

Original Article

Renal tumor with alpha b crystallin expression

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Abstract: In human, proximal convoluted tubules and thin limbs of Henle show expression of α B crystallin. Renal cell carcinoma also showed expression of α B crystallin in previous reports. We aimed to study the association between α B crystallin expression and renal cell carcinoma and urothelial carcinoma. Furthermore, we also investigated α B crystallin expression depending on the subtype of renal cell carcinoma and examined the relationship between α B crystallin expression and survival in patients with renal cell carcinoma. In our study, α B crystallin expression was different according to the type of tumor. A greater proportion of the clear cell type (52/77, 67.5%) and papillary type (4/4, 100%) showed reactivity compared to the chromophobe type (0/10, 0%). In the present study, a significantly greater number of renal cell carcinomas showed strong expression of α B crystallin (56/91, 61.5%) compared to urothelial carcinoma ($P=7.967e-07$). Therefore, α B crystallin might be a significant marker of renal cell carcinoma and might help to determine the type of renal tumor in cases of poorly differentiated kidney lesions and metastatic lesions. α B crystallin expression was not related to overall survival in univariate and multivariate models. In our study, alpha B crystallin could not be considered a prognostic marker of renal cell carcinoma.

Keywords: Alpha B crystallin, renal cell carcinoma, urothelial carcinoma, subtype, overall survival

Introduction

Renal cell carcinoma is estimated to account for 2-3% of adult malignancies. Approximately 80% of renal cell carcinomas are clear cell type, 15% are papillary type, and 5% are others [1]. α B crystallin is a small heat-shock protein that functions as a cytoprotective molecular chaperone. α B crystallin expression has been reported in renal epithelial tissues [2, 3]. In a previous report on immunohistochemical analysis of α B crystallin in rat nephrogenesis, α B crystallin expression was initially found to be focal in the capsular epithelium during the late embryonic stage. At birth, α B crystallin expression was observed in the thin limbs of the loop of Henle and the inner medullary collecting duct also became positive for expression of α B crystallin. At adult stages, the proximal convoluted tubules in the S3 segment, thin limbs of loop of Henle, and medullary collecting ducts showed strong positivity for α B crystallin. In the developing rat kidney, α B crystallin showed restricted distribution in tubules according to stage. The expression of α B crystallin was detected from the outer capsule to inner medulla and

increased in response to osmotic challenge. These findings indicate that α B crystallin has a function in the development and maintenance of tubules in the rat [4, 5]. In human, proximal convoluted tubules and thin limbs of Henle show expression of α B crystallin. [6] α B crystallin has been reported in several human epithelial tumors such as head and neck squamous cell carcinoma, breast cancer, and thyroid cancer [2, 7-9]. Renal cell carcinoma also showed expression of α B crystallin in previous reports [6, 10, 11]. Therefore we aimed to elucidate the association between α B crystallin expression and renal cell carcinoma and urothelial carcinoma. Furthermore, we also investigated α B crystallin expression depending on the subtype of renal cell carcinoma and examined the relationship between α B crystallin expression and survival in patients with renal cell carcinoma.

Materials and methods

Patients and tissue sampling

A total of 108 formalin-fixed paraffin-embedded kidney tissues were obtained from 91 patients

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Table 1. Clinical characteristics of 91 renal cell carcinoma patients and 17 urothelial carcinoma patients

Clinicopathologic feature	Renal cell carcinoma		Urothelial carcinoma			
	N	%	N	%		
Gender	Male	61	67	11	64.7	
	Female	30	33	6	35.3	
Age	≤60	52	57.1	4	23.5	
	>60	39	42.9	13	76.5	
T stage	T1a	31	34.1	Ta	2	11.8
	T1b	35	38.4	Tis	2	11.8
	T2a	6	6.6	T1	1	5.9
	T2b	3	3.3	2	3	17.6
	T3a	14	15.4	3	7	41.2
	T3b	0	0	4	2	11.8
	T3c	0	0			
N stage	N0	89	97.8	N0	16	94.1
	N1	2	2.2	N1	0	
				N2	1	5.9
				N3	0	
M	M0	91	100	M0	16	94.1
	M1	0	0	M1	1	5.9
Tumor type	Clear cell	77	84.6	Papillary	17	100
	Papillary	4	4.4			
	Chromophobe	10	11			

