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Rapid T2- and susceptometry-based CMRO2 quantification with interleaved TRUST (iTRUST)

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Abstract

Susceptometry-based oximetry (SBO) and T_2 -relaxation-under-spin-tagging (TRUST) are two promising methods for quantifying the cerebral metabolic rate of oxygen $(CMRO₂)$, a critical parameter of brain function. We present a combined method, interleaved TRUST (iTRUST), which achieves rapid, simultaneous quantification of both susceptometry- and T_2 -based CMRO₂ via insertion of a flow-encoded, dual-echo gradient-recalled echo (OxFlow) module within the *T*¹ recovery portion of the TRUST sequence. In addition to allowing direct comparison between SBO- and TRUST-derived venous oxygen saturation (Y_v) values, iTRUST substantially improves TRUST temporal resolution for $CMRO₂$ quantification and obviates the need for a separate blood flow measurement following TRUST acquisition. iTRUST was compared directly to TRUST and OxFlow alone in three resting subjects at baseline, exhibiting close agreement with the separate techniques and comparable precision. These baseline data as well as simulation results support the use of two instead of the traditional four T_2 preparation times for T_2 fitting, allowing simultaneous quantification of susceptometry- and T_2 -based Y_v (and CMRO₂) with three- and six-second temporal resolution, respectively. In 10 young healthy subjects, iTRUST was applied during a 5% $CO₂$ gas mixture-breathing paradigm. $T₂$ -based Y_v values were lower at baseline relative to susceptometry $(62.3 \pm 3.1 \text{ vs. } 66.7 \pm 5.1 \text{ %HbO}_2, P < 0.05)$, but increased more in response to hypercapnia. As a result, *T*₂-based CMRO₂ decreased from 140.4 ± 9.7 to 120.0 ± 9.5 μ Mol/100 g/ min, a significant – 14.6 ± 3.6% response (P < 0.0001), whereas susceptometry-based CMRO₂ changed insignificantly from 123.4 ± 18.7 to 127.9 ± 25.7 , a 3.3 ± 9.7 % response ($P = 0.31$). These differing results are in accord with previous studies applying the parent OxFlow or TRUST sequences individually, thus supporting the reliability of iTRUST but also strongly suggesting that a systematic bias exists between the susceptometry- and *T*2-based *Y*v quantification techniques.

Keywords

CMRO2; Blood oxygen saturation; TRUST; MR susceptometry; Phase-contrast MRI; Hypercapnia

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Introduction

The human brain comprises only 2% of total body mass, but accounts for approximately 20% of total body oxygen consumption (Rolfe and Brown, 1997). Because the brain is almost entirely dependent on aerobic metabolism to meet its energetic demands, irreversible ischemic damage will result in minutes if oxygen delivery is disrupted. Unlike surrogate markers of metabolism such as cerebral blood flow (CBF) or blood-oxygen-level dependent (BOLD) functional MRI (fMRI) signal, the cerebral metabolic rate of oxygen (CMRO $_2$) provides a direct measure of brain oxygen consumption. CMRO₂ changes significantly over the course of neonatal development (Liu et al., 2014a) and aging (Peng et al., 2014), and is altered in many of the most common neurologic diseases, including mild cognitive impairment (Liu et al., 2014b) and Alzheimer's disease (Ishii et al., 1996), Parkinson's disease (Borghammer et al., 2010), and multiple sclerosis (Ge et al., 2012). However, $CMRO₂$ is relatively stable across healthy subjects at baseline (Xu et al., 2009; Jain et al., 2010), and in response to physiologic challenges such as hypercapnia (Chen and Pike, 2010; Xu et al., 2011; Jain et al., 2011), hypoxia (Xu et al., 2012a), and apnea (Rodgers et al., 2013a). Thus, CMRO₂ is an important quantity for understanding brain function in health and disease.

The gold standard for $CMRO₂$ quantification is triple-oxygen positron emission tomography (PET) imaging (Ito et al., 2005), yet the technique is rarely applied in humans due to the radiation exposure and complexity of the protocol. Moreover, long scan times restrict PET to measuring resting-state CMRO2. MRI provides a non-invasive, non-contrast alternative. During the past two decades, BOLD fMRI has been applied extensively to study neuronal activation in health and disease (Matthews et al., 2006; Kim and Ogawa, 2012). However, BOLD signal does not provide a direct measure of brain oxygen metabolism, but rather reflects a complex interplay between CBF, cerebral blood volume, and tissue properties such blood vessel diameter, in addition to $CMRO₂$ (Blockley et al., 2013).

Recently, a number of MR-based approaches for direct quantification of cerebral venous oxygen saturation (*Y*v) have been proposed (Jain et al., 2010, 2014; Haacke et al., 1997; Fernandez-Seara et al., 2006; Fan et al., 2012, 2014; Wright et al., 1991; Golay et al., 2001; Lu and Ge, 2008; Chen and Pike, 2009; Bolar et al., 2011; Qin et al., 2011; Guo and Wong, 2012; Krishnamurthy et al., 2013; An and Lin, 2000; He and Yablonskiy, 2007; Bulte et al., 2012; Gauthier et al., 2012). In combination with phase-contrast MRI (PC-MRI) or arterial spin labeling (ASL) CBF quantification, these techniques allow for determination of CMRO2 via the Fick Principle (Kety and Schmidt, 1948):

$$
CMRO2=Ca · CBF · (Ya - Yv) (1)
$$

where C_a is the arterial oxygen content of blood in μ Mol/100 mL and Y_a the arterial oxygen saturation in percent hemoglobin oxygen saturation $(\%HbO₂)$, which can be measured with pulse oximetry. Total CBF (tCBF) is typically reported in units of mL blood/100 g brain tissue/minute, giving $CMRO₂$ in units of μ Mol/100 g/minute.

