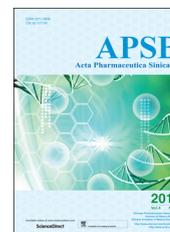




Chinese Pharmaceutical Association
Institute of Materia Medica, Chinese Academy of Medical Sciences

Acta Pharmaceutica Sinica B

www.elsevier.com/locate/apsb
www.sciencedirect.com



REVIEW

The pharmacological impact of ATP-binding cassette drug transporters on vemurafenib-based therapy



Chung-Pu Wu^{a,b,c}, Suresh V. Ambudkar^{d,*}

^aDepartment of Physiology and Pharmacology, College of Medicine, Chang Gung University, Tao-Yuan

^bMolecular Medicine Research Center, College of Medicine, Chang Gung University, Tao-Yuan

^cGraduate Institute of Biomedical Sciences, College of Medicine, Chang Gung University, Tao-Yuan

^dLaboratory of Cell Biology, Center for Cancer Research, National Cancer Institute, NIH, Bethesda, MD, USA

Received 1 November 2013; revised 2 December 2013; accepted 10 December 2013

KEY WORDS

ABC transporter;
Drug resistance;
Melanoma;
P-glycoprotein;
Vemurafenib

Abstract Melanoma is the most serious type of skin cancer and one of the most common cancers in the world. Advanced melanoma is often resistant to conventional therapies and has high potential for metastasis and low survival rates. Vemurafenib is a small molecule inhibitor of the BRAF serine-threonine kinase recently approved by the United States Food and Drug Administration to treat patients with metastatic and unresectable melanomas that carry an activating BRAF (V600E) mutation. Many clinical trials evaluating other therapeutic uses of vemurafenib are still ongoing. The ATP-binding cassette (ABC) transporters are membrane proteins with important physiological and pharmacological roles. Collectively, they transport and regulate levels of physiological substrates such as lipids, porphyrins and sterols. Some of them also remove xenobiotics and limit the oral bioavailability and distribution of many chemotherapeutics. The overexpression of three major ABC drug transporters is the most common mechanism for acquired resistance to anticancer drugs. In this review, we highlight some of the recent findings related to the effect of ABC drug transporters such as ABCB1 and ABCG2 on the oral bioavailability of vemurafenib, problems associated with treating

Abbreviations: ABC, ATP-binding cassette; AML, acute myeloid leukemia; BBB, blood–brain barrier; CNS, central nervous system; CSCs, cancer stem cells; GI, gastrointestinal; MAPK, mitogen-activated protein kinase; MDR, multidrug resistance; NBDs, nucleotide-binding domains; PFS, longer progression-free survival; PKIs, protein kinase inhibitors; TKIs, tyrosine kinase inhibitors; TMDs, transmembrane domains

*Corresponding author. Tel.: +1 301 402 4178; fax: +1 301 435 8188.

E-mail address: ambudkar@helix.nih.gov (Suresh V. Ambudkar).

Peer review under responsibility of Institute of Materia Medica, Chinese Academy of Medical Sciences and Chinese Pharmaceutical Association.



Production and hosting by Elsevier

melanoma brain metastases and the development of acquired resistance to vemurafenib in cancers harboring the BRAF (V600E) mutation.

© 2014 Chinese Pharmaceutical Association and Institute of Materia Medica, Chinese Academy of Medical Sciences. Production and hosting by Elsevier B.V. Open access under [CC BY-NC-ND license](#).

1. Introduction

Melanoma is the most serious type of skin cancer. It originates in pigment-producing melanocytes¹. Melanoma has become one of the most common cancers in the world. Due to its high potential for metastasis, individuals with this disease have a poor prognosis and low survival rates². Melanoma at advanced stages is often resistant to conventional radiation therapy and chemotherapy as a result of multiple mechanisms, including increased DNA repair and alterations of several key regulatory genes or proteins^{3,4}. Therefore, therapeutic approaches directed at specific signaling pathways or mutations in melanoma have been employed^{5,6}. One of the targets is the RAS-activated serine-threonine protein kinase B-raf (BRAF). It plays a central role in the regulation of the mitogen-activated protein kinase (MAPK) signaling pathway that regulates cell division, proliferation and differentiation in melanoma^{7,8}. The consequence of mutations is the constitutive activation of the BRAF kinase and downstream MAPK signaling that promotes unregulated cell proliferation and cell invasion. In melanoma patients, the BRAF(V600E; valine to glutamate) substitution is the most common mutation⁹, which is associated with poor clinical outcome¹⁰ and brain metastases¹¹. Since this mutation is found in approximately 40–60% of melanoma patients⁸, improved clinical outcome is expected for melanoma patients with inhibition of BRAF(V600E) signaling^{8–10}.

2. Vemurafenib treatment for BRAF (V600E) mutation patients with advanced or metastatic melanoma

Vemurafenib (PLX4032, Zelboraf[®]) is a small molecule inhibitor of the cytoplasmic BRAF serine-threonine kinase (chemical structure given in Fig. 1), which in 2011 was approved by the US Food and Drug Administration (FDA) for treatment of metastatic and unresectable melanomas that carry an activating BRAF(V600E) mutation^{12–14}. Moreover, in addition to treat unresectable BRAF(V600E) mutant melanomas¹², studies on evaluating the effectiveness of vemurafenib in brain metastases of melanoma (ClinicalTrials.gov identifier NCT01378975), colorectal cancer^{15,16} (ClinicalTrials.gov identifier NCT00405587) and thyroid cancer¹⁷ (ClinicalTrials.gov identifier NCT01709292) are ongoing. Unfortunately, acquired drug resistance to vemurafenib and relapse among patients were reported frequently within months of therapy^{12,14}. Identifying and overcoming mechanisms that lead to acquired clinical resistance to vemurafenib presents a significant therapeutic challenge¹⁸.

3. The impact of ATP-binding cassette transporter-mediated drug transport on cancer chemotherapy

Generally, the success of cancer chemotherapy depends on several key factors. For an anticancer agent to be effective, a sufficient amount of the drug must be distributed to the target site(s), which is dependent on the chemical and biological properties of the therapeutic agent, as well as the location of the target site(s). Cancer cells can often acquire resistance through adaptation or spontaneous

induction of certain key regulatory genes during the course of chemotherapy, which is dependent on the patient, cancer type, stage of the disease and treatment strategy^{4,19}. Collectively, drug absorption, distribution and acquired resistance may result in poor response to chemotherapy and unfavorable patient outcome. Among various adverse factors in cancer chemotherapy, energy dependent drug efflux and drug compartmentalization are the most common ways that cancer cells evade drug absorption and drug penetration^{20,21}.

