

# **HHS Public Access**

Rev Endocr Metab Disord. Author manuscript; available in PMC 2016 June 01.

Published in final edited form as:

Author manuscript

Rev Endocr Metab Disord. 2015 June ; 16(2): 141–148. doi:10.1007/s11154-015-9315-2.

## LRP Receptor Family Member Associated Bone Disease

## N Lara-Castillo and ML Johnson

UMKC School of Dentistry, Department of Oral and Craniofacial Sciences, Kansas City, MO 64108

## Abstract

A dozen years ago the identification of causal mutations in the low-density lipoprotein receptorrelated protein 5 (LRP5) gene involved in two rare bone disorders propelled research in the bone field in totally new directions. Since then, there have been an explosion in the number of reports that highlight the role of the Wnt/ $\beta$ -catenin pathway in the regulation of bone homeostasis. In this review we discuss some of the most recent reports (in the past 2 years) highlighting the involvement of the members of the LRP family (LRP5, LRP6, LRP4, and more recently LRP8) in the maintenance of bone and their implications in bone diseases. These reports include records of new single nucleotides polymorphisms (SNPs) and haplotypes that suggest variants in these genes can contribute to subtle variation in bone traits to mutations that give rise to extreme bone phenotypes. All of these serve to further support and reinforce the importance of this tightly regulated pathway in bone. Furthermore, we discuss provocative reports suggesting novel approaches through inhibitors of this pathway to treat rarer diseases such as Osteoporosis-Pseudoglioma Syndrome (OPPG), Osteogenesis Imperfecta (OI), and Sclerosteosis/Van Buchem disease. It is hoped that by understanding the role of each component of the pathway and their involvement in bone diseases that this knowledge will allow us to develop new, more effective therapeutic approaches for more common diseases such as post-menopausal osteoporosis, osteoarthritis, and rheumatoid arthritis as well as these rarer bone diseases.

#### Keywords

LRP; Wnt/β-catenin; bone related disease

## Introduction

In the mid-1990's two independent groups, working on the seemingly unrelated low bone mass disease, Osteoporosis Pseudoglioma Syndrome (OPPG) [1] and the High Bone Mass (HBM) kindred [2] identified a common linkage region harboring the causal gene/mutations, which raised the possibility that these were conditions were allelic variants of each other [1, 2]. A few years later, both groups were able to identify the low-density lipoprotein receptor related-protein 5 (*LRP5*) gene and the mutations responsible for these bone associated traits: presumed loss-of-function mutations in *LRP5* were causative for OPPG [3] and a single

Address Correspondence to: Nuria Lara, Ph.D. Research Associate, Department of Oral and Craniofacial Sciences, UMKC School of Dentistry, 650 East 25<sup>th</sup> Street, Kansas City, MO 64108, Iaran@umkc.edu, Fax: (816) 235-5524.

amino acid change in the first β-propeller module of LRP5 was causal for the HBM trait [4]. At the time these mutations were identified little was known about the function of LRP5. Other studies [5] however demonstrated that LRP5 was a co-receptor for Wnt ligands regulating the Wnt/β-catenin signaling pathway. After publication of these two groundbreaking LRP5 papers the field of Wnt signaling in bone related diseases has grown exponentially [6-14]. Nowadays, LRP5 is not the only LDL receptor family members that is known to play an important role in bone homeostasis, more and more data has been collected that demonstrate an equally important role for Lrp6 [15-17] and recently, Lrp4 and Lrp8 have emerged as others receptors from the family that play important roles in bone [18-21].

Members of this family regulate the Wnt/ $\beta$ -catenin signaling pathway. Hence, manipulating this pathway has become a major target for developing new therapeutics to treat patients with post-menopausal osteoporosis [22-28] and other bone related diseases (for review see [29, 30]). In this review we will present the most recent reports describing the involvement of members of the LRP family in bone homeostasis, discuss newly discovered mutations in these proteins that are causative of various bone diseases, as well as address the new discoveries further illustrating the importance of these molecules in the treatment of bone diseases. We will not discuss the growing literature involving targeting of Wnt signaling pathways that do not involve LRP family members (i.e. the non-canonical pathways) or other diseases outside of bone in which modulation of this pathway may have clinical applications, such as cancer.

## The LRP family of proteins

The low-density lipoprotein receptor (LDLR) family consists of several members. Proteins in this family have characteristic features: in the extracellular domains they contain ligand binding repeats,  $\beta$ -propeller motifs and epidermal growth factor-like repeats. In the intracellular domains they have several domains that are responsible for downstream signaling events by interacting with cytoplasmic adaptors and scaffolds (for review see [30]). LRP5 and LRP6 are structurally related proteins and share around 71% homology at the nucleotide level [31]. LRP5 and LRP6 are type I transmembrane receptors (C-terminus in cytosol); at the extracellular domain they have four YWTD  $\beta$ -propellers, four EGF-like domains, and LDLR type A domains and at the intracellular domain they have five PPPSP motifs. The structural organization of LRP4 and LRP8 is markedly different from Lrp5/6 [30]. Lrp4 is a type II transmembrane receptor (N-terminus in cytosol) and it belongs to the LRP subfamily III along with LRP5 and LRP6 [32]. Similar to LRP 5/6 it has four  $\beta$ propeller motifs and four epidermal growth factor-like repeats; unlike LRP 5/6 proteins, LRP4 has a NPxY motif in the cytosolic domain. LRP8 is also known as apolipoprotein E receptor 2, it belongs to the LRP subfamily I along with LDLR and VLDLR. The difference between LRP5/6 is that LRP8 only contains one  $\beta$ -propeller, and like LRP4, it has one NPxY motif in the intracellular domain [30].

Binding of the Wnts, ligands for these receptors, to LRP5 or LRP6 (LRP5/6) and their participating co-receptors, the frizzled (Fz) family of 7 transmembrane spanning proteins, results in a series of downstream intracelullar events [5], in particular the inhibition by

subsequent phosphorylation of GSK-3 $\beta$  [33]. GSK-3 $\beta$  is critical for the intracellular fate of  $\beta$ -catenin. Normally  $\beta$ -catenin is held in a "degradation complex" that includes a scaffolding protein, Axin, and dozens of other proteins including GSK-3 $\beta$  [34]. When GSK-3 $\beta$  is active it phosphorylates  $\beta$ -catenin, which leads to its ubiquitination and degradation by the 26S proteasome [35, 36]. Binding of Wnt ligand leads to both the inhibition of GSK-3 $\beta$  by phosphorylation and the collapse of the degradation complex, which allows free  $\beta$ -catenin to accumulate in the cytoplasm allowing it to translocate to the nucleus and activate targeted genes (for review see [37]).

