



HHS Public Access

Author manuscript

Cytokine. Author manuscript; available in PMC 2016 October 01.

Published in final edited form as:

Cytokine. 2015 October ; 75(2): 280–290. doi:10.1016/j.cyto.2015.05.005.

IL-1 family members in the pathogenesis and treatment of metabolic disease: Focus on adipose tissue inflammation and insulin resistance

Dov B Ballak^{1,3,4}, Rinke Stienstra^{1,2}, Cees J Tack¹, Charles A Dinarello^{1,4}, and Janna A van Diepen¹

¹Department of Medicine, Radboud University Medical Center, Nijmegen, The Netherlands

²Department of Human Nutrition, Wageningen University, Wageningen, The Netherlands

³Department of Integrative Physiology, University of Colorado Boulder, Boulder, CO, USA

⁴Department of Medicine, University of Colorado Denver, Aurora, CO, USA

Abstract

Obesity is characterized by a chronic, low-grade inflammation that contributes to the development of insulin resistance and type 2 diabetes. Cytokines and chemokines produced by immunocompetent cells influence local as well as systemic inflammation and are therefore critical contributors to the pathogenesis of type 2 diabetes. Hence, cytokines that modulate inflammatory responses are emerging as potential targets for intervention and treatment of the metabolic consequences of obesity. The interleukin-1 (IL-1) family of cytokines and receptors are key mediators of innate inflammatory responses and exhibit both pro- and anti-inflammatory functions. During the last decades, mechanistic insights into how the IL-1 family affects the initiation and progression of obesity-induced insulin resistance have increased significantly. Here, we review the current knowledge and understanding, with emphasis on the therapeutic potential of individual members of the IL-1 family of cytokines for improving insulin sensitivity in patients with diabetes.

Keywords

Interleukin-1 family; obesity; inflammation; insulin resistance; adipose tissue

Historical Context

Worldwide the incidence of obesity has increased dramatically. In 2013, the global proportion of overweight (defined as a Body Mass Index (BMI) above 25 kg/m²) or obese (BMI above 30 kg/m²) adults was estimated at 37.5% [1] Obesity is one of the main

Corresponding author: Dov Ballak, Radboud University Medical Center, Geert Grooteplein 8, 6525 GA, Nijmegen, The Netherlands. Tel 0031-243615395. duby.ballak@radboudumc.nl.

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

contributors for the development of insulin resistance, and strongly increases the risk for type 2 diabetes. Inflammatory mediators link obesity to insulin resistance [2]. Obesity itself results in a pro-inflammatory state in metabolic tissues, as illustrated by the upregulation of pro-inflammatory cytokines, such as Interleukin (IL)-1 β and TNF α , which can directly interfere with insulin signaling in adipocytes, hepatocytes, fibroblasts and myocytes [3-10].

The first observation revealing upregulation of TNF α in obese adipose tissue [11] that contributed to the development of insulin resistance [6], evoked interest in the role of inflammation during obesity. In 2003, it was reported that macrophages infiltrate adipose tissue in obesity [12, 13]. These cells produce pro-inflammatory cytokines, such as IL-1 β , TNF α , IL-6 and MCP-1 that contribute to the pathogenesis of obesity-induced insulin resistance. Since that publication, the number of studies investigating specific inflammatory processes in adipose tissue during the development of obesity and related diseases has increased dramatically. Reports reveal that both adaptive and innate immune responses contribute to the onset and progression of obesity-induced adipose tissue inflammation [14, 15].

Importantly, cytokines of the IL-1 family are critical regulators of innate immune responses. This family of cytokines comprises both pro- and anti-inflammatory members. IL-1 α and IL-1 β are classic pro-inflammatory cytokines; both are antagonized by the naturally occurring IL-1 receptor antagonist (IL-1Ra), also a member of the IL-1 family. IL-18, another member of the IL-1 cytokine family, is generally considered a pro-inflammatory cytokine [16]. At the beginning of this millennium, 7 other members of the IL-1 family have been identified. Of these novel members, IL-33, IL-36 α , IL-36 β and IL-36 γ are generally described as pro-inflammatory cytokines that contribute to inflammatory responses [17-19]. In contrast, IL-37 is an anti-inflammatory cytokine [20], similar to IL-36 Receptor antagonist (IL-36Ra) that specifically blocks IL-36 signaling [19]. The role of IL-38, also ascribed with receptor antagonizing properties, appears to have anti-inflammatory properties largely unknown [21, 22].

Growing interest in the role of IL-1 signaling was present after the observation that circulating IL-18 and IL-1Ra levels were increased in obese subjects and patients with type 2 diabetes, respectively [23, 24], and that adipose tissue is a major source of circulating IL-1Ra levels [25] (Figure 1). Early studies from the 1980s had already demonstrated that IL-1 β selectively damages the insulin-producing beta cells in the pancreatic islets of Langerhans [26, 27], suppresses food intake [28] and activates energy metabolism [29]. In 2002 the importance of the role of IL-1 β was highlighted by the observation that hyperglycemia, characteristic for type 2 diabetes, increased IL-1 β production from the β -cells themselves leading to impaired β -cell function and reduced insulin production [30].

In 2006, a causal relation between an IL-1 family member and insulin sensitivity was established in mice, as deficiency of IL-18 in mice appeared to induce hyperphagia, obesity and insulin resistance [31]. Since then, several studies have focused on deciphering the contribution of IL-1 family members to the progression of obesity-induced inflammation and the development of insulin resistance. So far, nearly all cytokines of the IL-1 family have been linked to obesity. Importantly, and in contrast to the pro-inflammatory properties

of the family, also insulin-sensitizing effects of anti-inflammatory family members have been described in obesity. This review addresses the emerging role of the IL-1 family members in metabolic inflammation and insulin resistance, with a specific focus on adipose tissue. In addition, the current and future therapeutic applications of cytokine blocking strategies as well as the use of anti-inflammatory cytokines from the IL-1 family to enhance insulin sensitivity will be presented.

IL-1 α , IL-1 β and IL-1 Receptor antagonist

IL-1 is a master regulator of innate immune responses by controlling numerous inflammatory processes [32]. 30 Years ago, two distinct forms of IL-1 were cloned [33, 34] and termed IL-1 α and IL-1 β [35]. Both IL-1 α and IL-1 β interact with the IL-1 receptor 1 (IL-1R1) and recruit the IL-1 receptor accessory protein (IL-1R3, formerly IL-1RAcP) to induce a downstream signal via several inflammatory kinases, such as Myd88, ERK, JNK and NF κ B, leading to transcription of inflammatory, catabolic genes [36-38]. Importantly, these signaling molecules overlap in that the IL-1 Receptor 1 (IL-1R1) and Tolllike receptors share the same signaling domain termed Toll-IL-Receptor (TIR) [39]. IL-1Ra on the other hand has anti-inflammatory effects by binding to the IL-1R1 and specifically inhibiting IL-1 signaling [40]. Endogenous IL-1Ra is increased during inflammation to oppose IL-1 signaling. Together, IL-1 α , IL-1 β and IL-1Ra are the most studied members of the IL-1 family and their respective biological activities are pivotal for inflammatory mechanisms in disease [41]. Indeed, IL-1 activity is linked to several auto-inflammatory diseases, including atherosclerosis and gout [42, 43].

There are considerable differences in the localization, regulation and function of IL-1 α and IL-1 β . First, whereas the IL-1 β precursor is produced by innate immune cells (i.e. monocytes, macrophages and dendritic cells), the IL-1 α precursor is constitutively expressed in resting cells in health [36]. Secondly, intracellular processing of the IL-1 β precursor is tightly controlled by inflammatory stimuli such as LPS, IL-1 α and IL-1 β [44]. This results in IL-1 β precursor activation by caspase-1, which, in turn, is activated by inflammasomes [45]. In contrast, extracellular activation of the IL-1 β precursor by other serine proteases (*i.e.* proteinase-3, elastase and cathepsin G) [46] is less tightly controlled and more dependent on presence of neutrophils. The precursor of IL-1 α on the other hand is present as a biologically active form. The IL-1 α precursor can be cleaved by calpain, and the biological function of the processed protein appears to be more active than the precursor. The IL-1 α precursor is released from necrotic cells and as such, may act as an alarmin, a signal released from dying cells that initiates an inflammatory response via IL-1R1 [36]. Moreover, IL-1 α is a dual-function cytokine, since the cytokine translocates to the nucleus as a pro-inflammatory transcription factor for genes, such as IL-1, IL-6 and IL-8 [32].

IL-1 α , IL-1 β and IL-1Ra in obesity and insulin resistance in humans

Evidence reveals that IL-1 activity is of importance in the pathology of type 2 diabetes by mediating obesity-induced inflammation and directly aggravating insulin resistance. In addition, IL-1 β contributes to the development of type 2 diabetes by inhibiting β -cell function and destroying β -cell mass, as reviewed elsewhere [47-50].

30 Years ago, IL-1 was reported to act as an anorexia-inducing cytokine during host responses to infection [28]. Indeed, known as the fever-inducing cytokine, IL-1 was shown to inhibit food intake [28] and stimulated resting energy expenditure during febrile conditions [29]. Thus, besides induction of inflammation and fever, IL-1 also affected metabolic pathways during infections. Only two decades later, the first relations between IL-1 and obesity were revealed. In obesity, peripheral blood mononuclear cells (PBMCs) showed increased IL-1 α production *ex vivo* [51] and elevated plasma levels of IL-1 α and IL-1 β were found [52, 53]. Notably, circulating levels of IL-1 α , IL-1 β , and antiinflammatory IL-1Ra were all positively correlated with obesity [24, 53].

Combined with the observation that adipose tissue appeared to be a major source of IL-1Ra production [25], the data prompted a considerable interest into the role of IL-1 in the development of obesity-induced inflammation and insulin resistance. More detailed analysis of adipose tissue revealed that human visceral adipose tissue (VAT), considered to be a major contributor to the increased levels of circulating inflammatory cytokines as well as the development of insulin resistance during obesity, contains higher levels of IL-1 α , IL-1 β and IL-1Ra as compared to subcutaneous adipose tissue (SAT) [54, 55]. Moreover, the inflammasome components NLRP3 and ASC, as well as caspase-1 were more active and more abundant in VAT compared to SAT, resulting in enhanced IL-1 β processing and release [54]. Notably, whereas the cellular origin of IL-1 β and IL-1Ra is found within the stromal vascular fraction (SVF) of the adipose tissue [56], the upstream IL-1 β activator caspase-1 is especially expressed in the adipocyte fraction [57]. This finding strengthens the hypothesis that cross-talk between products of resident macrophages and products of adipocytes affect each other and therefore may determine the net inflammatory trait of the adipose tissue and its output function [58]

In adipose tissue (VAT & SAT) of obese as compared to lean individuals, mRNA and protein levels of IL-1 β and IL-1Ra are increased [59, 60]. Moreover, weight loss markedly reduces mRNA expression of IL-1 β in adipose tissue [55, 61], suggesting that bodyweight or fat mass is a significant determinant for secretion of IL-1 β by adipose tissue. In addition, augmented activity of the NLRP3 inflammasome and IL-1 β in adipose tissue seems to distinguish metabolically unhealthy obese individuals (*i.e.* having several components of the metabolic syndrome), from metabolically healthy obese individuals [59].

Contributions of IL-1 β and IL-1Ra to the development of obesity and insulin resistance

Noticeably, until the beginning of the current millennium, the existence of causality between IL-1 β and development of type 2 diabetes in humans was unknown. A prospective study of 27,500 individuals addressed this issue. The investigators found that increased plasma IL-1 β as well as IL-6 levels augmented the risk to develop type 2 diabetes within a period of 2.3 years [62].

