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Mechanisms of Drug Resistance that Target the Androgen Axis in Castration Resistant Prostate Cancer (CRPC)

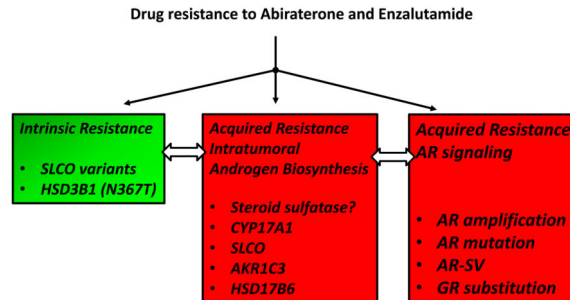
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Abstract

Castrate resistant prostate cancer (CRPC) is the fatal-form of prostate cancer and remains androgen dependent. The reactivation of the androgen axis occurs due to adaptive intratumoral androgen biosynthesis which can be driven by adrenal androgens and /or by changes in the androgen receptor (AR) including AR gene amplification. These mechanisms are targeted with P450c17 inhibitors e.g. Abiraterone acetate and AR super-antagonists e.g. Enzalutamide. Clinical experience indicates that with either agent an initial response is followed by drug resistance and the patient clinically progresses on these agents. This article reviews the mechanisms of intrinsic and acquired drug resistance that target the androgen axis and how this might be surmounted.

Graphical Abstract



Keywords

prostate cancer; steroidogenesis; androgen receptor; therapeutics

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1. Introduction

1. 1. Castration Resistant Prostate Cancer

Prostate cancer is the most common cancer in man and the second leading cause of cancer death in men per year in the US. Approximately 180,000 new cases are diagnosed and approximately 30,000 deaths occur annually [1]. Advanced localized prostate cancer is often treated with androgen deprivation therapy (ADT) which consists of leuprolide (a LH-RH agonist) [2], which is the equivalent to a chemical castration, often combined with an androgen receptor (AR) antagonist e.g. *R*-bicalutamide (Casodex), and or a dual type 1 and type 2 5 α -reductase inhibitor (dutasteride). Following this treatment there is a period of remission, however, the cancer invariably returns and is accompanied by an increase in serum prostatic-specific antigen (PSA). PSA is an androgen dependent gene and because this increase occurs in the presence of castrate levels of circulating androgens this from of the disease is referred to as castration resistant prostate cancer (CRPC). CRPC is uniformly fatal and occurs due to adaptive responses in the tumor that reactivate the androgen axis [3, 4]. The androgen axis is defined as activation of androgen receptor (AR) signaling that leads to androgen dependent gene expression. Reactivation of the axis can occur due to adaptive intratumoral androgen biosynthesis [5-9] or by changes in the AR (e.g. AR gene amplification and mutation) [10-16]. Both mechanisms are targeted by new drugs that have been approved by the US FDA e.g. abiraterone acetate (AA) [17-20], a P450c17 (17 α -hydroxylase/17,20-lyase) inhibitor which prevents the formation of adrenal androgens e.g. dehydroepiandrosterone-sulphate (DHEA-SO₄), DHEA and ⁴-androstene-3,17-dione (⁴-AD); and enzalutamide (ENZ) [12, 21], a AR super-antagonist. When used to treat metastatic CRPC (mCRPC), patients on these agents have an increased median survival of only 3-4 months before drug resistance occurs [22, 23].

1. 2. Experience with Abiraterone

AA is a P450c17 inhibitor that blocks both the 17 α -hydroxylase and 17,20-lyase activity of the enzyme. Inhibition of 17 α -hydroxylase prevents the production of cortisol in the adrenal and in turn blocks feed-back inhibition of the adrenal-pituitary axis so that the gland is hyperstimulated with ACTH. In an attempt to make cortisol in response to ACTH, desoxycorticosterone increases which is a potent mineralocorticoid which can lead to life-threatening hypertension. This side-effect is countered by the co-administration of prednisone, a synthetic glucocorticoid. Phase 1 and phase 3 clinical trials with AA showed significant benefit in terms of median time to PSA progression, median radiologic progression free survival and improved symptoms [12, 20, 24]. These trials also reported a > 90% decline in circulating DHEA providing a biochemical assessment of drug efficacy [24, 25].