Normally, the first line of cellular defense against xenobiotics is to rapidly reduce the intracellular concentration of xenobiotics by means of a transporter-mediated efflux system. Unfortunately, cancer cells can utilize the same protective mechanism by up-regulating some of the drug transporters that reduce drug sensitivity in patients, many of whom eventually relapse with multidrug-resistant forms of cancer¹⁹. One of the most common causes of acquired drug resistance in cancer is energy-dependent drug efflux by members of the human ATP-Binding Cassette (ABC) protein superfamily. Human ABC proteins are subdivided into seven families (ABCA-ABCG), based on structural and sequence similarities²⁰. Several ABC proteins are

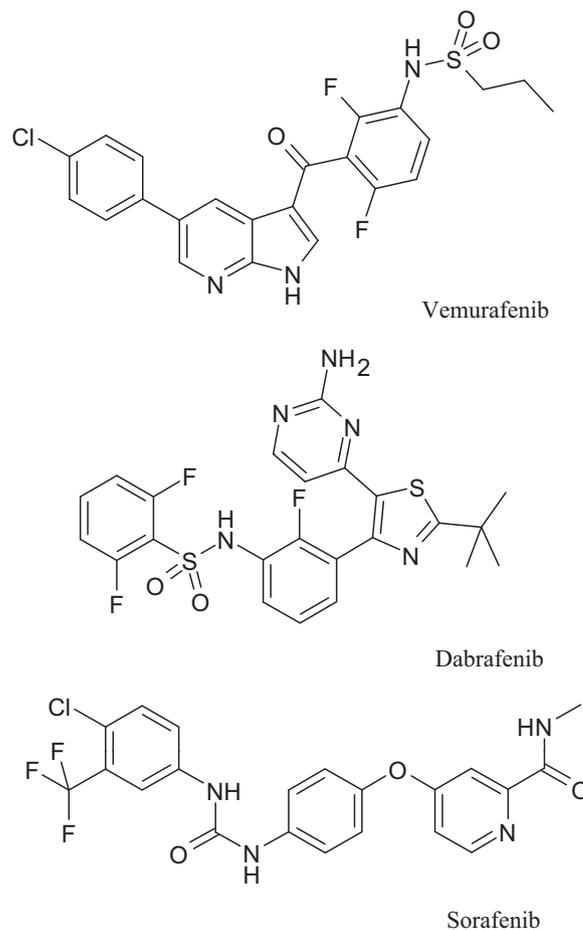


Figure 1 Chemical structures of vemurafenib, dabrafenib and sorafenib.

transporters that can utilize energy derived from ATP to mediate direct drug efflux. These ABC transporters are membrane proteins, consisting of transmembrane domains (TMDs) and distinctive nucleotide-binding domains (NBDs). The TMDs form substrate-binding pockets, while the NBDs generate energy from ATP hydrolysis to actively transport a wide range of substrates, including anticancer agents, across biological membranes, reducing intracellular drug concentration and eventually resulting in multidrug resistance (MDR)²². ABCA9, ABCB1, ABCB5, ABCB8, ABCC2, ABCD1 and ABCG2 are some of the ABC proteins that have been identified in melanoma cells^{23–28}. In this review, we focus mainly on the potential roles of ABCB1, ABCG2 and ABCB5 in limiting the absorption, distribution and penetration of vemurafenib, as well as in the development of resistance to this drug in cancer cells expressing a BRAF(V600E) mutation.

3.1. ABCB1

The 170 kDa cell membrane ABCB1 (also known as P-glycoprotein, P-gp) was the first member of the mammalian ABC protein family to be identified²⁹. ABCB1 consists of two transmembrane domains, each containing six α -helices, both linked to ATP-binding domains that provide energy by hydrolyzing ATP to transport drug substrate across cell membranes. A large number of classical anticancer agents including taxanes, *Vinca* alkaloids, etoposide, teniposide, camptothecins, methotrexate, colchicines, actinomycin D, anthracyclines and mitoxantrone are well-known drug substrates of ABCB1. More importantly, many of the newly developed targeted therapy drugs such as tyrosine kinase inhibitors (TKIs), have been identified as substrates of ABCB1 as well³⁰. ABCB1 is expressed in endothelial cells at the blood–brain barrier (BBB) sites in normal brain tissue and also in primary brain tumors, and its functions to limit penetration of the brain by many chemotherapeutics^{31,32}. In addition, ABCB1 is highly expressed in many normal tissues, including those of the liver and intestinal walls, signifying the physiological and pharmacological importance of ABCB1²⁰. Moreover, ABCB1 is known to be over-expressed in many types of cancer and is linked to the MDR phenotype³³. Considering the wide tissue distribution and substrate specificity of ABCB1, it is not surprising that ABCB1 plays a key role in limiting the oral bioavailability of anticancer drugs, preventing drug distribution and penetration through the blood–brain barrier and affecting therapeutic outcome in patients¹⁹. In terms of melanomas, endogenous ABCB1 mRNA has been detected in the melanoma cell lines SK-MEL-28, SK-MEL-5 and M16^{23,34}, as well as non-cutaneous melanomas^{35,36}. ABCB1 was also detected in a subpopulation of human melanoma cells that co-express ABCB5, hTERT, and Nanog, and has high self-renewal capacity, representing characteristics of melanoma stem cells³⁷. Interestingly, though the MDR phenotype has been shown in human BRO melanoma cells transfected with human ABCB1³⁸, the relevance of endogenous ABCB1 in conferring drug resistance in melanomas has not been demonstrated yet.