During the last decade, animal studies have further elucidated the causal role of IL-1 in suppressive metabolic health. In diet- or genetically induced obese mice, caspase-1 activity and protein levels of IL-1 β are increased in adipose tissue [57]. Moreover, mice lacking components of the inflammasome, IL-1 β or its receptor IL-1R1 are protected from the development of high-fat diet-induced inflammation and insulin resistance [63-65],

emphasizing the pathogenic role of inflammasome-mediated IL-1 β activity in the development of metabolic disease. Further support comes from studies revealing that mice deficient for neutrophil elastase, another regulator of IL-1 β activity, show reduced adipose tissue inflammation and improvement in insulin sensitivity [66, 67]. Moreover, treatment of obese mice with IL-1Ra and inhibitors of caspase-1 or neutrophil elastase led to an improvement in insulin sensitivity levels [57, 67, 68]. Thus, various mouse models characterized by reduced endogenous IL-1 β activity have shown protection from the detrimental effects of obesity on adipose tissue inflammation and insulin resistance. It would be worthwhile to establish whether inhibition of other known regulators of IL-1 β activity, such as PR3 and cathepsin G, have similar insulin-sensitizing effects.

In vitro studies have also revealed the pathogenic role of IL-1 β in the development of insulin resistance, and the possibilities that IL-1 β (signaling) inhibition to enhance insulin sensitivity. Indeed, IL-1 β treatment of adipocytes, disturbs insulin signaling via downregulation of insulin receptor substrate-1 expression, leading to a marked reduction of insulin-mediated GLUT-4 translocation [5, 69]. Consistent with this observation, macrophage-derived conditioned medium induces insulin resistance in primary human adipocytes, whereas inhibition of IL-1 β signaling using a neutralizing antibody or IL-1Ra reverses this effect [10].

Contributions of IL-1 α to the development of obesity and insulin resistance

Although there is ample evidence for the role of IL-1 β as a mediator for metabolic inflammation and insulin resistance during obesity, in contrast, the contribution of IL-1 α has been less studied. Plasma IL-1 α levels were found to be higher in obese mice, yet lower mRNA levels were measured in adipose tissue of obese compared to lean animals [70]. *In vitro*, IL-1 α inhibits adipocyte differentiation [70] and reduces insulin signaling in murine adipocytes [71, 72]. *In vivo*, injection of IL-1 α in mice increases plasma TG levels [70], but whether IL-1 α affects glucose metabolism remains to be determined. Nevertheless, a recent study demonstrated that endogenous oils derived from human adipose tissue activate and recruit neutrophils and macrophages via an IL-1 α dependent mechanism [73]. These data suggest that IL-1 α may recruit innate immune cells to adipose tissue in response to 'danger signals' released by necrotic adipocytes. Since (hypertrophic) adipocyte death is increased in adipose tissue during obesity [74], IL-1 α may control the initiation of adipose tissue inflammation during obesity.

IL-1 Receptor in the development of obesity and insulin resistance

The concept of causal relationship between increased obesity and increase IL-1 β is confounded by the observation that IL-1R1 knockout mice, characterized by absence of IL-1 signaling, paradoxically develop maturity-onset obesity and insulin resistance [75]. Furthermore IL-1Ra knockout mice, characterized by enhanced IL-1 signaling, are resistant to obesity [76]. However, these phenotypes are likely explained by the actions of IL-1 on food consumption and energy expenditure [77]. Suppression of appetite by leptin is dependent on IL-1 and can be reversed by injection of IL-1Ra [78]. This mechanism indicates that basal signaling via the IL-1 receptor is necessary to control central food intake in the brain, and moreover, that a delicate balance between IL-1 and IL-1Ra probably

determines energy balance and glucose homeostasis. The completely disrupted balance in knockout mice could possibly explain the paradoxical phenotypes of increased obesity in the absence of IL-1. Therefore, more subtle modulation of the ratio between IL-1 and IL-1Ra using specific inhibitors may have more relevance for total body physiology and pathology of obesity-induced metabolic disease, as no drug, no antibody and no inhibitor completely prevents IL-1 activity.

Given the large amount of evidence supporting a causal role of IL-1 β in the development of adipose tissue inflammation and insulin resistance, there is need to understand the role of IL-1 α in obesity, since neutralizing antibodies to IL-1 α have been used to reverse the negative nitrogen balance and loss of lean body mass in cancer cachexia as a consequence of metastatic sterile inflammation [79, 80]. Hypothetically, using IL-1 α blockade to reverse low-grade inflammation, could restore homeostasis and insulin sensitivity in metabolic inflammation. The governing effects of IL-1 α blockade on lean body mass are very interesting with respect to the importance of muscles in metabolic health. Therefore, future studies should study the effects of inhibiting IL-1 α in metabolic disease. Tissue distribution, cell type and mechanism for release of both cytokines suggest a different mode of action in low-grade sterile inflammation [81]. Within the context of obesity, IL-1 α may be secreted in response to adipocyte cell death or hypoxia, thereby initiating an inflammatory response. Subsequently, the resulting inflammatory signals may induce IL-1 β release and activation that further instigates and maintains the inflammatory trait, and inhibit peripheral insulin signaling. IL-1Ra on the other hand, is secreted in parallel to IL-1 β and/or IL-1 α and therefore related to obesity and insulin resistance. The increased levels of IL-1Ra thus serve as a circulating marker for endogenous IL-1 β or IL-1 α activity, especially since circulating levels of IL-1Ra are higher and thus easier to measure as compared to IL-1 β /IL-1 α . However, in contrast to IL-1 β or IL-1 α , whether increased IL-1Ra is only a marker for IL-1 activity or intended to reduce IL-1 signaling and thereby dampen the inflammatory response, remains unclear. Nevertheless, administration of IL-1Ra has shown to improve insulin sensitivity in animal models.

Therapy potential of blocking IL-1 signaling

Together, these studies have shown significant mechanistic insights into the contribution of IL-1 β in the pathogenesis of obesity-induced insulin resistance. Therefore, inhibition of IL-1 β (signaling) itself, or upstream activators of IL-1 β activity, such as caspase-1, may be an attractive therapeutic target. Indeed, several approaches aimed at modulating IL-1 signaling have been shown to be successful in improving insulin sensitivity in various obese mouse models, as discussed above. In humans, increased IL-1 β release can be treated with IL-1 blockade and is successfully applied in patients diagnosed with a broad spectrum of pro-inflammatory diseases [82]. Inhibition of IL-1 β actions can be achieved by using neutralizing monoclonal IL-1 β antibodies (e.g. gevokizumab or canakinumab). Alternatively, blockade of the IL-1R1 is effective using IL-1Ra (anakinra). Importantly, since this is receptor blockade, anakinra reduces the activities of both IL-1 β and IL-1 α .

These strategies have been used in several clinical studies to evaluate the therapeutic potential of IL-1 β inhibition to improve glycemic control in patients with diabetes, as it was

shown that reducing bioactivity of IL-1 with recombinant IL-1Ra improves β -cell function and reduce glucotoxicity in type 2 diabetes [30]. Indeed, trials revealed that inhibition of IL-1 β improves hyperglycemia in type 2 diabetes patients [83-87]. Of note, the reduction in HbA1c levels in these studies was most likely the result of improved β -cell function [85-87], since insulin sensitivity was unaltered in type 2 diabetes patients or subjects with the metabolic syndrome [84, 87, 88]. Thus, although *ex vivo* and *in vivo* studies provide strong evidence that IL-1 β is causally involved in obesity-induced insulin resistance, clinical evidence for an improvement of insulin sensitivity upon inhibition of IL-1 β is so far lacking. Of note, whereas preclinical studies are performed with genetically identical animals, outcomes of clinical studies may greatly be influenced by genetic variation. For example, circulating levels of IL-1Ra show great inter-individual variation [89]. Possibly, higher levels of IL-1Ra in certain individuals may limit beneficial effects of anti-IL-1 β treatment. Conversely, blocking anti-IL-1 β in individuals with levels of IL-1 signaling may prove more effective. Therefore, more personalized approaches that include immuno-phenotyping to identify individuals that particularly benefit from anti-inflammatory therapy may improve efficacy of blocking IL-1 signaling in patients with diabetes. Indeed, a more personalized approach is currently being tested in the CANTOS trial that investigates IL-1 β inhibition with canakinumab in the prevention of cardiovascular events. The study specifically includes patients with elevated chronic inflammation, as determined by hsCRP levels [90]. Within this large trial, the new onset of type 2 diabetes, determined by insulin sensitivity and β -cell function, will be evaluated as a secondary endpoint [90]. Based on preclinical studies that have been discussed above, new agents aimed at inhibiting upstream regulators of IL-1 β production and activity, such as caspase-1 or neutrophil elastase [57, 67] may hold promise.

IL-18

More than 20 years ago IL-18 was identified as gamma interferon (IFN- γ)-inducing factor [91, 92]. IL-18 primarily induces T-helper-1 responses due to the induction of IFN γ in combination with IL-12. However, in the absence of IL-12, there is a role for T-helper-2 [93] and NK-cell maturation, cytokine production and cytotoxicity [92, 94-96]. Similar to IL-1 β , the protein is synthesized as an inactive precursor and requires caspase-1 processing for activation [45]. IL-18 binds to the alpha chain of the IL-18 receptor and recruits the IL-18R beta chain, after which it signals through important immune regulatory proteins, such as Myd88, ERK, JNK and NF κ B that it shares with IL-1 [97] and Toll-like Receptor signaling [39, 98]. IL-18 forms a complex with the IL-18 Receptor α chain (IL-18R α) and the IL-18 Receptor β chain (IL-18R β) to exert its pro-inflammatory actions. Although IL-18R α is expressed by most cells, IL-18R β is not. IL-18 signaling is therefore dependent on IL-12-induced upregulation of IL-18R β . Moreover, IL-18 is inhibited by the natural occurring IL-18 binding protein, which binds free IL-18, but is not a competitor for the IL-18 receptor [99].

Furthermore, IL-18 is strongly associated with development of cardiovascular diseases [100-102], and a causal role for IL-18 in the development of atherosclerosis is supported by numerous experimental animal studies. Overexpressing IL-18 in atherosclerosis prone mouse models increased atherosclerotic plaque development [103-105], while development of atherosclerosis was reduced upon overexpression of IL-18bp [102], or absence of IL-18

[106]. The effects of IL-18 on atherosclerosis are mainly caused by the induction of IFN- γ , an important cytokine in the process of atherogenesis [107].

IL-18 in obesity and insulin resistance in humans

The first studies on the role of IL-18 in obesity and development of type 2 diabetes reported that plasma levels of IL-18 were elevated in obese individuals and subjects with type 2 diabetes [108-111]. Moreover, IL-18 levels were restored in individuals that lost weight after bariatric surgery [112]. Furthermore, several polymorphisms in the IL-18 gene were associated with risk factors for the metabolic syndrome, including increased blood pressure [110], insulin resistance [113, 114] impaired glucose regulation [111], the development of obesity [115] and associated with a higher prevalence of type 2 diabetes [111]. Interestingly, IL-18 was found to be secreted from adipose tissue at a higher rate in obese individuals as compared to lean controls [116, 117]. Moreover, IL-18 is higher expressed in [55] and secreted from [54] visceral as compared to subcutaneous adipose tissue.

Within adipose tissue, IL-18 mRNA is specifically expressed in stromal vascular cells [56, 118]. These observations indicated that increased levels of the pro-inflammatory cytokine IL-18 are associated with an elevated risk to develop metabolic disturbances during obesity. Notably, elevated plasma levels of IL-18 are independent of obesity or type 2 diabetes associated with insulin resistance [119]. Based on these observations, it was thought that elevated plasma levels of IL-18 could possibly play a pathophysiological role in the development of insulin resistance, similar to other pro-inflammatory cytokines. Since clinical studies could not elucidate the causative effects, preclinical experiments in animals were performed [31, 120]. Interestingly, these studies revealed a complex relation between IL-18 and metabolic disease, as discussed below. Of note, as compared to other IL-1 family members, circulating IL-18 plasma levels are substantially higher.