There are several extracellular inhibitors to the pathway including Sclerostin (the product of the *SOST* gene) [38], the Dickkopf family of proteins (Dkk1-4) [39-41] along with Kremen 1 and 2 [42, 41, 43], and the secreted frizzled related proteins (sFRPs) [44, 45]. In addition the Wise protein modulates the pathway in various contexts, mainly during development [46-48]. Sclerostin and Dkk-1 have a similar mode of inhibiting activation of the pathway; both bind to  $\beta$ -propeller motifs inhibiting the formation of Wnt/LRP/Fz trimolecular complex. The difference is the binding site: sclerostin binds to the first or second  $\beta$ -propeller whereas Dkk-1 binds to the third  $\beta$ -propeller. Kremen acts with high affinity for Dkk and forms a ternary complex with Dkk and LRP5/6 inducing endocytosis of this formed complex and resulting of the removal of LRP5/6 from plasma membrane [49, 41, 40]. sFRP-1 and sFRP-4 inhibit the pathway by binding Wnt ligand and preventing its binding to the Lrp5/6-frizzled co-receptor complex [44].

While the involvement of LRP5 and LRP6 in bone homeostasis has been established for the past dozen years, more recently MEGF4/LRP4 and LRP8 have been shown to play a role [50, 19, 20, 18, 21, 51] Global deletion of Lrp5 (Lrp5<sup>-/-</sup>) in mice resembles the phenotypes observed in patients with OPPG. These mice present low bone mass phenotype and a persistent embryonic eye vascularization through their adult life [52]. Kato and colleagues determined that the reason for the low bone mass phenotype in this animal model was due to a decreased osteoblast proliferation and a decrease in bone matrix deposition. They also determined that the persistent embryonic eye vascularization was due to a reduced level of macrophage-induced endothelial cell apoptosis [52]. A mouse model for the HBM trait was originally created by Babij and colleagues in 2003 [53]. These transgenic mice carry the point mutation (G171V) (HMB) in the human LRP5 cDNA under the expression of the 3.6kb rat type I collagen promoter. Recently Warman and colleagues have generated knock-in mutations in the mouse germline harboring the original G171V HBM mutation and the A214V mutation [54]. All of these HBM mouse lines resemble the human phenotype of increase bone mass. Opposite to the Lrp5<sup>-/-</sup> mouse model, the HBM mice had decreased osteoblast apoptosis and no difference in the osteoclast number. The fact that Lrp5 might play a role in osteoblast proliferation and apoptosis in osteoblasts cells has recently supported by Javaheri et al (2013) [55]. In their paper they observed a sex difference in the proliferating state of the cells in vitro: osteoblasts-like cells from HBM female proliferated faster and presented lower apoptosis than littermate controls; and osteoblasts-like cells from  $Lrp5^{-/-}$  females proliferated slower with higher apoptosis rates than littermate controls. The authors also reported that the expected increase in cell number in response to high levels of strain (3,400µE) was no different between genotypes [55]. Mutation of Lrp4 caused skeletal

abnormalities. Lrp4<sup>-/-</sup> mice are smaller, with retardation in growth and delay in ossification; in addition, digits are fused and shortened [21]. LRP8 plays a role in Wnt3a-induced osteoblastogenesis [51].

## Involvement of the different LRPs in bone homeostasis

The specific role of LRP5 and LRP6 in bone is not entirely clear; however recent studies have begun to tease out the subtle aspects of bone regulation controlled by these coreceptors. Even though both receptors are needed for normal bone development, they have distinct roles as well. In the past year it has been established that Lrp5 and Lrp6 control osteoblast differentiation at different stages. Lrp6 is needed for early stages of differentiation, whereas Lrp5 is required for late stages of differentiation. Riddle et al (2013) studied the deletion of Lrp5 and Lrp6 in osteoblasts [17]. In mice where Lrp5 was deleted specifically in osteoblasts (using an Osteocalcin Cre model) bones appeared normal up to 8 weeks of age (equal number of osteoblasts and osteoclasts). Bone accrual after this age started to present with decreased mineralizing surface per bone surface, decreases in bone formation and an increase in mineralization lag time. The authors concluded that Lrp5 function can be compensated during the period of rapid bone growth but it is required for proper osteoblast function later in postnatal life. Deleting Lrp6 in osteoblast had an impact on the trabecular compartment as at 4 month of age these mice did not reach peak trabecular bone mass. Both Lrp5 and Lrp6 mutants had decreased cortical bone area, cortical thickness and polar moment of inertia by 24 weeks of age. Deletion of both co-receptors led to an early death with mice suffering osteopenic phenotype. Osteoblasts isolated from calvaria differentiated into chondrocytes in *in vitro* osteoblast differentiation studies [17]. Conditional deletion of Lrp5 in osteocytes resulted in smaller and weaker bones with a compromised quality of cortical bone. It also resulted in diminished, but not abolished, new bone formation in response to mechanical loading [56]. Recently Lrp8 has been shown to play a role in Wnt3a-induced osteoblast differentiation (in vitro studies only), but no human related bone diseases have been reported at this point in time [51].

## LRP mutations in bone related diseases

#### LRP and Osteoporosis

Ever since its original description [3, 4] various research laboratories around the world have attempted to identify *LRP5* variants that can predict the incident of osteoporosis [6, 9-11, 13, 14]. Recently, Falcón-Ramirez et al (2013) studied the allelic and genotypic frequencies of four polymorphic markers (single nucleotide polymorphisms (SNPs) (C/T rs3736228, G/A rs4988321, T/C rs627174 and T/C rs901824) in the *LRP5* gene in 100 post-menopausal osteoporotic Mexican women. When genotype frequencies of these four SNPs were studied only the G/A polymorphism (rs4988321, Val667Met) showed differences between the osteoporotic and the control subjects. This led them to conclude that this SNP has a significant association with osteoporosis in this population. An equally interesting finding was that when the authors analyzed the haplotypes of these polymorphisms, there was a clear association between osteoporosis; and the TACT haplotype was significantly associated with higher risk of osteoporosis [57].