3.2. ABCG2

ABCG2 (also known as breast cancer resistance protein, BCRP; or placenta-specific ABC transporter, ABCP; or mitoxantrone resistance protein, MXR) was identified in 1998^{39,40}. In contrast to ABCB1, ABCG2 consists of a single ATP-binding domain followed by a transmembrane domain with six α -helices in a reverse orientation⁴¹. A functional unit of ABCG2 is a dimer or a multimer. Similar to ABCB1, ABCG2 is overexpressed in many cancers, and is linked to

reduced drug accumulation and to the development of MDR in patients with advanced non-small cell lung cancer or acute myeloid leukemia (AML)^{42,43}. ABCG2 is capable of transporting a large variety of anticancer agents such as etoposide, docetaxel, topotecan, CPT-11, SN-38, methotrexate, flavopiridol, anthracyclines, mitoxantrone, and similar to ABCB1, many tyrosine kinase inhibitors including imatinib, nilotinib, saracatinib and ponatinib^{30,44,45}. ABCG2 also has a physiological and pharmacological impact on drug bioavailability, drug distribution, protection of cells or tissues from xenobiotics and the transport of porphyrins and sterols³³. Similar to ABCB1, ABCG2 has been detected at the luminal membrane of brain capillaries and the BBB, protecting the brain from xenobiotics and chemotherapeutics^{46,47}. Studies have shown that both the protein expression and function of ABCG2 are up-regulated in neuro-epithelial tumors, restricting penetration of chemotherapeutics and leading to the development of MDR^{48,49}. ABCG2 is believed to play a protective role in cancer stem cells (CSCs) or “side population” cells, with self-renewal properties and critical roles in tumorigenesis, metastasis and relapse⁵⁰. Since ABCG2 is expressed in a wide range of human stem cells, it is considered as a biomarker for stem cells. ABCG2, along with CD133 and nestin, have been detected in melanomas^{25,51–53}, but the potential contribution of ABCG2 to chemoresistance in melanomas remains to be determined. Recently, ABCG2 has been linked to the disease gout, as mutations (for example Q141K) in this transporter result in decreased efflux of urate from kidney epithelial cells^{54,55}.

3.3. ABCB5

ABCB5 is predominantly expressed in pigment-producing (melanogenic) melanoma cells^{23,24}. The melanogenesis-related vesicles, called “melanosomes” are derived from lysosomes and represent a unique feature of melanomas^{56,57}. Structurally, ABCB5 has 73% sequence homology with ABCB1 protein^{24,26}. In contrast to ABCB1, which mediates drug efflux from cells, ABCB5 is thought to confer chemoresistance to cisplatin, doxorubicin and daunorubicin by intracellular drug sequestration^{16,24,57,58}. Furthermore, studies have reported that ABCB5 protein expression is up-regulated upon exposure to the chemotherapeutic drugs dacarbazine (DTIC) and doxorubicin^{28,59}. Both ABCB1 and ABCG2 are known to be present in cancer stem cells, and hence are used as stem cell markers⁶⁰. Similar cancer stem cell properties were discovered in metastatic melanoma cells, in which ABCB5 was present⁶¹. These ABCB5-positive melanoma stem cells are not only drug-resistant, but also possess self-renewal, differentiation and tumorigenic capabilities^{58,62}. Interestingly, a recent study showed that ABCB5-expressing cells are resistant to temozolomide, dacarbazine and vemurafenib, suggesting that ABCB5 may contribute to the drug resistance mechanism, and thus is a potential therapeutic target for melanoma chemotherapy²⁸. However, ABCB5-mediated transport of these drugs in melanoma patient samples has not yet been demonstrated.

4. The pharmacological impact of ABC drug transporters on the bioavailability and distribution of vemurafenib

Reports have shown a high incidence of melanoma metastases in the brain^{63,64}. Prior to the discovery of vemurafenib, a patient's response to the standard therapy of interleukin-2 and dacarbazine was extremely poor^{14,65}. However, in order for vemurafenib to be effective against brain metastases of melanoma, sufficient amounts of vemurafenib must first be absorbed in the gastrointestinal (GI)

tract (Fig. 2A), be distributed, and also penetrate the BBB and accumulate in the brain (Fig. 2B). The vasculature structure of the BBB consists of tightly sealed tight-junction protein complexes combined with overexpression of several ABC transporters that actively transport chemotherapeutics back into the bloodstream (Fig. 2B), making drug penetration of the brain a major obstacle in chemotherapy⁶⁶.

A recent study by Mohammed et al.⁶⁷ reported that the delivery of vemurafenib to the brain is restricted due to its direct transport by human ABCB1 and mouse Abcg2 at the blood–brain barrier. In their *in vitro* experiments, the intracellular accumulation of vemurafenib was reduced in MDCKII cells transfected with ABCB1

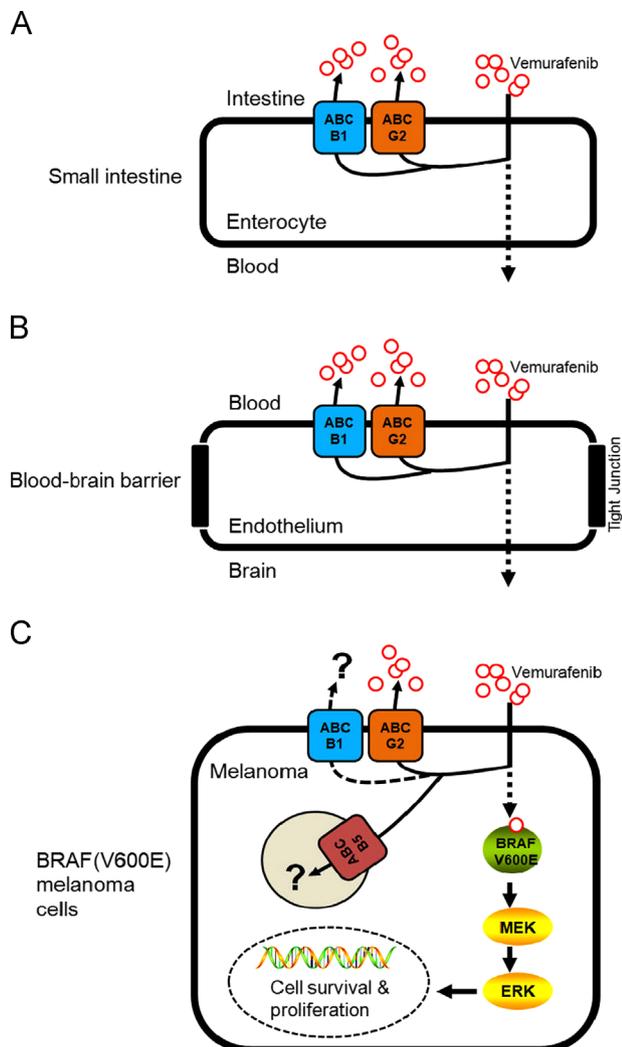


Figure 2 The potential role of multidrug resistance-associated ABC drug transporters in the oral bioavailability, brain penetration and therapeutic efficacy of vemurafenib in melanoma and other cancer cells harboring V600E mutation in BRAF kinase. (A) Highly active ABCB1 and ABCG2 transporters in intestinal epithelial cells can significantly limit the absorption of vemurafenib into the blood stream, reducing its bioavailability. (B) The presence of both ABCB1 and ABCG2 at the blood–brain barrier restricts vemurafenib penetration of the brain, reducing its effectiveness in patients with brain metastatic melanoma. (C) The presence of ABCG2 confers resistance to vemurafenib in BRAF(V600E) mutant A375 melanoma cells. The role of the ABCB5 transporter in melanoma remains to be evaluated.