Contributions of IL-18 to the development of obesity and insulin resistance

Studies with IL-18 and IL-18 Receptor knockout mice have shed light on the causal relation between IL-18, obesity and insulin resistance. Similar to humans, obesity increases IL-18 protein levels in adipose tissue from mice, an effect secondary to increased caspase-1 activity [121]. Paradoxically, however, both IL-18 knockout mice and IL-18 Receptor α chain knockout mice display increased bodyweight on normal chow diet and developed insulin resistance [31, 120]. The development of obesity in absence of IL-18 appeared to be due to hyperphagia [31, 120]. Indeed, injection of recombinant IL-18 reduced food intake and bodyweight gain in food deprived WT mice [120]. Together, these studies show that the presence of IL-18 in mice reduces food intake and improves insulin sensitivity.

Interestingly, acute injection of recombinant IL-18 in mice directly improves insulin sensitivity, suggesting that the effect of IL-18 on insulin sensitivity is not secondary to a reduction in food intake, but may directly govern insulin signaling [31]. Indeed, others have shown that IL-18 directly improves glucose uptake in murine 3T3 cells *in vitro*, and can counteract the insulin resistance induced by TNF α [122].

The mechanism by which IL-18 enhances insulin sensitivity is unclear. The knowledge that IL-18 signals through STAT3 [31], prompted others to investigate whether IL-18 is able to

activate AMPK, similar to other STAT3 activating proteins [123-125]. Interestingly, administration of IL-18 was shown to activate AMPK in muscle via the IL-18R and overexpression of IL-18 in muscle elicits positive metabolic effects by enhancing lipid oxidation and reducing weight gain in mice [126].

In summary, although increased circulating IL-18 levels are associated with symptoms of the metabolic syndrome and type 2 diabetes in humans, absence of IL-18 has detrimental effects on the development of obesity and insulin resistance in mice. Moreover, IL-18 increases rather than reduces insulin sensitivity *in vivo* and *in vitro*, at least partly via activation of AMPK. Thus, the enhanced IL-18 levels in obese individuals and type 2 diabetes may be more viewed upon as a compensatory response rather than a causal factor in the development of metabolic disturbances. Possibly, circulating IL-18 levels may simply reflect increased inflammasome-mediated caspase-1 activation, which has shown to be detrimental for obesity and insulin resistance, as described above. Alternatively, IL-18 may be secreted as a compensatory response to counteract the state of insulin resistance mediated by other pro-inflammatory cytokines, such as TNF α [122]. IL-18 resistance - the observation of reduced expression of the IL-18 receptor in PBMCs in type 2 diabetes patients [127] - possibly hampers these counteracting effects.

Therapeutic potential of IL-18

In summary, although experimental studies reveal a potential beneficial effect of IL-18 on the development obesity and insulin resistance, studies in human subjects are so far lacking and could be hampered by the presence of IL-18 resistance in type 2 diabetes patients. Importantly, the involvement of IL-18 in atherosclerotic plaque development and in other inflammatory conditions may seriously limit therapeutic options in humans for IL-18 within the context of metabolic disease.

IL-33

IL-33 (formerly IL-1F11), one of the novel IL-1 family members was identified in 2005 [128]. It signals through the ST-2 receptor and its co-receptor IL-1 receptor accessory protein (IL-1RAcP). Several isoforms of the ST2 receptor exist, including a transmembrane receptor (ST2L) and a soluble receptor (sST2), which can serve as a decoy receptor for IL-33 [129]. Full length IL-33 is biologically active, but cleavage by protease elastase, cathepsin G or proteinase 3, results in a 10-fold greater potency to activate ST2 [130, 131]. IL-33 is released by active secretion as well as after cell injury or necrosis, in the latter case acting as an alarmin and activating NF- κ B, p38 and JNK [18, 132]. IL-33 is expressed in multiple cells and organs, such as lungs, skin, lymph nodes, adipocytes and myocytes [17, 133]. By inducing T helper type 2 cell polarization and activation [134], IL-33 exacerbates inflammation [135-137]. In the past few years it has emerged as a cytokine playing an important role in several inflammatory diseases [138]: autoimmune diseases [139], rheumatoid arthritis, ulcerative colitis, asthma [140], sepsis [141] and allergy [142]. In addition, it has been ascribed with protective effects in atherosclerosis [143].

IL-33 in humans

The role of IL-33 in obesity, adipose tissue inflammation and insulin resistance has only recently been investigated. Expression of IL-33 was detected in human white adipose tissue as well as in human SGBS (Simpson-Golabi-Behmel-Syndrome) adipocytes and pre-adipocytes *in vitro* [144]. Hypoxia and TNF α , similar to IL-1 α and IL-1 β present in adipose tissue during obesity, induced IL-33 in SGBS (pre-) adipocytes [144]. Zeyda et al. subsequently observed that expression of IL-33 and its receptor ST-2 in adipose tissue were markedly enhanced in severely obese persons, and demonstrated that the main source for IL-33 in adipose tissue were endothelial cells, not adipocytes [145]. Plasma levels of IL-33 were similar between lean and obese individuals, indicating that in obesity a role for IL-33 may be limited to local effects. A recent study reported even higher IL-33 plasma levels in lean compared to non-lean individuals [146]. Since these observations were done in a relatively small number of individuals, larger studies are needed to further investigate this relationship. Nevertheless, there are reports of IL-33 function as an anti-inflammatory cytokine [18].

IL-33 in the development of obesity and insulin resistance

Similar to the human situation, IL-33 mRNA levels increase in the adipose tissue of diet-induced obese mice compared to WT-mice [145]. Interestingly, genetically obese (db/db) mice that lack leptin receptor signaling do not show an increase of IL-33 mRNA [145]. Notably, IL-33 expression in human adipose tissue significantly correlates with leptin expression [145], suggesting that IL-33 expression may be enhanced by leptin signaling. More evidence for this hypothesis was shown by the observation that leptin increased IL-33 expression in human smooth muscle cells [145]. As leptin levels are increased during obesity, this may be one of the mechanisms underlying the increase in IL-33 in adipose tissue during obesity.

The function of IL-33 in adipose tissue, especially during obesity, is not fully understood. Administration of IL-33 to genetically obese (ob/ob) mice that are leptin deficient reduces adipose tissue weight, fat mass and adipocyte size [147], without effects on total body weight. *In vitro*, recombinant IL-33 reduces adipocyte differentiation and lipid storage in adipocyte cultures. This suggests that IL-33 has potential interesting effects in obesity by reducing lipid storage and adipogenesis.

In addition, IL-33 may influence adipose tissue inflammation during obesity. IL-33 administration to ob/ob mice induces accumulation of T-helper 2 cytokines in serum and adipose tissue [147], which is known to associate with differentiation of macrophages towards an anti-inflammatory M2 phenotype. Indeed, IL-33 enhanced the accumulation of macrophages in adipose tissue, which were mainly the M2 macrophage phenotype. This effect was paralleled by lower fasting glucose levels without changes in insulin levels, suggesting increased insulin sensitivity [147]. Thus IL-33 shows potential as a therapy in obesity to skew adipose tissue macrophages from M1 to M2 phenotype, which may be beneficial for maintenance of insulin sensitivity. Of note, type 2 cytokine signaling has recently been linked to ameliorated adipose tissue inflammation and modulate biogenesis of beige fat [148], suggesting a role for IL-33 in browning of adipose tissue. Interestingly, it

has just been revealed that IL-33 indeed can activate UCP-1 in subcutaneous adipose tissue. Administration of IL-33 increased browning of white adipose tissue in mice and increased energy expenditure [149]. These observations likely explain the earlier observation of a reduced adipose tissue mass upon administration of IL-33 to mice [147]. Together, this shows that recombinant IL-33 may have protective effects concerning adipose tissue mass, adipose tissue inflammation and insulin resistance.

Compared to IL-1 and IL-18, only limited research has been performed concerning the role of IL-33 in metabolic disease. Future research should investigate whether IL-33 has ameliorating effects on insulin, glucose and lipid metabolism in more physiological models of obesity such as HFD-feeding, especially since the current observations are based on recombinant IL-33 treatment in ob/ob mice, which appear to have altered IL-33 signaling (as discussed above). In addition, the effects of recombinant IL-33 on other organs and tissues need to be determined. However, although IL-33 has beneficial effects during obesity, IL-33 as a potential treatment to enhance insulin sensitivity, may be hindered by the pro-inflammatory functions of this cytokine.

IL-37

IL-37 is a unique member of the IL-1 family (formerly IL-1F7). The cytokine broadly inhibits inflammation by reducing several signaling kinases and augmenting anti-inflammation genes [150]. Since 2010, an increasing number of studies have described the anti-inflammatory actions of this cytokine in models of disease [151]. During acute and chronic inflammation, IL-37 shifts the cytokine balance away from inflammation [151]. IL-37 binds to the IL-18 Receptor alpha chain but recruits the orphan IL-1R8 (Formerly, Single Ig IL-1-related Receptor, SIGIRR) rather than the IL-18 Receptor beta chain; thus IL-1R8 functions as the co-receptor for the complex. Despite binding to the IL-18R, IL-37 does not function as an IL-18 receptor antagonist. In fact, low concentrations of recombinant IL-37 are more effective in inhibiting innate inflammation than concentrations 100 fold higher [152], an observation inconsistent with receptor antagonist function. Signaling via IL-1R8 reduces NF- κ B and JNK activation, by preventing recruitment of receptor-proximal signaling component MyD88, which transduces signals from IL-1 and IL-18 as well as all TLRs [153, 154]. Since the TIR domain of IL-1R8 is mutated, MyD88 is sequestered to IL-1R8, depriving a proper signal from IL-1 or TLR ligands; as such, IL-1R8 acts as a decoy or sink for MyD88. Thus, IL-1R8 provides the extracellular actions of IL-37 with a mechanism to reduce IL-1 receptor signaling, but also signaling via receptors of the Toll-like receptor family [155].

In addition to an extracellular function, IL-37 has a nuclear function [156]. Similar to IL-1 α and IL-33, IL-37 translocates to the nucleus. In contrast to IL-1 α and IL-33, IL-37 downregulates the production of pro-inflammatory cytokines [156]. Caspase-1 is necessary for nuclear translocation [157], but appears to play a minor role for the processing and release of IL-37 [157]. Secretion of the IL-37 precursor takes place and provides the extracellular signal [152]. The IL-37 precursor is fully active [158]. Extracellular serine proteases, such as proteinase 3 or elastase, may play a role in processing IL-37 to a more active form.

IL-37 in obesity and insulin resistance in humans

Moschen et al. were the first to show that IL-37 mRNA was present in human adipose tissue [55]. As compared to liver, expression of IL-37 was significantly higher in human adipose tissue, particularly in the visceral fat depot [55]. Within the adipose tissue, the highest levels of IL-37 mRNA were present in the adipocytes, which was in contrast to IL-1 β , IL-1Ra, IL-18 and IL-33 that display higher expression levels in the stromal vascular fraction (SVF) [56]. These observations of IL-37 in adipose tissue are compatible with a modulatory role of this anti-inflammatory cytokine in obesity-induced inflammation and insulin resistance. IL-37 mRNA levels in human adipose tissue increase after marked weight loss in morbidly obese individuals [55], and higher IL-37 mRNA levels are associated with markers of enhanced insulin sensitivity and reduced adipose tissue inflammation [56]. No other studies have been performed to investigate the regulation of IL-37 during the development of obesity or type 2 diabetes in humans. In a Chinese cohort, IL-37 plasma levels were elevated in patients with coronary heart disease [159]. It will be interesting to evaluate whether (changes in) tissue or plasma levels of IL-37, or SNPs in the IL-37 gene [160], are associated with a change in the risk to develop metabolic diseases.

