

HHS Public Access

Author manuscript

Inorg Chem. Author manuscript; available in PMC 2015 September 01.

Published in final edited form as:

Inorg Chem. 2011 October 3; 50(19): 9213–9215. doi:10.1021/ic201615u.

[Ru(bpy)2(5CNU)2] 2+ as a Potential Dual Action PDT Agent

Robert N. Garner¥, **Judith C. Gallucci**¥, **Kim R. Dunbar**†,*, and **Claudia Turro**¥,* ¥Department of Chemistry, The Ohio State University, Columbus, OH 43210, USA

†Department of Chemistry, Texas A&M University, College Station, TX 77842, USA

Abstract

The cation *cis*-[$Ru(bpy)_{2}(5CNU)_{2}$]²⁺ (bpy = 2,2'-bipyridine, 5CNU = 5-cyanouracil) was synthesized and investigated for use as a potential dual-action photodynamic therapy agent. The complex undergoes efficient photoinduced 5CNU ligand exchange for solvent water molecules, thus simultaneously releasing biologically active 5CNU and generating $\text{[Ru(bpy)}_2(\text{H}_2\text{O})_2\text{]}^{2+}$. The latter binds covalently to ds-DNA, such that photolysis results in the generation of three potential therapeutic agents from a single molecule.

> The chemotherapy agent 5-flurouracil (5FU) has been in use for over 20 years for the treatment of malignancies including colorectal and breast cancers.¹ The mode of action of 5FU is multifaceted and involves its incorporation into DNA and RNA, as well as inhibition of thymidylate synthase, an enzyme important in DNA synthesis and repair.¹ Other derivitives of uracil also exhibit cellular activity including 5-cyanouracil (5CNU), which has been shown to inhibit pyrimidine catabolism *in vivo*. 2 Complexation of 5CNU to a photoactive transition metal complex represents a manner in which this drug may be caged in an inactive state until delivered to its target, at which time it can be released by irradiation with low energy light.

> In addition to the photoinduced release of the caged 5CNU drug, the transition metal portion of the complex itself may be biologically active. It has been shown by us and others that ruthenium and dirhodium complexes hold great potential for photodynamic thereapy (PDT) .³⁻⁶ In addition, we have also reported photoactive complexes inspired by the mode of action of cisplatin; in these cases, the complex is stable in the dark but undergoes ligand exchange and binds to DNA upon irradiation.⁷ These complexes include *cis*- $[Ru(bpy)₂(NH₃)₂]²⁺ (by = 2,2'-bipyridine)$ and *cis*- $[Rh₂(O₂ CCH₃)₂(CH₃CN)₆]²⁺,⁷$ where a 34-fold increase in toxicity towards Hs27 human skin fibroblasts was reported for the latter upon irradiation with visible light.^{7b} It should also be noted that ruthenium complexes are now recognized as promising anticancer drugs, with some being subjected to clinical and pre-clinical trials.⁸

^{*}Corresponding Author: To whom correspondence should be addressed: turro@chemistry.ohio-state.edu, dunbar@mail.chem.tamu.edu.

Supporting Information. Synthesis details, complete crystal structure and cyrstalligraphic data, and dark activity. This material is available free of charge via the Internet at <http://pubs.acs.org>

Nitriles undergo photoinduced ligand exchange more efficiently than other monodentate ligands when bound to Ru(II), includig pyridine and $NH₃$,^{9,10} As such, complexes with various nitrile ligands are promising PDT candidates. In this vein, we hypothesized that two 5CNU molecules coordinated to ruthenium through their nitrile substituents would efficiently undergo ligand exchange with solvent upon irradiation, thus releasing two equivalents of the drug and the diaqua Ru(II) complex. The present work focuses on the synthesis of the new complex cis -[Ru(bpy)₂(5CNU)₂]²⁺ (1) and the investigation of its photophysical properties and potential for its use as a dual-action photochemotherapeutic agent.

The reaction of 50 mg (0.1 mmol) of *cis*-Ru(bpy)₂Cl₂ with 2.2 eq AgCF₃SO₃ in 10 mL methanol at room temperature resulted in the precipation of AgCl and the formation of red $cis-Ru(bpy)_{2}(CF_{3}SO_{3})_{2}$. After filtration to remove AgCl, the solution was refluxed with a 5fold excess of 5CNU for 2 h to generate the yellow complex **1** (Supporting Information). The 1H NMR spectrum of **1** is consistent with two 5CNU molecules and two bpy ligands coordinated to the metal center (Supporting Information), also evidenced in the crystal structure of the product shown in Figure 1a. The Ru–N bond lengths to the 5CNU ligand are $2.034(3)$ and $2.025(3)$ Å, while those to the bpy nitrogen atoms positioned trans to $5CNU$ are 2.044(3) Å. Slightly longer Ru-N bonds were found for the two bpy nitrogen atoms positioned trans to another bpy ligand, 2.061(3) and 2.067(3) Å. These distances are similar to those previously reported for *cis*-[Ru(bpy)₂(CH₃CN)₂]²⁺ (**2**).¹¹