Measurement of Y_v poses the most significant technical challenge in CMRO₂ determination. Techniques for Y_v quantification can be categorized based on the contrast mechanism – venous blood magnetic susceptibility (Jain et al., 2010; Haacke et al., 1997; Fernandez-Seara et al., 2006; Fan et al., 2012, 2014), T_2 (Wright et al., 1991; Golay et al., 2001; Lu and Ge, 2008; Chen and Pike, 2009; Bolar et al., 2011; Qin et al., 2011; Guo and Wong, 2012; Jain et al., 2014; Krishnamurthy et al., 2013), *T*2′ (An and Lin, 2000; He and Yablonskiy, 2007), or T_2^* (BOLD) (Bulte et al., 2012; Gauthier et al., 2012) – as well as spatial specificity – large-vessel/whole-brain (Jain et al., 2010, 2014; Haacke et al., 1997; Fernandez-Seara et al., 2006; Fan et al., 2012; Wright et al., 1991; Golay et al., 2001; Lu and Ge, 2008; Chen and Pike, 2009; Qin et al., 2011), small-vessel/regional (Fan et al., 2014; Krishnamurthy et al., 2013), or parenchymal/voxel-wise (Bolar et al., 2011; Guo and Wong, 2012; An and Lin, 2000; He and Yablonskiy, 2007; Bulte et al., 2012; Gauthier et al., 2012). Regional and voxel-wise approaches are clearly desirable due to the heterogeneous nature of brain functional activation and pathology. However, these techniques have scan times on the order of several minutes, precluding dynamic measurements, and tend to suffer from low signal-to-noise ratio (SNR), requiring significant spatial averaging to achieve acceptable precision and thus negating the utility of regional or voxel-wise measurement. In comparison, techniques for whole-brain Y_v quantification are fast, robust, and easy to implement.

The two best-established methods for global Y_v quantification are T_2 -relaxation-under-spintagging (TRUST) (Lu and Ge, 2008) and susceptometry-based oximetry (SBO) (Jain et al., 2010). Both methods involve quantification of intravascular Y_v in the superior sagittal sinus (SSS), the largest cerebral venous drainage vessel, which, in combination with PC-MRI quantification of tCBF, can be used to determine $CMRO₂$ via Eq. (1). In the case of TRUST, tCBF measurement requires a separate PC-MRI acquisition (Xu et al., 2009). However, because PC-MRI and SBO are both gradient-recalled echo (GRE) sequences, they can be naturally combined into a single sequence, which we term OxFlow. This hybrid sequence was originally implemented via a two-slice interleaved approach with CMRO₂ quantification temporal resolution of 25 seconds (Jain et al., 2010). Recently, addition of view-sharing and SSS-based estimation of tCBF improved OxFlow temporal resolution to three seconds, allowing study of the regulation of $CMRO₂$ in response to dynamic physiologic paradigms such as breath-hold apnea (Rodgers et al., 2013a). Compared to OxFlow, TRUST has inherently lower temporal resolution, compounded by the need for a separate PC-MRI measurement to quantify CMRO2. Furthermore, the relationship between *T*2, *Y*v, and hematocrit (Hct) is non-linear, and must be calibrated to both pulse sequence parameters and field strength. However, unlike SBO, TRUST is vessel geometry independent, less sensitive to partial volume effects, and does not require background phase removal.

A particularly important application of $CMRO₂$ quantification is investigating the metabolic response to hypercapnia. Hypercapnia is relevant to a number of common diseases, including asthma, chronic obstructive pulmonary disease, obstructive sleep apnea, and congestive heart failure. Furthermore, knowledge of the $CMRO₂$ response to hypercapnia is of substantial importance to functional imaging, where hypercapnia is routinely used for

'calibrating' the fMRI signal (Davis et al., 1998), often under the assumption that hypercapnia is isometabolic (i.e., does not affect $CMRO₂$). However, the $CMRO₂$ response to a hypercapnic stimulus remains controversial (Yablonskiy, 2011), with previous studies reporting a wide range of results from reduced, to unchanged, to increased CMRO₂. An early MRI study using T_2 -based Y_v quantification reported an isometabolic response (Chen and Pike, 2010); however, $CMRO₂$ responses to mild and moderate hypercapnia were in different directions (5.0% and — 6.8%, respectively) and based on a calibration plot derived from room temperature blood samples (Chen and Pike, 2009), potentially impacting the accuracy of in vivo T_2 quantification (Spees et al., 2001). Subsequently, both OxFlow and TRUST have been applied to study the $CMRO₂$ response to hypercapnia using similar cohorts and experimental protocols involving a 5% $CO₂$ gas mixture delivery (Xu et al., 2011; Jain et al., 2011). While OxFlow data supported an isometabolic $CO₂$ response, the TRUST study found a significant $13.4 \pm 2.3\%$ (mean \pm standard error, $N = 14$) decrease in CMRO2. This discrepancy is disconcerting given both the importance of understanding the CMRO2 response to hypercapnia as well as the increasing application of TRUST and OxFlow in studying $CMRO₂$ responses to other stimuli and disease states. A recent study directly comparing resting TRUST- and SBO-derived Y_v values in the same cohort (Barhoum et al., 2014) found TRUST Y_v values to be slightly lower (mean \pm SD of 63.2 \pm 4.1 vs. 65.9 ± 3.3 %HbO₂, $P < 0.01$). However, this baseline difference does not by itself explain the discrepancy in the hypercapnia results, which depends on the relative change in *Y_v* in response to the stimulus.

In this work, we propose a combined technique – termed interleaved TRUST (iTRUST) – whereby an OxFlow module is interleaved within the T_1 recovery period of the TRUST sequence. This approach has two distinct benefits. First, it obviates the need for separate, non-simultaneous measurement of tCBF following the TRUST acquisition, substantially improving TRUST temporal resolution for CMRO₂ quantification. Second, it allows for direct comparison of Y_v quantified via magnetic susceptibility and T_2 measurement of blood. Further temporal acceleration compared to TRUST is achieved by using fewer tag-control image pairs for *T*2 fitting. Both the combination of the techniques as well as the use of fewer *T*2 fitting points are validated in simulations and in vivo. The sensitivity of the technique to detect dynamic changes is demonstrated in response to breath-hold apnea. Finally, iTRUST is applied in a cohort of young healthy subjects during a $CO₂$ gas mixture-breathing paradigm with the goal of further investigating the potential disagreement between the TRUST and OxFlow techniques with regard to the hypercapnic $CMRO₂$ response.

