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Ga_{i/o}-coupled receptor-mediated sensitization of adenylyl cyclase: 40 years later

Tarsis F. Brust, Jason M. Conley, and Val J. Watts

Department of Medicinal Chemistry and Molecular Pharmacology, College of Pharmacy, Purdue University. 575 Stadium Mall Drive, Lafayette – IN – 47907

Abstract

Heterologous sensitization of adenylyl cyclase (also referred to as superactivation, sensitization, or supersensitization of adenylyl cyclase) is a cellular adaptive response first described 40 years ago in the laboratory of Dr. Marshall Nirenberg. This apparently paradoxical cellular response occurs following persistent activation of $G\alpha_{i/o}$ -coupled receptors and causes marked enhancement in the activity of adenylyl cyclases, thereby increasing cAMP production. Since our last review in 2005, significant progress in the field has led to a better understanding of the relevance of, and the cellular biochemical processes that occur, during the development and expression of heterologous sensitization. In this review we will discuss the recent advancements in the field and the mechanistic hypotheses on heterologous sensitization.

Keywords

Heterologous sensitization; adenylyl cyclase; G protein-coupled receptor; superactivation; G protein subunit; protein kinase

1. Background

1.1 Introduction to heterologous sensitization

The history of and basic concepts involving heterologous sensitization of adenylyl cyclase were extensively discussed in our previous review (Watts and Neve, 2005). Thus, the present review will provide a limited introduction describing the history of this topic, and then incorporate recent findings into our current understanding of this paradoxical phenomenon, first described forty years ago in the laboratory of Dr. Marshall Nirenberg (Sharma et al., 1975).

Disclosures:

 $Corresponding \ author: \ Val \ J. \ Watts., \ Address: 575 \ Stadium \ Mall \ drive, \ West \ Lafayette, \ IN-47907-USA. \ Phone \ number: 1-765-496-3872, \ wattsv@purdue.edu.$

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G protein-coupled receptors (GPCRs) are seven transmembrane domain proteins involved in the transmission of extracellular signals into intracellular signaling cascades. Because of their diversity and prominent physiological relevance, GPCRs represent the most targeted class of proteins by FDA-approved drugs (Overington et al., 2006; Rask-Andersen et al., 2011). Various hormones, neurotransmitters, and drugs interact with GPCRs to elicit their cellular and physiological responses (Hanson and Stevens, 2009; Lefkowitz, 2004). Activation of GPCRs leads to conformational changes in the receptor that ultimately cause activation of heterotrimeric G proteins through the exchange of GDP for GTP in the Ga subunit (Nygaard et al., 2013; Rasmussen et al., 2011). The Ga subunit and $G\beta\gamma$ subunits are thought to then dissociate/undergo rearrangement, leading to diverse signaling events through a variety of effectors (Rasmussen et al., 2011). Following the activation of associated G proteins, the receptor can be phosphorylated by G protein-coupled receptor kinases (GRK), allowing for the recruitment of β -arrestin to the GPCR (Lefkowitz and Shenoy, 2005; Nygaard et al., 2013). Recruitment of β -arrestin can lead to receptor desensitization and receptor downregulation from the membrane, as well as diverse βarrestin signaling events (Lefkowitz and Shenoy, 2005).

There are four major types of Ga subunits of G proteins (Ga_s, Ga_{i/o}, Ga_{q/11}, and Ga_{12/13}) that lead to distinct cellular signaling events (Moreira, 2014). The Ga_s family of proteins activate adenylyl cyclases to increase intracellular cAMP concentrations (Rasmussen et al., 2011; Sunahara and Taussig, 2002). Intracellular cAMP levels are reduced following receptor activation of Ga_{i/o} proteins, which acutely inhibit adenylyl cyclases (figure 1A) (Sunahara and Taussig, 2002; Watts et al., 1998). In contrast, prolonged stimulation of Ga_{i/o}-coupled receptors leads to a cellular adaptive response known as heterologous sensitization of adenylyl cyclase (also referred to as superactivation, sensitization, or supersensitization of adenylyl cyclase) (figure 1B). This Ga_{i/o}-coupled receptor-mediated enhancement of cAMP signaling is most readily observed following the addition of receptor antagonist and subsequent activation of adenylyl cyclase (figure 2). Heterologous sensitization was first reported for the δ -opioid receptor in the laboratory of Dr. Marshall Nirenberg (Sharma et al., 1975). Since then, it has been demonstrated for a number of GPCRs and is believed to be a signaling event shared by most Ga_{i/o}-coupled receptors (Watts and Neve, 2005).

1.2 Adenylyl cyclases

Adenylyl cyclases are responsible for catalyzing the synthesis of cAMP from ATP and serve as the downstream effector for mediating the sensitized cAMP response. They are also acutely modulated by GPCRs that signal through $G\alpha_s$, $G\alpha_q/_{11}$, and $G\alpha_{i/o}$ proteins. The membrane-bound adenylyl cyclases share an overall similar structure, which is composed of three intracellular domains (N terminus, C1 domain, and C2 domain) and two transmembrane clusters (M1 and M2), each containing six transmembrane helixes (Cooper and Crossthwaite, 2006; Sadana and Dessauer, 2009; Wang et al., 2009). There are nine different isoforms of membrane-bound adenylyl cyclases (Cooper and Crossthwaite, 2006; Sadana and Dessauer, 2009), and the expression patterns and regulatory properties of the adenylyl cyclases are unique for each isoform. All membranous adenylyl cyclase isoforms are stimulated by $G\alpha_s$; however, the other regulatory properties allow them to be divided