The Ru^{III/II} couple of 1 is observed at +1.45 V *vs* SCE in CH₃CN (0.1 M Bu₄NPF₆), at a similar potential to that measured for 2 at $+1.44$ *vs* SCE,^{12,13} and related Ru(II) complexes.14,15 Two high energy transitions are observed in the electronic absorption spectrum of the chloride salt of **1** in water at 243 nm ($\varepsilon = 31,000 \text{ M}^{-1} \text{ cm}^{-1}$) and 284 nm (ε $= 48300 \text{ M}^{-1} \text{ cm}^{-1}$) assigned as ligand-centered (LC) $\pi \pi^*$ transitions of bpy, with maxima at 240 nm ($\varepsilon = 17$ 400 M⁻¹ cm⁻¹) and 283 nm ($\varepsilon = 52$ 500 M⁻¹ cm⁻¹) in **2** in CH₃CN. The shoulder at 315 nm ($\varepsilon = 21,300 \text{ M}^{-1} \text{ cm}^{-1}$) in **1** is not present in the absorption spectrum of **2** or in free 5CNU ($\lambda_{\text{max}} = 275$ nm, $\varepsilon = 11,700 \text{ M}^{-1} \text{ cm}^{-1}$ in water), and can therefore be assigned to a metal-to-ligand charge transfer (MLCT) transition from the Ru(II) center to the 5CNU ligand. The absorption maximum at 410 nm (ε = 7 800 M⁻¹ cm⁻¹) in **1** (H₂O) is consistent with the Ru→bpy MLCT transition observed at 425 nm ($\varepsilon = 8590$ M⁻¹ cm⁻¹) in 2 in CH₃CN.¹⁶

Density functional theory (DFT) calculations were perfromed on **1** in CH3CN. The minimized structure from the calculations is consistent with the crystal structure, as is evident from the Ru–N bond lenghts to the 5CNU ligands calculated to be 2.038 Å and shorter than the other Ru–N bonds in the complex (Table S2, Supporting Information). Moreover, the calculated orbital energies support the assignments of the electronic absorption transitions (details in Supporting Informatin). The molecular orbital (MO) diagram calculated for **1** is shown in Figure 1b. The electron density of the three highest occupied molecular orbitals (HOMOs) is localized on the metal center; these MOs are labeled $Ru(d\pi)$ in Figure 1b. The LUMO (lowest unoccupied molecular orbital) and LUMO +1 are bpy π^* orbitals, while the electron densities of the LUMO+2 and LUMO+3 are localized on the 5CNU ligands. Accordingly, inspection of Figure 1b reveals that Ru→bpy,

followed by Ru→5CNU, are expected to be the lowest energy transitions in **1**. It should also be noted that the Ru(σ*) orbitals lie at signficantly greater energies, ∼5.5 eV above the HOMO, and comprise the LUMO+10 and LUMO+11 (Figure 1).

Irradiation of the chloride salt of 1 in water with visible light $(\lambda_{irr} < 395 \text{ nm})$ results in spectral changes, with a decrease in the MLCT absorption at 410 nm, as shown in Figure 2. A species with a maximum at ∼450 nm forms within 2 min of irradiation (Figure 2, inset, bold line), attributed to the mono-aqua intermediate *cis*-[Ru(bpy)₂(5CNU)(H₂O)]²⁺ (3), similar to that previously reported for the photochemistry of **2**. ¹⁰ This peak deceases in intensity with continued photolysis with a concomitant increase of the product peak at 490 nm (Figure 2). The peak at 490 nm ($\varepsilon = 9300 \text{ M}^{-1} \text{ cm}^{-1}$) is known to correspond to the diaqua complex cis -[Ru(bpy)₂(H₂O)₂]²⁺ (4),¹⁷ consistent with the photoinduced exchange of each 5CNU ligand with a solvent water molelcule. The quantum yield for the disappearance of 1 to generate the mono-aqua intermediate 3 was measured to be 0.16 ± 4 ($\lambda_{rr} = 400$ nm, see Supporting Information for details), a slighly lower value than that reported for $2 (\Phi =$ 0.21, λ_{irr} = 400 nm).¹⁰ It should be noted that no spectral changes are observed when the sample is kept in the dark under similar experimental conditions.