In a study by Sassi et al. (2014) the incidence of Ala1330Val (rs3736228) and Val667Met (rs4988321) polymorphisms in Tunisian women with osteoporosis was examined. They found that Ala1330Val could be potentially associated with lower spine density in postmenopausal Tunisian women. They discussed that this was aligned with other researchers finding where they determined association of A1330V with lumbar spine BMD in Asians. However, in Caucasian populations the association was observed in both spine and femoral neck BMD [58]. Park SE et al (2014) found an association with LRP5 rs599083 and femoral neck BMD in Korean men [59]. Campos-Obando N. (2014) reported a single case of a 27year old patient who during her 7-month of pregnancy who obtained a mid-thoracic pain after lifting an object. DNA analysis determined two compound heterozygous missense mutations in the LRP5 gene [60]. This raises the question that although fractures and osteoporosis during pregnancy are not common, should patients with a family history of osteoporosis be genetically screened before conception? Recently Boudin and colleagues reported that mutation in the LRP4 was associated with osteoporosis. They found an association between rs2306029 and rs6485702 and BMD at all sites except in the lumbar spine [50].

#### Lrp5 and Osteoarthritis

Several genome-wide scans for osteoarthritis-susceptibility genes have shown that there is a linkage to chromosome 11q12-13 in the region where the LRP5 gene locus is located. This strongly suggested that LRP5 might be involved in the pathogenesis of osteoarthritis (OA) [61-63]. After that report several researchers have found mutations in LRP5 in patients with OA. In 2005 Smith et al reported that a common haplotype in the Lrp5 gene was associated with a 1.6 fold increase risk of knee OA in two Japanese cohorts [64]. In 2007 Urano et al reported a single polymorphism in the Lrp5 (Q89R) associated with spinal osteoarthritis in Japanese women. Yerges-Armstrong et al (2014) reported that the rs3736228 variant in the LRP5 gene (along with other 3 SNPs in other genes) was associated with increased risk of radiographic knee osteoarthritis in the Osteoarthritis Initiative and the Johnston County Osteoarthritis Project [65]. Lrp6 might be also playing an important role in OA progression. Joiner D et al (2013) presented data suggesting that deletion of one allele of Lrp6 can aggravate the progression of the condition [16]. Whether any of these LRP5/6 polymorphisms are causally related to the development of OA is unclear at this time. Global deletion of LRP5 (where LRP6 is still present) results in an increase in cartilage degradation in the destabilization of the medial meniscus (DMM) model. It is well known that LRP5<sup>-/-</sup> mutants have lower bone mass [66]. So results in this study were unexpected since in humans it has been reported that there is an inverse relationship between osteoarthritis and osteoporosis [67].

#### LRP5 in Rheumatoid Arthritis

Rheumatoid arthritis (RA) is a chronic autoimmune disease in which the etiology is still unknown. Recently the involvement of the canonical Wnt/ $\beta$ -catenin signaling has been described in the pathology of RA (for review see [68]). Diarra and collegues reported that Dkk-1 was involved in the bone-destructive pattern observed in a mouse model of rheumatoid arthritis. Furthermore, they identified tumor necrosis factor- $\alpha$  (TNF $\alpha$ ) as a key regulator of the Wnt pathway. In a search for variants in genes encoding DKK1, Sclerostin,

Kremen-1 and Lrp-5, de Rooy and colleagues screened 600 patients with RA in cohorts from Groningen (NL), Sheffield (UK) and Lund (Sweden) searching for single nucleotide polymorphisms (SNPs). In the Groningen (NL) cohort they found six DKK-1, three SOST, one Kremen-1 and ten LRP-5 SNPs were significantly associated with radiological progression of joint destruction. After performing a meta-analysis, three DKK-1 SNPs (rs1896368, rs1896367 and rs1528873) were associated with progression of joint damage and two Sost SNPs trended to significance (rs4792909 and rs6503475). They determined that RA patients bearing mutations in the DKK-1 gene had higher levels of DKK-1 and this was correlated with progression of the disease [69].

#### LRP5 and the Treatment of Bone Disease

Understanding the molecular aspects of the Wnt/ $\beta$ -catenin signaling pathway, and its regulation, has allowed scientists to develop new pharmaceutical approaches for treatment of low bone mass related diseases. Blockage of the natural inhibitors of the Wnt/ $\beta$ -catenin signaling pathway is receiving considerable attention and pharmaceutical developmental effort as a therapeutic option for the treatment for osteoporosis. Anti-sclerostin antibodies are the most advanced in the drug development pipeline and are being developed to increase bone formation in post-menopausal osteoporotic and osteopenic women [23, 26, 70]. In addition to the development of anti-sclerostin antibodies as a therapeutic, small molecule inhibitors that target sclerostin are being developed to circumvent the potential immunological reactions observed in some patients treated with the anti-sclerostin antibodies [30].

Arguably one of the more novel advancements in the field of pharmacological treatment of bone diseases by targeting the LRP5 signaling pathway in the past year has been attempts in animal models to treat diseases such as Osteoporosis-Pseudoglioma Syndrome (OPPG) and Osteogenesis Imperfecta (OI) with the anti-sclerostin antibody. OPPG is a genetic disease and it is well established that mutation in the LRP5 gene leads to this condition [1, 32, 4, 10, 14, 13, 7-9]. It is characterized by visual loss and severe juvenile osteoporosis. However, two independent groups have proposed new strategies for OPPG treatment [71, 72]. Kedlaya and colleagues (2013) showed that by crossing Lrp5<sup>-/-</sup> mouse (a model for OPPG) with the Sost<sup>-/-</sup> mouse (a model of sclerosteosis) they were able to improve bone density and quality of the bones in the Lrp5<sup>-/-</sup> mice. They also treated Lrp5<sup>-/-</sup> mice with an anti-sclerostin antibody for 3 weeks. Interestingly, the Lrp5<sup>-/-</sup> responded positively to the antibody treatment with increased bone mineral density, content and bone formation rates, which the authors attributed to a redundant level of function associated with the presence of Lrp6 in this animal model. The approach of regulating sclerostin function has further implications as a potential strategy for treating other sclerosing bone diseases.