In a neoadjuvant clinical trial we compared the effect of Leuprolide alone (12 weeks) followed by Leuprolide plus AA (12 weeks)-Arm 1, with patients that received Leuprolide plus AA (for all 24 weeks) [26]. Once patients were on AA they also received prednisone for the reasons stated above. We measured serum androgens at entry into the trial, at 12 weeks and 24 weeks using a novel validated stable isotope-dilution liquid chromatography tandem mass spectrometric (LC-MS/MS) assay, Table 1. We found that leuprolide alone

decreased circulating testosterone (T) levels to castrate levels but adrenal androgens (DHEA-SO₄, DHEA, ⁴-AD) were affected. Once AA was administered the adrenal androgens were decreased by > 90%. However, the amount of circulating DHEA-SO₄ that remains (20,000 ng/dL) exceeds the castrate levels of circulating testosterone (5 ng/dL) by more than 4,000-fold, suggesting that a significant reservoir of circulating DHEA-SO₄ remains to fuel intratumoral androgen biosynthesis. A significant difference was also observed in the serum DHT levels at 12 weeks versus the value at 24 weeks (p<0.001) in the first arm which may favor DHT formation via the backdoor pathway (see later) in the transition period from Leup to Leup + AA. The reservoir of DHEA-SO₄ that remains provides a mechanism for the clinical failure of all P450c17 inhibitors which could be exacerbated if the tumor has either intrinsic or acquired resistance phenotypes (see Section 2 for definitions).

This observation provides a mechanism for the clinical failure of all P450c17 inhibitors which could be exacerbated if the tumor has either intrinsic or acquired resistance phenotypes (see Section 2 for definitions).

1.3. Experience with Enzalutamide

ENZ is a potent AR antagonist that prevents nuclear localization of the receptor and promotes receptor degradation [21]. Side-effects seen with ENZ include CNS seizures mediated by an off-target effect by inhibiting GABA_A currents, and gynecomastia and fatigue [27]. The CNS side-effects can be overcome by substituting the second generation AR super-antagonist ARN-509 [28]. In the phase 1-2 study with ENZ the drug decreased PSA by > 50% or more in 56% of the patients, 22% of the patients showed reduced soft-tissue disease, 56% of the patients had stabilized bone disease and 49% of patients showed a favorable circulating tumor count. PET imaging of the AR by ¹⁸F-DHT showed a 20-100% reduction in AR occupancy, and the median time to progression was 47 weeks [29]. In the AFFIRM phase 3 clinical trial ENZ was found to increase survival in prostate cancer patients who received prior chemotherapy from 13.6 months to 18.4 months versus the placebo control group [30]. In the PREVAIL phase 3 clinical trial in which patients had not received prior chemotherapy ENZ showed significant benefit with respect to all secondary end points, including the time until the initiation of cytotoxic chemotherapy (hazard ratio, 0.35), the time until the first skeletal-related event (hazard ratio, 0.72), a complete or partial soft-tissue response (59% vs. 5%), the time until prostate-specific antigen (PSA) progression (hazard ratio, 0.17), and a rate of decline of at least 50% in PSA (78% vs. 3%) where p<0.001 for all the comparisons made [22].

2. Mechanisms of Drug Resistance

Multiple mechanisms exist to account for resistance to AA and ENZ observed clinically in mCRPC patients. These mechanisms may involve intrinsic or acquired resistance. Intrinsic resistance refers to a subset of individuals that will do poorly on these drugs due to germline or inherited mutations. Acquired resistance refers to either somatic mutations or changes in gene expression profiles that will surmount the effect of the drug. Acquired resistance can include adaptive changes in intratumoral androgen biosynthesis and /or AR signaling. These mechanisms are not mutually exclusive and may synergize with each other.

2.1. Intrinsic Drug Resistance

2. 1. 1. Inherited Mutations in Organic Anion Transporters—DHEA-SO₄ is the major source of adrenal androgens that can fuel intratumoral androgen biosynthesis even after AA treatment. For this conjugate to become bioavailable it must be transported by the solute carrier organic anion encoded membrane transporters (SLCO), also known as organic anion transporters (OATs). The members of this family involved in steroid uptake in the prostate include *SLCO1A2* and *SLCO2B1* [31]. The impact of *SLCO* genetic variation on steroid sequestration has been evaluated using *SLCO2B1* variants in prostate cancer cells. In cells transfected with the *SLCO2B1* SNP variant rs12422149 (935GA; Arg312Gln), the 935A variant exhibited higher maximal DHEA-SO₄ uptake when compared with either the wild type 935G allele or with mock-transfected cells [32]. In 538 patients with metastatic hormone-sensitive prostate cancer the median time to progression for patients with each of 3 *SLCO2B1* alleles: rs12422149 [935G>A; Arg312Gln, minor allelic frequency 21%]; rs1789693 [A/T intron variant with minor allelic frequency of 48%]; and rs1077858 [A/G intron variant with minor allelic frequency of 43%] was 10, 7, and 12 months shorter, respectively than when the wild type allele was present [32]. Data such as these indicate that *SLCO* gene variants will impact prostate cancer progression and response to ADT therapy.