When **1** (80 μM) is photolyzed for 90 s (Supporting Information), the majority of the product formed is the mono-aqua complex, **3**, with a maximum at ∼450 nm. If that solution is then stored in the dark, no further reactivity is observed (Figure S2), a fact that is indicative of the requirement of a photon for the conversion of **3** to the final product, **4**. This experiment clearly supports the conclusion that the formation of the bis-aqua product **4** from **1** requires the sequential absorption of two photons, one to effect the exchange of each 5CNU ligand.

The photolysis of 1 with visible light was also monitored by ¹H NMR spectroscopy in D₂O. Prior to irradiation, a peak corresponding to the C–H ring proton of the two bound 5CNU molecules is observed at 8.02 ppm. This peak decreases in intensity upon irradiation with the appearance of two new resonances at 8.00 ppm and 8.18 ppm, assigned to the same 5CNU proton in *cis*-[Ru(bpy)₂(5CNU)(D₂O)]²⁺ (3a) and free 5CNU in D₂O, respectively. Continued photolysis results in further decrease of the 5CNU peak corresponding to **3a** and greater intensity of the free 5CNU peak, consistent with the formation of the bis-aqua product. Spectral changes are also observed for the bpy protons, but overlapping resonances of the reactant, intermediate and product preclude assignments to be made.

The covalent binding of cisplatin to DNA results in decreased mobility of linearized plasmid in agrose gel electrophoresis. $7,18$ Our group has shown that certain complexes are capable of covalently binding to DNA when activated by light.⁷ Agrose mobility shift gels were conducted in order to test the ability of **1** to bind to DNA when irradiated. Photolysis of **1** (λ_{rr} = 395 nm, t_{irr} = 15 min) in the presence of linearized pUC18 plasmid results in a decrease of mobility of the ds-DNA as a function of increased complex concentration (Figure 3a).19 In contrast, when the plasmid is incubated in the dark for 20 min no change in the DNA mobility is observed (Figure 3b).

The work presented here demonstrates that, upon photolysis of **1** in aqueous media, complex **4** is formed which subsequently covelently binds to DNA. Moreover, two equivalents of the biologically active 5CNU molecule are released in the process, rendering this type of complex a potential dual-action photochemotherapy agent.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

C.T. and K.R.D. thank the National Science Foundation (CHE 0911354) for partial funding of this work and C.T. thanks the Ohio Supercomputer Center for their generous support. R.N.G. thanks the National Institutes of Health for a Ruth L. Kirschstein National Research Service Award/MARC Fellowship (GM 833552).