into four groups (Patel et al., 2001; Sunahara and Taussig, 2002). Group 1 adenylyl cyclases include AC1, AC3, and AC8 and are stimulated by calcium/calmodulin. AC2, AC4, and AC7 represent group 2 adenylyl cyclases, which are conditionally activated by $G\beta\gamma$ subunits. Group 3 adenylyl cyclases comprise AC5 and AC6, which are inhibited by calcium. AC9 is the lone adenylyl cyclase isoform in group 4 and is relatively insensitive to the small molecule adenylyl cyclase activator, forskolin (Hacker et al., 1998). These unique regulatory properties of the adenylyl cyclases provide isoform-specific mechanisms for the regulation of adenylyl cyclases, and also provide for group and isoform-specific mechanisms for the expression of heterologous sensitization (Watts and Neve, 2005). For example, prolonged activation of $Ga_{i/o}$ -coupled GPCRs elevates the calcium-stimulated activity of AC1 and AC8, but not AC3 (Avidor-Reiss et al., 1997; Cumbay and Watts, 2001; Nevo et al., 1998; Watts and Neve, 1996). Group 2 adenylyl cyclases are not sensitized to stimulation by $G\alpha_{e_s}$. but persistent dopamine D₂ receptor activation sensitizes AC2 to activation by PKC (Avidor-Reiss et al., 1997; Cumbay and Watts, 2001; Nevo et al., 1998; Rhee et al., 2000; Thomas and Hoffman, 1996; Watts and Neve, 1996). Other proteins that interact with and regulate activity of specific isoforms of adenylyl cyclase include calmodulin, A-kinaseanchoring-proteins (AKAPs), Snapin, Ric8a, protein phosphatase 2A (PP2A), and the protein associated with Myc (PAM) (Masada et al., 2012; Sadana and Dessauer, 2009; Wang et al., 2009). The regulatory properties and binding partners specific to each adenylyl cyclase isoform add to the complexity of heterologous sensitization and suggest that even though it is likely that some mechanistic features of sensitization are shared by all adenylyl cyclases, there are likely isoform-specific mechanisms as well.

2. Mechanistic insights on heterologous sensitization

2.1 Gai/o subunits

Heterologous sensitization is a phenomenon shared by numerous $Ga_{i/o}$ -coupled receptors and, because of this, is linked to activation of $Ga_{i/o}$ proteins. Mechanistically, inactivation of Gai/o with pertussis toxin treatment blunts receptor-mediated heterologous sensitization (Watts, 2002). Pretreatment with pertussis toxin leads to ADP-ribosylation of $Ga_{i/o}$ subunits, ultimately preventing their activation by receptors. Because pertussis toxin inhibits all isoforms of $G\alpha_i$ (i.e. $G\alpha_{i1}$, $G\alpha_{i2}$, $G\alpha_{i3}$), and $G\alpha_0$ ($G\alpha_{0a}$ and $G\alpha_{0b}$), pertussis toxin insensitive (PTXi) isoforms of inhibitory Ga have been employed to determine the isoforms involved in heterologous sensitization for a number of receptors (Clark et al., 2004; Taussig et al., 1992; Tso and Wong, 2000, 2001; Watts et al., 1998; Zhang et al., 2006). Despite receptor dependency, PTXi-G α_0 robustly rescued sensitization through the dopamine D₂ receptor and µ-opioid receptor.(Clark et al., 2004; Watts et al., 1998). These observations are consistent with studies of dopamine D_2 and μ -opioid receptor coupling to Ga_0 in the central nervous system (Chalecka-Franaszek et al., 2000; Jiang et al., 2001; Sternweis and Robishaw, 1984). On the other hand, there is also evidence that other inhibitory $G\alpha$ subunits (i.e. Ga_{i1} or Ga_{i2}) are also involved in heterologous sensitization (Tso and Wong, 2000, 2001; Zhang et al., 2006). This and other research (Clark et al., 2004; Watts et al., 1998) revealed only a partial rescue by a single PTXi Ga protein, suggesting that heterologous sensitization may involve multiple inhibitory Ga subunits.

There are two other inhibitory Ga subunits, Ga_t and Ga_z , of which only Ga_t is sensitive to inactivation by pertussis toxin (Wong et al., 1992; Yamaguchi et al., 1997). Expression of Ga_t blunts heterologous sensitization through μ and δ -opioid receptors (Avidor-Reiss et al., 1996; Rubenzik et al., 2001; Thomas and Hoffman, 1996), and heterologous sensitization mediated by Ga_z seems to be dependent on the isoform of adenylyl cyclase expressed in the cells. For example, following pertussis toxin treatment and overexpression of Ga_z , heterologous sensitization through the μ -opioid receptor was observed in cells expressing AC5, but not AC6 (Ammer and Christ, 2002).