or ABCG2, as a direct result of ABCB1 and ABCG2-mediated transport of vemurafenib. Moreover, the ABCB1 and ABCG2-mediated transport of vemurafenib can be inhibited by zosuquidar and Ko143, respectively. Furthermore, in their knockout mouse model, the brain-to-plasma ratios of vemurafenib were increased significantly when *Abcb1a/1b* and *Abcg2* were both absent. The authors concluded that vemurafenib is a substrate of both ABCB1 and ABCG2, and both transporters play a significant role in limiting the central nervous system (CNS) distribution of vemurafenib. The findings by Mohammed et al. were later supported by an independent group. Durmus et al.⁶⁸ reported that inhibition of both ABCB1 and ABCG2 could significantly improve the bioavailability (Fig. 2A) and brain penetration (Fig. 2B) of vemurafenib. In their *in vitro* experiments, vemurafenib transport mediated by either ABCB1 or ABCG2 was demonstrated by using MDCK II cells transfected with either human ABCB1 or ABCG2. The ABCB1- and ABCG2-mediated transport of vemurafenib was inhibited completely by the ABCB1 inhibitor zosuquidar and the ABCG2 inhibitor Ko143⁶⁸. *In vivo*, the dual *Abcb1a/1b* and *Abcg2* inhibitor elacridar significantly elevated the plasma levels of vemurafenib and brain accumulation in WT mice to the same levels as in *Abcb1a/1b*^{-/-}; *Abcg2*^{-/-} mice. Interestingly, Durmus et al.⁶⁸ found that *Abcg2* is responsible for reducing the intestinal uptake of vemurafenib, but limited to a lower oral dose. In contrast, *Abcb1a/1b* is accountable for reducing plasma levels of vemurafenib at later stages. This particular observation is in accordance with findings by Chapman et al.¹⁴, that in BRAF(V600E) mutant A375 melanoma cells, ABCG2 behaves as a high-affinity but low capacity transporter of vemurafenib.

5. The potential impact of ABC drug transporters on vemurafenib-based treatment of advanced or metastatic melanoma

Initial success at using vemurafenib to treat patients with metastatic and unresectable melanomas or other cancers that carry an activating BRAF(V600E) mutation was short lived. The rapid development of acquired resistance to vemurafenib is now becoming a major obstacle in the treatment of patients diagnosed with BRAF(V600E)-positive cancer^{12,14}. Multiple mechanisms involving the reactivation of the mitogen-activated protein kinase (MAPK) pathway have been reported in vemurafenib-resistant BRAF(V600E) mutant cancer cells. Up-regulation of CRAF^{69,70} and overexpression of Tpl2/COT⁶⁹, RAS activation^{38,71}, enhanced activation of the FGFR3/RAS pathway⁷², pathways that lead to reactivation of ERK signaling⁷³ and activation of RTK signaling pathways such as IGF-1R or PDGFR β ^{25,71,74} have all been shown to contribute to acquired resistance to vemurafenib, depending on the cancer type^{17,26}.

Recently, we have discovered that in addition to a RAF isoform switch and activation of various compensatory survival pathways^{25,38,69–74}, the overexpression of ABCG2 could also contribute to the development of acquired resistance to vemurafenib in BRAF(V600E) mutant cancer cells (Fig. 2C)⁶. This is not surprising since the overexpression of ABC transporters is one of the most common mechanisms of acquired resistance to anticancer agents³³. In our study, the interactions of vemurafenib with three major MDR-associated ABC drug transporters, ABCB1, ABCC1 and ABCG2 were investigated. Results showed that vemurafenib binds directly to the substrate binding pockets of ABCG2, inhibits its function and stimulates ATP hydrolysis. Similar interactions between vemurafenib and ABCB1 were observed, but the binding affinity and the

stimulation of ATP hydrolysis were significantly lower. We found that since vemurafenib binds to the drug binding site of human ABCG2 with relatively high affinity, it effectively inhibited ABCG2-mediated transport of other drug substrates. Moreover, at non-toxic concentrations, vemurafenib was able to restore chemosensitivity of ABCG2-overexpressing HEK293 cells to anticancer agents such as mitoxantrone and topotecan. Similarly, vemurafenib also restored the sensitivity of drug-resistant ABCG2-overexpressing and also expressing (V600E) mutant BRAF A375 melanoma cells to mitoxantrone⁶. In contrast, no interaction was detected between vemurafenib and ABCC1 protein. Moreover, 72 h of vemurafenib treatment had no significant effect on the expression of ABCB1, ABCC1 or ABCG2 protein in cancer cells expressing wild-type BRAF. Surprisingly, while overexpression of human ABCG2 had no effect on the chemosensitivity of wild-type BRAF cancer cells to vemurafenib, the ectopic expression of human ABCG2 led to vemurafenib resistance in A375 melanoma cells harboring the BRAF(V600E) mutation. We found that in A375 melanoma cells, BRAF kinase inhibition by vemurafenib was significantly reduced in the presence of functional ABCG2, implicating ABCG2-mediated efflux as a mechanism of resistance for vemurafenib⁶. Unfortunately, it is still unknown whether prolonged treatment with vemurafenib leads to overexpression of ABC drug transporters in BRAF(V600E) melanoma, thyroid or colorectal cancers. Furthermore, the potential impact of ABCB1 or ABCB5 or other MDR-associated ABC drug transporters on the therapeutic outcome using vemurafenib in melanomas or other cancers harboring the BRAF(V600E) mutation needs to be determined.