IL-37 in the development of obesity and insulin resistance

The causal effects of IL-37 in obesity-related adipose tissue inflammation and insulin sensitivity have been investigated in experimental studies. Since mice do not have a functional IL-37 gene, human IL-37 transgenic mice (IL-37tg) were generated and used for these experiments. High-fat feeding of IL-37tg mice resulted in less adipose tissue inflammation, as determined by a reduced number of adipose tissue macrophages and secretion of pro-inflammatory cytokines, as compared to high-fat feeding of control mice [56]. In addition, IL-37tg mice subjected to 16 weeks of HFD had better glucose tolerance and insulin sensitivity compared to WT mice. In line with these findings, silencing of IL-37 in hepatocytes *in vitro* enhances inflammation and induces insulin resistance. Similar to IL-33, IL-37 not only affects inflammatory pathways, but also directly affects adipogenesis and adiposity. *In vivo*, IL-37tg mice did not develop an obese phenotype in response to HFD-feeding and showed a marked reduction in adipose tissue mass. Moreover *in vitro*, recombinant IL-37 inhibited adipogenesis in human SGBS adipocytes [56]. This suggests that IL-37 exerts direct effects on adipose tissue lipid metabolism, which may be regulated via activation of AMPK. Indeed, IL-37 increases phosphorylation of AMPK in THP-1 cells [150], as well as murine 3T3 adipocytes [56]. AMPK is a central regulator of cellular metabolism [161] and has anti-obesity effects [162]. As AMPK is an important target for healthy metabolism and dampening of inflammation [163], the relation between IL-37 and AMPK provides an interesting target for further investigations.

In summary, the anti-inflammatory and metabolic actions of IL-37, such as activating AMPK and ameliorating insulin signaling, appear to counteract the harmful effects associated with obesity. However, the current studies have limitations and many questions remain. Physiological (human) models are necessary to further understand the role of IL-37 in the metabolic syndrome. It will be necessary to find the (metabolic) stimuli that trigger or suppress IL-37 production. Furthermore, the importance of nuclear versus the extracellular actions of IL-37 in alleviating chronic low-grade inflammation in obesity remains unclear.

Clearly, additional studies are warranted to reveal other biological effects of this cytokine, especially in humans, and to determine whether variation in IL-37 levels (*i.e.* caused by genetic variation) could possibly predict the risk for the development of metabolic diseases.

IL-36 α / β / γ , IL-36Ra and IL-38

IL-36 cytokines are new members of the IL-1 family, discovered a decade ago [164-169] and identified as IL-36 α , - β , - γ (formerly IL-1F6, -8, -9) and receptor antagonist IL-36Ra (formerly IL-1F5). IL-36 α , - β , - γ activate dendritic cells and polarize T-helper cell responses and play a significant role in the pathogenesis of skin diseases. As all pro-inflammatory IL-36 cytokines signal through the same receptor, they elicit similar effects. Thus, differences between these cytokines lie mainly in tissue or cell specific expression and therefore tissue-specific contributions of the various isoforms. The antagonist IL-36Ra blocks the signaling of the three pro-inflammatory IL-36 cytokines [19]. IL-36 cytokines need to be cleaved for full activity [170]. However, unlike IL-1 β , proteases other than caspase-1 are likely to be involved with processing of the inactive precursor to an active cytokine [19, 170].

IL-38 (formerly IL-1F10) was first described in 2001 as another novel IL-1 family member [171, 172]. However, the role of IL-38 in human disease is still largely unknown. IL-38 binds to the IL-36 receptor similar to IL-36 receptor antagonist [21]. This suggests that IL-38 may act as an anti-inflammatory cytokine. While other IL-1 family members are acknowledged to play a central role in the development of obesity-induced inflammation and insulin resistance, little is known about the IL-36 and IL-38 cytokines within the development of metabolic diseases.

IL-36 cytokines

Associated with pro-inflammatory effects, it may be expected that IL-36 α / β / γ can aggravate metabolic inflammation and that IL-36Ra could balance this, similar to the effects ascribed to the proinflammatory IL-1 β and antagonistic actions of IL-1Ra. However, expression of IL-1 β and IL-1Ra is abundant in metabolic tissues, such as adipose tissue, whereas for IL-36 cytokines expression is less clear. To date, IL-36 α , but not IL-36 β , is expressed in adipose tissue [173]. Moreover, expression of IL-36 α is relatively higher in the stromal vascular cells as compared to adipocytes. Whether the IL-36 cytokines affect metabolic inflammation or insulin sensitivity has not been evaluated. One report has described the effects of IL-36 cytokines on adipocytes *in vitro* [173]. Hence, both IL-36 α and IL-36 β induce modest IL-8 production from adipocytes, whereas only IL-36 α also induces IL-6 production and reduces adipogenesis. The effect on adipogenesis may be secondary to the induction of IL-6 by IL-36 α , since IL-6 is known to inhibit adipogenesis [174]. The difference between IL-36 α and IL-36 β in this model is remarkable, since both cytokines signal through the same receptor, suggesting that IL-36 α may possess a greater ability for induction of specific cytokine secretion. We are not aware of publications that have specifically investigated the role of IL-36 γ . Clearly, there is a gap in understand the role of IL-36 cytokines in obesity-induced inflammation and insulin resistance.

IL-38

Whether IL-38 has a role during the development of obesity and related diseases has not been investigated. Nevertheless, a single nucleotide polymorphism (SNP) in the IL-38 gene is associated with plasma C-reactive protein levels [175], a marker that is increased in serum of obese and type 2 diabetes patients. Interestingly, another SNP in a locus close to the IL-38 gene associates with increased circulating IL-1Ra levels [176]. Blocking IL-1 β activity with an antibody or with IL-1Ra is promising in treating type 2 diabetes [83, 87, 90]. In line with this, the IL-1Ra-increasing SNP in the IL-38 gene associated with lower fasting insulin levels and HOMA-IR [176]. Other SNPs that were found to associate with elevated IL-1Ra levels were not associated with insulin or HOMA-IR, suggesting that the association of the SNP in IL-38 with metabolic traits is independent of IL-1Ra levels. Clearly, more investigations are needed to elucidate whether IL-38 has any (direct) role in obesity-induced inflammation and type 2 diabetes.

To summarize, the role of the IL-36 subfamily and IL-38 in obesity and metabolic inflammation has not been sufficiently investigated and several questions remain. Is expression of IL-36 cytokines or IL-38 associated with obesity or insulin resistance; are these cytokines present in other metabolic organs, e.g. liver and muscles; are they activated or inhibited during the development obesity; are these cytokines causally involved in obesity-associated adipose tissue inflammation and insulin resistance? Future studies are warranted, before we can understand if there is a role for these members of the IL-1 family in obesity-induced inflammation or development of insulin resistance.

Conclusion

IL-1 family members are major players in the initiation and regulation of inflammation associated with obesity (Table 1). IL-1 β is well recognized for its role in insulin resistance during experimental obesity. Therefore, blocking endogenous IL-1 with IL-1Ra counteracts this effect in experimental settings, but can be extended to treatment of clinical insulin resistance with the known limitations, such as safety. However, reducing IL-1 activity to increase insulin sensitivity in human individuals is presently unproven. One advantage of blocking IL-1 is that it directly attacks the upstream inflammation, compared to oral downstream hypoglycemic agents. IL-18 appears to be beneficial for obesity and insulin resistance in animal models, but IL-18 is clearly involved in the pathogenesis of atherosclerosis, which may hamper possibilities as a therapy for metabolic disease. Recently, others and we have shown that IL-33 and IL-37, via anti-inflammatory properties modulate metabolism, and provide possible new targets for future research in combating obesity and type 2 diabetes. Therefore, both blocking pro-inflammatory or increasing (endogenous) anti-inflammatory members of the IL-1 family may be beneficial in metabolic disease. Additionally, ways to identify individuals that may benefit from anti-inflammatory treatment, for example those patients characterized by low endogenous levels of IL-1Ra, may also help to improve treatment outcomes. Other novel members of the IL-1 family (i.e. IL-36 $\alpha/\beta/\gamma$, IL-36Ra and IL-38) have not been studied in relation to metabolic diseases so far, but may be interesting cytokines for future research.

Acknowledgments

Grants: Ballak is supported by the Interleukin-1 foundation, Stienstra is supported by a Vidi grant of the Netherlands Organization for Scientific Research (NWO) (016.136.311). Dinarello is supported by NIH (grant AI-1561), Van Diepen is supported by a grant of the Dutch Diabetes Research Foundation (2013.81.1674).

References

1. Ng M, Fleming T, Robinson M, Thomson B, Graetz N, Margono C, et al. Global, regional, and national prevalence of overweight and obesity in children and adults during 1980-2013: a systematic analysis for the Global Burden of Disease Study 2013. *Lancet*. 2014; 384:766–81. [PubMed: 24880830]
2. Gregor MF, Hotamisligil GS. Inflammatory Mechanisms in Obesity. *Annu Rev Immunol*. 2011
3. Osborn O, Olefsky JM. The cellular and signaling networks linking the immune system and metabolism in disease. *Nat Med*. 2012; 18:363–74. [PubMed: 22395709]
4. Hotamisligil GS, Peraldi P, Budavari A, Ellis R, White MF, Spiegelman BM. IRS-1-mediated inhibition of insulin receptor tyrosine kinase activity in TNF- α - and obesity-induced insulin resistance. *Science*. 1996; 271:665–8. [PubMed: 8571133]
5. Jager J, Gremeaux T, Cormont M, Le Marchand-Brustel Y, Tanti JF. Interleukin-1 β -induced insulin resistance in adipocytes through down-regulation of insulin receptor substrate-1 expression. *Endocrinology*. 2007; 148:241–51. [PubMed: 17038556]
6. Hotamisligil GS, Murray DL, Choy LN, Spiegelman BM. Tumor necrosis factor α inhibits signaling from the insulin receptor. *Proc Natl Acad Sci U S A*. 1994; 91:4854–8. [PubMed: 8197147]
7. Feinstein R, Kanety H, Papa MZ, Lunenfeld B, Karasik A. Tumor necrosis factor α suppresses insulin-induced tyrosine phosphorylation of insulin receptor and its substrates. *J Biol Chem*. 1993; 268:26055–8. [PubMed: 8253716]
8. Begum N, Ragolia L. Effect of tumor necrosis factor- α on insulin action in cultured rat skeletal muscle cells. *Endocrinology*. 1996; 137:2441–6. [PubMed: 8641197]
9. Kroder G, Bossenmaier B, Kellerer M, Capp E, Stoyanov B, Muhlhofer A, et al. Tumor necrosis factor- α - and hyperglycemia-induced insulin resistance. Evidence for different mechanisms and different effects on insulin signaling. *J Clin Invest*. 1996; 97:1471–7. [PubMed: 8617880]
10. Gao D, Madi M, Ding C, Fok M, Steele T, Ford C, et al. Interleukin-1 β mediates macrophage-induced impairment of insulin signaling in human primary adipocytes. *Am J Physiol Endocrinol Metab*. 2014
11. Hotamisligil GS, Shargill NS, Spiegelman BM. Adipose expression of tumor necrosis factor- α : direct role in obesity-linked insulin resistance. *Science*. 1993; 259:87–91. [PubMed: 7678183]
12. Weisberg SP, McCann D, Desai M, Rosenbaum M, Leibel RL, Ferrante AW Jr. Obesity is associated with macrophage accumulation in adipose tissue. *J Clin Invest*. 2003; 112:1796–808. [PubMed: 14679176]
13. Xu H, Barnes GT, Yang Q, Tan G, Yang D, Chou CJ, et al. Chronic inflammation in fat plays a crucial role in the development of obesity-related insulin resistance. *J Clin Invest*. 2003; 112:1821–30. [PubMed: 14679177]
14. Cildir G, Akincilar SC, Tergaonkar V. Chronic adipose tissue inflammation: all immune cells on the stage. *Trends Mol Med*. 2013; 19:487–500. [PubMed: 23746697]
15. Sell H, Habich C, Eckel J. Adaptive immunity in obesity and insulin resistance. *Nat Rev Endocrinol*. 2012; 8:709–16. [PubMed: 22847239]
16. Dinarello CA. Interleukin-18, a proinflammatory cytokine. *Eur Cytokine Netw*. 2000; 11:483–6. [PubMed: 11203186]
17. Liew FY, Pitman NI, McInnes IB. Disease-associated functions of IL-33: the new kid in the IL-1 family. *Nat Rev Immunol*. 2010; 10:103–10. [PubMed: 20081870]
18. Miller AM. Role of IL-33 in inflammation and disease. *J Inflamm (Lond)*. 2011; 8:22. [PubMed: 21871091]