The acute activity of $Ga_{i/o}$ can also modulate the sensitized adenylyl cyclase response. Specifically, acute activation of the μ -opioid receptor reduced δ -opioid receptor-mediated heterologous sensitization (Levitt et al., 2011). Selective acute activation of δ -opioid, a_2 adrenergic, or nociception/orphanin FQ peptide receptors also reduced μ -opioid receptor-mediated heterologous sensitization (Levitt et al., 2011). These results suggest that even when sensitized, adenylyl cyclases are still subject to acute inhibition by $Ga_{i/o}$, and that during sensitization $Ga_{i/o}$ subunits retain their ability to be activated by receptors as well as inhibit cAMP. Such observations are consistent with divergent mechanisms for the acute inhibition response when compared to the sensitized response (Watts and Neve, 1996).

2.2 Gβγ subunits

Interaction with agonists promotes GPCR structural/conformational changes that lead to the activation of $G\alpha$ and also allow $G\beta\gamma$ subunits to modulate their effectors (Nygaard et al., 2013). Signaling through $G\beta\gamma$ subunits is diverse and can lead to a variety of cellular events including ERK phosphorylation, modulation of adenylyl cyclase isoforms (e.g. AC1 and AC2), conditional activation of glycine receptors, activation of phospholipase C $\beta 2/\beta 3$, GIRK (G protein-coupled inwardly rectifying potassium) and *N-type* calcium channels, phosphoinositide 3 kinase, and stabilization of microtubules (Cooper and Crossthwaite, 2006; Khan et al., 2013; Lin and Smrcka, 2011). Therefore, there are a number of different mechanisms through which $G\beta\gamma$ subunits may be involved with heterologous sensitization. To determine whether $G\beta\gamma$ subunits are required for heterologous sensitization, $G\beta\gamma$ scavengers have been employed. The C-terminus of G protein receptor kinase (BARK-Ct) and Ga_t (a-transducin) are $G\beta\gamma$ subunit sequestering-proteins. Overexpression of either of these prevented sensitization through CB1 cannabinoid, dopamine D₂, and µ-opioid receptors (Avidor-Reiss et al., 1996; Ejendal et al., 2012; Rhee et al., 2000; Rubenzik et al., 2001; Thomas and Hoffman, 1996). These data suggest that $G\beta\gamma$ subunits perform an essential role in the development of heterologous sensitization. That Gby subunits directly inhibit AC1 activity and conditionally activate AC2 (Cooper and Crossthwaite, 2006; Sunahara and Taussig, 2002) further suggests that the involvement of $G\beta\gamma$ subunits in heterologous sensitization may be dependent on the model system and adenylyl cyclase under analysis. For example, heterologous sensitization of AC2 by the dopamine D₂ receptor has been reported in HEK cells, but it has also been reported that chronic activation of the µ-opioid receptor in COS-7 cells leads to an inhibition of AC2 activity (Conley and Watts, 2013; Schallmach et al., 2006). It was also revealed that $G\beta\gamma$ scavengers partially rescued the reduced activity of AC2 following chronic activation of the μ -opioid receptor in COS-7 cells (Schallmach et al., 2006). Additionally, there are five $G\beta$ subunits and twelve

Gγ subunits, specific forms of which are associated with specific receptors and Gα proteins (Khan et al., 2013). It has further been hypothesized that different Gβγ dimers may have distinct functions (Dupre et al., 2009; Khan et al., 2013; McIntire et al., 2001), a hypothesis in agreement with the knockout of specific Gγ subunits resulting in unique phenotypes in mice (Moon et al., 2014; Schwindinger et al., 2012). Together, this research suggests that Gβγ subunits play an important role in the development of heterologous sensitization. That Gβγ scavengers inhibit sensitization of AC5, and that an AC5 mutant lacking interaction with Gβγ subunits still undergoes sensitization by the dopamine D₂ receptor (Ejendal et al., 2012; Sadana et al., 2009), suggest that one or more of the Gβγ effectors may be associated with sensitization. However, the possibility of dependence on a direct interaction is not excluded, especially for group 2 adenylyl cyclases, which are conditionally activated by Gβγ subunits through a direct interaction (Boran et al., 2011; Chen et al., 1995; Diel et al., 2008; Diel et al., 2006; Weitmann et al., 2001).

2.3 Gas subunits

 $G\alpha_s$ subunits activate all isoforms of adenylyl cyclases and, as several lines of evidence indicate, play an essential role in heterologous sensitization of specific isoforms of adenylyl cyclase. For example, heterologous sensitization has been associated with enhanced $G\alpha_s$ coupling to both receptors and adenylyl cyclases (Ammer and Schulz, 1995, 1996, 1998; Chakrabarti et al., 2005; Chen and Rasenick, 1995; Watts and Neve, 1996). It has been shown that persistent activation of the dopamine D₂ receptor leads to enhanced potency of forskolin and relative efficacy of isoproterenol (β -adrenergic receptor agonist) in C6 glioma cells (Watts and Neve, 1996). These findings are consistent with an enhancement in the activity of $G\alpha_s$ subunits, which can synergistically activate adenylyl cyclases in the presence of forskolin (Sunahara and Taussig, 2002). Accordingly, forskolin-stimulated sensitization of adenylyl cyclases has been reported in the presence of GTP (reflecting $G\alpha_s$ activity) in CHO cells following chronic activation of the adenosine A_3 receptor (Palmer et al., 1997). Notably, uncoupling of $G\alpha_s$ from adenylyl cyclases using manganese chloride blunted sensitization; furthermore, this enhanced adenylyl cyclase activity was not linked to changes in the expression levels of G proteins (Palmer et al., 1997).