6. Impact of ABC drug transporters on treatment with other BRAF inhibitors (dabrafenib and sorafenib)

Dabrafenib (GSK2118436) is a new BRAF inhibitor (Fig. 1) designed to target melanomas expressing V600E and V600K mutant BRAF. Good clinical response rates have been observed in metastatic melanoma (including brain metastases) patients receiving dabrafenib^{75,76}, but cases of acquired resistance to dabrafenib have also been reported^{77–79}. Although the link between ABC drug transporters and acquired resistance to dabrafenib is still lacking, a recent study using MDCKII cells indicated that dabrafenib is a substrate of both ABCB1 and ABCG2⁸⁰. Moreover, Mittapalli et al.⁸⁰ showed that in both *in vivo* and intact BBB models, the dabrafenib brain distribution is limited by the function of both ABCB1 and ABCG2. In contrast to vemurafenib and dabrafenib, sorafenib is a nonselective BRAF inhibitor (Fig. 1) that targets both BRAF and CRAF, and inhibits other multiple kinases⁸¹. A phase I/II clinical trial reported that in metastatic melanoma patients, combination therapy of sorafenib, carboplatin and paclitaxel demonstrated a better response rate and longer progression-free survival than with standard chemotherapy⁸². Like vemurafenib and dabrafenib, the interactions between sorafenib, ABCB1 and ABCG2 have been demonstrated by several independent groups. Other studies have reported that sorafenib is transported by both ABCB1^{83,84} and ABCG2, but more efficiently by ABCG2⁸⁴, and consistent with these findings the penetration of the brain by sorafenib was significantly higher in *Abcg2*^{-/-} mice than in WT^{84,85}.

7. Conclusions

Collectively, the actions of ABCB1 and ABCG2 in the GI tract and at the BBB contribute significantly to reduced oral bioavailability and limit the penetration of the brain by vemurafenib

(Fig. 2A and B), which is a major obstacle when treating patients with melanoma brain metastases. The clinical application of a dual ABCB1 and ABCG2 inhibitor such as elacridar could possibly provide a solution to increase the oral bioavailability and enhance brain penetration of vemurafenib in patients with brain metastatic melanoma⁸⁶. At the cellular level, the presence of MDR-associated ABC drug transporters may present new therapeutic challenges when treating cancers expressing the V600E mutant version of BRAF kinase. The ability of ABC drug transporters to effectively reduce the intracellular concentration of vemurafenib in cancer cells can potentially lead to acquired resistance to this drug (Fig. 2C). Moreover, the reported high affinity of vemurafenib for binding to ABCG2 suggests the potential use of vemurafenib as a chemosensitizer that would work alongside classical anticancer agents to treat ABCG2-positive MDR cancers. Consistent with these findings, vemurafenib was found to dock in the drug-binding pocket of the homology model of human ABCB1 and ABCG2 and also modulate the function of the ABCC10 (MRP 7) transporter⁸⁷. Thus, we propose that simultaneous administration of vemurafenib and protein kinase inhibitors targeting key signaling pathways that are involved in the development of acquired resistance to vemurafenib^{25,38,69–74}, as well as inhibiting the actions of ABC drug transporters in BRAF(V600E) mutant cancers³³, may offer great promise for effective treatment of melanoma patients.

Acknowledgments

CP Wu was supported by funds from National Science Council of Taiwan (Grant No. NSC102-2320-B-182-036). S.V. Ambudkar was supported by the Intramural Research Program of the National Institutes of Health, National Cancer Institute. National Cancer Institute, NIH, Center for Cancer Research. We thank George Leiman for editorial assistance.

References

- Shukla S, Skoumbourdis AP, Walsh MJ, Hartz AM, Fung KL, Wu CP, et al. Synthesis and characterization of a BODIPY conjugate of the BCR-ABL kinase inhibitor Tasigna (nilotinib): evidence for transport of Tasigna and its fluorescent derivative by ABC drug transporters. *Mol Pharm* 2011;**8**:1292–302.
- Balch CM, Gershenwald JE, Soong SJ, Thompson JF, Atkins MB, Byrd DR, et al. Final version of 2009 AJCC melanoma staging and classification. *J Clin Oncol* 2009;**27**:6199–206.
- Chin L, Garraway LA, Fisher DE. Malignant melanoma: genetics and therapeutics in the genomic era. *Genes Dev* 2006;**20**:2149–82.
- Soengas MS, Lowe SW. Apoptosis and melanoma chemoresistance. *Oncogene* 2003;**22**:3138–51.
- Tsao H, Atkins MB, Sober AJ. Management of cutaneous melanoma. *N Engl J Med* 2004;**351**:998–1012.
- Ahmadzadeh M, Johnson LA, Heemskerk B, Wunderlich JR, Dudley ME, White DE, et al. Tumor antigen-specific CD8 T cells infiltrating the tumor express high levels of PD-1 and are functionally impaired. *Blood* 2009;**114**:1537–44.
- Wolf A, Bauer B, Hartz AM. ABC transporters and the Alzheimer's disease enigma. *Front Psychiatry* 2012;**3**:54.
- Wan PT, Garnett MJ, Roe SM, Lee S, Niculescu-Duvaz D, Good VM, et al. Mechanism of activation of the RAF-ERK signaling pathway by oncogenic mutations of B-RAF. *Cell* 2004;**116**:855–67.
- Hartz AM, Miller DS, Bauer B. Restoring blood-brain barrier P-glycoprotein reduces brain amyloid-beta in a mouse model of Alzheimer's disease. *Mol Pharmacol* 2010;**77**:715–23.