19. Gresnigt MS, van de Veerdonk FL. Biology of IL-36 cytokines and their role in disease. *Semin Immunol.* 2013; 25:458–65. [PubMed: 24355486]
20. Boraschi D, Lucchesi D, Hainzl S, Leitner M, Maier E, Mangelberger D, et al. IL-37: a new anti-inflammatory cytokine of the IL-1 family. *Eur Cytokine Netw.* 2011; 22:127–47. [PubMed: 22047735]
21. van de Veerdonk FL, Stoeckman AK, Wu G, Boeckermann AN, Azam T, Netea MG, et al. IL-38 binds to the IL-36 receptor and has biological effects on immune cells similar to IL-36 receptor antagonist. *Proc Natl Acad Sci U S A.* 2012; 109:3001–5. [PubMed: 22315422]
22. Garlanda C, Dinarello CA, Mantovani A. The interleukin-1 family: back to the future. *Immunity.* 2013; 39:1003–18. [PubMed: 24332029]
23. Esposito K, Pontillo A, Ciotola M, Di Palo C, Grella E, Nicoletti G, et al. Weight loss reduces interleukin-18 levels in obese women. *J Clin Endocrinol Metab.* 2002; 87:3864–6. [PubMed: 12161523]
24. Meier CA, Bobbioni E, Gabay C, Assimakopoulos-Jeannot F, Golay A, Dayer JM. IL-1 receptor antagonist serum levels are increased in human obesity: a possible link to the resistance to leptin? *J Clin Endocrinol Metab.* 2002; 87:1184–8. [PubMed: 11889184]
25. Juge-Aubry CE, Somm E, Giusti V, Pernin A, Chicheportiche R, Verdumo C, et al. Adipose tissue is a major source of interleukin-1 receptor antagonist: upregulation in obesity and inflammation. *Diabetes.* 2003; 52:1104–10. [PubMed: 12716739]
26. Bendtzen K, Mandrup-Poulsen T, Nerup J, Nielsen JH, Dinarello CA, Svenson M. Cytotoxicity of human p17 interleukin-1 for pancreatic islets of Langerhans. *Science.* 1986; 232:1545–7. [PubMed: 3086977]
27. Mandrup-Poulsen T, Bendtzen K, Nerup J, Dinarello CA, Svenson M, Nielsen JH. Affinity-purified human interleukin 1 is cytotoxic to isolated islets of Langerhans. *Diabetologia.* 1986; 29:63–7. [PubMed: 3514344]
28. McCarthy DO, Kluger MJ, Vander AJ. Suppression of food intake during infection: is interleukin-1 involved? *Am J Clin Nutr.* 1985; 42:1179–82. [PubMed: 3907325]
29. Tocco-Bradley R, Georgieff M, Jones CT, Moldawer LL, Dinarello CA, Blackburn GL, et al. Changes in energy expenditure and fat metabolism in rats infused with interleukin-1. *Eur J Clin Invest.* 1987; 17:504–10. [PubMed: 3123250]
30. Maedler K, Sergeev P, Ris F, Oberholzer J, Joller-Jemelka HI, Spinas GA, et al. Glucose-induced beta cell production of IL-1beta contributes to glucotoxicity in human pancreatic islets. *J Clin Invest.* 2002; 110:851–60. [PubMed: 12235117]
31. Netea MG, Joosten LA, Lewis E, Jensen DR, Voshol PJ, Kullberg BJ, et al. Deficiency of interleukin-18 in mice leads to hyperphagia, obesity and insulin resistance. *Nat Med.* 2006; 12:650–6. [PubMed: 16732281]
32. Dinarello CA. Immunological and inflammatory functions of the interleukin-1 family. *Annu Rev Immunol.* 2009; 27:519–50. [PubMed: 19302047]
33. Auron PE, Webb AC, Rosenwasser LJ, Mucci SF, Rich A, Wolff SM, et al. Nucleotide sequence of human monocyte interleukin 1 precursor cDNA. *Proc Natl Acad Sci U S A.* 1984; 81:7907–11. [PubMed: 6083565]
34. Lomedico PT, Gubler U, Hellmann CP, Dukovich M, Giri JG, Pan YC, et al. Cloning and expression of murine interleukin-1 cDNA in *Escherichia coli*. *Nature.* 1984; 312:458–62. [PubMed: 6209582]
35. March CJ, Mosley B, Larsen A, Cerretti DP, Braedt G, Price V, et al. Cloning, sequence and expression of two distinct human interleukin-1 complementary DNAs. *Nature.* 1985; 315:641–7. [PubMed: 2989698]
36. Rider P, Carmi Y, Voronov E, Apte RN. Interleukin-1alpha. *Semin Immunol.* 2013; 25:430–8. [PubMed: 24183701]
37. Dinarello CA. A clinical perspective of IL-1beta as the gatekeeper of inflammation. *Eur J Immunol.* 2011; 41:1203–17. [PubMed: 21523780]
38. Cooney RN, Shumate M. The inhibitory effects of interleukin-1 on growth hormone action during catabolic illness. *Vitam Horm.* 2006; 74:317–40. [PubMed: 17027521]

39. Akira S, Takeda K, Kaisho T. Toll-like receptors: critical proteins linking innate and acquired immunity. *Nat Immunol.* 2001; 2:675–80. [PubMed: 11477402]
40. Hannum CH, Wilcox CJ, Arend WP, Joslin FG, Dripps DJ, Heimdal PL, et al. Interleukin-1 receptor antagonist activity of a human interleukin-1 inhibitor. *Nature.* 1990; 343:336–40. [PubMed: 2137200]
41. Dinarello CA. Biologic basis for interleukin-1 in disease. *Blood.* 1996; 87:2095–147. [PubMed: 8630372]
42. Gabay C, Lamacchia C, Palmer G. IL-1 pathways in inflammation and human diseases. *Nat Rev Rheumatol.* 2010; 6:232–41. [PubMed: 20177398]
43. Sheedy FJ, Moore KJ. IL-1 signaling in atherosclerosis: sibling rivalry. *Nat Immunol.* 2013; 14:1030–2. [PubMed: 24048132]
44. Dinarello CA. Interleukin-1 in the pathogenesis and treatment of inflammatory diseases. *Blood.* 2011; 117:3720–32. [PubMed: 21304099]
45. Lamkanfi M, Dixit VM. Mechanisms and Functions of Inflammasomes. *Cell.* 2014; 157:1013–22. [PubMed: 24855941]
46. Netea MG, van de Veerdonk FL, van der Meer JW, Dinarello CA, Joosten LA. Inflammasome-Independent Regulation of IL-1-Family Cytokines. *Annu Rev Immunol.* 2014
47. Dinarello CA, Donath MY, Mandrup-Poulsen T. Role of IL-1beta in type 2 diabetes. *Curr Opin Endocrinol Diabetes Obes.* 2010; 17:314–21. [PubMed: 20588114]
48. Donath MY, Dalmas E, Sauter NS, Boni-Schnetzler M. Inflammation in obesity and diabetes: islet dysfunction and therapeutic opportunity. *Cell Metab.* 2013; 17:860–72. [PubMed: 23747245]
49. Donath MY, Schumann DM, Faulenbach M, Ellingsgaard H, Perren A, Ehses JA. Islet inflammation in type 2 diabetes: from metabolic stress to therapy. *Diabetes Care.* 2008; 31(2):S161–4. [PubMed: 18227479]
50. Mandrup-Poulsen T. The role of interleukin-1 in the pathogenesis of IDDM. *Diabetologia.* 1996; 39:1005–29. [PubMed: 8877284]
51. Raymond NC, Dysken M, Bettin K, Eckert ED, Crow SJ, Markus K, et al. Cytokine production in patients with anorexia nervosa, bulimia nervosa, and obesity. *Int J Eat Disord.* 2000; 28:293–302. [PubMed: 10942915]
52. Um JY, Chung HS, Song MY, Shin HD, Kim HM. Association of interleukin-1beta gene polymorphism with body mass index in women. *Clin Chem.* 2004; 50:647–50. [PubMed: 14981033]
53. Di Renzo L, Bigioni M, Del Gobbo V, Premrov MG, Barbini U, Di Lorenzo N, et al. Interleukin-1 (IL-1) receptor antagonist gene polymorphism in normal weight obese syndrome: relationship to body composition and IL-1 alpha and beta plasma levels. *Pharmacol Res.* 2007; 55:131–8. [PubMed: 17174563]
54. Koenen TB, Stienstra R, van Tits LJ, Joosten LA, van Velzen JF, Hijmans A, et al. The inflammasome and caspase-1 activation: a new mechanism underlying increased inflammatory activity in human visceral adipose tissue. *Endocrinology.* 2011; 152:3769–78. [PubMed: 21862623]
55. Moschen AR, Molnar C, Enrich B, Geiger S, Ebenbichler CF, Tilg H. Adipose and liver expression of interleukin (IL)-1 family members in morbid obesity and effects of weight loss. *Mol Med.* 2011; 17:840–5. [PubMed: 21394384]
56. Ballak DB, van Diepen JA, Moschen AR, Jansen HJ, Hijmans A, Groenhof GJ, et al. IL-37 protects against obesity-induced inflammation and insulin resistance. *Nat Commun.* 2014; 5:4711. [PubMed: 25182023]
57. Stienstra R, Joosten LA, Koenen T, van Tits B, van Diepen JA, van den Berg SA, et al. The inflammasome-mediated caspase-1 activation controls adipocyte differentiation and insulin sensitivity. *Cell Metab.* 2010; 12:593–605. [PubMed: 21109192]
58. Suganami T, Nishida J, Ogawa Y. A paracrine loop between adipocytes and macrophages aggravates inflammatory changes: role of free fatty acids and tumor necrosis factor alpha. *Arterioscler Thromb Vasc Biol.* 2005; 25:2062–8. [PubMed: 16123319]