Theory

Susceptometry-based quantification of Yv (SBO)

Susceptometry-based oximetry exploits the relative paramagnetism of deoxygenated versus oxygenated hemoglobin, which causes the susceptibility of blood relative to surrounding tissue, χ , to be linearly related to venous oxygen saturation:

$$
\Delta \chi = \text{Hct} \cdot (\Delta \chi_{\text{do}}(1 - Y_v) + \Delta \chi_{\text{oxy}}) \quad (2)
$$

where χ_{do} and χ_{oxy} are the experimentally determined volume susceptibility differences between fully oxygenated and deoxygenated erythrocytes and fully oxygenated erythrocytes and water, respectively. Values of $4\pi \times 0.273$ and $4\pi \times 0.008$ p.p.m. (SI units) are used for χ_{do} and χ_{oxy} , based on ex vivo calibration experiments (Spees et al., 2001; Jain et al., 2012).

Blood susceptibility induces a local field offset, \overline{B} , which can be measured with a field mapping sequence as:

$$
\Delta B = \Delta \phi / \gamma \Delta TE \quad (3)
$$

where φ is the difference in phase accrual between echoes spaced apart by TE in the blood versus surrounding reference tissue. By modeling the vessel of interest as an infinitely long, circular cylinder, the relationship between B and χ can be calculated analytically:

$$
\Delta B = \frac{1}{6} \Delta \chi B_0 (3 \cos^2 \theta - 1) \quad (4)
$$

where θ is the vessel angle with respect to the main magnetic field, B₀. Combining Eqs. (2)– (4) allows for determination of Y_v by measurement of φ .

The SSS, the largest cerebral venous drainage vessel, is relatively long and straight when the subject is lying supine in the scanner, and can therefore be effectively approximated by the infinite cylinder model, despite its non-circular cross-section (Jain et al., 2010; Li et al., 2012). The SSS has also been shown to have a Y_v nearly identical to that in the internal jugular vein (Xu et al., 2009; Jain et al., 2014), making it an excellent surrogate for global venous *Y*v. Furthermore, while field mapping of the internal jugular vein is complicated by the presence of trachea-induced susceptibility artifacts, the field local to the SSS is relatively homogenous.

Combination of SBO and PC-MRI for CMRO2 quantification (OxFlow)

SBO can be combined with PC-MRI blood flow quantification to allow simultaneous measurement of Y_v , tCBF, and, therefore, CMRO₂, from a single sequence. By adding flow encoding to the same dual-echo GRE used for Y_v quantification, SSS blood flow (SSSBF) and Y_v can be quantified from data acquired in the same TR period. SSSBF can then be retrospectively up-scaled to tCBF based on a single measurement of the SSSBF:tCBF ratio at baseline (Rodgers et al., 2013a).

In this study, OxFlow was implemented with a BRISK k-space sampling scheme, with onequarter k-space acquired at each time point (Doyle et al., 1995). BRISK provides reduced motion sensitivity compared to previous view-sharing implementations of OxFlow using Keyhole k-space sampling (Rodgers et al., 2013b; Van Vaals et al., 1993). BRISK images were reconstructed by interpolating across time points using the nearest acquired data at each k-space segment, effectively resulting in a sliding window reconstruction with minimum window width of three seconds (inner 1/8th of k-space) and maximum window

width of 60 seconds (outer 5/8th of k-space). Other OxFlow sequence parameters include: TR/TE₁/TE₂ = 14.2/6.5/11.5 ms, VENC = 40 cm/s, reconstructed matrix = 192 \times 192, and resolution = $1.0 \times 1.0 \times 5.0$ mm.

TRUST and interleaved TRUST (iTRUST)

The TRUST pulse sequence uses a non-selective MLEV-16 CPMG T_2 preparation of varying effective echo time (eTE) – 0, 40, 80, and 160 ms – following either an 8 ms adiabatic hyperbolic secant pulse (bandwidth $= 2214$ Hz, thickness $= 100$ mm) to invert the blood magnetization (tag), or application of an equivalent off-resonance pulse without gradient (control). Similar in principle to ASL, tag-control subtraction of each eTE image pair isolates the venous blood signal. A nonselective 90° spoiler RF pulse is applied to reset the magnetization before each tag-control module (Xu et al., 2012b). A two-compartment exchange model is used to relate Y_v to T_2 :

$$
1/T_2 = A + B \cdot (1 - Y_v) + C \cdot (1 - Y_v)^2
$$
 (5)

where A , B , and C are Hct- and CPMG spacing (t_{CPMG}) -dependent constants which have been determined from ex vivo blood samples (Lu et al., 2012). T_2 is quantified by monoexponential fitting of SSS tag-control difference signals versus eTE as:

$$
\Delta S = S_0 e^{\text{E} \cdot (1/T_1 - 1/T_2)} \quad (6)
$$

where S_0 is the difference signal at $eTE = 0$ and a T_1 value of 1.613 seconds is assumed for venous blood (Xu et al., 2012b).