Other groups have studied the effect of LRP5 manipulation treatment in Osteogenesis Imperfecta (OI)[73, 74]. OI is a genetic disorder in which a defective collagen is produced that ultimately leads to brittle bones. There are several types (degrees) of OI. Current lines of treatment include bisphosphonate, monoclonal RANKL antibody treatment, and rPTH (1-34) (teriparatide) (for review see [75]). Children with moderate to severe phenotype are currently treated with bisphosphonates. Clinicians normally follow this treatment since this

class of drugs inhibits osteoclast activity, increases bone mass, and therefore decreases bone fracture. The unfortunate aspect of bisphosphonate therapy in children is the risk of developing bisphosphonate-related osteonecrosis of the jaw [76, 77]. In addition, inhibiting osteoclast activity only means that the osteoclasts are not removing bone; while osteoblasts are still making defective collagen. In the past year, a new approach for treatment has been tested and it was determined that activation of LRP5 signaling pathway can overcome mild malfunctions of collagen assembly [73, 78, 79]. Sinder and colleagues [78] injected antisclerostin antibodies for two weeks in 8-week old mice that were heterozygous for the col1a1 G349C point mutation (Brtl/+ mice). They reported that this short treatment led to increased osteocalcin levels in serum, increased periosteal bone formation rates, increased mineral apposition rates, and a slight increase in bone mass (measured by microCT). However local micromechanical properties (measured by nanoindentation) were not significantly different among groups, suggesting that sclerostin antibody treatment does not improve the quality of the bone [78]. In 2014, the same group reported that treatment of 6 month old Brtl/+ mice with anti-sclerostin antibody (for 5 weeks) produced an anabolic effect and the bones were stronger [79]. Jacobsen CM. et. al. (2014) also tested the same concept but used a different approach. In their study two mouse models were crossed: an OI mice model (Col1a2<sup>+/p.G610C</sup>) and a HBM mouse model (Lrp5<sup>+/p.A214V</sup>). By doing this cross the bone properties of OI mice were highly improved. Furthermore, when OI mice were treated with anti-sclerostin antibody bone mass and strength were improved significantly [73]. However, the anti-sclerostin antibody treatment didn't have the same effect on another mouse model that had advanced (high severity) disease [74]. The difference might be due to the mouse model used as well as the length, frequency of the treatment and the source of the antibody. Roschger and colleagues used the Colla1<sup>rt/+</sup> mice and they used the anti-sclerostin antibody from Novartis (Sost-ab BPS804) (100mg/kg body weight) once per week for 4 weeks. Whereas Jacobsen et al used the anti-sclerostin antibody from Amgen (Scl-AbIII) (25mg/kg) twice a week per 6 weeks. Snider et al injected the antisclerostin antibody from Amgen (Scl-AbVI) (25mg/kg) two times a week for 5 weeks (in the adult mice studies).

## **Conclusions and Prospectives**

The increasing reports describing mutations of the members of the LRP family and their relation to bone reinforces the important role of these genes/proteins in the pathogenesis of devastating diseases such as osteoporosis, osteoarthritis, rheumatoid arthritis, Osteoporosis Pseudoglioma Syndrome and Osteogenesis Imperfecta. Figure 1 aims to summarize the important role of LRP family members in bone related disease; mutations in these proteins can lead to increased or decreased bone mass as well as the points of possible pharmaceutical interventions for treatment of diseases. The field is advancing considerably but more research is needed to fully understand the LRPs/Wnt/ $\beta$ -catenin pathway in order to develop new therapeutic approaches to successfully improve the lives of patients afflicted with these diseases. Treatment with anti-sclerostin antibody is a promising approach on the near horizon; however, it is important to keep in mind that development of new drugs to control this pathway need to be bone tissue specific as the Wnt/LRPs/ $\beta$ -catenin is present and tightly regulated in nearly all cell types in the body. The anti-sclerostin antibody shows

great promise in this regard owing to the expression of sclerostin mainly in bone, which confers a high degree of bone cell targeting of this particular pharmaceutical agent. The past dozen years since the original description of the LRP5 mutations causal for OPPG and HBM has seen significant advance in our understanding of how bone mass is (mis)regulated and brought us to the doorstep of a new era of bone anabolic agents. Hopefully the next dozen years will bring us full circle to complete treatment for many bone diseases through targeting of the LRP receptor family mediated pathway.

#### Acknowledgments

This work is funded by grants from the NIH NIAMS RO1 AR053949 and RC2 AR058962, NIA P01 AG039355, a grant from the UMKC CEMT through the Missouri Life Science Research Board, and a University of Missouri Research Board Faculty Grant.

