okamoto T & Smith TK (2013). Important role of mucosal serotonin in colonic propulsion and peristaltic reflexes: *in vitro* analyses in mice lacking tryptophan hydroxylase 1. *J Physiol* **591**, 5939–5957.
- Hu XQ & Peoples RW (2008). The 5-HT<sub>3B</sub> subunit confers spontaneous channel opening and altered ligand properties of the 5-HT<sub>3</sub> receptor. *J Biol Chem* **283**, 6826–6831.
- Kadowaki M, Wade PR & Gershon MD (1996). Participation of 5-HT<sub>3</sub>, 5-HT<sub>4</sub>, and nicotinic receptors in the peristaltic reflex of guinea pig distal colon. *Am J PhysiolGastrointestLiver Physiol* **271**, G849–857.
- Keating DJ & Spencer NJ (2010). Release of 5-hydroxytryptamine from the mucosa is not required for the generation or propagation of colonic migrating motor complexes. *Gastroenterology* **138**, 659–670.
- Kunze WA, Clerc N, Furness JB & Gola M (2000). The soma and neurites of primary afferent neurons in the guinea-pig intestine respond differentially to deformation. *J Physiol* **526**, 375–385.
- Li Z, Chalazonitis A, Huang YY, Mann JJ, Margolis KG, Yang QM, Kim DO, Cote F, Mallet J & Gershon MD (2011). Essential roles of enteric neuronal serotonin in gastrointestinal motility and the development/survival of enteric dopaminergic neurons. *J Neurosci* **31**, 8998–9009.
- Monro RL, Bertrand PP & Bornstein JC (2004). ATP participates in three excitatory postsynaptic potentials in the submucous plexus of the guinea pig ileum. *J Physiol* **556**, 571–584.
- Nurgali K, Furness JB & Stebbing MJ (2003). Analysis of purinergic and cholinergic fast synaptic transmission to identified myenteric neurons. *Neuroscience* **116**, 335–347.
- Nurgali K, Stebbing MJ & Furness JB (2004). Correlation of electrophysiological and morphological characteristics of enteric neurons in the mouse colon. *J Comp Neurol* **468**, 112–124.
- Sia TC, Flack N, Robinson L, Kyloh M, Nicholas SJ, Brookes SJ, Wattoo DA, Dinning P, Oliver J & Spencer NJ (2013a). Is serotonin in enteric nerves required for distension-evoked peristalsis and propulsion of content in guinea-pig distal colon? *Neuroscience* **240**, 325–335.
- Sia TC, Whiting M, Kyloh M, Nicholas SJ, Oliver J, Brookes SJ, Dinning PG, Wattoo DA & Spencer NJ (2013b). 5-HT<sub>3</sub> and 5-HT<sub>4</sub> antagonists inhibit peristaltic contractions in guinea-pig distal colon by mechanisms independent of endogenous 5-HT. *Front Neurosci* **7**, 136.
- Spencer NJ, Hennig GW & Smith TK (2003). Stretch-activated neuronal pathways to longitudinal and circular muscle in guinea pig distal colon. *Am J Physiol Gastrointest Liver Physiol* **284**, G231–241.
- Spencer NJ, Nicholas SJ, Robinson L, Kyloh M, Flack N, Brookes SJ, Zagorodnyuk VP & Keating DJ (2011). Mechanisms underlying distension-evoked peristalsis in guinea-pig distal colon: is there a role for enterochromaffin (EC) cells? *Am J Physiol Gastrointest Liver Physiol* **301**, G519–527.
- Spencer NJ & Smith TK (2004). Mechanosensory S-neurons rather than AH-neurons appear to generate a rhythmic motor pattern in guinea-pig distal colon. *J Physiol* **558**, 577–596.
- Wade PR & Wood JD (1988). Synaptic behavior of myenteric neurons in guinea pig distal colon. *Am J PhysiolGastrointestLiver Physiol* **255**, G184–190.
- Wardell CF, Bornstein JC & Furness JB (1994). Projections of 5-hydroxytryptamine-immunoreactive neurons in guinea-pig distal colon. *Cell Tissue Res* **278**, 379–387.
- Yadav VK, Balaji S, Suresh PS, Liu XS, Lu X, Li Z, Guo XE, Mann JJ, Balapure AK, Gershon MD, Medhamurthy R, Vidal M, Karsenty G & Ducy P (2010). Pharmacological inhibition of gut-derived serotonin synthesis is a potential bone anabolic treatment for osteoporosis. *Nat Med* **16**, 308–312.
- Zagorodnyuk VP & Spencer NJ (2011). Localization of the sensory neurons and mechanoreceptors required for stretch-evoked colonic migrating motor complexes in mouse colon. *Front Physiol* **2**, 98.
- Zhou X & Galligan JJ (1999). Synaptic activation and properties of 5-hydroxytryptamine<sub>3</sub> receptors in myenteric neurons of guinea pig intestine. *J PharmacolExpTher* **290**, 803–810.

## Additional information

### Competing interests

None declared.

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