The TRUST sequence used in the present work follows that described in recent literature (Xu et al., 2012b), with a TR of three seconds used to provide an optimal tradeoff between scan duration, accuracy, and precision, allowing a single Y_v value to be quantified every 24 seconds. Important differences relative to the published sequence include insertion of a slice-selective saturation pulse 200 ms before EPI readout (prior to T_2 preparation) in order to better suppress static tissue signal (Qin et al., 2011; Jain et al., 2014), and use of a flowcompensated EPI readout with TE of 8 ms (5/8th partial Fourier readout). Flow compensation prevents flow velocity-dependent signal variations between tag and control images, which could lead to errors in the difference signals, especially in situations of rapidly changing flow (Xu et al., 2012b). An alternative approach to avoiding these effects is use of a shorter TE achieved via parallel imaging (Xu et al., 2012b), though this reduces SNR Other TRUST sequence parameters include: $t_{CPMG} = 10$ ms, reconstructed matrix = 64 \times 64, and resolution = 3.4 \times 3.4 \times 5.0 mm.

More than half of the duration of the TRUST sequence consists of dead time, required to allow blood signal to undergo sufficient T_1 recovery following global saturation before the next T_2 preparation. In iTRUST, this time is utilized to run an OxFlow module at the same location as the TRUST readout slice (Fig. 1), beginning 350 ms after the saturation in order to capture the tissue signal at approximately its steady-state longitudinal magnetization. Besides the added OxFlow module, iTRUST is otherwise identical to TRUST.

It is important to note that the RF pulses played out during the OxFlow module only affect spins in the imaging slice, whereas spins relevant to T_2 -quantification are located outside the imaging slice in the labeling slab. Furthermore, because the OxFlow module is run during both tag and control, any effect on spins in the subsequently acquired EPI images used for T_2 quantification should be identical, and hence removed by tag-control subtraction. Likewise, the OxFlow GRE acquisition itself is unaffected by the TRUST sequence because it is acquired only after global magnetization reset.

Materials and methods

Human subject protocols

All human subject imaging protocols were approved by the University of Pennsylvania's Institutional Review Board, and subjects provided written informed consent prior to participation. Studies were performed on 10 healthy subjects (age 29 ± 5 years, range $24-42$, six males and four females) using a 3 T Siemens Tim Trio system (Siemens Medical Solutions, Erlangen, Germany) with a 12-channel (validation study and apnea study) or 32 channel (hypercapnia study) receive-only head coil. A vendor provided time-of-flight axial localizer scan was used for slice selection, and retrospectively to determine θ in Eq. (4). Before each OxFlow or iTRUST acquisition, a two-slice interleaved PC-MRI pulse sequence was run at the level of the internal carotid and vertebral arteries in the neck and the SSS in the head in order to determine the subject's SSSBF:tCBF ratio. OxFlow, TRUST, and iTRUST pulse sequences were programmed in SequenceTree (Magland and Wehrli, 2006).

At the end of each scanning session, a 1-mm-isotropic 3D *T*1-weighted MPRAGE (Mugler and Brookeman, 1990) data set was acquired so that tCBF could be normalized per unit brain mass in each subject. Total brain volume was obtained using the BET tool in FSL (Smith, 2002), and converted to mass based on an average brain density of 1.05 g/mL (Herscovitch and Raichle, 1985). Total intracranial mass (gray matter, white matter, and cerebrospinal fluid (CSF)) rather than total parenchymal mass (gray matter and white matter) was used for normalization to facilitate comparison of $CMRO₂$ values with prior studies that did the same (Xu et al., 2009; Jain et al., 2010). It has recently been shown that inclusion of CSF volumes inflow normalization may bias toward underestimation of $CMRO₂$ in older individuals (Peng et al., 2014), though this is not a concern in the present study due to the relatively young age of the subjects.

Validation study

To test whether the combination of OxFlow and TRUST causes a bias in the measurements of either sequence, equivalent OxFlow, TRUST, and iTRUST sequences were run back to back for four minutes each in three subjects (age 29 ± 3 years, range 26–34, two males and one female). This protocol corresponds to 10 repetitions of TRUST and iTRUST with 24 second temporal resolution, and 80 repetitions of OxFlow with three-second temporal resolution. For the OxFlow only sequence, TR was increased to 31.25 ms to use the entire three-second time frame with sequence parameters otherwise equal to the iTRUST-inserted OxFlow module.

For each subject, T_2 -based Y_v (Y_v - T_2) was derived from TRUST and iTRUST data, and SBO-based Y_V (Y_V -SBO) and tCBF from iTRUST and OxFlow data. Differences in parameter values across subjects were compared between techniques – TRUST vs. iTRUST for Y_v - T_2 , OxFlow vs. iTRUST for Y_v -SBO and tCBF – to determine any potential bias in the interleaved approach relative to the separate techniques. Further, T_2 values obtained from the iTRUST data were recalculated using only the 0 and 80 ms eTE image pairs to determine any bias caused by using fewer eTEs. T_2 fitting with two eTEs has previously been demonstrated at 7 T field strength (Krishnamurthy et al., 2014), where the short *T*² value of blood precludes the use of longer T_2 preparations.

Simulations

The use of fewer eTE images pairs was further explored by simulating TRUST difference signals with a blood T_2 value of 72 ms, corresponding to typical physiologic values of Y_v = 65% HbO₂ and Hct = 0.40, with noise added corresponding to the typically observed SNR range of our acquired TRUST data (SNR = 20-80). This SNR range is similar to that reported in previous studies (Xu et al., 2012b). Exponential fitting was performed and *Y*^v values were determined from the published calibration curve (Lu et al., 2012) using all four (0, 40, 80, 160 ms), three (0, 40, and 80 ms), or two (0 and 80 ms) eTEs. Root-mean-square error (RMSE) relative to the true Y_v of 65 %HbO₂ was quantified as a function of SNR and number of eTEs used.

Apnea study

To evaluate the sensitivity of the iTRUST technique to detect dynamic changes inflow, *Y*v-SBO, and Y_v-T_2 , a breath-hold challenge was conducted in one healthy subject (age 28 years, male). iTRUST was run with two eTEs (0 and 80 ms) during a paradigm consisting of two minutes baseline, one-minute breath hold after inhalation, and two minutes recovery. Y_v -SBO and tCBF were quantified every three seconds. Y_v - T_2 values were quantified with sliding-window reconstruction using all adjacent difference image pairs, yielding six-second temporal resolution from 12-second data windows. The mean and standard deviation of the difference between time matched Y_v-T_2 and Y_v -SBO values was quantified across all time points, and compared by paired two-sample Student's *t*-tests.