10. Arkenau HT, Kefford R, Long GV. Targeting BRAF for patients with melanoma. *Br J Cancer* 2011;**104**:392–8.
11. Berghoff AS, Capper D, Preusser M. Lack of BRAF V600E protein expression in primary central nervous system lymphoma. *Appl Immunohistochem Mol Morphol* 2013;**21**:351–3.
12. Bollag G, Hirth P, Tsai J, Zhang J, Ibrahim PN, Cho H, et al. Clinical efficacy of a RAF inhibitor needs broad target blockade in BRAF-mutant melanoma. *Nature* 2010;**467**:596–9.
13. Facchetti F, Monzani E, La Porta CA. New perspectives in the treatment of melanoma: anti-angiogenic and anti-lymphangiogenic strategies. *Recent Pat Anticancer Drug Discov* 2007;**2**:73–8.
14. Chapman PB, Hauschild A, Robert C, Haanen JB, Ascierto P, Larkin J, et al. Improved survival with vemurafenib in melanoma with BRAF V600E mutation. *N Engl J Med* 2011;**364**:2507–16.
15. Hartz AM, Bauer B. ABC transporters in the CNS—an inventory. *Curr Pharm Biotechnol* 2011;**12**:656–73.
16. Cheung PF, Cheng CK, Wong NC, Ho JC, Yip CW, Lui VC, et al. Granulin-epithelin precursor is an oncofetal protein defining hepatic cancer stem cells. *PLoS One* 2011;**6**:e28246.
17. Rouzaud F, Costin GE, Yamaguchi Y, Valencia JC, Berens WF, Chen KG, et al. Regulation of constitutive and UVR-induced skin pigmentation by melanocortin 1 receptor isoforms. *FASEB J* 2006;**20**:1927–9.
18. Das Thakur M, Salangsang F, Landman AS, Sellers WR, Pryer NK, Levesque MP, et al. Modelling vemurafenib resistance in melanoma reveals a strategy to forestall drug resistance. *Nature* 2013;**494**:251–5.
19. Esiobu N, Green M, Echeverry A, Bonilla TD, Stinson CM, Hartz A, et al. High numbers of *Staphylococcus aureus* at three bathing beaches in South Florida. *Int J Environ Health Res* 2013;**23**:46–57.
20. Gottesman MM, Fojo T, Bates SE. Multidrug resistance in cancer: role of ATP-dependent transporters. *Nat Rev* 2002;**2**:48–58.
21. Calcagno AM, Salcido CD, Gillet JP, Wu CP, Fostel JM, Mumau MD, et al. Prolonged drug selection of breast cancer cells and enrichment of cancer stem cell characteristics. *J Nat Cancer Inst* 2010;**102**:1637–52.
22. Higgins CF. ABC transporters: from microorganisms to man. *Annu Rev Cell Biol* 1992;**8**:67–113.
23. Szakacs G, Annereau JP, Lababidi S, Shankavaram U, Arciello A, Bussey KJ, et al. Predicting drug sensitivity and resistance: profiling ABC transporter genes in cancer cells. *Cancer Cell* 2004;**6**:129–37.
24. Chen KG, Szakacs G, Annereau JP, Rouzaud F, Liang XJ, Valencia JC, et al. Principal expression of two mRNA isoforms (ABCB 5alpha and ABCB 5beta) of the ATP-binding cassette transporter gene ABCB 5 in melanoma cells and melanocytes. *Pigment Cell Res* 2005;**18**:102–12.
25. Facchetti F, Monzani E, Cavallini G, Bergamini E, La Porta CA. Effect of a caloric restriction regimen on the angiogenic capacity of aorta and on the expression of endothelin-1 during ageing. *Exp Gerontol* 2007;**42**:662–7.
26. Frank NY, Pendse SS, Lapchak PH, Margaryan A, Shlain D, Doeing C, et al. Regulation of progenitor cell fusion by ABCB5 P-glycoprotein, a novel human ATP-binding cassette transporter. *J Biol Chem* 2003;**278**:47156–65.
27. Elliott AM, Al-Hajj MA. ABCB8 mediates doxorubicin resistance in melanoma cells by protecting the mitochondrial genome. *Mol Cancer Res* 2009;**7**:79–87.
28. Chartrain M, Riond J, Stenvein A, Vandenberghe I, Gomes B, Lamant L, et al. Melanoma chemotherapy leads to the selection of ABCB5-expressing cells. *PLoS One* 2012;**7**:e36762.
29. Juliano RL, Ling V. A surface glycoprotein modulating drug permeability in Chinese hamster ovary cell mutants. *Biochim Biophys Acta* 1976;**455**:152–62.
30. Szakacs G, Paterson JK, Ludwig JA, Booth-Genthe C, Gottesman MM. Targeting multidrug resistance in cancer. *Nat Rev* 2006;**5**:219–34.
31. Thiebaut F, Tsuruo T, Hamada H, Gottesman MM, Pastan I, Willingham MC. Immunohistochemical localization in normal tissues of different epitopes in the multidrug transport protein P170: evidence for localization in brain capillaries and crossreactivity of one antibody with a muscle protein. *J Histochem Cytochem* 1989;**37**:159–64.