#### References

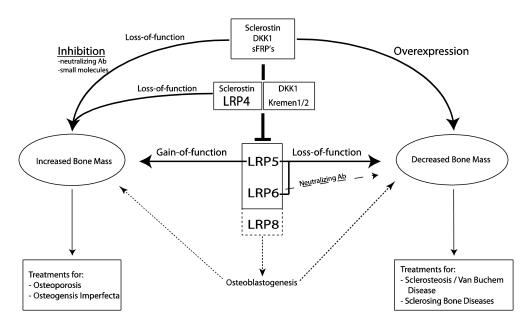
- Gong Y, Vikkula M, Boon L, Liu J, Beighton P, Ramesar R, et al. Osteoporosis-Pseudoglioma Syndrome, a Disorder Affecting Skeletal Strength and Vision, is Assigned to Chromosome Region 11q12-13. American journal of human genetics. 1996; 59:146–51. [PubMed: 8659519]
- Johnson ML, Gong G, Kimberling W, Recker SM, Kimmel DB, Recker RB. Linkage of a gene causing high bone mass to human chromosome 11 (11q12-13). American journal of human genetics. 1997; 60(6):1326–32. doi:S0002-9297(07)64224-4 [pii]. [PubMed: 9199553]
- Gong Y, Slee RB, Fukai N, Rawadi G, Roman-Roman S, Reginato AM, et al. LDL receptor-related protein 5 (LRP5) affects bone accrual and eye development. Cell. 2001; 107(4):513–23. doi:S0092-8674(01)00571-2 [pii]. [PubMed: 11719191]
- 4. Little RD, Carulli JP, Del Mastro RG, Dupuis J, Osborne M, Folz C, et al. A mutation in the LDL receptor-related protein 5 gene results in the autosomal dominant high-bone-mass trait. Am J Hum Genet. 2002; 70(1):11–9.10.1086/338450 [PubMed: 11741193]
- Tamai K, Semenov M, Kato Y, Spokony R, Liu C, Katsuyama Y, et al. LDL-receptor-related proteins in Wnt signal transduction. Nature. 2000; 407(6803):530–5.10.1038/35035117 [PubMed: 11029007]
- 6. Balemans W, Devogelaer JP, Cleiren E, Piters E, Caussin E, Van Hul W. Novel LRP5 Missense Mutation in a Patient With a High Bone Mass Phenotype Results in Decreased DKK1-Mediated Inhibition of Wnt Signaling\*. Journal of Bone and Mineral Research. 2007; 22(5):708–16.10.1359/ jbmr.070211 [PubMed: 17295608]
- Barros E, Dias da Silva M, Kunii I, Hauache O, Lazaretti-Castro M. A novel mutation in the *LRP5* gene is associated with osteoporosis-pseudoglioma syndrome. Osteoporosis International. 2007; 18(7):1017–8.10.1007/s00198-007-0360-x [PubMed: 17437160]
- Marques-Pinheiro A, Levasseur R, Cormier C, Bonneau J, Boileau C, Varret M, et al. Novel LRP5 gene mutation in a patient with osteoporosis-pseudoglioma syndrome. Joint Bone Spine. 2010; 77(2):151–3. [PubMed: 20096619]
- Narumi S, Numakura C, Shiihara T, Seiwa C, Nozaki Y, Yamagata T, et al. Various types of LRP5 mutations in four patients with osteoporosis-pseudoglioma syndrome: Identification of a 7.2-kb microdeletion using oligonucleotide tiling microarray. American Journal of Medical Genetics Part A. 2010; 152A(1):133–40.10.1002/ajmg.a.33177 [PubMed: 20034086]
- Okubo M, Horinishi A, Kim DH, Yamamoto TT, Murase T. Seven Novel Sequence Variants in the Human Low Density Lipoprotein Receptor Related Protein 5 (LRP5) Gene. Human mutation. 2002; 19:186–8. [PubMed: 11793484]
- Saarinen A, Valimaki VV, Valimaki MJ, Loyttyniemi E, Auro K, Uusen P, et al. The A1330V Polymorhpism of the Low-Density Lipoprotein Receptor-Related Protein 5 Gene (LRP5) Associates with Low Peak Bone Mass in Young Healthy Men. Bone. 2007; 40:1006–12. [PubMed: 17223614]

- Streeten EA, McBride DJ, Lodge AL, Pollin TI, Stinchcomb DG, Agarwala R, et al. Reduced incidence of hip fracture in the Old Order Amish. J Bone Miner Res. 2004; 19(2):308–13.10.1359/ JBMR.0301223 [PubMed: 14969401]
- Streeten EA, Puffenberger E, Morton H, McBride D. Osteoporosis Pseudoglioma Syndrome: 4 Siblings with a Compound Heterozygote LRP5 Mutation. J Bone Miner Res. 2004; 19(Suppl 1):S182.
- 14. Van Wesenbeeck L, Cleiren E, Gram J, Beals RK, Benichou O, Scopelliti D, et al. Six novel missense mutations in the LDL receptor-related protein 5 (LRP5) gene in different conditions with an increased bone density. Am J Hum Genet. 2003; 72(3):763–71.10.1086/368277 [PubMed: 12579474]
- Joeng KS, Schumacher CA, Zylstra-Diegel CR, Long F, Williams BO. Lrp5 and Lrp6 redundantly control skeletal development in the mouse embryo. Dev Biol. 2011; 359(2):222–9. doi:10.1016/ j.ydbio.2011.08.020 S0012-1606(11)01220-6 [pii]. [PubMed: 21924256]
- 16. Joiner DM, Less KD, Van Wieren EM, Hess D, Williams BO. Heterozygosity for an inactivating mutation in low-density lipoprotein-related receptor 6 (Lrp6) increases osteoarthritis severity in mice after ligament and meniscus injury. Osteoarthritis and cartilage / OARS, Osteoarthritis Research Society. 2013; 21(10):1576–85.10.1016/j.joca.2013.05.019
- Riddle RC, Diegel CR, Leslie JM, Van Koevering KK, Faugere MC, Clemens TL, et al. Lrp5 and Lrp6 exert overlapping functions in osteoblasts during postnatal bone acquisition. PLoS One. 2013; 8(5):e63323. doi:10.1371/journal.pone.0063323 PONE-D-13-01527 [pii]. [PubMed: 23675479]
- Choi HY, Dieckmann M, Herz J, Niemeier A. Lrp4, a Novel Receptor for Dickkopf 1 and Sclerostin, Is Expressed by Osteoblasts and Regulates Bone Growth and Turnover *In Vivo*. PLoS ONE. 2009; 4(11):e7930. [PubMed: 19936252]
- Kumar J, Swanberg M, McGuigan F, Callreus M, Gerdhem P, Åkesson K. LRP4 association to bone properties and fracture and interaction with genes in the Wnt-and BMP signaling pathways. Bone. 2011; 49(3):343–8. [PubMed: 21645651]
- 20. Li Y, Pawlik B, Elcioglu N, Aglan M, Kayserili H, Yigit G, et al. LRP4 Mutations Alter Wnt/ [beta]-Catenin Signaling and Cause Limb and Kidney Malformations in Cenani-Lenz Syndrome. The American Journal of Human Genetics. 2010; 86(5):696–706. [PubMed: 20381006]
- 21. Simon-Chazottes D, Tutois S, Kuehn M, Evans M, Bourgade F, Cook S, et al. Mutations in the gene encoding the low-density lipoprotein receptor LRP4 cause abnormal limb development in the mouse. Genomics. 2006; 87(5):673–7. [PubMed: 16517118]
- 22. Experimental drug may change the treatment of osteoporosis. Romosozumab appears to increase bone mineral density and help rebuild the skeleton at the same time. DukeMedicine healthnews. 2014; 20(4):7.
- Costa AG, Bilezikian JP, Lewiecki EM. Update on romosozumab : a humanized monoclonal antibody to sclerostin. Expert opinion on biological therapy. 2014; 14(5):697– 707.10.1517/14712598.2014.895808 [PubMed: 24665957]
- 24. Evenepoel P, D'Haese P, Brandenburg V. Romosozumab in postmenopausal women with osteopenia. The New England journal of medicine. 2014; 370(17):1664.10.1056/ NEJMc1402396#SA1 [PubMed: 24758631]
- McClung MR, Grauer A. Romosozumab in postmenopausal women with osteopenia. The New England journal of medicine. 2014; 370(17):1664–5.10.1056/NEJMc1402396 [PubMed: 24758630]
- 26. McClung MR, Grauer A, Boonen S, Bolognese MA, Brown JP, Diez-Perez A, et al. Romosozumab in postmenopausal women with low bone mineral density. The New England journal of medicine. 2014; 370(5):412–20.10.1056/NEJMoa1305224 [PubMed: 24382002]
- McColm J, Hu L, Womack T, Tang CC, Chiang AY. Single- and multiple-dose randomized studies of blosozumab, a monoclonal antibody against sclerostin, in healthy postmenopausal women. J Bone Miner Res. 2014; 29(4):935–43.10.1002/jbmr.2092 [PubMed: 23996473]
- Recker R, Benson C, Matsumoto T, Bolognese M, Robins D, Alam J, et al. A randomized, doubleblind phase 2 clinical trial of blosozumab, a sclerostin antibody, in postmenopausal women with low bone mineral density. J Bone Miner Res. 201410.1002/jbmr.2351