Hypercapnia study

In 10 subjects, iTRUST comprising only two eTEs (0 and 80 ms) was applied during a hypercapnia paradigm to determine whether differences exist in the CMRO₂ as determined via T_2 - versus susceptometry-based quantification of Y_v . A two-way non-rebreathing Tvalve (2700 Series, Hans Rudolph, Inc., Kansas City, MO, USA) was used to deliver 5% CO2 in room air for five minutes via a 100 L Douglas bag. Room air was delivered five minutes before and after hypercapnia, and MRI data were collected continuously for the entire 15 minutes. *Y*_a and heart rate (HR) were monitored with pulse oximetry, and end-tidal $CO₂$ (EtCO₂) and respiratory rate (RR) with capnography (Expression, Invivo Research Inc., Orlando, FL, USA).

tCBF, *Y*_v-SBO, and SBO-based CMRO₂ (CMRO₂-SBO) parameter values were determined from the OxFlow data at three-second temporal resolution, and *Y*^a values were sampled at

three-second intervals to match the MRI data. Y_v-T_2 values were quantified every six seconds from the EPI data with 12-second sliding-window reconstruction. tCBF and *Y*^a values were interpolated to the corresponding Y_v-T_2 time points to determine T_2 -based $CMRO₂ (CMRO₂-T₂)$ values every six seconds. For each parameter, means and standard deviations were quantified across the baseline (0-5 minutes) and steady-state hypercapnia (7.5–10 minutes) periods, and used to determine percent changes in response to hypercapnia. Changes in $CMRO₂-T₂$ and $CMRO₂-SBO$ in response to hypercapnia were evaluated with one-sample Student's *t*-tests.

Image analysis

All image reconstructions were performed with in-house-written Matlab (Mathworks, Natick, MA) scripts. BRISK-sampled raw OxFlow data, whether acquired alone or as part of an iTRUST sequence, were first reordered to create full k-space images corresponding to each three-second time point. To determine tCBF, the phase difference between positive gradient-moment flow-encoded and flow-compensated images acquired at $TE₁$ were used to generate velocity maps, and SSSBF was obtained by integrating velocity across the vessel cross-sectional area. Data from the two-slice interleaved PC-MRI sequence used to determine the SSSBF:tCBF ratio were processed analogously. This ratio was then used to upscale the dynamically acquired SSSBF data to determine tCBF.

For *Y*v-SBO determination, a raw phase difference map was generated from images acquired at TE_1 and TE_2 of the flow-compensated OxFlow interleave. Low spatial frequency bulk susceptibility effects were removed via second-order polynomial fitting of the induced field in the surrounding brain tissue (Langham et al., 2009). The average phase difference, φ , was determined between pixels entirely within the SSS (i.e., without any tissue partial voluming) and pixels in a reference region of brain tissue surrounding the SSS approximately one vessel-radius in width and located one vessel-radius from the SSS border, allowing determination of Y_v -SBO from Eqs. (2)–(4).

TRUST or iTRUST EPI data for *T*2-determiation were first reconstructed and corrected for N/2 ghosting. Difference images were produced for each eTE via tag-control subtraction. As previously described (Lu and Ge, 2008), the four brightest pixels in the SSS were selected for *T*2 fitting, using a weighted least-squares fit calculated by the Matlab function lscov.m.

Results

Across the three subjects scanned at baseline, quantified Y_v and tCBF values were consistent with previous reports (Xu et al., 2009; Jain et al., 2010), and mean absolute bias between TRUST and iTRUST $Y_v - T_2$ (Fig. 2A) and between OxFlow and iTRUST Y_v -SBO (Fig. 2B) and tCBF (Fig. 2C) values were small. These values likely represent an upper bound on any true bias, as they also include contributions from measurement noise and true physiologic variation over the scan duration. Standard deviations of the parameter values varied across subjects, but were similar between techniques, suggesting precision of the combined iTRUST sequence to be comparable to the separate TRUST and OxFlow sequences.

In Fig. 3, iTRUST Y_v-T_2 values are shown based on T_2 fitting using all four (0, 40, 80, and 160 ms) or just two (0 and 80 ms) eTE difference signals. The mean \pm SD difference between the two sets of values was small at 0.2 ± 1.8 %HbO₂ ($P = 0.65$). The 95% confidence interval for the linear least-squares regression line includes the line of identity, further indicating that no significant bias is introduced by using two instead of four eTEs. *Y*v-*T*2 variability was slightly larger when using two versus four eTEs (subject-averaged SDs of 2.6 and 1.6 %HbO₂, respectively). However, this difference is largely eliminated if RMSEs are scan-time normalized, that is, after multiplying by $\sqrt{\text{number e} \cdot \text{TEs}}$ used for fitting. These data support the use of two eTEs in subsequent iTRUST experiments.

Fig. 4 shows RMSEs for Y_v-T_2 values across the typical TRUST SNR range, both absolute (Fig. 4A) and scan-time normalized (Fig. 4B). Even before normalization, three and two eTEs result in less error than four eTEs. Normalized for scan time, both three and two eTEs perform significantly better than four eTEs, with ≈30%–45% reduction in RMSE across the SNR range.

iTRUST with two eTEs was evaluated in response to breath-hold apnea to test the ability of the technique to detect dynamic physiologic processes. A time-course plot of the extracted parameter values (Fig. 5) demonstrates the expected apneic response of increased Y_v and tCBF (Rodgers et al., 2013a). Y_v -SBO and Y_v - T_2 values match closely, with Y_v -SBO values higher by an average of 1.5 ± 3.0 %HbO₂ (*P* < 0.01).