The *HSD3B1* (N367T) mutation is the first to be identified in a steroidogenic enzyme that may affect drug response to AA [33]. HSD3B1 is the principal enzyme in the prostate involved in either the conversion of DHEA to ⁴-androstene-3,17-dione (⁴-AD) or the conversion of ⁵-androstene-3 β 17 β -diol (⁵-Adiol) to T via its dual 3 β -hydroxysteroid dehydrogenase/ketosteroid isomerase activities. The germline N367T mutation makes the enzyme more stable and CRPC patients harboring this mutation will be more prone to evade ADT by being able to convert residual DHEA into potent androgens more readily. Tumors bearing the *HSD3B1* N367T mutation more rapidly progress to CRPC in xenograft models than in tumors bearing the wild-type allele [33]. The population frequency of the mutated allele is about 22%. It is likely that inherited SNPs in other steroidogenic enzymes may also impact drug response and hence mediate intrinsic resistance phenotypes to AA or ENZ but these have yet to be documented.

2. 2. Acquired Drug Resistance

2. 2. 1. Denovo synthesis of Androgens—Nelson and colleagues have made the case that prostate tumors catalyze *denovo* synthesis of active androgens [7, 8]. The conversion of [¹⁴C]-acetate into DHT was observed *ex vivo* in the LNCaP xenograft model and formation of cholesterol and cholesteryl esters was also observed. This mechanism could be an adaptive response to first and second line ADT. However, such studies do not take into account the large excess of DHEA-SO₄ that remains after AA treatment in patients. *De novo* synthesis is unlikely to make a significant contribution to intratumoral androgen biosynthesis, when such a large depot of DHEA-SO₄ exists in the circulation after AA treatment. Nevertheless increases in *CYP11A* (side-chain cleavage enzyme) and *CYP17A1* transcripts have been observed in response to AA treatment and possibly contribute to acquired drug resistance [5].

2. 2. 2. Bioavailability of DHEA-SO₄—The presence of high circulating DHEA-SO₄ after P450c17 inhibition can be exploited by prostate cancer tumors if there is high expression of SLCOs and steroid sulfatase (STS) to liberate free DHEA, Fig .1. SLCO1A2 is implicated in DHEA-SO₄-induced prostate cancer cell growth in androgen-depleted media. SLCO1A2-transfected LNCaP and 22RV1 cells showed increased DHEA-SO₄ stimulated growth when compared to *SLCO1A2*-knockdown cells, which were insensitive to DHEA-SO₄ stimulated growth [34] In addition SLCO1A2 mRNA expression was higher in PCa cell lines grown under androgen-depleted conditions (and was suppressed by addition of DHT). These data suggest that expression of SLCO1A2 is up-regulated by androgen deprivation and may lead to acquired drug resistance, by providing more residual DHEA-SO₄ for intratumoral androgen biosynthesis. Less is known about adaptive responses in *STS* expression following ADT.

2. 2. 3. Overexpression of Type 5 17 β -HSD or AKR1C3—AKR1C3 (type 5 17 β -hydroxysteroid dehydrogenase) plays an essential role in the formation of T and DHT in the prostate irrespective of the pathway used [35-38]. In the canonical pathway: DHEA \rightarrow ⁴-AD \rightarrow T \rightarrow DHT, AKR1C3 catalyzes the reduction of ⁴-AD \rightarrow T. In the alternative pathway: DHEA \rightarrow ⁴-AD \rightarrow 5 α -androstane-3,17-adione (Adione) \rightarrow DHT, AKR1C3 catalyzes the reduction of Adione to DHT; in the backdoor pathway: Progesterone \rightarrow 5 α -dihydroprogesterone \rightarrow allopregnanolone \rightarrow androsterone \rightarrow 5 α -androstane-3 β ,17 β -diol (3 α -diol) \rightarrow DHT, AKR1C3 catalyzes the conversion of androsterone to 3 α -diol; and in the ⁵-Adiol pathway: DHEA \rightarrow ⁵-Adiol \rightarrow T \rightarrow DHT, AKR1C3 catalyzes the conversion of DHEA to ⁵-Adiol. Thus overexpression of AKR1C3 in CPRC would provide a mechanism to divert trace androgens that remain after ADT to potent androgens via these three pathways within the tumor. Studies have shown that AKR1C3 is overexpressed in prostate cancer cell lines 10-16 fold and up to 3-fold in androgen responsive and androgen independent prostate cancer cell xenografts upon androgen deprivation when measured by qRT-PCR, Table 2 [6, 39-43]. By contrast AKR1C3 expression is repressed by androgens e.g. R1881 [44]. These studies suggest that AKR1C3 upregulation is an adaptive response to ADT, and could contribute to drug resistance observed with AA or ENZ. Investigation of the mechanism by which AKR1C3 is upregulated by ADT identified ERG as binding to the *AKR1C3* gene promoter. TMPRSS2-ERG is a fusion protein, a biomarker of advanced prostate cancer and is co-expressed with AKR1C3 in advanced disease. Since ERG is induced by androgens, intratumoral production of DHT by AKR1C3 would increase ERG expression and create a feed-forward mechanism to sustain AKR1C3 expression and androgen biosynthesis. In this manner TMPRSS2-ERG overrides the repressive effect of the AR on the *AKR1C3* gene promoter. TMPRSS2-ERG k/d in VCaP cells led to a reduction in AKR1C3 expression and a decrease in the ability to convert Adione to DHT supporting this mechanism [45]. It is noteworthy that analysis of gene expression in 25 mCRPC tumors obtained from the Gene Expression Omnibus revealed a significant correlation ($p < 0.0001$ $r = 0.69$) between AKR1C3 and ERG co-expression where AKR1C3 increased expression ranged from 0- 100-fold and ERG expression ranged from 0-250-fold over non-expressing tumors [45].

