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Author manuscript *Ophthalmology*. Author manuscript; available in PMC 2016 October 01.

Published in final edited form as:

Ophthalmology. 2015 October; 122(10): 2142–2145. doi:10.1016/j.ophtha.2015.04.026.

## Varicella Zoster Virus in Ischemic Optic Neuropathy

Liliya Golas, MD<sup>1</sup>, Jeffrey L. Bennett, MD, PhD<sup>1,2</sup>, Teresa M. White, BS<sup>2</sup>, Barry Skarf, MD, PhD<sup>3</sup>, Robert Lesser, MD<sup>4</sup>, Maria A. Nagel, MD<sup>2</sup>, and Don Gilden, MD<sup>2,5</sup>

<sup>1</sup>Department of Ophthalmology, University of Colorado School of Medicine, Aurora, Colorado.

<sup>2</sup>Department of Neurology, University of Colorado School of Medicine, Aurora, Colorado.

<sup>3</sup>Department of Neurology, Henry Ford Hospital, Detroit, Michigan.

<sup>4</sup>Department of Ophthalmology and Neurology, Yale University School of Medicine, New Haven, Connecticut.

<sup>5</sup>Department of Immunology and Microbiology, University of Colorado School of Medicine, Aurora, Colorado.

Anterior ischemic optic neuropathy (AION), is classified as arteritic (giant cell arteritis; GCA) or non-arteritic (NAION). Varicella zoster virus (VZV) vasculopathy causes symptoms and signs of AION.<sup>1-4</sup> Because VZV is present in temporal arteries (TAs) of most patients with GCA<sup>1,5</sup> and ischemic optic nerve and retinal pathologies,<sup>2,3</sup> we examined GCA-negative TAs from seven AION patients for VZV.

#### Case 1

79 year-old man developed acute vision loss right eye (OD) and worsened vision left eye (OS), preceded by jaw claudication. Visual acuity (VA) was no light perception (NLP) OD and count fingers (CF) OS (baseline 20/50 OD, 20/150 OS), with a right afferent pupillary defect (APD), bilateral disc edema, right disc hemorrhage and left peripapillary cotton wool spot (CWS). Erythrocyte sedimentation rate (ESR) was normal and C-reactive protein (CRP) was 78 (normal <4.9 mg/dL). Oral prednisone was started and TA biopsy (day-8) was GCA-negative. Valacyclovir was started at one month with prednisone taper. VA remained unchanged. Optic nerves developed pallor. TA biopsy was VZV-negative.

No conflicting relationship exists for any author.

**Corresponding author and address for reprints:** Jeffrey L. Bennett, MD, PhD, Department of Neurology, University of Colorado School of Medicine, 12700 E. 19<sup>th</sup> Avenue, Box B182, Aurora, CO 80045. Jeffrey.bennett@ucdenver.edu Telephone: 303-724-4312; fax: 303-724-4329.

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Conflict of Interest:

#### Case 2

78 year-old woman suffered sudden painless vision loss OS preceded by jaw claudication. VA was 20/25-2 OD, 20/30 OS with left inferior visual field (VF) defect, left APD and left disc edema with CWS. ESR was normal; CRP was 25.9 (normal <10 mg/dL). Solumedrol was infused for 3 days with prednisone taper. Bilateral TA biopsies were GCA-negative; immunohistochemistry revealed VZV antigen (Fig 1B). CRP elevated during prednisone taper, and valacyclovir was added; CRP normalized 1 week later.

#### Case 3

53 year-old woman developed sudden painless vision loss OS. Five months later, VA was 20/25 OD, CF OS, with a left APD and diffuse left disc pallor with gliosis. ESR was 31; CRP was normal. TA biopsy was GCA-negative, but VZV-positive (Fig 1C).

#### Case 4

68 year-old woman noted painless vision loss OD with associated polymyalgia rheumatica, frontal headaches, jaw tightness, diplopia and VF change OD. ESR was 79; CRP was 14 (normal <1 mg/dL). TA biopsy was GCA-negative after 2 months of prednisone. Six months later during prednisone taper, she experienced further vision loss OD. Examination showed VA 20/150 OD, 20/40 OS, with right a APD and right disc edema with hemorrhages. ESR and CRP were normal. Re-evaluation of earlier TA biopsy revealed VZV antigen (Fig 1G) with adjacent GCA histopathology.

### Case 5

81 year-old woman with history of sudden painless vision loss OS three years earlier, experienced new acute vision loss OD. Examination showed VA 20/100 OD, CF OS (baseline 20/40 OD and CF OS) with a left APD, right pallid disc edema with hemorrhage, and left disc pallor. ESR and CRP were normal. TA biopsy after one week of prednisone was GCA-negative but VZV-positive (Fig 1H). Oral acyclovir was added to prednisone taper, but vision did not improve.