The above findings suggest that during the sensitization process there is an enhancement of the interactions between Ga_s subunits and adenylyl cyclases, leading to enhanced activity of the adenylyl cyclases. These observations, the fact that all isoforms of adenylyl cyclases are stimulated by Ga_s , and that heterologous sensitization has been demonstrated for most isoforms of adenylyl cyclases, prompted us to hypothesize that heterologous sensitization is a result of enhanced interactions between Ga_s and adenylyl cyclases (Avidor-Reiss et al., 1997; Cooper and Crossthwaite, 2006; Watts and Neve, 2005; Watts et al., 2001; Watts and Neve, 1996). In order to test this hypothesis, two complementary approaches were employed. The first used a small series of mutant adenylyl cyclases (i.e. AC1 and AC5), which were insensitive to stimulation by Ga_s (Zimmermann et al., 1998). The three Ga_s -insensitive AC5 mutants failed to support dopamine D_2 receptor-induced sensitization, which was consistent with our hypothesis (Watts et al., 2001). On the other hand, the Ga_s -insensitive mutants of AC1 could still be sensitized by the persistent activation of the dopamine D_2 receptor, although the magnitude of sensitization was reduced compared to

wild-type AC1 (Vortherms et al., 2004). The second complementary approach used mouse embryonic fibroblasts that were $G\alpha_s$ deficient (Bastepe et al., 2002). In agreement with the previous studies, AC5 was not sensitized by the dopamine D_2 receptor in $G\alpha_s$ -deficient cells; moreover, sensitization was rescued by expression of $G\alpha_s$ (Vortherms et al., 2006). The results with AC1 were also consistent with the previous report using $G\alpha_s$ -insensitive mutants of AC1 (Vortherms et al., 2004). Specifically, chronic activation of the dopamine D_2 receptor led to a significant sensitized response of AC1 in the $G\alpha_s$ -deficient cells that was less than the response in the presence of $G\alpha_s$ (Vortherms et al., 2006). These data suggest that there are both $G\alpha_s$ -dependent and -independent mechanisms of heterologous sensitization, as well as adenylyl cyclase isoform-specific requirements for the expression of the sensitized response. It is also worth noting the effects of prolonged activation of $G\alpha_{i/o}$ coupled receptors on the $G\alpha_s$ -stimulated AC2 response. It has been reported that chronic activation of such receptors leads to either superinhibition or has no effects on the $G\alpha_s$ mediated AC2 response (Avidor-Reiss et al., 1997; Nevo et al., 1998; Rhee et al., 2000; Schallmach et al., 2006; Thomas and Hoffman, 1996).

The mechanisms responsible for the $G\alpha_s$ -dependent enhancements of adenylyl cyclase activity in heterologous sensitization have not yet been elucidated. However, posttranslational modifications on both $G\alpha_s$ and adenylyl cyclases, as well as membrane compartmentalization of Gas have been suggested (for more discussion see Watts and Neve, 2005). One of our previous hypotheses was that a change in palmitoylation of $G\alpha_s$ might lead to greater colocalization and interaction of Gas with adenylyl cyclase. However, $G\alpha_s$ -deficient cells overexpressing a palmitoylation-deficient $G\alpha_s$ mutant underwent sensitization to the same extent as cells expressing wild type Ga_s , suggesting that Ga_s palmitoylation is not required for heterologous sensitization (Ejendal et al., 2012). The interaction between $G\alpha_s$ and $G\beta\gamma$ subunits has also been investigated. These experiments revealed that a G $\beta\gamma$ binding-deficient G α_s mutant can also lead to sensitization of AC5 by the dopamine D₂ receptor in the same model mentioned above (Ejendal et al., 2012). Because sequestration of $G\beta\gamma$ subunits with β ARKct blunts sensitization of AC5 by the dopamine D_2 receptor, it seems that the mechanism by which $G\beta\gamma$ subunits participate in heterologous sensitization does not involve direct coupling to Ga_s (Ejendal et al., 2012). It has been shown that inhibition of small GTPases involved in signalosome assembly, such as Sar1, inhibited heterologous sensitization of AC5 (Ejendal et al., 2012). Because inhibition of Sar1 does not interfere with the interaction of AC5 with Gby subunits, these data suggest that $G\alpha_s$ and $G\beta\gamma$ subunits play their roles at different steps or signaling complexes of heterologous sensitization (Dupre et al., 2007; Dupre et al., 2009; Ejendal et al., 2012).

2.4 AGS proteins

Regulation of G protein activity is important for the modulation of acute GPCR signaling and may also be associated with heterologous sensitization. One group of proteins that regulate G protein activity is the AGS (activator of G protein signaling) proteins, which interact with G α subunits and were first identified in a yeast-based genetic screen as activators of G $\beta\gamma$ signaling (Cismowski et al., 2001; Cismowski et al., 1999; Takesono et al., 1999). Although AGS1 enhances GTP γ S binding to G α_i in vitro (Cismowski et al., 1999), in intact HEK cells, overexpression of AGS1 had no effect on acute dopamine D₂ receptor-

mediated inhibition of AC1 activity (Nguyen and Watts, 2005). It seems that the effects of AGS1 on inhibitory GPCRs are linked to $G\beta\gamma$ subunits. For example, overexpression of AGS1 leads to an inhibition of M₂ muscarinic receptor-mediated activation of GIRK channels (Luscher and Slesinger, 2010; Takesono et al., 2002). Additionally, for both the dopamine D₂ and the formyl peptide receptors, overexpression of AGS1 inhibited ERK phosphorylation (Graham et al., 2002; Nguyen and Watts, 2005). Consistent with the inhibitory effects on the signaling of G $\beta\gamma$ subunits, overexpression of AGS1 inhibited dopamine D₂ receptor-mediated sensitization of AC1 in HEK cells (Nguyen and Watts, 2005).