59. Esser N, L'Homme L, De Roover A, Kohnen L, Scheen AJ, Moutschen M, et al. Obesity phenotype is related to NLRP3 inflammasome activity and immunological profile of visceral adipose tissue. *Diabetologia*. 2013; 56:2487–97. [PubMed: 24013717]
60. Juge-Aubry CE, Somm E, Chicheportiche R, Burger D, Pernin A, Cuenod-Pittet B, et al. Regulatory effects of interleukin (IL)-1, interferon-beta, and IL-4 on the production of IL-1 receptor antagonist by human adipose tissue. *J Clin Endocrinol Metab*. 2004; 89:2652–8. [PubMed: 15181037]
61. Vandanmagsar B, Youm YH, Ravussin A, Galgani JE, Stadler K, Mynatt RL, et al. The NLRP3 inflammasome instigates obesity-induced inflammation and insulin resistance. *Nat Med*. 2011; 17:179–88. [PubMed: 21217695]
62. Spranger J, Kroke A, Möhlig M, Hoffmann K, Bergmann MM, Ristow M, et al. Inflammatory cytokines and the risk to develop type 2 diabetes: results of the prospective population-based European Prospective Investigation into Cancer and Nutrition (EPIC)-Potsdam Study. *Diabetes*. 2003; 52:812–7. [PubMed: 12606524]
63. Stienstra R, Tack CJ, Kanneganti TD, Joosten LA, Netea MG. The inflammasome puts obesity in the danger zone. *Cell Metab*. 2012; 15:10–8. [PubMed: 22225872]
64. McGillicuddy FC, Harford KA, Reynolds CM, Oliver E, Claessens M, Mills KH, et al. Lack of interleukin-1 receptor I (IL-1RI) protects mice from high-fat diet-induced adipose tissue inflammation coincident with improved glucose homeostasis. *Diabetes*. 2011; 60:1688–98. [PubMed: 21515850]
65. Wen H, Gris D, Lei Y, Jha S, Zhang L, Huang MT, et al. Fatty acid-induced NLRP3-ASC inflammasome activation interferes with insulin signaling. *Nat Immunol*. 2011; 12:408–15. [PubMed: 21478880]
66. Talukdar S, Oh da Y, Bandyopadhyay G, Li D, Xu J, McNelis J, et al. Neutrophils mediate insulin resistance in mice fed a high-fat diet through secreted elastase. *Nat Med*. 2012; 18:1407–12. [PubMed: 22863787]
67. Mansuy-Aubert V, Zhou QL, Xie X, Gong Z, Huang JY, Khan AR, et al. Imbalance between neutrophil elastase and its inhibitor alpha1-antitrypsin in obesity alters insulin sensitivity, inflammation, and energy expenditure. *Cell Metab*. 2013; 17:534–48. [PubMed: 23562077]
68. Sauter NS, Schulthess FT, Galasso R, Castellani LW, Maedler K. The antiinflammatory cytokine interleukin-1 receptor antagonist protects from high-fat diet-induced hyperglycemia. *Endocrinology*. 2008; 149:2208–18. [PubMed: 18239070]
69. Lagathu C, Yvan-Charvet L, Bastard JP, Maachi M, Quignard-Boulangé A, Capeau J, et al. Long-term treatment with interleukin-1beta induces insulin resistance in murine and human adipocytes. *Diabetologia*. 2006; 49:2162–73. [PubMed: 16865359]
70. Um JY, Rim HK, Kim SJ, Kim HL, Hong SH. Functional polymorphism of IL-1 alpha and its potential role in obesity in humans and mice. *PLoS One*. 2011; 6:e29524. [PubMed: 22216303]
71. He J, Usui I, Ishizuka K, Kanatani Y, Hiratani K, Iwata M, et al. Interleukin-1alpha inhibits insulin signaling with phosphorylating insulin receptor substrate-1 on serine residues in 3T3-L1 adipocytes. *Mol Endocrinol*. 2006; 20:114–24. [PubMed: 16150868]
72. Uno T, He J, Usui I, Kanatani Y, Bukhari A, Fujisaka S, et al. Long-term interleukin-1alpha treatment inhibits insulin signaling via IL-6 production and SOCS3 expression in 3T3-L1 adipocytes. *Horm Metab Res*. 2008; 40:8–12. [PubMed: 18085494]
73. Tynan GA, Hearnden CH, Oleszycka E, Lyons CL, Coutts G, O'Connell J, et al. Endogenous Oils Derived From Human Adipocytes Are Potent Adjuvants That Promote IL-1alpha-Dependent Inflammation. *Diabetes*. 2014; 63:2037–50. [PubMed: 24458363]
74. Cinti S, Mitchell G, Barbatelli G, Murano I, Ceresi E, Faloia E, et al. Adipocyte death defines macrophage localization and function in adipose tissue of obese mice and humans. *J Lipid Res*. 2005; 46:2347–55. [PubMed: 16150820]
75. Garcia MC, Wernstedt I, Berndtsson A, Enge M, Bell M, Hultgren O, et al. Mature-onset obesity in interleukin-1 receptor I knockout mice. *Diabetes*. 2006; 55:1205–13. [PubMed: 16644674]
76. Somm E, Henrichot E, Pernin A, Juge-Aubry CE, Muzzin P, Dayer JM, et al. Decreased fat mass in interleukin-1 receptor antagonist-deficient mice: impact on adipogenesis, food intake, and energy expenditure. *Diabetes*. 2005; 54:3503–9. [PubMed: 16306368]

77. Yeh SS, Schuster MW. Geriatric cachexia: the role of cytokines. *Am J Clin Nutr.* 1999; 70:183–97. [PubMed: 10426694]
78. Luheshi GN, Gardner JD, Rushforth DA, Loudon AS, Rothwell NJ. Leptin actions on food intake and body temperature are mediated by IL-1. *Proc Natl Acad Sci U S A.* 1999; 96:7047–52. [PubMed: 10359836]
79. Dinarello CA. Interleukin-1alpha neutralisation in patients with cancer. *Lancet Oncol.* 2014; 15:552–3. [PubMed: 24746840]
80. Hong DS, Hui D, Bruera E, Janku F, Naing A, Falchook GS, et al. MABp1, a first-in-class true human antibody targeting interleukin-1alpha in refractory cancers: an open-label, phase 1 dose-escalation and expansion study. *Lancet Oncol.* 2014; 15:656–66. [PubMed: 24746841]
81. Rider P, Carmi Y, Guttman O, Braiman A, Cohen I, Voronov E, et al. IL-1alpha and IL-1beta recruit different myeloid cells and promote different stages of sterile inflammation. *J Immunol.* 2011; 187:4835–43. [PubMed: 21930960]
82. Dinarello CA, Simon A, van der Meer JW. Treating inflammation by blocking interleukin-1 in a broad spectrum of diseases. *Nat Rev Drug Discov.* 2012; 11:633–52. [PubMed: 22850787]
83. Hensen J, Howard CP, Walter V, Thuren T. Impact of interleukin-1beta antibody (canakinumab) on glycaemic indicators in patients with type 2 diabetes mellitus: results of secondary endpoints from a randomized, placebo-controlled trial. *Diabetes Metab.* 2013; 39:524–31. [PubMed: 24075453]
84. Cavelti-Weder C, Babians-Brunner A, Keller C, Stahel MA, Kurz-Levin M, Zayed H, et al. Effects of gevokizumab on glycemia and inflammatory markers in type 2 diabetes. *Diabetes Care.* 2012; 35:1654–62. [PubMed: 22699287]
85. Ridker PM, Howard CP, Walter V, Everett B, Libby P, Hensen J, et al. Effects of interleukin-1beta inhibition with canakinumab on hemoglobin A1c, lipids, C-reactive protein, interleukin-6, and fibrinogen: a phase IIb randomized, placebo-controlled trial. *Circulation.* 2012; 126:2739–48. [PubMed: 23129601]
86. Sloan-Lancaster J, Abu-Raddad E, Polzer J, Miller JW, Scherer JC, De Gaetano A, et al. Double-blind, randomized study evaluating the glycemic and anti-inflammatory effects of subcutaneous LY2189102, a neutralizing IL-1beta antibody, in patients with type 2 diabetes. *Diabetes Care.* 2013; 36:2239–46. [PubMed: 23514733]
87. Larsen CM, Faulenbach M, Vaag A, Volund A, Ehses JA, Seifert B, et al. Interleukin-1-receptor antagonist in type 2 diabetes mellitus. *N Engl J Med.* 2007; 356:1517–26. [PubMed: 17429083]
88. van Asseldonk EJ, Stienstra R, Koenen TB, Joosten LA, Netea MG, Tack CJ. Treatment with Anakinra improves disposition index but not insulin sensitivity in nondiabetic subjects with the metabolic syndrome: a randomized, double-blind, placebo-controlled study. *J Clin Endocrinol Metab.* 2011; 96:2119–26. [PubMed: 21508140]
89. Matteini AM, Li J, Lange EM, Tanaka T, Lange LA, Tracy RP, et al. Novel gene variants predict serum levels of the cytokines IL-18 and IL-1ra in older adults. *Cytokine.* 2014; 65:10–6. [PubMed: 24182552]
90. Ridker PM, Thuren T, Zalewski A, Libby P. Interleukin-1beta inhibition and the prevention of recurrent cardiovascular events: rationale and design of the Canakinumab Anti-inflammatory Thrombosis Outcomes Study (CANTOS). *Am Heart J.* 2011; 162:597–605. [PubMed: 21982649]
91. Nakamura K, Okamura H, Wada M, Nagata K, Tamura T. Endotoxin-induced serum factor that stimulates gamma interferon production. *Infect Immun.* 1989; 57:590–5. [PubMed: 2492265]
92. Okamura H, Tsutsi H, Komatsu T, Yutsudo M, Hakura A, Tanimoto T, et al. Cloning of a new cytokine that induces IFN-gamma production by T cells. *Nature.* 1995; 378:88–91. [PubMed: 7477296]
93. Nakanishi K, Yoshimoto T, Tsutsui H, Okamura H. Interleukin-18 regulates both Th1 and Th2 responses. *Annu Rev Immunol.* 2001; 19:423–74. [PubMed: 11244043]
94. Yoshimoto T, Takeda K, Tanaka T, Ohkusu K, Kashiwamura S, Okamura H, et al. IL-12 up-regulates IL-18 receptor expression on T cells, Th1 cells, and B cells: synergism with IL-18 for IFN-gamma production. *J Immunol.* 1998; 161:3400–7. [PubMed: 9759857]
95. Dao T, Mehal WZ, Crispe IN. IL-18 augments perforin-dependent cytotoxicity of liver NK-T cells. *J Immunol.* 1998; 161:2217–22. [PubMed: 9725214]

96. Micallef MJ, Ohtsuki T, Kohno K, Tanabe F, Ushio S, Namba M, et al. Interferon-gamma-inducing factor enhances T helper 1 cytokine production by stimulated human T cells: synergism with interleukin-12 for interferon-gamma production. *Eur J Immunol.* 1996; 26:1647–51. [PubMed: 8766574]
97. Thomassen E, Bird TA, Renshaw BR, Kennedy MK, Sims JE. Binding of interleukin-18 to the interleukin-1 receptor homologous receptor IL-1Rrp1 leads to activation of signaling pathways similar to those used by interleukin-1. *J Interferon Cytokine Res.* 1998; 18:1077–88. [PubMed: 9877452]
98. Seki E, Tsutsui H, Nakano H, Tsuji N, Hoshino K, Adachi O, et al. Lipopolysaccharide-induced IL-18 secretion from murine Kupffer cells independently of myeloid differentiation factor 88 that is critically involved in induction of production of IL-12 and IL-1beta. *J Immunol.* 2001; 166:2651–7. [PubMed: 11160328]
99. Novick D, Kim S, Kaplanski G, Dinarello CA. Interleukin-18, more than a Th1 cytokine. *Semin Immunol.* 2013; 25:439–48. [PubMed: 24275602]
100. Jefferis BJ, Papacosta O, Owen CG, Wannamethee SG, Humphries SE, Woodward M, et al. Interleukin 18 and coronary heart disease: prospective study and systematic review. *Atherosclerosis.* 2011; 217:227–33. [PubMed: 21481392]
101. Gerdes N, Sukhova GK, Libby P, Reynolds RS, Young JL, Schonbeck U. Expression of interleukin (IL)-18 and functional IL-18 receptor on human vascular endothelial cells, smooth muscle cells, and macrophages: implications for atherogenesis. *J Exp Med.* 2002; 195:245–57. [PubMed: 11805151]
102. Mallat Z, Heymes C, Corbaz A, Logeart D, Alouani S, Cohen-Solal A, et al. Evidence for altered interleukin 18 (IL)-18 pathway in human heart failure. *FASEB J.* 2004; 18:1752–4. [PubMed: 15371332]
103. de Nooijer R, von der Thusen JH, Verkleij CJ, Kuiper J, Jukema JW, van der Wall EE, et al. Overexpression of IL-18 decreases intimal collagen content and promotes a vulnerable plaque phenotype in apolipoprotein-E-deficient mice. *Arterioscler Thromb Vasc Biol.* 2004; 24:2313–9. [PubMed: 15472128]
104. Whitman SC, Ravisankar P, Daugherty A. Interleukin-18 enhances atherosclerosis in apolipoprotein E(-/-) mice through release of interferon-gamma. *Circ Res.* 2002; 90:E34–8. [PubMed: 11834721]
105. Tenger C, Sundborger A, Jawien J, Zhou X. IL-18 accelerates atherosclerosis accompanied by elevation of IFN-gamma and CXCL16 expression independently of T cells. *Arterioscler Thromb Vasc Biol.* 2005; 25:791–6. [PubMed: 15604417]
106. Elhage R, Jawien J, Rudling M, Ljunggren HG, Takeda K, Akira S, et al. Reduced atherosclerosis in interleukin-18 deficient apolipoprotein E-knockout mice. *Cardiovasc Res.* 2003; 59:234–40. [PubMed: 12829194]
107. McLaren JE, Ramji DP. Interferon gamma: a master regulator of atherosclerosis. *Cytokine Growth Factor Rev.* 2009; 20:125–35. [PubMed: 19041276]
108. Escobar-Morreale HF, Botella-Carretero JI, Villuendas G, Sancho J, San Millan JL. Serum interleukin-18 concentrations are increased in the polycystic ovary syndrome: relationship to insulin resistance and to obesity. *J Clin Endocrinol Metab.* 2004; 89:806–11. [PubMed: 14764799]
109. Moriwaki Y, Yamamoto T, Shibutani Y, Aoki E, Tsutsumi Z, Takahashi S, et al. Elevated levels of interleukin-18 and tumor necrosis factor-alpha in serum of patients with type 2 diabetes mellitus: relationship with diabetic nephropathy. *Metabolism.* 2003; 52:605–8. [PubMed: 12759891]
110. Evans J, Collins M, Jennings C, van der Merwe L, Soderstrom I, Olsson T, et al. The association of interleukin-18 genotype and serum levels with metabolic risk factors for cardiovascular disease. *Eur J Endocrinol.* 2007; 157:633–40. [PubMed: 17984243]
111. Huang Y, Xu M, Hong J, Gu W, Bi Y, Li X. -607 C/A polymorphism in the promoter of IL-18 gene is associated with 2 h post-loading plasma glucose level in Chinese. *Endocrine.* 2010; 37:507–12. [PubMed: 20960175]