#### Case 6

63 year-old man developed painless superior VF loss OD. Examination showed: VA 20/25 OD, 20/25-2 OS, bilateral superior arcuate scotomas, and bilateral inferior disc edema with hemorrhages. ESR and CRP were normal. Two months later, examination revealed progressive VF loss and additional right superior disc edema with hemorrhages. Six months later, examination showed progressive bilateral superior VF loss, bilateral disc pallor and left superior peripapillary subretinal hemorrhage. In four months, he lost inferior VF OS and developed left APD. TA biopsy was GCA-negative, but VZV-positive (Fig 1I).

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#### Case 7

49 year-old man developed sudden loss of vision OD. Examination revealed VA: hand motion OD, 20/20 OS, with a right APD and right disc edema. ESR and CRP were normal. After intravenous corticosteroids, VA was 20/200 OD. Eleven months later, he developed sudden vision loss OS with VA 20/200 bilaterally and left disc edema with hemorrhage. Four months later, he developed bitemporal headache and vesicular lip lesions, and was treated with acyclovir. VA was now 20/200 OD, 20/60 OS. Three months later, TA biopsy was GCA-negative and VZV-negative.

Table summarizing cases available: www.aaojournal.org.

GCA-negative TA biopsies contained VZV antigen in 5/7 (71%) patients with AION. Importantly, most patients with VZV-positive biopsies had atypical AION features (vascular gliosis at optic disc, subretinal hemorrhage and progressive vision loss), suggesting that VZV vasculopathy in AION produces a wider range of ischemia than in classical GCA. Due to the multifocal nature of VZV vasculopathy and the rich innervation of the vascular supply to optic nerve and retina, VZV vasculopathy may produces a spectrum of ischemic injuries: anterior and posterior ION, retinal necrosis and central retinal artery occlusion.<sup>2-4</sup> Thus, searching for VZV in atypical NAION cases with elevated cup/disc ratios, pain, retinal pathology or slow progression may identify individuals who could benefit from antiviral treatment. Additional prospective randomized controlled trials are needed to evaluate effects of antiviral therapy in these patients.

Two VZV-positive, biopsy-negative AION cases had clinical features consistent with GCA. In one patient, GCA pathology was subsequently found adjacent to VZV antigen. We recently reported a strong correlation between GCA pathology and VZV.<sup>5</sup> Examining TA biopsies for VZV together with histopathology may increase diagnosis of GCA and prevent irreversible vision loss. Antiviral treatment may prevent future ischemic events in the fellow eye and brain by shortening exposure to long-term corticosteroids that potentiate VZV infection. In our study, it seems unlikely that corticosteroids confounded the results of VZV antigen testing. Of the 5 patients with VZV-positive TA biopsies, 2 received corticosteroids for 1 week, one for 2 months and the other 2 patients had no recent steroid exposure at the time of biopsy.

Overall, virological examination of TA biopsies in AION patients with or without clinical and laboratory findings for GCA may reveal VZV infection and should be included in the standard evaluation of AION patients. Extensive serial sections of TA biopsies should be examined for both GCA pathology and for VZV antigen, since patients whose TAs contain VZV may benefit from antiviral treatment.

#### **Supplementary Material**

Refer to Web version on PubMed Central for supplementary material.

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#### Acknowledgments

Financial Support:

J.L.B. supported by the Guthy-Jackson Charitable Foundation and research grant EYO22936 from the National Institutes of Health; M.A.N. and D.G. supported by research grant AG032958 from the National Institutes of Health.

The funding organization had no role in the design or conduct of this research.

#### Abbreviations and Acronyms

AION	anterior ischemic optic neuropathy
AMD	age-related macular degeneration
APD	afferent pupillary defect
CF	count finger
CRP	C-reactive protein
ESR	erythrocyte sedimentation rate
GCA	giant cell arteritis
NAION	non-arteritic anterior ischemic optic neuropathy
NLP	no light perception
OD	right eye
OS	left eye
ТА	temporal artery
VA	visual acuity
VF	visual field
VZV	varicella zoster virus

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#### Figure 1.

Varicella zoster virus (VZV) in temporal arteries from patients with anterior ischemic optic neuropathy (AION). One-hundred 5-µm sections from each TA were cut and baked for 1 hour at 60°C. Every other slide (50 sections/TA) was deparaffinized and immunostained with either mouse anti-VZV glycoprotein E (a late viral protein indicative of productive infection) IgG1 antibody or control mouse IgG1 antibody (Dako) and examined by light microscopy; if VZV antigen was detected, adjacent sections were stained with hematoxylin and eosin and examined histologically, as described.<sup>5</sup> VZV was detected in 5 temporal arteries from patients with AION. VZV antigen was seen in the adventitia of a positive control VZV-infected cadaveric cerebral artery (**A**, pink) 14 days post-infection *in vitro*. Note VZV antigen in cells adjacent to the internal elastic lamina of patient 2 (**B**), in the adventitia of patient 3 (**C**), in the adventitia and media of patient 4 (**G**), in the adventitia, media and intima of patient 5 (**H**), and in the media of patient 6 (**I**). No staining was seen in sections adjacent to those containing VZV antigen when mouse IgG1 antibody was substituted for mouse anti-VZV gE IgG1 antibody (**D-F**, **J-L**). Magnification 600X.

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