AGS3 is another AGS protein that is expressed throughout the brain (Blumer et al., 2005). ASG3 interacts preferentially with $Ga_{i/0}$ proteins in the GDP bound state and inhibits GTPyS binding (Blumer et al., 2005; Cismowski et al., 2000; Cismowski et al., 2001), and has been implicated in withdrawal following administration of drugs of abuse. For instance, Bowers et al. found increased levels of expression of AGS3 in the nucleus accumbens of rodents in cocaine and ethanol withdrawal models (2003; 2008). In these studies, knockdown of AGS3 reduced ethanol-seeking behavior during ethanol withdrawal (Bowers et al., 2008) and could also inhibit the behaviors associated with cocaine withdrawal (Bowers et al., 2003). Consistent with the animal models, in rat primary nucleus accumbens/ striatal neurons, morphine withdrawal caused an increase in the expression of AGS3 (Fan et al., 2009). The effects of AGS3 on acute and sensitized signaling of $Ga_{i/o}$ -coupled GPCRs have also been studied in cell models. In HEK cells, overexpression of AGS3 led to an enhancement of both acute dopamine D₂ receptor-mediated inhibition of AC1 and the potency of dopamine D_2 receptor-mediated conditional activation of AC2, a signaling event that is attributed to activation of $G\beta\gamma$ subunits (Conley and Watts, 2013; Federman et al., 1992; Watts and Neve, 1997). For heterologous sensitization it has been shown that AGS3 attenuates α_2 adrenergic receptor-mediated sensitization in CHO cells (Sato et al., 2004). It has also been shown that overexpression of AGS3 inhibits dopamine D₂ receptor-mediated sensitization of AC1 and AC2 in HEK cells (Conley and Watts, 2013). In contrast, it was reported that a siRNA-mediated reduction of AGS3 suppressed morphine-induced sensitization in nucleus accumbens/striatal neurons (Fan et al., 2009). These studies suggest a potential role for AGS proteins in the development of heterologous sensitization, although their functions appear to be both isoform- and model-specific.

2.5 RGS proteins

Another group of important modulators of G protein activity that may have a role in heterologous sensitization are the RGS (regulator of G protein signaling) proteins. This group of proteins regulate the kinetics of GPCR signaling by accelerating the GTPase activity of G α proteins (Neubig and Siderovski, 2002), and can also regulate GPCR signaling by modulating the interaction of G α proteins with their effectors independent of their GTPase accelerating activity (Hooks et al., 2008). Moreover, RGS proteins can also interact with additional signaling proteins, and may be part of larger signaling complexes with scaffolding proteins, receptors and other signaling proteins, including adenylyl cyclases and G proteins (Abramow-Newerly et al., 2006; Hooks et al., 2008; Roy et al., 2006).

Because RGS proteins interact with several components of the signaling cascade of GPCRs, they may play a role in heterologous sensitization. It has been shown that overexpression of $Ga_{i/o}$ -selective RGS proteins reduces receptor-mediated inhibition of cAMP production (Chatterjee et al., 1997; Huang et al., 1997; Wang et al., 2002), but also that RGS proteins may positively regulate signaling of $Ga_{i/o}$ -coupled receptors in cell lines (Boutet-Robinet et al., 2003). Additionally, overexpression of RGS2 and $G\alpha_s$ with different isoforms of adenylyl cyclase in HEK cells causes a decrease in basal cAMP production by AC1, AC2, AC3, AC4, AC5, and AC6, compared with control cells that were not transfected with RGS2 (Roy et al., 2006). In rats, several drugs of abuse (i.e. amphetamine, morphine, and cocaine), when administered acutely, reduce the levels of RGS4 mRNA in the locus coeruleus, whereas chronic treatments lead to increased RGS4 mRNA levels (Bishop et al., 2002; Gold et al., 2003). In contrast, it was shown that in the nucleus accumbens of mice, acute morphine leads to an increase in the levels of RGS9-2, whereas chronic morphine treatments lead to a decrease in the levels of RGS9-2 (Zachariou et al., 2003). These findings indicate that different isoforms of RGS proteins are distinctly regulated by drugs of abuse. Individual isoforms of RGS proteins may therefore have different roles in heterologous sensitization, possibly due to interactions with distinct signaling proteins (Hooks et al., 2008). Indeed, studies indicate a role for RGS proteins in heterologous sensitization. For example, expression of RGS-insensitive Ga_0 leads to enhanced potency and efficacy of heterologous sensitization through the μ -opioid receptor in C6 glioma cells (Clark et al., 2004). Furthermore, there is evidence that RGS9-2/G β 5 complexes inhibit μ opioid receptor-mediated heterologous sensitization of AC5 in mouse striatal neurons and HEK cells stably expressing the µ-opioid receptor and AC5 (Xie et al., 2012). Activation of RGS proteins could lead to disinhibition of adenylyl cyclases through regulation (i.e. GTPase enhancement) of $Ga_{i/0}$ proteins; however, studies to date suggest an inhibitory function for RGS proteins in heterologous sensitization, which is consistent with the inhibitory acute effects on isoforms of adenylyl cyclases (Clark et al., 2004; Roy et al., 2006; Xie et al., 2012).