All 10 subjects were able to successfully complete the hypercapnia paradigm. Average brain volume, Hct, and SSS angle (θ) were 1468 \pm 77 mL, 0.43 \pm 0.04, and 15.2° \pm 5.0°, respectively. On average, the SSSBF:tCBF ratio was 0.48 ± 0.03 , in line with previous studies (Rodgers et al., 2013a). Subject-averaged time-course plots of physiologic parameters measured via pulse oximetry (Y_a, HR) and capnography (ECO_2, RR) are displayed in Fig. 6.

Fig. 7 displays a representative subject time-course plot of all MRI-derived parameters (and *Y*a) in absolute physiologic units (Fig. 7A), subject-averaged plots of both absolute parameter values (Fig. 7B) and baseline-normalized parameter values (Fig. 7C), and a scatter plot comparing Y_v -SBO and Y_v - T_2 values across all subjects and time points (Fig. 7D). Parameter values were observed to reach a steady state after approximately 2.5 minutes of hypercapnia. Average baseline (0–5 minutes) and steady-state hypercapnia (7.5–10 minutes) values are displayed in Table 1. EtCO₂, tCBF, Y_v -SBO, and Y_v - T_2 all increased significantly in response to hypercapnia (*P* < 0.0001). Subject-averaged cerebrovascular reactivity was $4.6 \pm 0.9\%$ tCBF/mm Hg EtCO₂, in line with previous results (Chen and Pike, 2010; Jain et al., 2011).

*Y*v-*T*2, although lower than *Y*v-SBO at baseline, increased more during hypercapnia. As a result, in response to hypercapnia CMRO₂-SBO did not change significantly $(3.3 \pm 9.7\%)$, *P* $= 0.31$), whereas CMRO₂-*T*₂ decreased substantially (− 14.6 ± 3.6%, *P* < 0.0001). Following cessation of apnea, tCBF and Y_v undershot before gradually returning to baseline. $CMRO₂$ values during the end-recovery period (12.5–15 minutes) were not significantly

different from baseline values ($P = 0.36$ and $P = 0.33$ for CMRO₂-SBO and CMRO₂-T₂, respectively).

Discussion

Validation of iTRUST

Because changes inflow and Y_v tend to oppose each other both at baseline and in response to stimuli, it is critical to measure these two quantities simultaneously to most accurately determine CMRO₂. This is especially important during physiologic stimuli, where temporal mismatch between Y_v and flow quantification could lead to significant errors. iTRUST makes such simultaneous measurement, previously achievable only with susceptometrybased CMRO₂ approaches, possible for *T*₂-based CMRO₂ quantification as well.

Combination of the TRUST and OxFlow techniques in iTRUST did not significantly impact the accuracy or precision of the quantified parameters (Fig. 2). This is expected, as the OxFlow and *T*₂-quantificaiton portions of the pulse sequence are separated in such a way that they should not affect one another's spin histories. While less time is available for OxFlow measurement in iTRUST than OxFlow alone for a given temporal resolution (1420 ms versus 3000 ms in this study), this did not appear to impact the precision of the OxFlow data as evidenced by similar standard deviations for iTRUST and OxFlow derived parameters (Fig. 2B and C).

T2-based CMRO2 temporal resolution

Previous implementations of TRUST had a temporal resolution for CMRO₂ quantification of several minutes (Liu et al., 2013), compared to as little as three seconds for OxFlow (Rodgers et al., 2013a). This is partially due to the usual acquisition of three TRUST averages (requiring $3 \times 24 = 72$ seconds) and measurement of each arterial inflow vessel with a separate 30 seconds PC-MRI measurement, which has been demonstrated to produce accurate and reproducible $CMRO₂$ measurements (Liu et al., 2013). While this approach is optimal when a single $CMRO₂$ measure of baseline physiology is the objective, it does not allow for quantification of dynamic changes in T_2 -based $Y_v/CMRO_2$.

iTRUST improves $CMRO₂-T₂$ temporal resolution to as little as six seconds via insertion of flow quantification within the T_1 recovery period and use of two eTEs with sliding window reconstruction. These modifications may also improve measurement precision. For instance, rapid measurement of SSSBF is achieved more easily than quantification of tCBF in the neck arteries, due to the sagittal sinus' larger size, less pulsatile flow, and fixed position in the scanner even during swallowing, breath hold, or gas mixture breathing manipulations that can complicate flow quantification in the neck arteries. Upscaling this dynamically acquired SSSBF to tCBF only requires a single high-quality PC-MRI acquisition before or after accelerated SSSBF-only measurement, since the SSSBF:tCBF ratio has ben observed to remain fixed in response to blood flow changes (Rodgers et al., 2013a).

Simulation results (Fig. 4) suggest that inclusion of a 160 ms eTE difference image actually reduces T_2 estimation precision due to its relatively low SNR. For iTRUST at 3 T, T_2 measurement based on two eTEs performs slightly better than three eTEs after

normalization for scan time differences. This is because an eTE of 80 ms most closely matches the physiologic T_2 range (60-100 ms). In vivo measurements at baseline suggest a slightly greater Y_v variation when retrospectively using two vs. four eTEs for T_2 fitting (SDs of 2.6 versus 1.6 %HbO2, respectively). However, this greater variation likely reflects some degree of sensitivity to true physiologic fluctuations – absent in the simulation data – which is more significantly removed through averaging when using all four eTEs for fitting. One limitation of using only two eTEs is that confidence intervals for the exponential fitting (and therefore $Y_{\rm v}$) cannot be derived based on the regression of the exponential fit.