with stage I to IV renal cell carcinoma and 17 patients with stage I to III urothelial carcinoma of the renal pelvis. All patients underwent radical nephrectomy at Samsung Changwon hospital between 2003 and 2012. All clinical information was acquired through medical records. None of the patients received adjuvant treatment. For patients with renal cell carcinoma, 61 were male and 30 were female, and 52 (57.1%) were aged 60 or younger. Patients with suspicion of distant metastasis at the time of nephrectomy were excluded from this study. The proportions of T/N stages were as follows: T1a: 31 patients (34.1%), T1b: 35 (38.4%), T2a: 6 (6.6%), T2b: 3 (3.3%), T3a: 14 (15.4%), T4: 2 (2.2%)/N0: 89 (97.8%), N1: 2 (2.2%). The stages of the tumor were determined according to the TNM system of the American Joint Committee on Cancer (AJCC), 7th edition. Renal cell carcinoma subtyping was performed based on previous reports. The reviews were conducted by two experienced pathologists (E. H. Lee and H. W. Lee). The tumor subtypes were categorized as follows: clear cell, 77 (84.6%); papillary, 4 (4.4%); chromophobe, 10 (11%). The

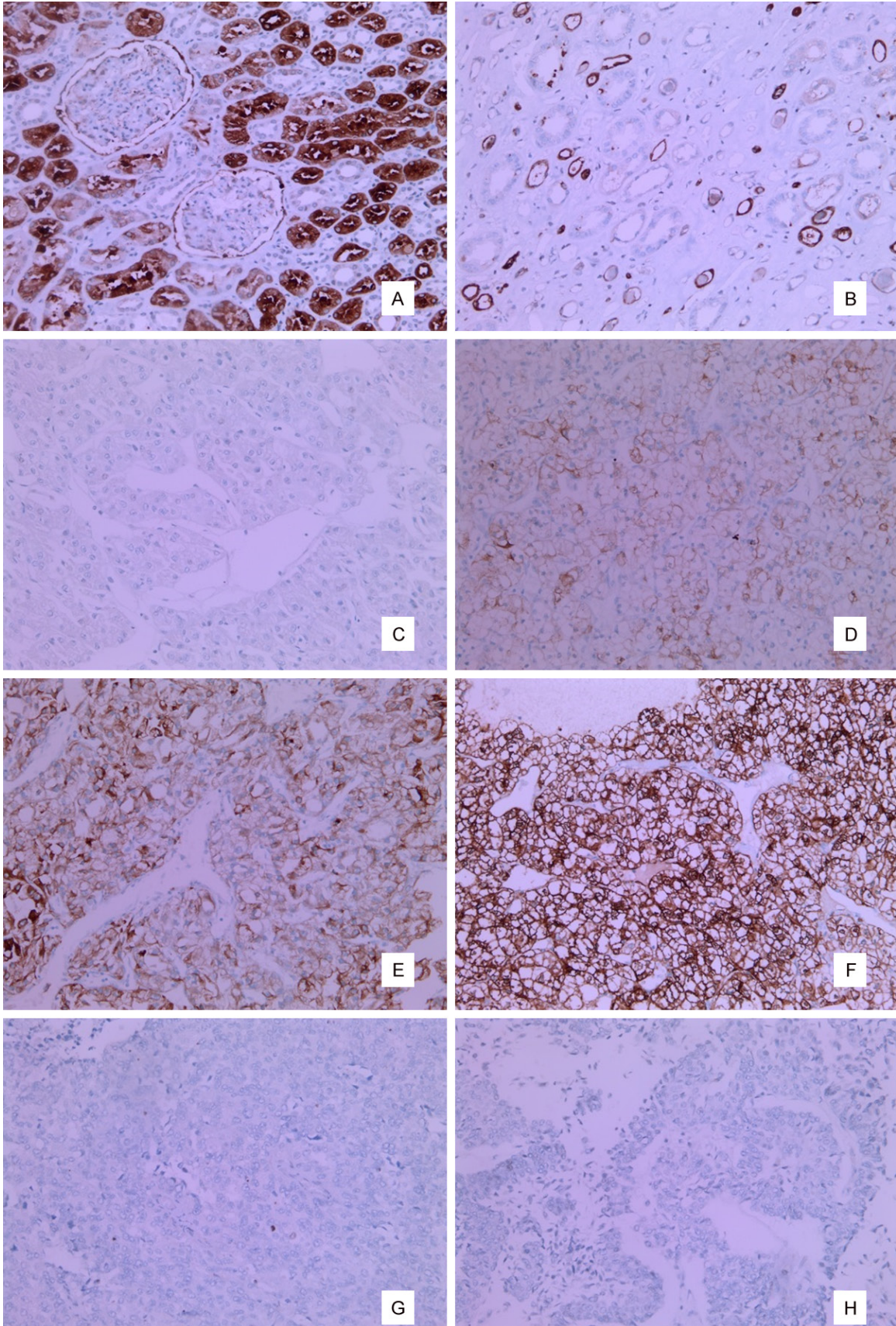
mean follow-up duration was 1,830.2 days. The median overall survival was 1,591 days and mean overall survival was 1,830.198 days. The clinical characteristics of 91 patients with renal cell carcinoma and 17 patients with urothelial carcinoma of pelvis are summarized in **Table 1**.

