

Significance of Diagnosing Parasitic Infestation in Evaluation of Unexplained Eosinophilia

VINAY KHANNA¹, KRITI TILAK², CHIRANJAY MUKHOPADHYAY³, RUCHEE KHANNA⁴

ABSTRACT

Background: The evaluation of unexplained eosinophilia in an asymptomatic individual has always been a diagnostic challenge and requires understanding about a wide range of probable causative agents. Helminthic infestation and schistosomiasis are the main parasitic causes of eosinophilia. Therefore, the availability of simple and accurate diagnostic tests for detection of parasitic infections can prove to be valuable in early diagnosis and solving the mystery of unexplained eosinophilia.

Materials and Methods: In the present study we attempt to find an association between relative eosinophilia and parasitic infections and also to find the parasites responsible for eosinophilia in a substantial number of cases. A retrospective

study for the presence of eosinophilia was done on 621 cases positive for parasitic infestation.

Results: Among a total of 621 cases of parasitic infestation only 66 (10.6%) cases were found to have relative eosinophilia. The parasites found to be responsible for eosinophilia were *Trichuris trichiura*, *Ascaris lumbricoides*, *Strongyloides stercoralis*, filarial worm and hook worm.

Conclusion: From the study it is concluded that eosinophilia is not a universal finding in cases with parasitic infestation. Although presence of eosinophilia can serve as one of the many diagnostic clues to look for the presence of helminthic infestation if other non-infectious causes of eosinophilia are ruled out.

Keywords: *Ascaris lumbricoides*, Helminths, Karnataka, *Strongyloides stercoralis*, *Trichuris trichiura*

INTRODUCTION

It appears well proven that the presence of absolute eosinophilia is attributable, in a high percentage of cases, to the presence of helminthic infection [1-5]. Though, the implication of relative eosinophilia has not been reported. The assessment of unexplained eosinophilia in an asymptomatic individual is a perplexing problem that requires understanding about an extensive range of probable pathogens and their global distribution. Nevertheless, the prevention of morbidity by the diagnosis and early treatment of helminth infection is an essential task, and it requires abundant acumen to challenge them. The main parasitic causes related to relative eosinophilia are geohelminthic diseases (specifically, hookworms) and schistosomiasis.

Eosinophilia develops as an immunologically mediated response in connotation with varied processes. It is found in association with various atopic diseases, drug-related hypersensitive reactions, collagen vascular diseases, and malignancies. Eosinophilia is also particularly associated with helminth infections and especially during that phase of development when they invade the host tissues [6]. Infections are usually chronic and have high re-infection rates and are characteristically over dispersed, with patterns of intense infections being restricted to minorities within the common population [7,8].

The pattern and degree of eosinophilia in parasitic infections is determined by the development, migration, maturation, burden and distribution of the parasite within the host as well as by the host's immune response. Parasites tend to provoke marked eosinophilia when they or their products interact with immune effect or cells in tissues, chiefly during migration (e.g. trichinosis, ascariasis, gnathostomiasis, filarial parasites). Provocation of eosinophils in blood is usually absent when there is a mechanical hurdle between the parasite and the host (e.g. adult tapeworms that are solely intraluminal or hydatid cysts that are enclosed in a cystic structure) [9,10].

In their study, Nokes et al., [11] showed that even children with moderately intense *Trichuris trichiura* infection had improvements in their cognitive function (i.e., attentiveness, auditory short-term memory, and long-term memory) after receiving treatment. Therefore, the availability of dependable and simple diagnostic tests for detection of helminthic infections could provide important tools for patient care.

OBJECTIVES

1. To find an association between relative eosinophilia and parasitic infestation.
2. To find the parasites responsible for eosinophilia in substantial number of cases.