32. Albino AP, Nanus DM, Mentle IR, Cordon-Cardo C, McNutt NS, Bressler J, et al. Analysis of ras oncogenes in malignant melanoma and precursor lesions: correlation of point mutations with differentiation phenotype. *Oncogene* 1989;**4**:1363–74.
33. Rouzaud F, Hearing VJ. Regulatory elements of the melanocortin 1 receptor. *Peptides* 2005;**26**:1858–70.
34. Maellaro E, Pacenti L, Del Bello B, Valentini MA, Mangiavacchi P, de Felice C, et al. Different effects of interferon-alpha on melanoma cell lines: a study on telomerase reverse transcriptase, telomerase activity and apoptosis. *Br J Dermatol* 2003;**148**:1115–24.
35. Dunne BM, McNamara M, Clynes M, Shering SG, Larkin AM, Moran E, et al. MDR1 expression is associated with adverse survival in melanoma of the uveal tract. *Hum Pathol* 1998;**29**:594–8.
36. McNamara M, Clynes M, Dunne B, NicAmhloibh R, Lee WR, Barnes C, et al. Multidrug resistance in ocular melanoma. *Br J Ophthalmol* 1996;**80**:1009–12.
37. Keshet GI, Goldstein I, Itzhaki O, Cesarkas K, Shenhav L, Yakirevitch A, et al. MDR1 expression identifies human melanoma stem cells. *Biochem Biophys Res Commun* 2008;**368**:930–6.
38. Lincke CR, van der Blik AM, Schuurhuis GJ, van der Velde-Koerts T, Smit JJ, Borst P. Multidrug resistance phenotype of human BRO melanoma cells transfected with a wild-type human mdr1 complementary DNA. *Cancer Res* 1990;**50**:1779–85.
39. Doyle LA, Yang W, Abruzzo LV, Krogmann T, Gao Y, Rishi AK, et al. A multidrug resistance transporter from human MCF-7 breast cancer cells. *Proc Natl Acad Sci USA* 1998;**95**:15665–70.
40. Allikmets R, Schriml LM, Hutchinson A, Romano-Spica V, Dean M. A human placenta-specific ATP-binding cassette gene (ABCP) on chromosome 4q22 that is involved in multidrug resistance. *Cancer Res* 1998;**58**:5337–9.
41. McDevitt CA, Collins RF, Conway M, Modok S, Storm J, Kerr ID, et al. Purification and 3D structural analysis of oligomeric human multidrug transporter ABCG2. *Structure* 2006;**14**:1623–32.
42. Yoh K, Ishii G, Yokose T, Minegishi Y, Tsuta K, Goto K, et al. Breast cancer resistance protein impacts clinical outcome in platinum-based chemotherapy for advanced non-small cell lung cancer. *Clin Cancer Res* 2004;**10**:1691–7.
43. Wu CP, Hsieh CH, Wu YS. The emergence of drug transporter-mediated multidrug resistance to cancer chemotherapy. *Mol Pharm* 2011;**8**:1996–2011.
44. Dohse M, Scharenberg C, Shukla S, Robey RW, Volkman T, Deeken JF, et al. Comparison of ATP-binding cassette transporter interactions with the tyrosine kinase inhibitors imatinib, nilotinib, and dasatinib. *Drug Metab Dispos* 2010;**38**:1371–80.
45. Dai CL, Tiwari AK, Wu CP, Su XD, Wang SR, Liu DG, et al. Lapatinib (Tykerb, GW572016) reverses multidrug resistance in cancer cells by inhibiting the activity of ATP-binding cassette subfamily B member 1 and G member 2. *Cancer Res* 2008;**68**:7905–14.
46. Hori S, Ohtsuki S, Hosoya K, Nakashima E, Terasaki T. A pericyte-derived angiopoietin-1 multimeric complex induces occludin gene expression in brain capillary endothelial cells through Tie-2 activation *in vitro*. *J Neurochem* 2004;**89**:503–13.
47. Cooray HC, Blackmore CG, Maskell L, Barrand MA. Localisation of breast cancer resistance protein in microvessel endothelium of human brain. *Neuroreport* 2002;**13**:2059–63.
48. Bleau AM, Hambarzumyan D, Ozawa T, Fomchenko EI, Huse JT, Brennan CW, et al. PTEN/PI3K/Akt pathway regulates the side population phenotype and ABCG2 activity in glioma tumor stem-like cells. *Cell Stem Cell* 2009;**4**:226–35.
49. Ginguene C, Champier J, Maallem S, Strazielle N, Jouvett A, Fevre-Montange M, et al. P-glycoprotein (ABCB1) and breast cancer resistance protein (ABCG2) localize in the microvessels forming the blood-tumor barrier in ependymomas. *Brain Pathol* 2010;**20**:926–35.
50. Visvader JE, Lindeman GJ. Cancer stem cells in solid tumours: accumulating evidence and unresolved questions. *Nat Rev* 2008;**8**:755–68.
51. Dou J, Pan M, Wen P, Li Y, Tang Q, Chu L, et al. Isolation and identification of cancer stem-like cells from murine melanoma cell lines. *Cell Mol Immunol* 2007;**4**:467–72.