Immunohistochemical staining

Formalin-fixed paraffin-embedded (FFPE) tissue samples from patient tumors were collected retrospectively. To construct the tissue microarray for 91 renal cell carcinomas, evaluation of primary H&E-stained slides was conducted by two experienced pathologists (E. H. Lee and H. W. Lee). The most representative tumor areas were sampled. Each tumor core was 2 tissue cores, 6 mm in diameter. A total of 20 TMA blocks were created. For 17 urothelial carcinoma samples, a representative area was selected by H&E slide review by two experienced pathologists (E. H. Lee and H. W. Lee) without TMA construction. Representative

sections from the standard tissue blocks and TMA blocks were cut with a microtome at 4- μ m thickness and dried overnight at 37°C on a salinized slide. Immunohistochemical staining was performed using a Benchmark XT slide stainer (Ventana, Inc.) according to the manufacturer's instructions. The antibody used for immunohistochemical staining of α B crystallin was mouse IgG1 monoclonal antibody, clone 1B6.1-3G4 (Enzo Life Sciences, Inc.; 1:200 dilution, 1 hour incubation at room temperature). To evaluate α B crystallin protein expression, the intensity was scored using a scoring system from 0-3 (0: negative; 1: weak; 2: moderate; 3: strong) and multiplied by the percentage of positive cells. The total score range was therefore 0-300. For α B crystallin expression, cytoplasmic and membranous staining was considered positive and nuclear staining was excluded from the scoring. Human pilocytic astrocytoma tissue was used as a positive control for α B crystallin. The immunohistochemical staining was evaluated by experienced pathologists (E. H. Lee and H. W. Lee) who were blinded to the patient information.

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Figure 1. A. In immunohistochemical staining, α B crystallin expression was positive in the proximal convoluted tubules of the normal kidney. B. α B crystallin expression was positive thin limbs of Henle of the normal kidney. C. Negative expression of α B crystallin in renal cell carcinoma. D. Weak (1+) expression of α B crystallin in renal cell carcinoma. E. Moderate (2+) expression of α B crystallin in renal cell carcinoma. F. Strong (3+) expression of α B crystallin in renal cell carcinoma. G, H. Negative expression of α B crystallin in urothelial carcinoma.