AKR1C3 is also one of the most overexpressed steroidogenic genes in CRPC patients [46]. A 5-10 fold increase in expression levels over primary tumors was observed using Affymetrix microarray data and validated by qRT-PCR. AKR1C3 was also overexpressed in 11/19 soft-tissue metastasis from mCRPC patients by IHC [44, 46][46] and in bone metastasis by qRT-PCR and IHC [47]. Analysis of several studies suggest that AKR1C3 is upregulated in subset of mCRPC patients where estimates are that it is overexpressed at the RNA and protein level in at least one third of all mCRPC patients [39, 43, 44, 47]. These estimates are likely to be conservative since whether patients had received prior second-line ADT therapy was not reported when measuring transcript levels. Data obtained from the Oncomine database for 363 prostate cancer tumor specimens when used to construct Kaplan-Meier survival plots showed that the median survival time for patients with tumors that were low AKR1C3 expressors and low TMPRSS2-ERG expressors was 12-13 years, but patients whose tumors were high AKR1C3 expressors and high TMPRSS2-ERG expressors had median survival times of only 7-10 years [45]. In this study high and low expressors fell either side of the median value, however, it is not clear as to whether the prostate cancer specimens were derived from radical prostatectomy samples, needle biopsies, or transurethral prostate tissue specimens. Others have since confirmed the high level of AKR1C3 transcripts in bone metastasis of CRPC patients and found that this was due to overexpression rather than by increased copy number variation [47].

2. 2. 4. Overexpression of HSD17B6—HSD17B6 is the major 3 α -hydroxysteroid dehydrogenase in human prostate that catalyzes the back conversion of 3 α -diol to DHT [48-50]. Long-term culture of LNCaP or VCaP cells in androgen-deprived media increased transcriptomic expression *HSD17B6*. In 42 prostate cancer patients undergoing androgen deprivation therapy (ADT) HSD17B6 was about 2-fold higher than that in tissues of 100 untreated individuals. In men receiving ADT, patients showing biochemical progression had a higher *HSD17B6* score than those without progression [51]. These results suggest that 3 α / β -diol also represent potential precursors of DHT, and the back conversion to DHT from androgen derivatives catalyzed by HSD17B6 may be a promising target to counter ADT resistance. HSD17B6 is also upregulated by TMPRSS2-ERG and when this transcription factor is silenced in VCaP cells there is a dramatic reduction in HSD17B6 expression but only a modest effect on the conversion of 3 α -diol to DHT suggesting that AKR1C3 may be the major source of potent androgens in these cells [45].

2. 2. 5. Androgen Receptor Splice Variants—Resistance to AA and ENZ, is also associated with increased expression of androgen receptor splice variants (AR-SVs) [52]. The two most common AR-SV's are AR-V7 and ARes567 (the exome skipping variant), Fig. 2. These variants have lost their ligand-binding domain and are constitutively active without ligand [53, 54]. Originally it was felt that they would heterodimerize with full-length AR (AR-FL), but it now appears that AR-SVs are transcriptionally active in their own right [55]. Microarray analysis shows that the AR-SV transcriptionally activates a gene set that differs from AR-FL and which may make the tumor more aggressive [56]. AR-V7 may be the most important splice variant and is implicated in AA and ENZ resistance and is detected in circulating tumor cells (CTCs) of CRPC patients [57]. In this study 39% of the patients resistant to ENZ and 31% of the patients resistant to AA had detectable AR-V7. By

contrast in AA resistant xenografts of VCaP cells AR-V7 mRNA was expressed as < 1% of the mRNA for AR-FL. AR-V7 is clearly associated with disease progression and the drug resistant phenotype but its exact contribution to resistance mechanisms still needs to be established [58]. The exact mechanism by which AA and ENZ result in AR-SVs is unknown, but could involve genomic rearrangement of the AR gene and/or alternative splicing of the AR pre-messenger RNA (mRNA).