MATERIALS AND METHODS

Fecal samples of patients attending tertiary care hospital of Karnataka state, India, being clinically suspected of parasitic infestations were sent to the Parasitology section of the Department of Microbiology, Kasturba Medical College, Manipal, India. The samples were investigated for the presence of parasites using direct wet mount or stool concentration methods. All the cases found positive for parasitic infestation were included in the study. Cases with eosinophilia due to any reason other than parasitic infestations were excluded from the study. A retrospective study was done on a total of 621 cases found positive for parasitic infestation, over a study period of five years from year 2005 to 2010, for the presence of eosinophilia. The data was analysed using SPSS software ver. 16.

RESULTS

Out of total 621 cases positive for parasitic infection, 443 (71.3%) were males while 178 (28.7%) were females. Sixteen seven (10.8%)

Organisms/ Infection	Infected patients with relative eosinophil count (%)		Total no. of cases positive for parasitic infections
	<5	>5	
<i>Plasmodium falciparum</i>	141	1 (0.7%)	142
<i>Plasmodium vivax</i>	170	0 (0%)	170
Mixed infection (<i>Plasmodium falciparum</i> and <i>Plasmodium vivax</i>)	32	1 (3%)	33
<i>Plasmodium ovale</i>	2	0 (0%)	2
Cysticercosis	25	0 (0%)	25
<i>Echinococcus</i> spp.	33	0 (0%)	33
<i>Toxoplasma gondii</i>	14	0 (0%)	14
Filariasis	12	14 (53.8%)	26
<i>Ascaris lumbricoides</i>	1	7 (87.5%)	8
Scabies	17	6 (26.1%)	23
Hookworm	20	11 (35.5%)	31
<i>Trichuris trichiura</i>	1	8 (88.88%)	9
<i>Leishmania donovani</i>	2	0 (0%)	2
<i>Pneumocystis carinii</i>	2	0 (0%)	2
<i>Blastocystis hominis</i>	15	2 (11.8%)	17
<i>Trichomonas</i>	4	0 (0%)	4
<i>Blastocystis hominis</i> & <i>Trichomonas</i> coinfection	1	0 (0%)	1
<i>Giardia lamblia</i>	2	1 (33.3%)	3
<i>Cyclospora</i>	4	1 (20%)	5
<i>Cryptosporidium parvum</i>	16	1 (5.9%)	17
<i>Entamoeba histolytica/dispar</i>	10	6 (37.5%)	16
<i>Cryptosporidium parvum</i> & <i>Entamoeba histolytica/dispar</i> coinfection	1	0 (0%)	1
<i>Entamoeba coli</i>	17	0 (0%)	17
<i>Isospora belli</i>	6	0 (0%)	6
<i>Strongyloides stercoralis</i>	4	7 (63.6%)	11
<i>Isospora belli</i> & <i>Strongyloides stercoralis</i> coinfection	3	0 (0%)	3
Total	555	66 (10.6%)	621

[Table/Fig-1]: Number of cases positive for parasitic infections having eosinophilia

belonged to age group 0-16 years, 420 (67.6%) belonged to age group 17-50 years and 134 (21.6%) were above 50 years of age. 468 cases belonged to Karnataka state, out of which 147 (31.4%) cases were resident of Udipi district which accounts for the maximum prevalence of cases in Karnataka state. Only 66 (10.6%) cases were found to have relative eosinophilia. Cases whose samples were positive for *Plasmodium vivax*, *Plasmodium ovale*, *Taenia solium*, *Echinococcus* spp., *Toxoplasma gondii*, *Leishmania donovani*, *Pneumocystis carinii*, *Trichomonas* spp. and *Entamoeba coli* had normal relative eosinophil count. However, *Trichuris trichiura*, *Ascaris lumbricoides*, *Strongyloides stercoralis*, filarial worm and hookworm accounted for eosinophilia in 88.9%, 87.5%, 63.6%, 53.8% and 35.5% of the cases respectively [Table/Fig-1].

DISCUSSION

Eosinophils are a striking feature of many parasitic diseases. Helminthic infections are the most common parasitic diseases that produce eosinophilia. Nematode infections account for the majority of patients with eosinophilia in tropical countries, especially in areas where filariasis, ascariasis and hookworm infection are endemic.