2.6 Protein kinases and phosphatases

A role for phosphorylation in heterologous sensitization can be supported by the fact that the activity of different isoforms of adenylyl cyclases can be regulated by direct phosphorylation (Sadana and Dessauer, 2009; Watts and Neve, 2005). Moreover, protein kinases and phosphatases can also indirectly regulate the activity of adenylyl cyclases (Defer et al., 2000; Sadana and Dessauer, 2009). It has been shown that direct phosphorylation of AC5 and AC6 by protein kinase A (PKA) has inhibitory effects on the activity of these adenylyl cyclase isoforms (Chen et al., 1997; Iwami et al., 1995). Adenylyl cyclases can also be modulated by different isoforms of protein kinase C (PKC), which display stimulatory effects on AC1, AC2, AC3, AC5, and AC7, and inhibitory effects on AC4, AC6, and AC9 (Defer et al., 2000; Sadana and Dessauer, 2009). Adenylyl cyclases can also be regulated by calmodulin kinases (CaMK); CaMK IV has inhibitory effects on AC1 and CaMK II inhibits AC3 (Wayman et al., 1996; Wei et al., 1996). Furthermore, activation of the calcium-activated phosphatase calcineurin leads to inhibition of AC9 (Antoni et al., 1995). Together, these data suggest that a mechanism for heterologous sensitization may occur through the

modulation of the phosphorylation state of adenylyl cyclases or its regulatory proteins (figure 3).

Activation of $Ga_{i/o}$ -coupled receptors leads to downstream activation of protein kinases, some of which can regulate the activity of adenylyl cyclases, such as PKA, PKC, and Raf-1 kinase (figure 3) (Gordon et al., 2001; Johnston et al., 2002; Kotecha et al., 2002; Liu et al., 2003; Oak et al., 2001; Varga et al., 2003a; Varga et al., 2003b; Yoon et al., 2011). Prolonged activation of $Ga_{i/o}$ -coupled receptors sensitizes the response of AC2 and AC5 to PKC activation in cellular models, indicating that the phosphorylated state of these adenylyl cyclases can also be potentiated, and that there are additional mechanisms other than PKCmediated phosphorylation of adenylyl cyclases for these specific examples of heterologous sensitization (Avidor-Reiss et al., 1997; Conley and Watts, 2013; Cumbay and Watts, 2001; Watts and Neve, 1996). Notably, it has been shown that chronic activation of opioid receptors leads to increased phosphorylation of adenylyl cyclases, and this phosphorylated state is associated with enhanced cAMP production (Chakrabarti et al., 1998; Varga et al., 2002; Varga et al., 2003a; Varga et al., 1999). These observations suggest that phosphorylation of adenylyl cyclases might regulate heterologous sensitization through at least two different mechanisms, one in which phosphorylation is part of the acute stimulatory signal, and another in which phosphorylation occurs during the development of sensitization to increase the activity of the adenylyl cyclases. One protein that might be involved in phosphorylation of adenylyl cyclases during sensitization is Raf-1. In CHO cells, heterologous sensitization via the δ -opioid receptor leads to phosphorylation of AC6, and inhibition of Raf-1 attenuates heterologous sensitization (Varga et al., 2002; Varga et al., 1999; Yue et al., 2006). Moreover, prolonged morphine treatments in primary rat dorsal root ganglion neurons lead to enhanced cAMP production, which is prevented by inhibition of Raf-1 (Yue et al., 2008). It has also been shown that Raf-1 can phosphorylate AC2 and AC5, but not AC1 (Ding et al., 2004). Notably, Raf-1-mediated phosphorylation of adenylyl cyclases has been linked to the activity of receptors tyrosine kinases (Beazely et al., 2005; Ding et al., 2004; Tan et al., 2001; Varga et al., 1998; Varga et al., 2003a).

Several lines of evidence suggest that GPCRs can mediate transactivation of receptors tyrosine kinases (Oak et al., 2001; Wang et al., 2005; Yoon et al., 2011). In CHO cells that stably express either the dopamine D_2 or D_4 receptor, trans-activation of platelet-derived growth factor receptors seems to be dependent on G proteins, since it can be completely inhibited by pertussis toxin and partially inhibited by β ARKct (Oak et al., 2001). Transactivation of epidermal growth factor receptors by Gai/o-coupled dopamine receptors is a required event for quinpirole-mediated ERK phosphorylation in primary striatal neurons (Wang et al., 2005). Recent studies have suggested a mechanism for heterologous sensitization involving a signaling switch from G proteins to receptor tyrosine kinase-like signaling. This mechanism has been shown for the µ-opioid receptor in HEK cells, where prolonged activation of the receptor caused Src kinase-mediated receptor phosphorylation at a tyrosine residue. This results in the recruitment and activation of growth factor receptorbound protein/son of sevenless, further leading to the activation of Ras/Raf-1, which in turn phosphorylates the adenylyl cyclases and causes enhanced cAMP production (Zhang et al., 2013; Zhang et al., 2009). These studies suggest that the conversion of a $G\alpha_{i/o}$ -coupled receptor into a receptor tyrosine kinase-like signaling complex may be associated with the

development of heterologous sensitization. However, because inhibition of Ras and disruption of the receptor tyrosine kinase-like signaling complex did not fully inhibit heterologous sensitization, it is likely that there are additional events underlying the development of sensitization (Zhang et al., 2013). Further, AC1 is not phosphorylated by Raf-1, which supports the existence of isoform-dependent mechanisms (Ding et al., 2004).