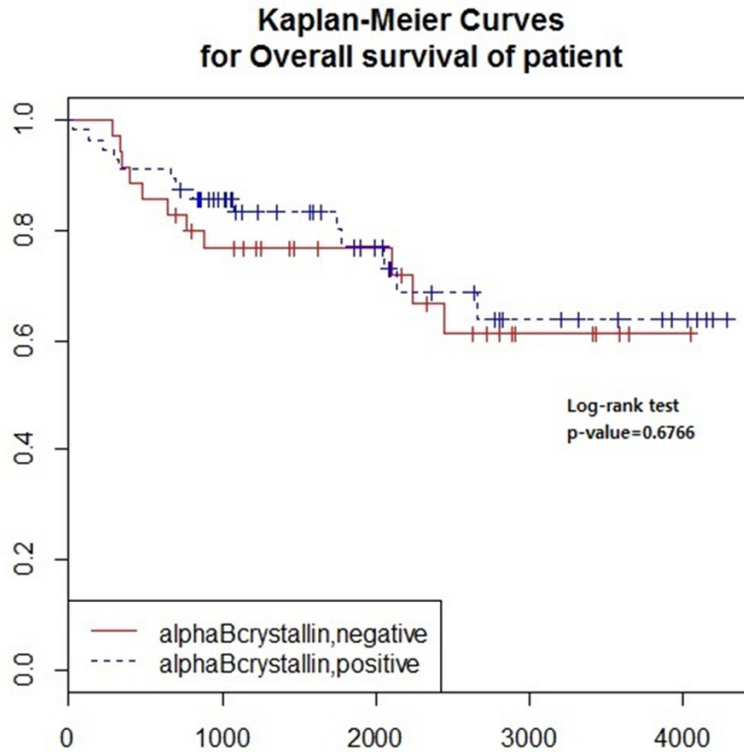


Figure 2. Kaplan-Meier curves and Log rank tests for overall survival according to α B crystallin expression were performed. α B crystallin showed no relationship to overall survival in renal cell carcinoma ($P=0.6766$).

and urothelial carcinoma, we used Fisher's exact test. And to analyze α B crystallin expression according to type of renal cell carcinoma, we used Fisher's exact test. To determine relationships between survival and α B crystallin expression with respect to clinicopathologic characteristics, we performed analysis by univariate and multivariate Cox regression models. Kaplan-Meier analysis was performed to estimate the overall survival of the α B crystallin positive group and negative group. Log rank test was used to compare the survival distribution of the two groups (positive vs. negative α B crystallin expression). A P value < 0.05 was considered statistically significant (**Figure 2**).

Results

Immunohistochemical staining of α B crystallin

Table 2. Results of immunohistochemical staining in renal cell carcinoma

Score (0-3)	1 (weak)	2 (moderate)	3 (strong)
0 (no)	0 (0%)	9 (9.9%)	73 (80.2%)
Proportion (%)	0-25	26-50	51-75
	20 (22%)	11 (12.1%)	19 (20.8%)
		75-100	41 (45.1%)

Statistical analysis

Statistical analysis was conducted using R software (version 3.1.0) with packages named "prettyR" and "survival". To identify associations between α B crystallin and clinicopathologic characteristics, we used Fisher's exact test and t-test. To examine difference of α B crystallin expression in renal cell carcinoma

In immunohistochemical staining, α B crystallin expression was positive in the proximal convoluted tubules (**Figure 1A**) and thin limbs of Henle of the normal kidney (**Figure 1B**). The intensity of staining in renal carcinomas was found to be negative (**Figure 1C**), weak (**Figure 1D**), moderate (**Figure 1E**), and strong (**Figure 1F**). All cases of urothelial carcinoma of pelvis were clearly negative (**Figure 1G** and **1H**). The intensity score multiplied by the percentage of positive cells in the tumor yielded a mean total score of 154.336 and median total score of 180. The cut-off value was determined by the median score (180) and the study population was divided into a positive group and negative group on the basis of median score. The percentage of stained cells in the positive group was 60-100%. The results of staining are summarized in **Table 2**.