Hypercapnia study

Parameter values quantified from the hypercapnia data were in good agreement with previous studies using TRUST or OxFlow independently, both in terms of resting state values (Xu et al., 2009; Jain et al., 2010) and changes in response to hypercapnia (Xu et al., 2011; Jain et al., 2011). Specifically, hypercapnia caused significant reduction in CMRO2- T_2 (− 14.6 ± 3.6%, mean ± SD), similar to the original TRUST study (Xu et al., 2011) (− 13.4 \pm 8.6%, mean \pm SD, calculated from the reported standard error with *N* = 14), and also a non-significant change in CMRO₂-SBO (3.3 \pm 9.7%, mean \pm SD), similar to the original OxFlow study (Jain et al., 2011). It was suggested (Jain et al., 2011) that the negative hypercapnic response observed with TRUST could have been biased due to flow measurement in the SSS, rather than in the neck arteries as was done with OxFlow. However, the present study used only SSS-based flow quantification, yet achieved results consistent with both previous studies (Xu et al., 2011; Jain et al., 2011). This consistency lends additional support to the use of SSS-based quantification of tCBF, a critical requirement for obtaining high temporal resolution $CMRO₂$ quantification with OxFlow and iTRUST. It also strengthens confidence in the accuracy of the iTRUST technique for Y_v-T_2 quantification, including use of two eTEs for T_2 fitting. Most strikingly, it implies that the observed bias between Y_v - T_2 and Y_v -SBO values – both in terms of the baseline offset and relative changes in response to hypercapnia – is not due to random error, differences in experimental protocols, or differences in subject populations, but rather a systematic bias between the techniques.

Average baseline $Y_v - T_2$ values were observed to be significantly lower than Y_v -SBO values $(62.3 \pm 3.1 \text{ vs. } 66.7 \pm 5.1 \text{ %HbO}_2)$, respectively, $P < 0.05$), consistent with another recent study (Barhoum et al., 2014). Longer TRUST EPI readouts have been shown to cause a systematic underestimation of T_2 (and therefore Y_v), especially at lower SNR (Xu et al., 2012b). This effect was hypothesized to be caused by variations in blood flow, and led the authors to recommend use of a shorter (3 ms) EPI TE via application of parallel imaging. While the present study used a longer TE (8 ms), the slice-select gradient was first momentcompensated. This should prevent signal differences due to varying degrees of intravoxel dephasing in tag/control images acquired at different blood flow velocities. Furthermore, if a flow velocity-dependent bias did exist, the proposed Y_v-T_2 underestimation would be expected to get worse at higher Y_v values due to the accompanying CBF and heart rate increase during hypercapnia. In fact, the opposite trend was observed, with Y_v - T_2 values rising significantly more than *Y*_v-SBO values during the hypercapnic stimulus, regardless of the baseline offset between Y_v-T_2 and Y_v -SBO. This is illustrated by the subject specific

regression lines in Fig. 7D, all of which had slopes significantly greater than unity (β = 1.47 \pm 0.20, *P* < 0.0001 for H₀: β = 1).

As recently described by Xu et al. (2014), a flow-dependent error in *Y*_v-SBO values could potentially arise due to phase accumulation as venous blood travels through an inhomogeneous B_0 field. This flow-dependent phase accumulation will increase quadratically with echo time and linearly with the dot product between the flow velocity and the background field gradient. Were the background field gradient direction similar in each subject, it could cause a systematic bias toward flow-dependent over- or under-estimation of Y_v -SBO. However, this effect alone cannot explain the observed bias between Y_v - T_2 and Y_v -SBO values, as it would predict the bias to increase in magnitude with increasing flow velocity, whereas the observed bias reverses direction between the low flow (baseline) and high flow (hypercapnia) states. Detailed investigation of this potential source of error will be explored in future work by applying a quadratic phase model – the "adaptive-quadratic fit" as described by Xu et al. (2014) – to an SBO sequence with several rephased echoes and longer echo times.

In addition to the aforementioned flow effects, another likely source of the observed discrepancy is an error in the calibration of one or both techniques – that is, the values of the constants in the model equations. However, the susceptibility model (Eq. (2)) is considerably simpler, with only two calibration constants – χ_{do} and χ_{oxy} – defining a linear relationship between measured phase, Hct, and *Y*v. The values of these constants have been validated theoretically (Spees et al., 2001) and experimentally (Spees et al., 2001; Jain et al., 2012) with excellent agreement. In contrast, TRUST requires calibration of a quadratic equation (Eq. (5)) with six linear coefficients (Lu et al., 2012). This calibration equation is based on a two-compartment exchange model, which may be less appropriate than an alternative diffusion-based model (Gardener et al., 2010). Furthermore, unlike SBO, *T*2-based *Y*v quantification has a complex dependence on field strength and pulse sequence parameters (e.g., RF inversion pulse, t_{CPMG}).

Applications of iTRUST

In this work, we were interested in directly comparing T_2 - and susceptometry-based Y_v CMRO2 values; however, iTRUST could also be used specifically as a high temporal resolution T_2 -based CMRO₂ quantification technique. In this case, a single rather than dualecho PC-MRI sequence module could be used, allowing for reduction in the required viewsharing factor. Such a technique could be applied to $CMRO₂$ quantification in the jugular vein, which is less well suited to SBO because of trachea-induced susceptibility artifacts.

A potential clinical application of iTRUST is the assessment of cerebro-vascular reserve (CVR), the ability of the brain to dynamically increase flow in response to a vasodilatory challenge such as hypercapnia or breath hold apnea. Reduced CVR is strongly correlated with increased stroke risk (Gupta et al., 2012) and associated with lower cognitive performance in subjects with mild cognitive impairment and Alzheimer's disease (Richiardi et al., 2014). While CVR has typically been assessed in terms of blood flow changes only, iTRUST and similar techniques for rapid CMRO2 quantification (Rodgers et al., 2013a) allow multi-parametric assessment of the brain's response to stimuli. Because CMRO₂ is a

more direct reflection of oxygen supply and demand, CVR assessed in terms of $CMRO₂$ may provide a more meaningful index of neurovascular dysfunction than traditional flowbased CVR.