- Montagnani A. Bone anabolics in osteoporosis: Actuality and perspectives. World journal of orthopedics. 2014; 5(3):247–54.10.5312/wjo.v5.i3.247 [PubMed: 25035827]
- Rey JP, Ellies DL. Wnt modulators in the biotech pipeline. Developmental dynamics : an official publication of the American Association of Anatomists. 2010; 239(1):102–14.10.1002/dvdy.22181 [PubMed: 20014100]
- 31. Brown SD, Twells RC, Hey PJ, Cox RD, Levy ER, Soderman AR, et al. Isolation and Characterization of LRP6, a Novel Member of the Low Density Lipoprotein Receptor Gene Family. Biochem Biophys Res Commun. 1998; 248:879–88. [PubMed: 9704021]
- 32. Li Y, Cam J, Bu G. Low-density lipoprotein receptor family: endocytosis and signal transduction. Molecular neurobiology. 2001; 23(1):53–67.10.1385/MN:23:1:53 [PubMed: 11642543]
- Zeng X, Tamai K, Doble B, Li S, Huang H, Habas R, et al. A dual-kinase mechanism for Wnt coreceptor phosphorylation and activation. Nature. 2005; 438(7069):873–7.10.1038/nature04185 [PubMed: 16341017]
- 34. Tamai K, Zeng X, Liu C, Zhang X, Harada Y, Chang Z, et al. A mechanism for Wnt coreceptor activation. Molecular cell. 2004; 13(1):149–56. [PubMed: 14731402]
- Aberle H, Bauer A, Stappert J, Kispert A, Kemler R. beta-catenin is a target for the ubiquitinproteasome pathway. The EMBO journal. 1997; 16(13):3797–804.10.1093/emboj/16.13.3797 [PubMed: 9233789]
- 36. Yanagawa S, Matsuda Y, Lee JS, Matsubayashi H, Sese S, Kadowaki T, et al. Casein kinase I phosphorylates the Armadillo protein and induces its degradation in Drosophila. The EMBO journal. 2002; 21(7):1733–42.10.1093/emboj/21.7.1733 [PubMed: 11927557]
- 37. Nusse R. WNT targets. Repression and activation. Trends in genetics : TIG. 1999; 15(1):1–3. [PubMed: 10087922]
- Li X, Zhang Y, Kang H, Liu W, Liu P, Zhang J, et al. Sclerostin binds to LRP5/6 and antagonizes canonical Wnt signaling. The Journal of biological chemistry. 2005; 280(20):19883–7.10.1074/ jbc.M413274200 [PubMed: 15778503]
- 39. Fedi P, Bafico A, Nieto Soria A, Burgess WH, Miki T, Bottaro DP, et al. Isolation and biochemical characterization of the human Dkk-1 homologue, a novel inhibitor of mammalian Wnt signaling. The Journal of biological chemistry. 1999; 274(27):19465–72. [PubMed: 10383463]
- 40. Bao J, Zheng JJ, Wu D. The structural basis of DKK-mediated inhibition of Wnt/LRP signaling. Science signaling. 2012; 5(224):pe22.10.1126/scisignal.2003028 [PubMed: 22589387]
- 41. Mao B, Wu W, Davidson G, Marhold J, Li M, M, Mechler B, et al. Kremen Proteins are Dickkopf Receptors that Regulate Wnt/β-Catenin Signalling. Nature. 2002; 417:664–7. [PubMed: 12050670]
- Nakamura T, Aoki S, Kitajima K, Takahashi T, Matsumoto K, Nakamura T. Molecular cloning and characterization of Kremen, a novel kringle-containing transmembrane protein. Biochimica et biophysica acta. 2001; 1518(1-2):63–72. [PubMed: 11267660]
- 43. Cselenyi CS, Lee E. Context-dependent activation or inhibition of Wnt-beta-catenin signaling by Kremen. Science signaling. 2008; 1(8):pe10.10.1126/stke.18pe10 [PubMed: 18314504]
- 44. Galli LM, Barnes T, Cheng T, Acosta L, Anglade A, Willert K, et al. Differential inhibition of Wnt-3a by Sfrp-1, Sfrp-2, and Sfrp-3. Developmental dynamics : an official publication of the American Association of Anatomists. 2006; 235(3):681–90.10.1002/dvdy.20681 [PubMed: 16425220]
- Surana R, Sikka S, Cai W, Shin EM, Warrier SR, Tan HJ, et al. Secreted frizzled related proteins: Implications in cancers. Biochimica et biophysica acta. 2014; 1845(1):53–65.10.1016/j.bbcan. 2013.11.004 [PubMed: 24316024]
- Guidato S, Itasaki N. Wise retained in the endoplasmic reticulum inhibits Wnt signaling by reducing cell surface LRP6. Dev Biol. 2007; 310(2):250–63.10.1016/j.ydbio.2007.07.033 [PubMed: 17765217]
- Lintern KB, Guidato S, Rowe A, Saldanha JW, Itasaki N. Characterization of wise protein and its molecular mechanism to interact with both Wnt and BMP signals. The Journal of biological chemistry. 2009; 284(34):23159–68.10.1074/jbc.M109.025478 [PubMed: 19553665]