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Table 3. α B crystallin expression in renal cell carcinoma and urothelial carcinoma

α B crystallin	Renal cell carcinoma (mean score)			Urothelial carcinoma (mean score)	p value
Positive	56 (259.7321429)			0 (NA)	7.967e-07
Negative	35 (61.4)			17 (0)	
α B crystallin	Clear cell (mean score)	Papillary (mean score)	Chromophobe (mean score)	Urothelial carcinoma (mean score)	1.639e-11
Positive	52 (257.5)	4 (288.75)	0 (NA)	0 (NA)	
Negative	25 (79)	0 (NA)	10 (17.4)	17 (0)	

Table 4. α B crystallin expression in renal cell carcinoma according to clinicopathologic characteristics

		α B crystallin		95% confidence interval		P value
		Positive	Negative	Lower	Upper	
Age	Mean	57.80357	56.65714	-6.762666	4.469809	0.686
Sex	Male	32 (52%)	29 (48%)	0.0816431	0.8321968	0.0125
	Female	24 (80%)	6 (20%)			
T stage	1a	24 (77%)	7 (23%)	0.04431		
	1b	21 (60%)	14 (40%)			
	2a	4 (67%)	2 (33%)			
	2b	2 (67%)	1 (33%)			
	3a	4 (29%)	10 (71%)			
	3b	0 (NA)	0 (NA)			
	3c	0 (NA)	0 (NA)			
	4	1 (50%)	1 (50%)			
N stage	0	55 (62%)	34 (38%)	0.02002464	129.11977469	1
	1	1 (50%)	1 (50%)			
Type	Clear cell	52 (68%)	25 (32%)	1.426e-05		
	Papillary	4 (100%)	0 (0%)			
	Chromophobe	0 (0%)	10 (100%)			
Overall survival (days)	Mean	1837.661	1818.257	-514.4767	475.6696	0.9381

α B crystallin expression in renal cell carcinoma and urothelial carcinoma

A greater proportion of renal cell carcinoma samples showed strong reactivity to α B crystallin (56/91, 61.5%) compared to urothelial carcinoma and this difference was statistically significant ($P=7.967e-07$). α B crystallin expression was different according to the type of tumor. A greater proportion of the clear cell type (52/77, 67.5%) and papillary type (4/4, 100%) showed reactivity compared to the chromophobe type (0/10, 0%) and urothelial carcinoma (0/17, 0%). This difference was statistically significant ($P=1.639e-11$). These results are summarized in **Table 3**.

Univariate analysis of α B crystallin expression of renal cell carcinoma and clinicopathologic characteristics

Results of univariate analysis of α B crystallin expression and clinicopathologic characteristics

are shown in **Table 4**. Sex, cancer type, and T stage were associated with α B crystallin expression. A total of 56 cases showed strong α B crystallin expression. α B crystallin overexpression was observed significantly more frequently in the clear cell type and papillary type compared to the chromophobe type (clear cell: 52/77 [68%]; papillary: 4/4 [100%]; chromophobe: 0/10 [25%]; $P=1.426e-05$). The proportion of alpha B crystallin expression was different between T stages ($P=0.04431$) and was higher in females than in males (male: 32/61 [52%], female: 24/30 [80%]).