That protein kinases can also negatively regulate the activity of adenylyl cyclases suggests that protein phosphatases may also have a function in heterologous sensitization. However, to our knowledge phosphatases have not been directly linked to heterologous sensitization thus far. The current data indicates that phosphorylation has a function in heterologous sensitization; nevertheless, it is likely that this role depends on both the isoform of adenylyl cyclase and the protein kinase/phosphatase under investigation.

3. Heterologous sensitization in animals

Demonstration in animal models was an important step in the establishment of heterologous sensitization as a more physiologically relevant receptor response following prolonged activation of $Ga_{i/o}$ -coupled receptors. For example, in 1988 Nestler and Tallman observed that following a five-day chronic treatment of rats with subcutaneous morphine pellets, there was an increase in the amounts of active PKA in the locus coeruleus. These results are consistent with the known outcomes of heterologous sensitization. Also in 1988, Duman et al. used a similar approach to show that chronic morphine treatments in rats resulted in increased cAMP levels in the locus coeruleus. Surprisingly, both manuscripts reported no change in cAMP/PKA activity in other brain regions known to express opioid receptors, such as the striatum (Duman et al., 1988; Nestler and Tallman, 1988).

Heterologous sensitization has long been associated with drug tolerance and dependence (Sharma et al., 1975). However, a report by Bohn et al. in 2000 further investigated these phenomena and provided evidence that the recruitment of β -arrestin was associated with opioid tolerance while heterologous sensitization appeared to be related to dependence (Bohn et al., 2000). Using β -arrestin knockout mice, they first showed that those mice did not display the signs of tolerance that are observed in wild type mice following chronic morphine treatments. On the other hand, the knockout mice still displayed behaviors consistent with drug dependence. Notably, neurons from the striatum of both wild type and β-arrestin knockout mice underwent heterologous sensitization following chronic treatment of those animals with morphine (Bohn et al., 2000). These data suggest that β -arrestin is not required for heterologous sensitization and that heterologous sensitization appears to be associated with drug dependence in mice (Bohn et al., 2000). These results are consistent with a recent report by our group that used quantitative bias analyses to show that the agonist-mediated recruitment of β -arrestin to the dopamine D₂ receptor is not correlated with heterologous sensitization in CHO cells (Brust et al., 2015). The G protein-coupled receptor kinases (GRK) that phosphorylate GPCRs allowing for the recruitment of β -arrestins (Lefkowitz and Shenoy, 2005) were also implicated in heterologous sensitization. It has been shown that overexpression of GRK2 and GRK3 inhibit D₂ dopamine receptormediated heterologous sensitization in HEK cells (Namkung et al., 2009). Nevertheless, this effect was associated with sequestration of $G\beta\gamma$ subunits (both GRK2 and GRK3 bind $G\beta\gamma$

subunits) and not with enhanced β -arrestin recruitment, since overexpression of GRK5 and GRK6 (which do not bind G $\beta\gamma$ subunits) had no significant effects (Namkung et al., 2009).

Agonists of the μ -opioid receptor are commonly used in clinics for their analgesic effects (Dworkin et al., 2007; Pasternak, 2014). It has been recently shown that tissue injury in mice results in increased μ -opioid receptor constitutive activity, which inhibits signaling events associated with nociception in neurons of the spinal cord (Corder et al., 2013). In the injured mice, inhibition of the constitutive activity of the μ -opioid receptor with naltrexone in the post-hyperalgesia state results in pain reinstatement, induction of behaviors consistent with opioid dependence, and also causes heterologous sensitization of AC1 in the spinal cord (Corder et al., 2013). Consistent with previous reports, these studies also showed a link between heterologous sensitization and the appearance of behaviors analogous to opioid dependence in mice. Furthermore, the results suggest that injury-induced physiological concentrations of endogenous opioids or constitutive receptor activity are enough to develop μ -opioid receptor-mediated heterologous sensitization of AC1 in the spinal cord of mice (Corder et al., 2013).

Heterologous sensitization has also been shown in animals through the activation of dopamine receptors. For instance, chronic i.p. injections of the dopamine D_2 receptor agonist quinpirole results in an enhancement of the cAMP response of striatal neurons to activation of dopamine D_1 receptors ($G\alpha_s$ -coupled) in hamsters (Chester et al., 2006). Additionally, chronic treatment of mice with quinpirole or pramipexole for five weeks sensitizes neurons from the nucleus accumbens to dopamine-activated cAMP production (Aloisi et al., 2011). These studies indicate that prolonged activation of the dopamine D_2 receptor-mediated activation of adenylyl cyclases. Together with the reports of heterologous sensitization through opioid receptors, these findings suggest that the cellular adaptive responses observed in cell models following prolonged activation of $G\alpha_{i/o}$ -coupled receptors (i.e. heterologous sensitization) are also manifested in animals.