- Ellies DL, Economou A, Viviano B, Rey JP, Paine-Saunders S, Krumlauf R, et al. Wise regulates bone deposition through genetic interactions with Lrp5. PLoS One. 2014; 9(5):e96257.10.1371/ journal.pone.0096257 [PubMed: 24789067]
- Balemans W, Piters E, Cleiren E, Ai M, Van Wesenbeeck L, Warman ML, et al. The binding between sclerostin and LRP5 is altered by DKK1 and by high-bone mass LRP5 mutations. Calcified tissue international. 2008; 82(6):445–53.10.1007/s00223-008-9130-9 [PubMed: 18521528]
- Boudin E, Steenackers E, de Freitas F, Nielsen TL, Andersen M, Brixen K, et al. A common LRP4 haplotype is associated with bone mineral density and hip geometry in men-data from the Odense Androgen Study (OAS). Bone. 2013; 53(2):414–20.10.1016/j.bone.2013.01.014 [PubMed: 23321396]
- Zhang J, Zhang X, Zhang L, Zhou F, van Dinther M, Ten Dijke P. LRP8 mediates Wnt/betacatenin signaling and controls osteoblast differentiation. J Bone Miner Res. 2012; 27(10):2065– 74.10.1002/jbmr.1661 [PubMed: 22589174]
- 52. Kato M, Patel MS, Levasseur R, Lobov I, Chang BHJ, Glass DA, et al. Cbfa 1-Independent Decrease in Osteoblast Proliferation, Osteopenia, and Persistent Embryonic Eye Vascularization in Mice Deficient in Lrp5, a Wnt Coreceptor. The Journal of cell biology. 2002; 157:303–14. [PubMed: 11956231]
- Babij P, Zhao W, Small C, Kharode Y, Yaworsky PJ, Bouxsein ML, et al. High bone mass in mice expressing a mutant LRP5 gene. J Bone Miner Res. 2003; 18(6):960–74.10.1359/jbmr. 2003.18.6.960 [PubMed: 12817748]
- 54. Niziolek PJ, Warman ML, Robling AG. Mechanotransduction in bone tissue: The A214V and G171V mutations in Lrp5 enhance load-induced osteogenesis in a surface-selective manner. Bone. 2012; 51(3):459–65.10.1016/j.bone.2012.05.023 [PubMed: 22750014]
- 55. Javaheri B, Sunters A, Zaman G, Suswillo RF, Saxon LK, Lanyon LE, et al. Lrp5 is not required for the proliferative response of osteoblasts to strain but regulates proliferation and apoptosis in a cell autonomous manner. PLoS One. 2012; 7(5):e35726.10.1371/journal.pone.0035726 [PubMed: 22567110]
- 56. Zhao L, Shim JW, Dodge TR, Robling AG, Yokota H. Inactivation of Lrp5 in osteocytes reduces young's modulus and responsiveness to the mechanical loading. Bone. 2013; 54(1):35–43. doi: 10.1016/j.bone.2013.01.033 S8756-3282(13)00050-1 [pii]. [PubMed: 23356985]
- Falcon-Ramirez E, Casas-Avila L, Cerda-Flores RM, Castro-Hernandez C, Rubio-Lightbourn J, Velazquez-Cruz R, et al. Association of LRP5 haplotypes with osteoporosis in Mexican women. Mol Biol Rep. 2013; 40(3):2705–10.10.1007/s11033-012-2357-6 [PubMed: 23242660]
- 58. Sassi R, Sahli H, Souissi C, El Mahmoudi H, Zouari B, Ben Ammar ElGaaied A, et al. Association of LRP5 genotypes with osteoporosis in Tunisian post-menopausal women. BMC Musculoskelet Disord. 2014; 15:144. doi:10.1186/1471-2474-15-144 1471-2474-15-144 [pii]. [PubMed: 24885293]
- 59. Park SE, Oh KW, Lee WY, Baek KH, Yoon KH, Son HY, et al. Association of osteoporosis susceptibility genes with bone mineral density and bone metabolism related markers in Koreans: The Chungju Metabolic Disease Cohort (CMC) study. Endocrine journal. 2014
- Campos-Obando N, Oei L, Hoefsloot LH, Kiewiet RM, Klaver CC, Simon ME, et al. Osteoporotic vertebral fractures during pregnancy: be aware of a potential underlying genetic cause. The Journal of clinical endocrinology and metabolism. 2014; 99(4):1107–11.10.1210/jc.2013-3238 [PubMed: 24423337]
- 61. Chapman K, Mustafa Z, Dowling B, Southam L, Carr A, Loughlin J. Finer linkage mapping of primary hip osteoarthritis susceptibility on chromosome 11q in a cohort of affected female sibling pairs. Arthritis and rheumatism. 2002; 46(7):1780–3.10.1002/art.10412 [PubMed: 12124861]
- Chapman K, Mustafa Z, Irven C, Carr AJ, Clipsham K, Smith A, et al. Osteoarthritis-susceptibility locus on chromosome 11q, detected by linkage. Am J Hum Genet. 1999; 65(1):167– 74.10.1086/302465 [PubMed: 10364529]
- 63. Shi XW, Guo X, Lv AL, Kang L, Zhou YL, Zhang YZ, et al. Heritability estimates and linkage analysis of 23 short tandem repeat loci on chromosomes 2, 11, and 12 in an endemic osteochondropathy in China. Scandinavian journal of rheumatology. 2010; 39(3):259– 65.10.3109/03009740903270599 [PubMed: 20166850]