112. Schernthaner GH, Kopp HP, Kriwanek S, Krzyzanowska K, Satler M, Koppensteiner R, et al. Effect of massive weight loss induced by bariatric surgery on serum levels of interleukin-18 and monocyte-chemoattractant-protein-1 in morbid obesity. *Obes Surg.* 2006; 16:709–15. [PubMed: 16756729]
113. Smart MC, Dedoussis G, Yiannakouris N, Grisoni ML, Dror GK, Yannakoulia M, et al. Genetic variation within IL18 is associated with insulin levels, insulin resistance and postprandial measures. *Nutr Metab Cardiovasc Dis.* 2011; 21:476–84. [PubMed: 20227263]
114. Presta I, Andreozzi F, Succurro E, Marini MA, Laratta E, Lauro R, et al. IL-18 gene polymorphism and metabolic syndrome. *Nutr Metab Cardiovasc Dis.* 2009; 19:e5–6. [PubMed: 19176284]
115. Thompson SR, Sanders J, Stephens JW, Miller GJ, Humphries SE. A common interleukin 18 haplotype is associated with higher body mass index in subjects with diabetes and coronary heart disease. *Metabolism.* 2007; 56:662–9. [PubMed: 17445542]
116. Skurk T, Kolb H, Muller-Scholze S, Rohrig K, Hauner H, Herder C. The proatherogenic cytokine interleukin-18 is secreted by human adipocytes. *Eur J Endocrinol.* 2005; 152:863–8. [PubMed: 15941925]
117. Leick L, Lindegaard B, Stensvold D, Plomgaard P, Saltin B, Pilegaard H. Adipose tissue interleukin-18 mRNA and plasma interleukin-18: effect of obesity and exercise. *Obesity (Silver Spring).* 2007; 15:356–63. [PubMed: 17299108]
118. Bruun JM, Stallknecht B, Helge JW, Richelsen B. Interleukin-18 in plasma and adipose tissue: effects of obesity, insulin resistance, and weight loss. *Eur J Endocrinol.* 2007; 157:465–71. [PubMed: 17893261]
119. Fischer CP, Perstrup LB, Berntsen A, Eskildsen P, Pedersen BK. Elevated plasma interleukin-18 is a marker of insulin-resistance in type 2 diabetic and non-diabetic humans. *Clin Immunol.* 2005; 117:152–60. [PubMed: 16112617]
120. Zorrilla EP, Sanchez-Alavez M, Sugama S, Brennan M, Fernandez R, Bartfai T, et al. Interleukin-18 controls energy homeostasis by suppressing appetite and feed efficiency. *Proc Natl Acad Sci U S A.* 2007; 104:11097–102. [PubMed: 17578927]
121. Membrez M, Ammon-Zufferey C, Philippe D, Aprikian O, Monnard I, Mace K, et al. Interleukin-18 protein level is upregulated in adipose tissue of obese mice. *Obesity (Silver Spring).* 2009; 17:393–5. [PubMed: 19039317]
122. Yang YS, Li XY, Hong J, Gu WQ, Zhang YF, Yang J, et al. Interleukin-18 enhances glucose uptake in 3T3-L1 adipocytes. *Endocrine.* 2007; 32:297–302. [PubMed: 18247160]
123. Carey AL, Steinberg GR, Macaulay SL, Thomas WG, Holmes AG, Ram m G, et al. Interleukin-6 increases insulin-stimulated glucose disposal in humans and glucose uptake and fatty acid oxidation in vitro via AMP-activated protein kinase. *Diabetes.* 2006; 55:2688–97. [PubMed: 17003332]
124. Minokoshi Y, Kim YB, Peroni OD, Fryer LG, Muller C, Carling D, et al. Leptin stimulates fatty-acid oxidation by activating AMP-activated protein kinase. *Nature.* 2002; 415:339–43. [PubMed: 11797013]
125. Watt MJ, Dzamko N, Thomas WG, Rose-John S, Ernst M, Carling D, et al. CNTF reverses obesity-induced insulin resistance by activating skeletal muscle AMPK. *Nat Med.* 2006; 12:541–8. [PubMed: 16604088]
126. Lindegaard B, Matthews VB, Brandt C, Hojman P, Allen TL, Estevez E, et al. Interleukin-18 activates skeletal muscle AMPK and reduces weight gain and insulin resistance in mice. *Diabetes.* 2013
127. Zilverschoon GR, Tack CJ, Joosten LA, Kullberg BJ, van der Meer JW, Netea MG. Interleukin-18 resistance in patients with obesity and type 2 diabetes mellitus. *Int J Obes (Lond).* 2008; 32:1407–14. [PubMed: 18645574]
128. Schmitz J, Owyang A, Oldham E, Song Y, Murphy E, McClanahan TK, et al. IL-33, an interleukin-1-like cytokine that signals via the IL-1 receptor-related protein ST2 and induces T helper type 2-associated cytokines. *Immunity.* 2005; 23:479–90. [PubMed: 16286016]
129. Iwahana H, Yanagisawa K, Ito-Kosaka A, Kuroiwa K, Tago K, Komatsu N, et al. Different promoter usage and multiple transcription initiation sites of the interleukin-1 receptor-related

- human ST2 gene in UT-7 and TM12 cells. *Eur J Biochem.* 1999; 264:397–406. [PubMed: 10491084]
130. Bae S, Kang T, Hong J, Lee S, Choi J, Jhun H, et al. Contradictory functions (activation/termination) of neutrophil proteinase 3 enzyme (PR3) in interleukin-33 biological activity. *J Biol Chem.* 2012; 287:8205–13. [PubMed: 22270365]
131. Lefrancais E, Roga S, Gautier V, Gonzalez-de-Peredo A, Monsarrat B, Girard JP, et al. IL-33 is processed into mature bioactive forms by neutrophil elastase and cathepsin G. *Proc Natl Acad Sci U S A.* 2012; 109:1673–8. [PubMed: 22307629]
132. Martin MU. Special aspects of interleukin-33 and the IL-33 receptor complex. *Semin Immunol.* 2013; 25:449–57. [PubMed: 24230466]
133. Mirchandani AS, Salmond RJ, Liew FY. Interleukin-33 and the function of innate lymphoid cells. *Trends Immunol.* 2012; 33:389–96. [PubMed: 22609147]
134. Oboki K, Ohno T, Kajiwara N, Arae K, Morita H, Ishii A, et al. IL-33 is a crucial amplifier of innate rather than acquired immunity. *Proc Natl Acad Sci U S A.* 2010; 107:18581–6. [PubMed: 20937871]
135. Xu D, Jiang HR, Li Y, Pushparaj PN, Kurowska-Stolarska M, Leung BP, et al. IL-33 exacerbates autoantibody-induced arthritis. *J Immunol.* 2010; 184:2620–6. [PubMed: 20139274]
136. Stolarski B, Kurowska-Stolarska M, Kewin P, Xu D, Liew FY. IL-33 exacerbates eosinophil-mediated airway inflammation. *J Immunol.* 2010; 185:3472–80. [PubMed: 20693421]
137. Verri WA Jr, Souto FO, Vieira SM, Almeida SC, Fukada SY, Xu D, et al. IL-33 induces neutrophil migration in rheumatoid arthritis and is a target of anti-TNF therapy. *Ann Rheum Dis.* 2010; 69:1697–703. [PubMed: 20472598]
138. Liew FY. IL-33: a Janus cytokine. *Ann Rheum Dis.* 2012; 71(Suppl 2):i101–4. [PubMed: 22460136]
139. Pei C, Barbour M, Fairlie-Clarke KJ, Allan D, Mu R, Jiang HR. Emerging role of interleukin-33 in autoimmune diseases. *Immunology.* 2014; 141:9–17. [PubMed: 24116703]
140. Palmer G, Gabay C. Interleukin-33 biology with potential insights into human diseases. *Nat Rev Rheumatol.* 2011; 7:321–9. [PubMed: 21519352]
141. Alves-Filho JC, Sonogo F, Souto FO, Freitas A, Verri WA Jr, Auxiliado ra-Martins M, et al. Interleukin-33 attenuates sepsis by enhancing neutrophil influx to the site of infection. *Nat Med.* 2010; 16:708–12. [PubMed: 20473304]
142. Ohno T, Morita H, Arae K, Matsumoto K, Nakae S. Interleukin-33 in allergy. *Allergy.* 2012; 67:1203–14. [PubMed: 22913600]
143. Miller AM, Xu D, Asquith DL, Denby L, Li Y, Sattar N, et al. IL-33 reduces the development of atherosclerosis. *J Exp Med.* 2008; 205:339–46. [PubMed: 18268038]
144. Wood IS, Wang B, Trayhurn P. IL-33, a recently identified interleukin-1 gene family member, is expressed in human adipocytes. *Biochem Biophys Res Commun.* 2009; 384:105–9. [PubMed: 19393621]
145. Zeyda M, Wernly B, Demyanets S, Kaun C, Hammerle M, Hantusch B, et al. Severe obesity increases adipose tissue expression of interleukin-33 and its receptor ST2, both predominantly detectable in endothelial cells of human adipose tissue. *Int J Obes (Lond).* 2012
146. Hasan A, Al-Ghimlas F, Warsame S, Al-Hubail A, Ahmad R, Bennakhi A, et al. IL-33 is negatively associated with the BMI and confers a protective lipid/metabolic profile in non-diabetic but not diabetic subjects. *BMC Immunol.* 2014; 15:19. [PubMed: 24886535]
147. Miller AM, Asquith DL, Hueber AJ, Anderson LA, Holmes WM, McKenzie AN, et al. Interleukin-33 induces protective effects in adipose tissue inflammation during obesity in mice. *Circ Res.* 2010; 107:650–8. [PubMed: 20634488]
148. Qiu Y, Nguyen KD, Odegaard JI, Cui X, Tian X, Locksley RM, et al. Eosinophils and type 2 cytokine signaling in macrophages orchestrate development of functional beige fat. *Cell.* 2014; 157:1292–308. [PubMed: 24906148]
149. Lee MW, Odegaard JI, Mukundan L, Qiu Y, Molofsky AB, Nussbaum JC, et al. Activated Type 2 Innate Lymphoid Cells Regulate Beige Fat Biogenesis. *Cell.* 2015; 160:74–87. [PubMed: 25543153]