2. 2. 6. Androgen Receptor Mutation and Substitute Steroid Receptors—

Clinical experience with AR-antagonists such as *R*-bicalutamide showed that overtime the AR acquired somatic mutations that enabled the antagonist to work as an agonist and also make the AR ligand permissive [11, 15]. One of the most common mutations is T877A. This now appears to be the clinical experience with AA and ENZ as well [57, 59]. The T877A mutant can use among other steroids progesterone. Missense AR exon 8 mutations were detected in 11/62 patients (18%), including the first reported case of an F876L mutation in an ENZ resistant patient [60]. In addition, the H874Y and T877A mutations were detected in 7 AA-resistant patients. In patients (n=30) that switched from AA onto ENZ, AR gene amplification and or exon 8 mutations in CTCs predicted adverse outcomes including lower rates of PSA decline 30% (P=0.013, χ^2) and shorter time to radiographic/clinical progression (p < 0.010) [60]. The ability of DHT to enhance CRPC cell growth regardless of whether the AR is expressed has been reported in a mechanism that involves activation of STAT-5 via the hijacking of the glucocorticoid receptor (GR) [61]. Preclinical prostate cancer models also point to the possibility that ENZ resistance may be mediated by the GR [62, 63]. These results are provocative since AA is often co-administered with prednisone. However, the immediate benefit of this drug combination in CRPC may not support this assertion, [64]. It is conceivable that the long term administration of a prednisone could make the tumor more dependent on the GR or act as ligand for the mutated AR.

3. 0. Biomarkers of Drug Resistance

Non-invasive biomarkers of AA and ENZ drug resistance would be ideal for precision therapy of CRPC. Thus far attention has focused on CTCs as a source of biomarkers. While CTCs report the expression of AR-SVs [57], they have not been routinely examined for expression changes in steroidogenic enzymes that may contribute to drug resistance. As yet the ability of CTC derived biomarkers to have sufficient specificity and sensitivity to predict prognostic outcome has yet to be demonstrated. Molecular profiling of tumor biopsies for transcript or protein levels of *SLCO*, *SULT*, *HSD3B1*, *AKR1C3*, *HSD17B6*, *AR-SV* could interrogate pathways in the androgen axis expected to cause drug resistance which would be actionable if therapeutics existed to act on each of these targets.

4. 0. Approaches to Counter Drug Resistance

4. 1. Combination Trials

Patients that progress on AA or ENZ could be switched to the other drug to prolong survival. However, patient experience with sequential therapy of this type has shown limited benefit and likely results from overlapping mechanisms of resistance shared by each agent [65]. An alternative would be to give both AA and ENZ concurrently and clinical trials are

ongoing to determine whether there is additional benefit from such a combination. However, drug resistance is still anticipated to emerge. Thus there is a need for better approaches to counter the drug resistance phenotype.

4. 2. Sulfatase Inhibitors

If DHEA-SO₄ contributes to the drug resistance phenotype this could be surmounted by the administration of a STS inhibitor 667 COUMATE (Irosustat) or STX213 [66]. Additionally, EM-1913 has been found effective in blocking 80% of the growth of the rat ventral prostate when stimulated with DHEA-SO₄ [67]. Although this approach appears promising, incomplete inhibition of P450c17 still results in a significant amount of free DHEA and 4-AD that would be available for the synthesis of potent androgens.

4. 3. Bifunctional P450c 17 inhibitors and AR Antagonists/Degraders

New second generation inhibitors of P450c17 e.g. galaterone inhibit the 17/20-lyase activity and also antagonize and degrade the AR T877A mutant [68]. By targeting both P45017c and AR these agents could supersede combination AA and ENZ therapy and target resistance to either therapy and shows promise.

4.4. AKR1C3 Inhibitors

Based on the central role of AKR1C3 in intratumoral androgen biosynthesis and its up-regulation by ADT a series of AKR1C3 inhibitors have been developed based on repurposing non-steroidal anti-inflammatory drugs (NSAIDs) [69-72]. Three series of compounds have been reported and include: *N*-phenylaminobenzoates (which are fenamate derivatives); *N*-naphthylaminobenzoates; and *N*-benzoyl-indoles (which are derived from indomethacin). Lead compounds in each library no longer inhibit the NSAID target COX-1 or COX-2. Each of the leads have nanomolar potency for AKR1C3 and are effective in cell-based models in blocking the conversion of 4-AD to T in LNCaP-AKR1C3 cells, and block the production of PSA expression driven by the AKR1C3 substrate 4-AD in VCaP cells [4]. The *N*-naphthylaminobenzoates have the added advantage that they also act as AR antagonists and represent “first-in-class” bifunctional AKR1C3 inhibitors and AR antagonists [73]. Recently, CD-2B cells, a subline of LNCaP cells were grown in the presence of ENZ to generate a ENZ resistant cell line (C4-2B MDVR). This cell line over expressed AKR1C3 40-fold when compared to the control cell line, and this difference was observed at the mRNA and protein levels. In addition the C4-2B MDVR cells produced large amounts of T (131 pg per 5 × 10⁶ cells) and DHT (18 pg per 5 × 10⁶ cells) versus the parental cells which produced (0.15 pg T per 5 × 10⁶ cells; and 0 pg DHT per 5 × 10⁶ cells); and grew in the presence of ENZ. Indomethacin an AKR1C3 inhibitor was able to arrest the growth of these cells in cologenic assays. Xenografts of CW22Rv1 cells which are also ENZ resistant and express AKR1C3 gave rise to tumors in castrate SCID mice which were not responsive to ENZ but could be prevented from growing in the presence of indomethacin. These experiments provide important *in vivo* “proof-of principle” to use an AKR1C3 inhibitor to surmount ENZ drug resistance [41].