52. Grichnik JM, Burch JA, Schulteis RD, Shan S, Liu J, Darrow TL, et al. Melanoma, a tumor based on a mutant stem cell? *J Invest Dermatol* 2006;**126**:142–53.
53. Klein WM, Wu BP, Zhao S, Wu H, Klein-Szanto AJ, Tahan SR. Increased expression of stem cell markers in malignant melanoma. *Mod Pathol* 2007;**20**:102–7.
54. Woodward OM, Tukaye DN, Cui J, Greenwell P, Constantoulakis LM, Parker BS, et al. Gout-causing Q141K mutation in ABCG2 leads to instability of the nucleotide-binding domain and can be corrected with small molecules. *Proc Natl Acad Sci USA* 2013;**110**:5223–8.
55. Matsuo H, Takada T, Ichida K, Nakamura T, Nakayama A, Ikebuchi Y, et al. Common defects of ABCG2, a high-capacity urate exporter, cause gout: a function-based genetic analysis in a Japanese population. *Sci Transl Med* 2009;**1**:5ra11.
56. Berens W, van den Bossche K, Yoon TJ, Westbroek W, Valencia JC, Out CJ, et al. Different approaches for assaying melanosome transfer. *Pigment Cell Res* 2005;**18**:370–81.
57. Chen KG, Valencia JC, Lai B, Zhang G, Paterson JK, Rouzaud F, et al. Melanosomal sequestration of cytotoxic drugs contributes to the intractability of malignant melanomas. *Proc Natl Acad Sci USA* 2006;**103**:9903–7.
58. Frank NY, Margaryan A, Huang Y, Schatton T, Waaga-Gasser AM, Gasser M, et al. ABCB5-mediated doxorubicin transport and chemoresistance in human malignant melanoma. *Cancer Res* 2005;**65**:4320–33.
59. Johannessen CM, Boehm JS, Kim SY, Thomas SR, Wardwell L, Johnson LA, et al. COT drives resistance to RAF inhibition through MAP kinase pathway reactivation. *Nature* 2010;**468**:968–72.
60. de Grouw EP, Raaijmakers MH, Boezeman JB, van der Reijden BA, van de Locht LT, de Witte TJ, et al. Preferential expression of a high number of ATP binding cassette transporters in both normal and leukemic CD34⁺CD38⁻ cells. *Leukemia* 2006;**20**:750–4.
61. Sigalotti L, Covre A, Zabierowski S, Himes B, Colizzi F, Natali PG, et al. Cancer testis antigens in human melanoma stem cells: expression, distribution, and methylation status. *J Cell Physiol* 2008;**215**:287–91.
62. Schatton T, Murphy GF, Frank NY, Yamaura K, Waaga-Gasser AM, Gasser M, et al. Identification of cells initiating human melanomas. *Nature* 2008;**451**:345–9.
63. Johnson JD, Young B. Demographics of brain metastasis. *Neurosurg Clin N Am* 1996;**7**:337–44.
64. Fife KM, Colman MH, Stevens GN, Firth IC, Moon D, Shannon KF, et al. Determinants of outcome in melanoma patients with cerebral metastases. *J Clin Oncol* 2004;**22**:1293–300.
65. Atkins MB, Lotze MT, Dutcher JP, Fisher RI, Weiss G, Margolin K, et al. High-dose recombinant interleukin 2 therapy for patients with metastatic melanoma: analysis of 270 patients treated between 1985 and 1993. *J Clin Oncol* 1999;**17**:2105–16.
66. Chanda P, Yuhki N, Li M, Bader JS, Hartz A, Boerwinkle E, et al. Comprehensive evaluation of imputation performance in African Americans. *J Hum Genet* 2012;**57**:411–21.
67. Mohammed RL, Echeverry A, Stinson CM, Green M, Bonilla TD, Hartz A, et al. Survival trends of *Staphylococcus aureus*, *Pseudomonas aeruginosa*, and *Clostridium perfringens* in a sandy South Florida beach. *Mar Pollut Bull* 2012;**64**:1201–9.
68. Durmus S, Sparidans RW, Wagenaar E, Beijnen JH, Schinkel AH. Oral availability and brain penetration of the B-RAFV600E inhibitor vemurafenib can be enhanced by the P-glycoprotein (ABCB1) and breast cancer resistance protein (ABCG2) inhibitor elacridar. *Mol Pharm* 2012;**9**:3236–45.
69. Hartz AJ, Sherr BF, Sherr EB. Photoresponse in the heterotrophic marine dinoflagellate *Oxyrrhis marina*. *J Eukaryot Microbiol* 2011;**58**:171–7.
70. O'Neill L, Hartz AJ. Lower mortality rates at cardiac specialty hospitals traceable to healthier patients and to doctors' performing more procedures. *Health Aff (Millwood)* 2012;**31**:806–15.
71. Nazarian R, Shi H, Wang Q, Kong X, Koya RC, Lee H, et al. Melanomas acquire resistance to B-RAF(V600E) inhibition by RTK or N-RAS upregulation. *Nature* 2010;**468**:973–7.
72. Smith FO, Klapper JA, Wunderlich JR, Rosenberg SA, Dudley ME. Impact of a recombinant fowlpox vaccine on the efficacy of adoptive cell therapy with tumor infiltrating lymphocytes in a patient with metastatic melanoma. *J Immunother* 2009;**32**:870–4.
73. Valencia JC, Watabe H, Chi A, Rouzaud F, Chen KG, Vieira WD, et al. Sorting of Pmel17 to melanosomes through the plasma membrane by API and AP2: evidence for the polarized nature of melanocytes. *J Cell Sci* 2006;**119**:1080–91.
74. Hartz AM, Bauer B, Soldner EL, Wolf A, Boy S, Backhaus R, et al. Amyloid-beta contributes to blood-brain barrier leakage in transgenic human amyloid precursor protein mice and in humans with cerebral amyloid angiopathy. *Stroke* 2012;**43**:514–23.
75. Falchook GS, Long GV, Kurzrock R, Kim KB, Arkenau TH, Brown MP, et al. Dabrafenib in patients with melanoma, untreated brain metastases, and other solid tumours: a phase 1 dose-escalation trial. *Lancet* 2012;**379**:1893–901.
76. Ascierto PA, Gogas HJ, Grob JJ, Algarra SM, Mohr P, Hansson J, et al. Adjuvant interferon alfa in malignant melanoma: an interdisciplinary and multinational expert review. *Crit Rev Oncol Hematol* 2013;**85**:149–61.
77. Flach EH, Rebecca VW, Herlyn M, Smalley KS, Anderson AR. Fibroblasts contribute to melanoma tumor growth and drug resistance. *Mol Pharm* 2011;**8**:2039–49.
78. Carlino MS, Saunders CA, Haydu LE, Menzies AM, Martin Jr. CC, Lebowitz PF, et al. (18)F-labelled fluorodeoxyglucose-positron emission tomography (FDG-PET) heterogeneity of response is prognostic in dabrafenib treated BRAF mutant metastatic melanoma. *Eur J Cancer* 2013;**49**:395–402.
79. Wilmott JS, Tembe V, Howle JR, Sharma R, Thompson JF, Rizos H, et al. Intratumoral molecular heterogeneity in a BRAF-mutant, BRAF inhibitor-resistant melanoma: a case illustrating the challenges for personalized medicine. *Mol Cancer Ther* 2012;**11**:2704–8.
80. Mittapalli RK, Vaidhyanathan S, Dudek AZ, Elmquist WF. Mechanisms limiting distribution of the threonine-protein kinase B-RaFV600E inhibitor dabrafenib to the brain: implications for the treatment of melanoma brain metastases. *J Pharmacol Exp Ther* 2013;**344**:655–64.
81. Wilhelm SM, Carter C, Tang L, Wilkie D, McNabola A, Rong H, et al. BAY 43-9006 exhibits broad spectrum oral antitumor activity and targets the RAF/MEK/ERK pathway and receptor tyrosine kinases involved in tumor progression and angiogenesis. *Cancer Res* 2004;**64**:7099–109.
82. Flaherty KT, Schiller J, Schuchter LM, Liu G, Tuveson DA, Redlinger M, et al. A phase I trial of the oral, multikinase inhibitor sorafenib in combination with carboplatin and paclitaxel. *Clin Cancer Res* 2008;**14**:4836–42.
83. Abraham J, Edgerly M, Wilson R, Chen C, Rutt A, Bakke S, et al. A phase I study of the P-glycoprotein antagonist tariquidar in combination with vinorelbine. *Clin Cancer Res* 2009;**15**:3574–82.
84. Lagas JS, Fan L, Wagenaar E, Vlaming ML, van Tellingen O, Beijnen JH, et al. P-glycoprotein (P-gp/Abcb1), Abcc2, and Abcc3 determine the pharmacokinetics of etoposide. *Clin Cancer Res* 2010;**16**:130–40.
85. Agarwal S, Hartz AM, Elmquist WF, Bauer B. Breast cancer resistance protein and P-glycoprotein in brain cancer: two gatekeepers team up. *Curr Pharm Des* 2011;**17**:2793–802.
86. Hyafil F, Vergely C, Vignaud DP, Grand-Perret T. *In vitro* and *in vivo* reversal of multidrug resistance by GF120918, an acridonecarboxamide derivative. *Cancer Res* 1993;**53**:4595–602.
87. Vispute SG, Chen JJ, Sun YL, Sodani KS, Singh S, Pan Y, et al. Vemurafenib (PL4032, Zelboraf[®]), a BRAF inhibitor, modulates ABCB1-, ABCG2-, and ABC10-mediated multidrug resistance. *J Cancer Res Updates* 2013;**2**:306–17.