4. Conclusions and future perspectives

Heterologous sensitization is characterized by a seemingly paradoxical increase in the activity of adenylyl cyclases following prolonged activation of $Ga_{i/o}$ -coupled receptors. Increasing evidence suggests that several proteins are involved in the development and expression of heterologous sensitization of adenylyl cyclase isoforms. Moreover, there appear to be adenylyl cyclase-isoform specific mechanisms. The reports discussed here suggest that during persistent activation of $Ga_{i/o}$ -coupled receptors, either an activation or a disinhibition (or perhaps both) of adenylyl cyclases drives the sensitized response. Despite progress in the last forty years, the precise mechanism of this receptor-mediated response remains unknown. It is also important to note that 1) the specificity of the mechanistic clues reported for heterologous sensitization in terms of type of receptor and adenylyl cyclase isoform and 2) methodological and model system differences likely influence the overall outcomes of the experiments and contribute to the difficulties associated with drawing a generalized mechanism for heterologous sensitization.

In addition to G proteins and kinases, several adenylyl cyclase-interacting proteins have been identified (Sadana and Dessauer, 2009; Wang et al., 2009). One relatively simple hypothesis for the general mechanism of heterologous sensitization is that the observed enhanced adenylyl cyclase activity results from a protein interaction that is induced (or inhibited) by prolonged receptor activation. These interactions can be part of a larger signaling complex and may involve translocation of signaling components or even a rearrangement of pre-existing signaling complexes; these could influence important signaling features, such as the presence of adenylyl cyclases in microdomains and cellular compartmentalization (Wang et al., 2009; Willoughby and Cooper, 2007). We are exploring approaches to examine the heterologous sensitization interactome using a bimolecular fluorescence complementation (BiFC) screening approach. BiFC is a method used to measure and visualize protein-protein interactions. In BiFC, proteins are fused with opposite termini of a fluorescent protein (e.g. N- and C-termini of Green Fluorescent Protein). Upon physical interaction between the fusion proteins, the two termini of the fluorescent protein complement to form a full fluorescent protein that can be detected by a fluorescence microscope, a flow cytometer, or a fluorescence plate reader (Hu and Kerppola, 2003; Vidi et al., 2010). Adenylyl cyclase isoforms tagged with a BiFC terminus and a library of proteins tagged with the complementary BiFC terminus can be employed for the screen. To further study their roles in heterologous sensitiziation, we anticipate identifying proteins that have increased/decreased interactions with adenylyl cyclases as a result of persistent activation of $Ga_{i/o}$ -coupled receptors. These studies may also be useful to identify additional adenylyl cyclase isoform-specific interacting partners and understand where in the receptor signaling cascade heterologous sensitization branches out and becomes specific for individual isoforms of adenylyl cyclase. A second strategy to achieve this and discover additional proteins involved in heterologous sensitization is siRNA library screening. We have recently developed a high-throughput assay platform to examine the effects of siRNAmediated knockdown of proteins on heterologous sensitization (Conley et al., 2014). These methods can be adapted to screen a siRNA genome-wide library, pursuing genes that modulate heterologous sensitization of different isoforms of adenylyl cyclase. It is anticipated that these experiments will lead to the identification of genes and proteins that are shared by different adenylyl cyclase isoforms, as well as those that are specific modulators of individual isoforms of adenylyl cyclases.

Heterologous sensitization of adenylyl cyclases is associated with the physiological responses observed following chronic treatment with agonists of $Ga_{i/o}$ -coupled receptors. For example, the mechanisms may be relevant to the side effects of pharmacological treatments that lead to increased $Ga_{i/o}$ -coupled receptor activity. It is anticipated that continued research to unveil the molecular mechanisms of heterologous sensitization in cell and animal models will lead to a better understanding of the molecular basis for adaptive pathological changes associated with drug dependence and neurological disorders.

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Figure 1.

Differential regulation of adenylyl cyclase by $G\alpha_{i/o}$ -coupled receptors. **A**. Acute activation of $G\alpha_{i/o}$ -coupled receptors leads to activation of $G\alpha_{i/o}$ proteins, which inhibit adenylyl cyclases and, therefore, decrease cAMP production. **B**. In heterologous sensitization persistent activation of $G\alpha_{i/o}$ -coupled receptors causes a cellular adaptive response that enhances drug-stimulated adenylyl cyclase activity and increased cAMP accumulation.



Figure 2.

Temporal relationship of cAMP accumulation as a result of the activation of $Ga_{i/o}$ -coupled receptors. Initial activation of $Ga_{i/o}$ -coupled receptors leads to a decrease in cAMP production, however, persistent activation leads to heterologous sensitization and a subsequent enhancement in cAMP production that is readily observed by removing the $Ga_{i/o}$ agonist and stimulating the adenylyl cyclase.



Figure 3.

Isoform-specific regulation of adenylyl cyclase by protein kinases that are activated by $G\alpha_{i/o}$ -coupled GPCRs. One of the proposed mechanisms for heterologous sensitization is that the phosphorylation state of the adenylyl cyclase is modified due to prolonged activation of $G\alpha_{i/o}$ -coupled receptors. Activation of inhibitory GPCRs leads to active $G\alpha_{i/o}$ and $G\beta\gamma$ subunits that can activate PKC, which phosphorylates and activates AC1, AC2, and AC5. Activation of $G\alpha_{i/o}$ inhibits PKA, leading to disinhibition of AC5 and AC6. $G\beta\gamma$ subunits can also activate Raf-1, which phosphorylates and activates AC2, AC5, and AC6.