Both GTx-therapeutics and Astellas have developed AKR1C3 inhibitors as potential therapeutics for CRPC. GTX-560, a potent competitive inhibitor of AKR1C3, revealed for

the first time that this compound could block the hitherto unrecognized AR co-activator function of AKR1C3 suggesting that the compound would have added benefit in the treatment of CRPC patients [74]. GTX-560 was also found to have antitumor activity in prostate cancer cell xenograft models of CRPC. Astellas took their lead compound ASP9521 into man following favorable preclinical development, including absorption, distribution, metabolism, elimination and toxicological properties, and *in vivo* proof-of-principle studies to block tumor growth in prostate cancer cell xenograft models [75]. The compound was found to be orally bioavailable and well tolerated but without efficacy where the clinical endpoint was a decrease in serum PSA in mCRPC patients [76]. However, the drug was inadequately tested, 6/13 mCRPC patients withdrew from the trial and patients were not preselected for AKR1C3 status and there was no measurement of intratumoral androgens. The authors also concluded that the lack of effect might have been due to the possibility that AKR1C3 was not expressed at a higher enough level to see a beneficial effect since the patients had not received second line ADT.

4.5. Coactivator Competitive peptides

An entirely novel approach would be to annotate the co-regulators that interact with the AR-SVs in CRPC and then develop peptido-mimetics that could compete for the co-regulatory domain of the AR-SV. Progress in this area has been limited to date [77], but some “proof-of-principle” using peptides to disrupt the interaction between p160 and AR in castration-resistant AR expressing prostate cancer cells has been reported [78].

Conclusions—Drug resistance to the standard treatment-of-care for mCRPC involving AA or ENZ represents a a major clinical problem. Co-administration of both drugs is likely to prolong survival but drug resistance will prevail. Intrinsic drug resistance results from allelic variants in *SLCO* and *HSD3B1*. Acquired drug resistance involves adaptive changes in the androgen axis and several mechanisms may combine to exacerbate drug resistance e.g. the incomplete inhibition of P450c17 coupled with the overexpression of AKR1C3 may create a “perfect-storm” so that intratumoral androgen biosynthesis can occur. There is an urgent need for new agents that do not target P450c17 or the AR with a focus on targeting the mechanisms of therapeutic drug resistance. Promising approaches include the development of HSD3B1 and AKR1C3 inhibitors and peptido-mimetics that could disrupt the interaction between AR-FL, AR-SV’s with their steroid receptor co-regulators.

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Abbreviations

ADMET	absorption distribution, metabolism: elimination and toxicology
⁴-AD	4-androstene-3,17-dione
⁵-Adiol	5-androstene-3 β ,17 β -diol
3α-Adiol	5 α -androstane-3 α ,17 β -diol

Adione	5 α -androstane-3,17-dione
ADT	androgen deprivation therapy (ADT)
AR	androgen receptor
AR-FL	androgen receptor full length
AR-SV	androgen receptor splice variant
CRPC	castration resistant prostate cancer
DHEA	dehydroepiandrosterone
DHT	5 α -dihydrotestosterone
3α-diol	5 α -androstane-3 α ,17 β -diol
HSD	hydroxysteroid dehydrogenase
SLCO	solute carrier organic anion transporter
STS	steroid sulfatase
T	testosterone

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Publication Highlights

- Abiraterone and enzalutamide in castration resistant prostate cancer leads to drug resistance
- Intrinsic drug resistance arises due to allelic variants in organic anion transporters and *HSD3B1*
- Acquired drug resistance arises due to intratumoral androgen biosynthesis and changes in the AR
- Acquired resistance can involve the induction of AKR1C3 and emergence of AR-splice variants
- AKR1C3 inhibitors or peptidomimetics that block AR-splice variants may surmount drug resistance