Eosinophilia was present in only 32.2% (47 out of 146) of all helminth infections in our total patient population. This indicates that helminth infestation may not be accompanied with rise in eosinophil count in blood. The finding can be explained by the fact that eosinophilia

occurs only when the parasites invade the host tissues [12-17]. In a study from the United Kingdom, blood eosinophilia was present at the time of diagnosis in only 44% of 1107 travelers and immigrants with schistosomiasis [18]. In population of non-immigrants, as in our study, this value is even lower.

In a study conducted by Javier Pardo et al., [1], among 161 eosinophilic cases 116 (54.5%) had 1 parasite, 30 (14.1%) had 2, and 15 (7.0%) had >3. Filariae (n = 63, 29.6%) were the most frequently isolated parasite, followed by schistosomes (n = 37, 17.4%), hookworms (n = 36, 16.8%), and *Trichuris* spp. (n = 18, 8.4%). This is consistent with the results of our study. There was a statistically significant association (p<0.05) between the country of origin and the final diagnosis: filariasis was diagnosed in 77% of the patients with eosinophilia from Cameroon, 63% of the patients from Mali had schistosomiasis, and 30.8% of the patients from Nigeria had hookworm infection.

In studies conducted on immigrants with eosinophilia the etiologic agent was identified in 15% to 64% of cases (depending on the population, the selected eosinophil count, and the methods) [19].

In a rare case report from Turkey [20], a young patient of parasitic infestation presented with eosinophilic ascites. The patient was treated with Albendazole for three months which successfully brought the eosinophilic cell count to normal. Thus, emphasizing the importance of excluding parasitic infestation in all patients with eosinophilic ascites.

In another case report from Korea [21], a patient of Eosinophilic Gastroenteritis presented as acute pancreatitis or a pancreatic mass. It was discussed that duodenal oedema or thickening caused by eosinophilic infiltration are common findings in these patients. EGE may be considered in the differential diagnosis of unexplained acute pancreatitis, especially in a patient with duodenal oedema on an imaging study or peripheral eosinophilia.

CONCLUSION

We conclude that the diagnostic importance of blood eosinophilia in patients is limited. Nevertheless, blood eosinophilia in these patients is only one of the many diagnostic hints, and it is recommended to independently channelize the extent and course of the diagnostic work-up by supplementary data on risk and exposure, clinical signs and symptoms, and other laboratory results.

REFERENCES

- [1] Pardo J, Carranza C, Muro A, Moreno AA, Martin AM, Martin T, et al. Helminth-related eosinophilia in African immigrants, Gran Canaria. *Emerg Infect Dis*. 2006;12:1587-89.
- [2] Libman MD, MacLean JD, Gyorkos TW. Screening for schistosomiasis, filariasis, and strongyloidiasis among expatriates returning from the tropics. *Clin Infect Dis*. 1993;17:353-39.
- [3] Nutman TB, Ottesen EA, Ieng S, Samuels J, Kimball E, Lutkoski M, et al. Eosinophilia in Southeast Asian refugees: evaluation at a referral center. *J Infect Dis*. 1987;155:309-13.
- [4] Schulte C, Krebs B, Jelinek T, Nothdurft HD, von Sonnenburg F, Loscher T. Diagnostic significance of blood eosinophilia in returning travelers. *Clin Infect Dis*. 2002;34:407-11.
- [5] Seybolt LM, Christiansen D, Barnett ED. Diagnostic evaluation of newly arrived asymptomatic refugees with eosinophilia. *Clin Infect Dis*. 2006;42:363-67.
- [6] Guerrant RL, Walker DH, Weller PF, Wilson ME. Eosinophilia. In: Guerrant RL, Walker DH, Weller PF, editors. *Tropical infectious diseases: principles, pathogens, and practice*. 1st edition. Philadelphia: Churchill Livingstone; 1999. pp. 1400-19.
- [7] Behnke JM, Barnard CJ, Wakelin D. Understanding chronic nematode infections: evolutionary considerations, current hypotheses, and the way forward. *Int J Parasitol*. 1992;22:861-907.
- [8] Pawlowski ZS, Cooper ES, Bundy DAP. Trichuriasis. In: Pawlowski ZS, editor. *Bailliere's clinical tropical medicine and communicable diseases*. 2nd edition. London: Bailliere Tindall Limited; 1987. pp. 629-43.
- [9] Moore TA, Nutman TB. Eosinophilia in the returning traveler. *Infect Dis Clin North Am*. 1998;12:503.
- [10] Weller PF. Eosinophilia in travelers. *Med Clin North Am*. 1992;76:1413.
- [11] Nokes C, Grantham-McGregor SM, Sawyer AW, Cooper ES, Robinson BA, Bundy DA. Moderate to heavy infections to *Trichuris trichiura* affect cognitive function in Jamaican children. *Parasitology*. 1992;104:539-47.
- [12] Aziz E. *Strongyloides stercoralis* infestation: review of the literature and report of 33 cases. *South Med J* 1969;62:806-10.
- [13] Davidson RA. Strongyloidiasis: a presentation of 63 cases. *NC Med J*. 1965;42:23.
- [14] Genta RM, Weesner R, Douce RW, Huitger-O'Connor T, Walzer PD. Strongyloidiasis in US veterans of the Vietnam and other wars. *JAMA*. 1987;258:49-52.