- 64. Smith AJ, Gidley J, Sandy JR, Perry MJ, Elson CJ, Kirwan JR, et al. Haplotypes of the low-density lipoprotein receptor-related protein 5 (LRP5) gene: are they a risk factor in osteoarthritis? Osteoarthritis and cartilage / OARS, Osteoarthritis Research Society. 2005; 13(7):608–13.10.1016/ j.joca.2005.01.008
- Yerges-Armstrong LM, Yau MS, Liu Y, Krishnan S, Renner JB, Eaton CB, et al. Association analysis of BMD-associated SNPs with knee osteoarthritis. J Bone Miner Res. 2014; 29(6):1373– 9.10.1002/jbmr.2160 [PubMed: 24339167]
- 66. Lodewyckx L, Luyten FP, Lories RJ. Genetic deletion of low-density lipoprotein receptor-related protein 5 increases cartilage degradation in instability-induced osteoarthritis. Rheumatology (Oxford). 2012; 51(11):1973–8. doi:10.1093/rheumatology/kes 178kes178[pii]. [PubMed: 22850184]
- Dequeker J, Aerssens J, Luyten FP. Osteoarthritis and osteoporosis: clinical and research evidence of inverse relationship. Aging clinical and experimental research. 2003; 15(5):426–39. [PubMed: 14703009]
- Miao CG, Yang YY, He X, Li XF, Huang C, Huang Y, et al. Wnt signaling pathway in rheumatoid arthritis, with special emphasis on the different roles in synovial inflammation and bone remodeling. Cellular signalling. 2013; 25(10):2069–78.10.1016/j.cellsig.2013.04.002 [PubMed: 23602936]
- 69. de Rooy DP, Yeremenko NG, Wilson AG, Knevel R, Lindqvist E, Saxne T, et al. Genetic studies on components of the Wnt signalling pathway and the severity of joint destruction in rheumatoid arthritis. Annals of the rheumatic diseases. 2013; 72(5):769–75.10.1136/ annrheumdis-2012-202184 [PubMed: 23041840]
- Padhi D, Allison M, Kivitz AJ, Gutierrez MJ, Stouch B, Wang C, et al. Multiple doses of sclerostin antibody romosozumab in healthy men and postmenopausal women with low bone mass: A randomized, double-blind, placebo-controlled study. Journal of clinical pharmacology. 201310.1002/jcph.239
- Chang MK, Kramer I, Keller H, Gooi JH, Collett C, Jenkins D, et al. Reversing LRP5-dependent osteoporosis and SOST deficiency-induced sclerosing bone disorders by altering WNT signaling activity. J Bone Miner Res. 2014; 29(1):29–42.10.1002/jbmr.2059 [PubMed: 23901037]
- 72. Kedlaya R, Veera S, Horan DJ, Moss RE, Ayturk UM, Jacobsen CM, et al. Sclerostin inhibition reverses skeletal fragility in an Lrp5-deficient mouse model of OPPG syndrome. Science translational medicine. 2013; 5(211):211ra158.10.1126/scitranslmed.3006627
- 73. Jacobsen CM, Barber LA, Ayturk UM, Roberts HJ, Deal LE, Schwartz MA, et al. Targeting the LRP5 Pathway Improves Bone Properties in a Mouse Model of Osteogenesis Imperfecta. J Bone Miner Res. 2014; 29(10):2297–306.10.1002/jbmr.2198 [PubMed: 24677211]
- Roschger A, Roschger P, Keplingter P, Klaushofer K, Abdullah S, Kneissel M, et al. Effect of sclerostin antibody treatment in a mouse model of severe osteogenesis imperfecta. Bone. 2014; 66:182–8.10.1016/j.bone.2014.06.015 [PubMed: 24953712]
- Lindahl K, Langdahl B, Ljunggren O, Kindmark A. Treatment of osteogenesis imperfecta in adults. European journal of endocrinology / European Federation of Endocrine Societies. 2014; 171(2):R79–90.10.1530/EJE-14-0017 [PubMed: 24760541]
- 76. Ngan KK, Bowe J, Goodger N. The risk of bisphosphonate-related osteonecrosis of the jaw in children. A case report and literature review. Dental update. 2013; 40(9):733–4. 6–8. [PubMed: 24386765]
- 77. Christou J, Johnson AR, Hodgson TA. Bisphosphonate-related osteonecrosis of the jaws and its relevance to children- -a review. International journal of paediatric dentistry / the British Paedodontic Society [and] the International Association of Dentistry for Children. 2013; 23(5): 330–7.10.1111/ipd.12047
- 78. Sinder BP, Eddy MM, Ominsky MS, Caird MS, Marini JC, Kozloff KM. Sclerostin antibody improves skeletal parameters in a Brtl/+ mouse model of osteogenesis imperfecta. J Bone Miner Res. 2013; 28(1):73–80.10.1002/jbmr.1717 [PubMed: 22836659]
- 79. Sinder BP, White LE, Salemi JD, Ominsky MS, Caird MS, Marini JC, et al. Adult Brtl/+ mouse model of osteogenesis imperfecta demonstrates anabolic response to sclerostin antibody treatment with increased bone mass and strength. Osteoporos Int. 2014; 25(8):2097–107.10.1007/ s00198-014-2737-y [PubMed: 24803333]



#### Figure 1.

The roles of various key members of the LRP receptor family; LRP4, LRP5, LRP6 and LRP8 in bone related traits and bone mass regulation. Gain-of-function mutations in LRP5 give rise to high bone mass phenotypes in humans and mice, whereas loss-of-function mutations in LRP5 and LRP6 give rise to decreased bone mass. Knockout or loss of function mutations of the inhibitors of the Wnt/ $\beta$ -catenin signaling pathway such as sclerostin, DKK1 and the sFRPs give rise to increased bone mass and overexpression of these proteins results in decreased bone mass. Sclerostin inhibition of LRP5/6 appears to be LRP4 dependent. Loss-of-function mutations in LRP4 lead to increased bone mass. Likewise, Dkk1 inhibitory function is mediated through Kremen1/2 and loss of either of these binding partners results in increased bone mass. At present it is unclear whether LRP8 is inhibited by sclerostin or Dkk1. Recent studies have shown that neutralizing antibodies to LRP6 can reduce bone mass. Treatment with neutralizing antibodies to sclerostin or Dkk1 as well as small molecule inhibitors results in increased bone mass. LRP5/6 and LRP8 appear to have roles in regulating osteoblastogenesis and therefore have the potential through this axis to either increase or decrease bone mass. Diseases such as Osteoporosis and Osteogenesis Imperfecta are envisioned as candidates diseases for treatment with agents that lead to increased bone mass, whereas diseases such as Sclerosteosis/Van Buchem's and potentially other sclerosing bone diseases are candidates for approaches that lead to decreased bone mass. See text for references