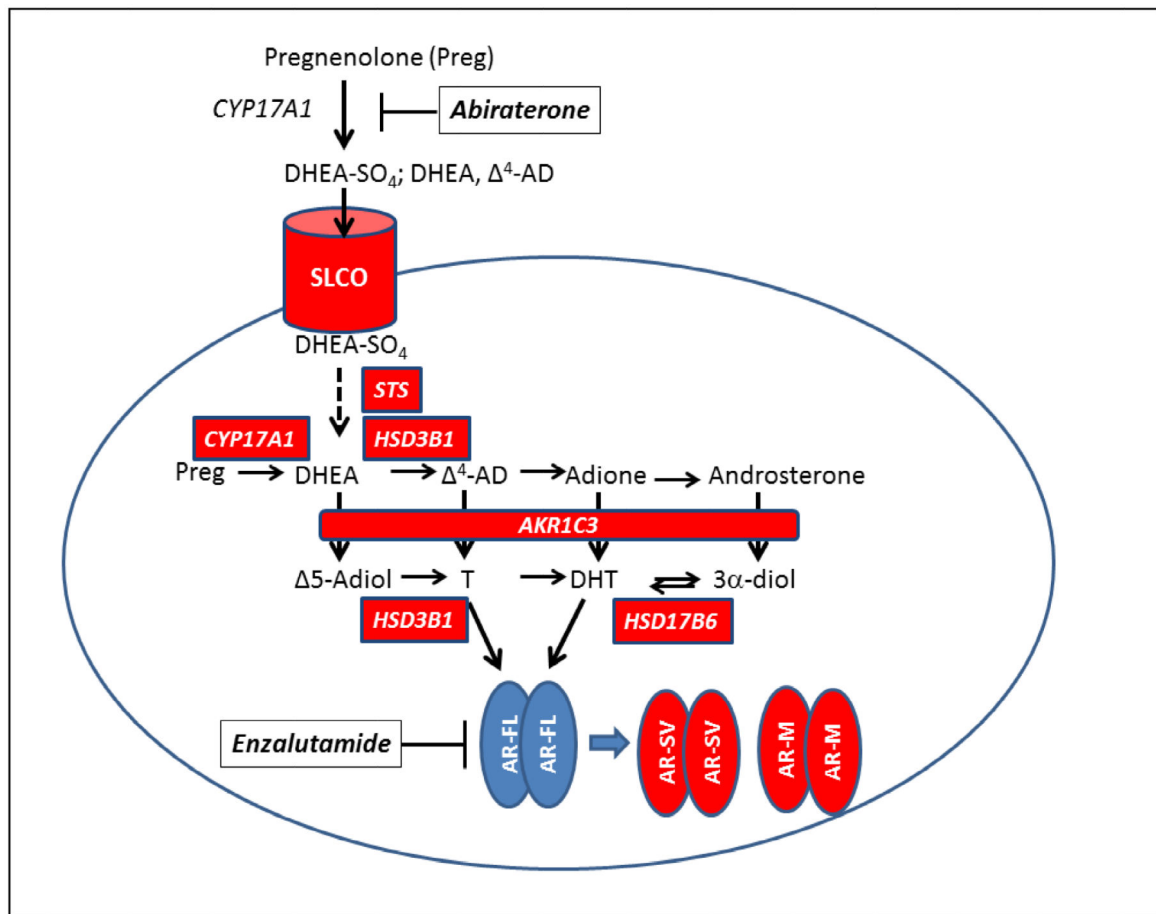


Figure 1. Composite Mechanisms of Drug Resistance in Castration Resistant Prostate Cancer. Mechanisms indicated in red boxes; arrow between AR-FL and AR-SV and AR-SM indicates selection pressure to produce new AR subtypes; where AR-FL, androgen receptor full length; AR-SV, androgen receptor splice variant; AR-M, mutated androgen receptor. *italics = gene names*; AKR1C3, aldo-keto reductase 1C3; AR-SV, = androgen receptor splice variant; AR-M, = mutated androgen receptor; CYP17A1, DHEA = dehydroepiandrosterone; Δ^4 -AD, 4-androstene-3,17-dione; Adione, = 5 α -androstane-3,17-dione; Δ^5 -Adiol, 5-androstene-3 β ,17 β -diol; DHT, 5 α -dihydrotestosterone; 3 α -Adiol, 5 α -androstane-3 α ,17 β -diol; HSD3B1; 3 β -hydroxysteroid dehydrogenase type 1; P450c17 (17 α -hydroxylase,17/20-lyase); T =testosterone; SLCO; = solute carrier organic anion transporter;

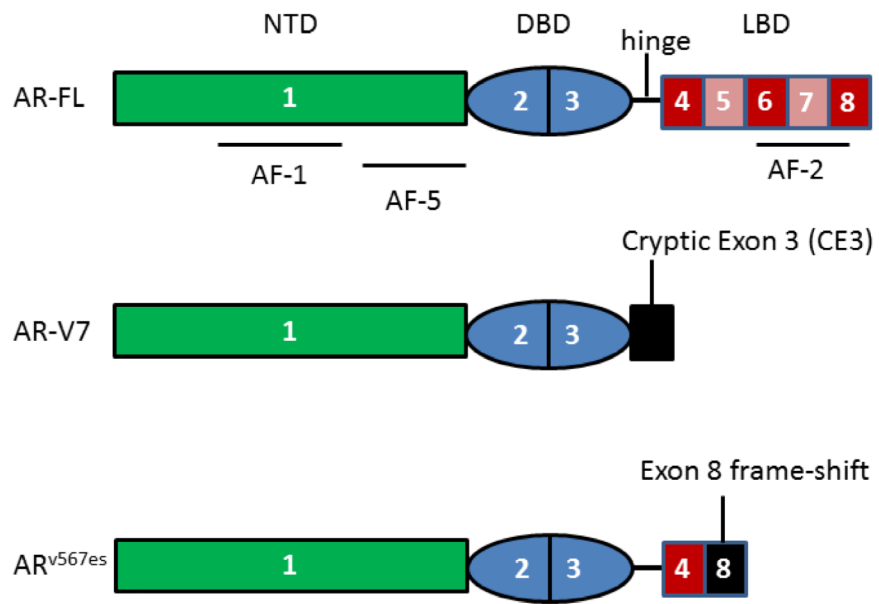


Figure 2.
Domains of AR-SVs. Numbers refer to AR gene exons.