150. Nold MF, Nold-Petry CA, Zepp JA, Palmer BE, Bufler P, Dinarello CA. IL-37 is a fundamental inhibitor of innate immunity. *Nat Immunol.* 2011; 11:1014–22. [PubMed: 20935647]
151. Dinarello CA, Bufler P. Interleukin-37. *Semin Immunol.* 2013; 25:466–8. [PubMed: 24275599]
152. Li S, Neff CP, Barber K, Hong J, Luo Y, Azam T, et al. Extracellular forms of IL-37 inhibit innate inflammation in vitro and in vivo but require the IL-1 family decoy receptor IL-1R8. *Proc Natl Acad Sci U S A.* 2015
153. Wald D, Qin J, Zhao Z, Qian Y, Naramura M, Tian L, et al. SIGIRR, a negative regulator of Toll-like receptor-interleukin 1 receptor signaling. *Nat Immunol.* 2003; 4:920–7. [PubMed: 12925853]
154. Qin J, Qian Y, Yao J, Grace C, Li X. SIGIRR inhibits interleukin-1 receptor-and toll-like receptor 4-mediated signaling through different mechanisms. *J Biol Chem.* 2005; 280:25233–41. [PubMed: 15866876]
155. Garlanda C, Riva F, Bonavita E, Mantovani A. Negative regulatory receptors of the IL-1 family. *Semin Immunol.* 2013; 25:408–15. [PubMed: 24239046]
156. Sharma S, Kulk N, Nold MF, Graf R, Kim SH, Reinhardt D, et al. The IL-1 family member 7b translocates to the nucleus and down-regulates proinflammatory cytokines. *J Immunol.* 2008; 180:5477–82. [PubMed: 18390730]
157. Bulau AM, Nold MF, Li S, Nold-Petry CA, Fink M, Mansell A, et al. Role of caspase-1 in nuclear translocation of IL-37, release of the cytokine, and IL-37 inhibition of innate immune responses. *Proc Natl Acad Sci U S A.* 2014
158. Li S, H J, Nold MF, Nold-Petry PA, Azam T, Garlanda C, Bufler P, Kim SH, Mantovani A, Dinarello CA. Recombinant IL-37 inhibits LPS induced inflammation in a SIGIRR- and MAPK-dependent manner. *Cytokine.* 2013; 63:281.
159. Ji Q, Zeng Q, Huang Y, Shi Y, Lin Y, Lu Z, et al. Elevated plasma IL-37, IL-18, and IL-18BP concentrations in patients with acute coronary syndrome. *Mediators Inflamm.* 2014; 2014:165742. [PubMed: 24733959]
160. Pei B, Xu S, Liu T, Pan F, Xu J, Ding C. Associations of the IL-1F7 gene polymorphisms with rheumatoid arthritis in Chinese Han population. *Int J Immunogenet.* 2013; 40:199–203. [PubMed: 23171316]
161. Hardie DG, Ross FA, Hawley SA. AMPK: a nutrient and energy sensor that maintains energy homeostasis. *Nat Rev Mol Cell Biol.* 2012; 13:251–62. [PubMed: 22436748]
162. Ceddia RB. The role of AMP-activated protein kinase in regulating white adipose tissue metabolism. *Mol Cell Endocrinol.* 2013; 366:194–203. [PubMed: 22750051]
163. O'Neill LA, Hardie DG. Metabolism of inflammation limited by AMPK and pseudo-starvation. *Nature.* 2013; 493:346–55. [PubMed: 23325217]
164. Dunn E, Sims JE, Nicklin MJ, O'Neill LA. Annotating genes with potential roles in the immune system: six new members of the IL-1 family. *Trends Immunol.* 2001; 22:533–6. [PubMed: 11574261]
165. Kumar S, McDonnell PC, Lehr R, Tierney L, Tzimas MN, Griswold DE, et al. Identification and initial characterization of four novel members of the interleukin-1 family. *J Biol Chem.* 2000; 275:10308–14. [PubMed: 10744718]
166. Smith DE, Renshaw BR, Ketchem RR, Kubin M, Garka KE, Sims JE. Four new members expand the interleukin-1 superfamily. *J Biol Chem.* 2000; 275:1169–75. [PubMed: 10625660]
167. Barton JL, Herbst R, Bosisio D, Higgins L, Nicklin MJ. A tissue specific IL-1 receptor antagonist homolog from the IL-1 cluster lacks IL-1, IL-1ra, IL-18 and IL-18 antagonist activities. *Eur J Immunol.* 2000; 30:3299–308. [PubMed: 11093146]
168. Busfield SJ, Comrack CA, Yu G, Chickering TW, Smutko JS, Zhou H, et al. Identification and gene organization of three novel members of the IL-1 family on human chromosome 2. *Genomics.* 2000; 66:213–6. [PubMed: 10860666]
169. Mulero JJ, Pace AM, Nelken ST, Loeb DB, Correa TR, Drmanac R, et al. IL1HY1: A novel interleukin-1 receptor antagonist gene. *Biochem Biophys Res Commun.* 1999; 263:702–6. [PubMed: 10512743]

170. Towne JE, Renshaw BR, Douangpanya J, Lipsky BP, Shen M, Gabel CA, et al. Interleukin-36 (IL-36) ligands require processing for full agonist (IL-36alpha, IL-36beta, and IL-36gamma) or antagonist (IL-36Ra) activity. *J Biol Chem.* 2011; 286:42594–602. [PubMed: 21965679]
171. Bensen JT, Dawson PA, Mychaleckyj JC, Bowden DW. Identification of a novel human cytokine gene in the interleukin gene cluster on chromosome 2q12-14. *J Interferon Cytokine Res.* 2001; 21:899–904. [PubMed: 11747621]
172. Lin H, Ho AS, Haley-Vicente D, Zhang J, Bernal-Fussell J, Pace AM, et al. Cloning and characterization of IL-1HY2, a novel interleukin-1 family member. *J Biol Chem.* 2001; 276:20597–602. [PubMed: 11278614]
173. van Asseldonk EJ, Stienstra R, Koenen TB, van Tits LJ, Joosten LA, Tack CJ, et al. The Effect of the Interleukin-1 Cytokine Family Members IL-1F6 and IL-1F8 on Adipocyte Differentiation. *Obesity (Silver Spring).* 2010
174. Ohsumi J, Sakakibara S, Yamaguchi J, Miyadai K, Yoshioka S, Fujiwara T, et al. Troglitazone prevents the inhibitory effects of inflammatory cytokines on insulin-induced adipocyte differentiation in 3T3-L1 cells. *Endocrinology.* 1994; 135:2279–82. [PubMed: 7956951]
175. Dehghan A, Dupuis J, Barbalic M, Bis JC, Eiriksdottir G, Lu C, et al. Meta-analysis of genome-wide association studies in >80 000 subjects identifies multiple loci for C-reactive protein levels. *Circulation.* 2011; 123:731–8. [PubMed: 21300955]
176. Herder C, Nuotio ML, Shah S, Blankenberg S, Brunner EJ, Carstensen M, et al. Genetic determinants of circulating interleukin-1 receptor antagonist levels and their association with glycemic traits. *Diabetes.* 2014

Highlights

- Cytokines of the IL-1 family play an important role in metabolic inflammation
- The development of obesity affects IL-1 α , IL-1 β , IL-1Ra, IL-18, IL-33 and IL-37 activity
- IL-1 α , IL-1 β , IL-1Ra, IL-18, IL-33 and IL-37 alter insulin signaling routes
- The IL-1 family of cytokines holds multiple targets to combat metabolic diseases

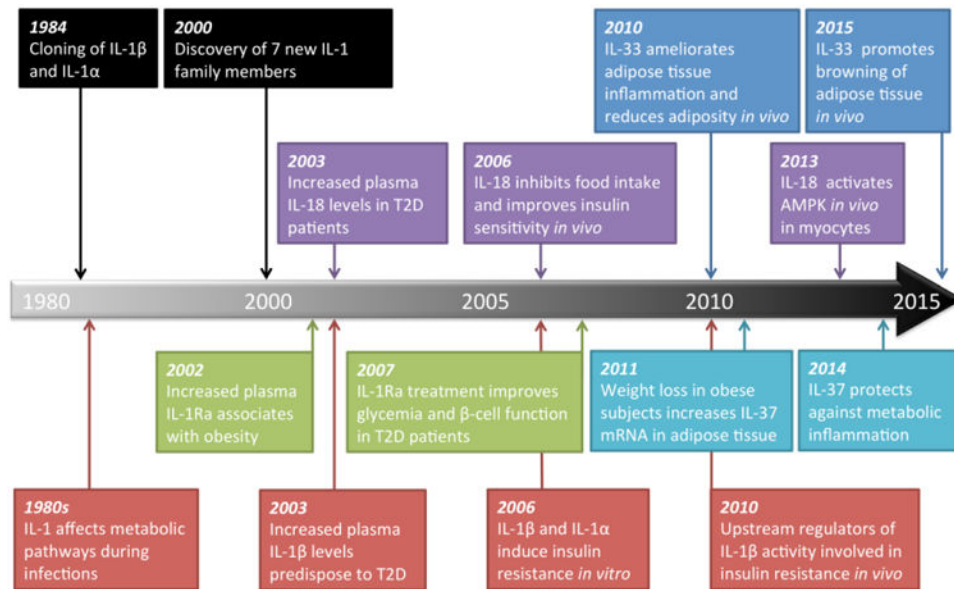


Figure 1. Timeline of the discovery and specific contribution of IL-1 family members to the development of obesity, inflammation and insulin resistance.

Table 1
Metabolic and inflammatory effects of the IL-1 family of cytokines

Cytokine	Primary Property	Metabolic effects (human)	Metabolic effects (animal) ¹	Mechanism of action
IL-1α	<i>Pro-inflammatory</i>	↑ circulating levels in obesity	↑ plasma TG levels	Inhibits insulin receptor signalling and induces inflammatory gene transcription
IL-1β	<i>Pro-inflammatory</i>	SNPs associated with obesity and T2D ↑ circulating levels augment risk to develop T2D	↓ insulin sensitivity (<i>in vitro</i>) ↓ adipogenesis	Inhibits insulin receptor signalling and induces inflammatory gene transcription
IL-1Ra	<i>Antagonist / Anti-inflammatory</i>	SNPs associated with obesity and T2D ↑ (β-cell function & insulin secretion)	↓ insulin sensitivity ↑ insulin sensitivity	Blocks receptor and prevents IL-1 α and IL-1 β activities
IL-18	<i>Pro-inflammatory</i>	↔ insulin sensitivity SNPs associated with obesity and T2D ↑ circulating levels in obese, T2D patients	↑ increases appetite ↓ food intake	activates AMPK
IL-33	<i>Pro-inflammatory</i>	levels associate with insulin resistance SNPs associated with obesity and T2D ↔ circulating levels in obese	↓ obesity ↑ insulin sensitivity ↓ adipogenesis	reduces food intake increases atherosclerosis development promotes browning of adipose tissue
IL-37	<i>Anti-inflammatory</i>	Adipose tissue mRNA associates with enhanced insulin sensitivity ↑ mRNA in adipose tissue by weight loss in morbidly obese subjects	↓ fasting glucose ↓ adipogenesis	activates AMPK
IL-36α/β/γ	<i>Pro-inflammatory</i>	unknown	↑ insulin sensitivity	inhibits adipogenesis <i>in vitro</i>
IL-36Ra	<i>Antagonist / Anti-inflammatory</i>	unknown	unknown	
IL-38	<i>Antagonist?</i>	IL-38 SNP associates with ↓ insulin and ↓ HOMA-IR	unknown	

Italic font indicates evidence obtained from knock-out or transgenic models. **Bold font** indicates evidence obtained from experiments with recombinant molecules.