- [15] Grove DJ. Strongyloidiasis in Allied ex-prisoners of war in Southeast Asia. *BMJ*. 1980;280:598-601.
- [16] Gelpi AP, Mustafa A. Seasonal pneumonitis with eosinophilia: a study of larval ascariasis in Saudi Arabia. *Am J Trop Med Hyg*. 1967;16:646-57.
- [17] Fryatt RJ, Teng J, Harries AD, Siorvanes L, Hall AP. Intestinal helminthiasis in expatriates returning to Britain from the tropics: a controlled study. *Trop Geogr Med*. 1990;42:119-22.
- [18] Whitty CJ, Mabey DC, Armstrong M, Wright SG, Chiodini PL. Presentation and outcome of 1107 cases of schistosomiasis from Africa diagnosed in a non-endemic country. *Trans R Soc Trop Med Hyg*. 2000;94:531-34.
- [19] Harries AD, Myers B, Bhattacharya D. Eosinophilia in Caucasians returning from the tropics. *Trans R Soc Trop Med Hyg*. 1986;80:327-28.
- [20] Oncu K, Yazgan Y, Kaplan M, et al. An Extremely Uncommon Case of Parasitic Infection Presenting as Eosinophilic Ascites in a Young Patient. *Case Reports in Gastroenterology*. 2011;5(1):139-143. doi:10.1159/000326927
- [21] Baek MS, Mok YM, Han WC, Kim YS. A Patient with Eosinophilic Gastroenteritis Presenting with Acute Pancreatitis and Ascites. *Gut and Liver*. 2014;8(2):224-27. doi:10.5009/gnl.2014.8.2.224

PARTICULARS OF CONTRIBUTORS:

1. Associate Professor, Department of Microbiology, Kasturba Medical College, Manipal University, Manipal, Karnataka, India.
2. Post Graduate Student, Department of Microbiology, Kasturba Medical College, Manipal University, Manipal, Karnataka, India.
3. Professor, Department of Microbiology, Kasturba Medical College, Manipal University, Manipal, Karnataka, India.
4. Assistant Professor, Department of Pathology, Kasturba Medical College, Manipal University, Manipal, Karnataka, India.

NAME, ADDRESS, E-MAIL ID OF THE CORRESPONDING AUTHOR:

Dr. Vinay Khanna,
Associate Professor, Department of Microbiology, Kasturba Medical College,
Manipal University, Manipal, Karnataka-576104, India.
E-mail: drvinay2004@yahoo.com

Date of Submission: **Nov 21, 2014**Date of Peer Review: **Jan 28, 2015**Date of Acceptance: **Apr 21, 2015**Date of Publishing: **Jul 01, 2015****FINANCIAL OR OTHER COMPETING INTERESTS:** None.