Table 1
Serum Androgen Measurements-Neoadjuvant Study Luprolide vs Luprolide plus Abiraterone Acetate

Serum Androgen Levels in Abi + Leuprolide Neoadjuvant Clinical Trial	24 weeks Leuprolide/ 12 weeks AA + Prednisone			24 weeks Leuprolide/ 24 weeks AA + Prednisone			p Value
	Baseline (n= 28)	Week 12 (n =28)	Week 24 (n = 27)	Baseline (n =27)	Week 12 (n =29)	Week 24 (n =27)	
Median serum androgen (ng/dL)							
Testosterone	429	17	5	425	7	16	0.0003
DHT	29	13	17	40	13	6	0.0176
DHEA	242	201	19	176	26	30	< 0.0001
DHEA-gluconide	1901	1508	515	1522	415	292	0.0003
DHEA-sulfate	231K	200K ^a	22K ^b	195K ^a	16K ^b	17K ^b	<0.0001 ^a vs ^b
Androsterone	11	5	0.6	9	1	0.8	0.003
4-Androstene-3, 17-dione	76	52	7	59	9	8	<0.001

Taken from Ref: [26]

a and b superscripts refer to the groups that were compared to reject the null hypothesis. Other comparisons are at 12 weeks versus baseline based on log-transformed data

Table 2
Overexpression of AKR1C3 in Prostate Cancer Patients and Prostate Cancer Cell Lines

	Patient Samples			Cell Lines					
	Affymetrix Microarray	qRT-PCR	IHC	LNCaP	DuCaP	VCaP	22Rv1	PC-3	DU-145
Stanbrough et al. [46]	32 CRPC patients 5.3 fold increase vs primary tumors	10-fold higher in 32 CRPC patients vs primary tumors	11/19 CRPC recurrent prostate cancer & soft tissue mets vs 1/18 primary tumors						
Hofland et al [40]		10-fold higher in CRPC (n=10; p<0.001) vs normal prostate			Up regulated in CS-FBS 9-fold by qRT-PCR	Up regulated in CS-FBS 5-fold by qRT-PCR			
Pfeiffer et al., [42]		CRPC : low grade (n=21;20) 2.3 fold			Up regulated in CS-FBS 5-fold by qRT-PCR				
Jernberg et al, [47]		5-fold higher in bone mets vs hormone naive patients (n=51; p<0.01)	Bone mets. n =51 (rs with RNA 0.73; p<0.0000001)						
Hamid et al., [39]		3-fold Higher AKR1C3 (n =41) than in BPH (n=9 samples p<0.001)	41 samples IHC staining (Rs with RNA 0.39; <0.0001)	-ive IB	+ ive IB	+++ ive IB	++ ive IB AR7V	+ ive IB	-ive IB
Tian et al., [43]			3-fold higher in 40 needle biopsy GS=6, GS=7, GS=8, GS= 9 vs low grade GS (n=10 group; p<0.05). Positive correlation with Gleason Score						
Mitsiades et al., [44]	6/19 mets upregulated	Upregulated 3-fold in 30% mCRPC (n = 19) vs normal prostate (n=19) and in 5 patients CTCs		AKR1C3 o/exp upon ADT CS-FBS (16-fold) or MDV3100 (6-fold) by qRT- PCR					

	Patient Samples			Cell Lines					
	Affymetrix Microarray	qRT-PCR	IHC	LNCaP	DuCaP	VCaP	22Rv1	PC-3	DU-145
Frankhauser et al. [79]	20 samples: NS Enrichment of <i>HSD17B</i> genes as a group q value <0.1	2-fold higher in hormone refractory cancer vs BPH (n=4) p<0.012 Taylor cohort mCRPC 153% increase vs BPH (n= 41) p <0.0001		Repressed by R1881 (16-fold) by qRT- PCR					
Powell et al. [42].	25 mCRPC tumors by microarray show high correlation between AKR1C3 expression and ERG rs = 0.69	Up to 100-fold increase in AKR1C3 and 250-fold increase in ERG expression							

CS-FBS = charcoal stripped FBS; BPH= benign prostatic hyperplasia; GS = Gleason Grade; IB = immunoblot (+ to +++ intensity of staining within the same experiment);