The described approach of inserting a fast imaging sequence within a longitudinal signal recovery period has applications beyond iTRUST. T_2 -relaxation-under-phase-contrast (TRU-PC), which uses phase-contrast rather than tag-control isolation of venous blood (Krishnamurthy et al., 2013), and which can probe vessels as small as 1 mm, contains an equivalent signal waiting period as in TRUST. Addition of flow quantification within TRU-PC would provide a means of quantifying oxygen flux rather than simply oxygen saturation in small regional vessels not suitable to SBO. An interleaved approach similar to iTRUST has been used to quantify perfusion, intravascular venous oxygen saturation (Y_y) , and T_2^* (termed PIVOT) via insertion of a multi-echo GRE within the post-label delay of a pulsed ASL sequence (Englund et al., 2013). The technique allowed simultaneous measurement of all three parameters with two-second temporal resolution during a reactive hyperemia paradigm in the leg. Such combination of perfusion and Y_v quantification may also provide a method for improved BOLD fMRI calibration, as suggested in recent work by Driver et al. (2012).

Conclusions

We presented a novel technique, iTRUST, for combined susceptometry-and T_2 -based quantification of $CMRO₂$ at high temporal resolution. Simulations and in vivo evaluations demonstrate that iTRUST has comparable precision and accuracy relative to the traditional uncombined methods. In addition, iTRUST provides significantly improved temporal resolution for T_2 -based CMRO₂ quantification. In summary, iTRUST is a promising method for dynamic assessment of CMRO₂, and offers a unique approach for evaluating and comparing susceptometry- and T_2 -based CMRO₂ quantification techniques.

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Abbreviations

Fig. 1.

iTRUST pulse sequence and example images. (A) TRUST sequence diagram with (B) inset depicting the OxFlow module inserted within the T_1 recovery period of the TRUST sequence. (C) Sagittal scout image indicating the relative positions of the labeling slab (red) and imaging slice (blue). (D) Magnitude image with square region of interest indicating the position of the SSS. (E) Velocity map and (F) phase difference map of the SSS ROI from (D). (G) TRUST difference images for each eTE. Note that the spin histories of the OxFlow module and TRUST sequence should not interact as they are isolated by the global spin reset and the spatial separation of the imaging slice and labeling slab.

Fig. 2.

TRUST, OxFlow, and iTRUST parameter values acquired sequentially for four minutes each in three resting subjects. (A) TRUST vs. iTRUST Y_v - T_2 values. (B) OxFlow vs. iTRUST *Y*v-SBO values. (C) OxFlow vs. iTRUST tCBF values. Mean absolute bias is the absolute value of the bias between techniques, averaged across subjects. Error bars indicate \pm 1 SD across the *N* = 10 (A) or *N* = 80 (B and C) data points collected in each four-minute acquisition.

Fig. 3.

Scatter plot of iTRUST Y_v-T_2 values fitted using all four (0, 40, 80, and 160 ms) vs. only two (0 and 80 ms) eTEs from the same data. The 30 data points represent 10 repeat measures from each of three subjects. The linear least-squares regression line for all data points (solid line) is shown along side the line of identity (dotted line). 95% confidence intervals for the slope [0.93, 1.25] and intercept [− 16.4, 4.3] of the linear fit contain 1 and 0, respectively, indicating no statistically significant bias between the four and two eTE Y_v-T_2 values.

Fig. 4.

Simulation of expected Y_v-T_2 error vs. the number of eTEs used for T_2 fitting. (A) Rootmean-square error (RMSE) in Y_v-T_2 vs. TRUST difference signal SNR for four, three, or two eTEs. (B) RMSE normalized to acquisition time. Simulations were performed for *N* = 1000 virtual images for each SNR value, incremented by an SNR value of 1.

Fig. 5.

iTRUST parameter values in response to a 60-second breath-hold in a single subject. T_2 fitting with two eTEs and application of sliding window reconstruction yields Y_v-T_2 temporal resolution of six seconds. Y_v -SBO and tCBF temporal resolution is three seconds. Gray shading indicates the apnea period.

Fig. 6.

Subject-averaged time-course plots of physiologic parameters measured via pulse oximetry (Y_a, HR) and capnography (ECO_2, RR). Gray shading indicates the hypercapnia period. Error bars indicate standard errors (*N*=10). Comparing average baseline (0–5minutes) and steady-state hypercapnia (7.5–10minutes) values across subjects, significant increases were observed in EtCO₂ ($P < 0.0001$), Y_a ($P < 0.01$), and HR ($P < 0.05$). RR did not show a significant change $(P = 0.64)$.

Fig. 7.

iTRUST-derived parameter values inresponse to five minutes of 5% $CO₂$ gas mixture breathing. (A) Time-course plots of absolute parameter values from (A) a representative subject and (B) averaged across all 10 subjects. (C) Subject-averaged parameter values normalized to average baseline values, with error bars indicating standard errors (*N*=10) at each time point. The arteriovenous oxygen saturation difference $(AVO₂D)$ is equal to Y_a – *Y*_v. In all time-course plots, tCBF, *Y*_a, *Y*_v-SBO, AVO₂D-SBO, and CMRO₂-SBO temporal resolution is three seconds, and Y_v-T_2 , AVO_2D-i_2 , and $CMRO_2-T_2$ temporal resolution is six seconds. Gray shading indicates the hypercapnia period. (D) Scatter plot of time-matched Y_v -SBO and Y_v -*T*₂ values across all subjects and time points ($N = 1490$), with different symbols/colors denoting individual subjects. Linear least-squares regression lines are plotted for each subject (solid lines), as well as the line of identity (dotted line). Mean slope and *R* 2 values of the regression lines across subjects are $\beta = 1.47 \pm 0.20$ and $R^2 = 0.90 \pm 0.02$.

Table 1

Summary of hypercapnia paradigm parameter values derived from pulse oximetry, capnography, and iTRUST MRI in 10 subjects.

Parentheses indicate the standard deviations of parameter values across subjects. *P*-values are based on one-sample Student's *t*-tests of the percent changes from baseline to hyper-capnia. AVO₂D is the arteriovenous oxygen difference, equal to $Y_a - Y_V$.