References

- 1. Longley DB, Harkin DP, Johnston PG. Nature Reviews Cancer. 2003; 3:330–338. [PubMed: 12724731]
- 2. (a) Gentry GA, Morse PA, Dorsett MT. Cancer Research. 1971; 31:909–912. [PubMed: 5088491] (b) Porter DJT, Chestnut WG, Merrill BM, Spector T. J Biol Chem. 1992; 267:5236–5242. [PubMed: 1544906]
- 3. (a) Angeles-Boza AM, Bradley PM, Fu PKL, Wicke SE, Bacsa J, Dunbar KR, Turro C. Inorg Chem. 2004; 43:8510–8519. [PubMed: 15606200] (b) Bradley PM, Angeles-Boza AM, Dunbar KR, Turro C. Inorg Chem. 2004; 43:2450–2452. [PubMed: 15074956] (c) Angeles-Boza AM, Bradley PM, Fu PKL, Shatruk M, Hilfiger MG, Dunbar KR, Turro C. Inorg Chem. 2005; 44:7262–7264. [PubMed: 16212337] (d) Aguirre JD, Angeles-Boza AM, Chouai A, Pellois JP, Turro C, Dunbar KR. J Am Chem Soc. 2009; 131:11353–11360. [PubMed: 19624128] (e) Joyce LE, Aguirre JD, Angeles-Boza AM, Chouai A, Fu PKL, Dunbar KR, Turro C. Inorg Chem. 2010; 49:5371–5376. [PubMed: 20496907]
- 4. (a) Liu Y, Hammitt R, Lutterman DA, Joyce LE, Thummel RP, Turro C. Inorg Chem. 2009; 48:375–385. [PubMed: 19035764] (b) Zhao R, Hammitt R, Thummel RP, Liu Y, Turro C, Snapka RM. Dalton Trans. 2009; 48:10926–10931. [PubMed: 20023923] (c) Sun Y, Joyce LE, Dickson NM, Turro C. Chem Comm. 2010; 46:2426–2428. [PubMed: 20379547] (d) Sun Y, Joyce LE, Dickson NM, Turro C. Chem Comm. 2010; 46:6759–6761. [PubMed: 20717583]
- 5. (a) Jain A, Wang J, Mashack ER, Winkel BSJ, Brewer K. J Inorg Chem. 2009; 48:9077–9084.(b) Prussin AJ II, Zhao S, Jain A, Winkel BSJ, Brewer KJ. J Inorg Bio-chem. 2009; 103:427–431.(c) Higgins SLH, White TA, Winkel BSJ, Brewer K. J Inorg Chem. 2011; 50:463–470.
- 6. (a) Farrer NJ, Salassa L, Sadler P. J Dalton Trans. 2009; 48:10690–10701.(b) Salassa L, Ruiu T, Garino C, Pizarro AM, Bardelli F, Gianolio D, Westendorf A, Bednarski PJ, Lamberti C, Gobetto R, Sadler PJ. Organometallics. 2010; 29:6703–6710.(c) Farrer NJ, Woods JA, Salassa L, Zhao Y, Robinson KS, Clarkson G, Mackay FS, Sadler PJ. Angew Chemie, Int Ed. 2010; 49:8905–8908.(d) Farrer NJ, Woods JA, Munk VP, Mackay FS, Sadler P. J Chem Res Tox. 2010; 23:413–421.(e) Ronconi L, Sadler P. J Dalton Trans. 2011; 40:262–268.(e) Liu HK, Sadler P. J Acc Chem Res. 2011; 44:348–359.
- 7. (a) Singh TN, Turro C. Inorg Chem. 2004; 43:7260–7262. [PubMed: 15530069] (b) Lutterman DA, Fu PKL, Turro C. J Am Chem Soc. 2006; 128:738–739. [PubMed: 16417361]
- 8. (a) Antonarakis ES, Emadi A. Cancer Chemother Pharmacol. 2010; 66:1–9. [PubMed: 20213076] (b) Ang WH, Dyson PJ. Eur J Inorg Chem. 2006:4003–4018.
- 9. (a) Pinnick DV, Durham B. Inorg Chem. 1984; 23:1440–1445.(b) Cruz AJ, Kirgan R, Siam K, Heiland P, Rillema DP. Inorg Chim Acta. 2010; 363:2496–2505.
- 10. Liu Y, Turner DB, Singh TN, Angeles-Boza AM, Chouai A, Dunbar KR, Turro C. J Am Chem Soc. 2009; 131:26–27. [PubMed: 19072048]

- 11. Chattopadhyay SK, Mitra K, Biswas S, Naskar S, Mishra D, Adhikary B. Trans Metal Chem. 2004; 29:1–6.
- 12. Steel PJ, Lahousse F, Lerner D, Marzin C. Inorg Chem. 1983; 22:1488–1493.
- 13. Walsh JL, Durham B. Inorg Chem. 1982; 21:329–332.
- 14. Caspar JV, Meyer T. J Inorg Chem. 1983; 22:2444–2453.
- 15. Juris A, Balzani V, Barigelletti F, Campagna S, Belser P, Von Zelewsky A. Coord Chem Rev. 1988; 84:85–277.
- 16. Brown GM, Callahan RW, Meyer TJ. Inorg Chem. 1975; 8:1915–1921.
- 17. Durham B, Wilson SR, Hodgson DJ, Meyer TJ. J Am Chem Soc. 1980; 102:600–607.
- 18. Fang Z, Swavey S, Holder A, Winkel B, Brewer KJ. Inorg Chem Comm. 2002; 5:1078–1081.
- 19. Linearization of 100 μM pUC18 plasmid was carried out with 50 units of *SmaI* (Invitrogen Life Technologies) restriction enzyme in 100 mM Tris, pH = 7.6, 150 mM NaCl at 37 °C for 1 hr, followed by deactivation at 65 °C for 5 min

Figure 1.

(a) Thermal ellipsoid plot (drawn with 30% probability ellipsoids; hydrogen atoms removed for the sake of clarity) and (b) calculated molecular orbital diagram of **1**.

Figure 2.

Changes to the electronic absorption spectrum of 77 μ M **1** upon irradiation in H₂O at t_{irr} = 0, 5, 7.5, 10, 12.5, and 15 min; inset: 0, 1, 2, and 3 min (λ_{irr} > 395 nm).

Figure 3.

Imaged ethidium bromide stained agarose gel of 50 μM linearized pUC18 plasmid (10 mM phophate buffer, pH = 8.3) in the presence of various concentrations of **1**. Lanes 1 and 8, 1kb DNA molecular weight standard; lanes 2 and 7, linearized plasmid alone; lanes 3 – 6, 25 μM, 75 μM, 150 μM, and 250 μM complex (a) irradiated with λ_{irr} > 395 nm (t_{irr} = 15 min) and (b) incubated in dark for 15 min at 298 K.