Analysis of overall survival according to α B crystallin expression and clinicopathologic characteristics of renal cell carcinoma by Cox-regression model

Univariate and multivariate Cox-regression models of overall survival with respect to α B crystallin expression and clinicopathologic

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Table 5. Overall survival according to clinicopathologic characteristics

Independent variable	Reference	Univariate Cox regression		Multivariate Cox regression	
		Hazard ratio (95% CI)	<i>p</i> value	Hazard ratio (95% CI)	<i>p</i> value
αB crystallin = positive	Negative	0.8451 (0.3828-1.865)	0.677	1.48075 (0.5466-4.011)	0.4401
Age		1.04533 (1.01-1.082)	0.0127	1.04269 (1.0017-1.085)	0.0409
Sex = female	Male	0.6914 (0.2885-1.657)	0.408	0.73627 (0.2826-1.918)	0.5309
Papillary	Clear cell	0.8624 (0.1156-6.432)	0.885	1.20829 (0.1515-9.636)	0.8582
Chromophobe		1.1569 (0.3448-3.882)	0.813	1.52048 (0.3790-6.101)	0.5544
T stage	T1	1.4565 (1.171-1.812)	0.000734	1.35175 (1.0306-1.773)	0.0294
N stage	NO	23.6984 (4.716-119.1)	0.000122	2.68901 (0.2997-24.125)	0.3769

characteristics are summarized in **Table 5**. αB crystallin expression was not related to overall survival in the univariate and multivariate model (univariate $P=0.677$; multivariate, $P=0.4401$). N stage showed an association with overall survival only in the univariate model ($P=0.000122$). However, age and T stage were associated with overall survival in both the univariate and multivariate model (T stage, $P=0.000734$ and 0.0294 ; age, $P=0.0127$ and 0.0409).

Discussion

Heat shock proteins act as molecular chaperones to protect the kidneys against stress, such as heat, ischemia, hypertension, inflammation, and drugs [13, 14]. αB crystallin is one of the small heat-shock proteins and has an antiapoptotic function. In vitro, αB crystallin induced resistance to oxidative stress-induced apoptosis in rabbit lens epithelial cells [12]. αB crystallin expression was reported during renal tubulogenesis and probably has a cytoprotective function in kidney development [5, 6]. αB crystallin was also found in rat renal tubules, including the pars recta of proximal tubules, thin limbs of loops of Henle, and inner medullary collecting ducts [5, 6]. In humans, αB crystallin was detected in the proximal tubules and thin limbs of Henle, but not in the distal tubules and glomerular components [6]. Renal cell carcinoma originates in the renal tubules and accounts for 80-85% of malignant renal tumors. A pathological classification of renal cell carcinomas was proposed in 1986 [13]. Approximately 80% of renal cell carcinomas are clear cell type, 15% are papillary type, and 5% are others. Clear cell carcinoma is thought to have originated in the proximal convoluted tubule. Papillary renal cell carcinomas are derived from proximal convoluted tubule and

distal convoluted tubule. And chromophobe renal cell carcinomas are derived from intercalated cell of cortical collecting duct [14]. In our study, αB crystallin expression was different according to the type of tumor. A greater proportion of the clear cell type (52/77, 67.5%) and papillary type (4/4, 100%) showed reactivity compared to the chromophobe type (0/10, 0%). These findings probably due to cell origin of renal cell carcinoma. In human kidney, proximal convoluted tubule shows reactivity of αB crystallin so clear cell type and papillary type renal cell carcinoma have a tendency of αB crystallin expression. And in human kidney, collecting duct shows negative reactivity of αB crystallin. Chromophobe type is derived from intercalated cell of cortical collecting duct. So all chromophobe type shows negative reactivity for αB crystallin in our study. In the present study, a significantly greater number of renal cell carcinomas showed strong expression of αB crystallin (56/91, 61.5%) compared to urothelial carcinoma ($P=7.967e-07$). Therefore, αB crystallin might be a significant marker of renal cell carcinoma subtype and our findings might help to determine the type of renal tumor in cases of poorly differentiated kidney lesions and metastatic lesions. αB crystallin expression was not related to overall survival in univariate and multivariate models. In our study, alpha B crystallin was not related to overall survival and could not be considered a prognostic marker of renal cell carcinoma. Our study has limitations of the small number of cases and the use of only immunohistochemical staining as a protein detection method.

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Disclosure of conflict of interest

None.

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