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Research Article

Further Studies on Arcanobacterium phocisimile: a Novel Species of Genus Arcanobacterium

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Arcanobacterium phocisimile, a newly described species with the type strain A. phocisimile 2698^T isolated from a vaginal swab of a harbour seal and four additional A. phocisimile strains also isolated from four harbour seals could reliably be identified by phenotypic properties, by matrix assisted laser desorption ionization time of flight mass spectrometry (MALDI-TOF MS), and by sequencing the genomic targets 16S rDNA and 16S-23S rDNA intergenic spacer region and the genes rpoB and gap. The A. phocisimile strains investigated in the present study were isolated together with several other bacterial species indicating that the pathogenic importance of A. phocisimile remains unclear. However, the detection of peptidic spectra by MALDI-TOF MS and the presented phenotypic and genotypic approach might help to identify A. phocisimile in future.

1. Introduction

Genus Arcanobacterium comprises the species Arcanobacterium haemolyticum, Arcanobacterium hippocoleae, Arcanobacterium pluranimalium, and Arcanobacterium phocae [1]. More recently A. canis and A. phocisimile were described as novel species of this genus [2, 3]. Arcanobacterium pyogenes together with Arcanobacterium bernardiae, Arcanobacterium bonasi, and Arcanobacterium bialowiezense was reclassified to the newly described species Trueperella [1]. The original description of A. phocisimile was based on physiological and biochemical characteristics, chemotaxonomic analysis, and 16S rDNA sequencing results of two strains isolated with several other bacterial species from a vaginal swab and an anal swab of two free living harbour seals of the German North Sea [3].

In the present study both initially described *A. phocisimile* strains and three additional strains obtained from three harbour seals were identified and further characterized phenotypically by MALDI-TOF MS analysis and genotypically by amplification and sequencing of various molecular targets.

2. Materials and Methods

2.1. Bacterial Strains. The A. phocisimile strains used in the present study included the previously described type strains A. phocisimile 2698^{T} (LMG 27073^{T} ; CCM 8430^{T}) and A. phocisimile 4112 [3]. Additionally investigated A. phocisimile 3047 was isolated (post mortem) together with Bacillus spp., Enterococcus spp., Erysipelothrix rhusiopathiae, and α -haemolytic streptococci in the year 2005 from the lung of a female harbour seal with bronchopneumonia and perforation

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[7]

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(10) C3130R

(rpoB primer)

| Oligonucleotide primers | Sequence | Program* | Expected size of PCR product (bp) | References |
|---|--|----------|-----------------------------------|------------|
| (1) 16S rDNA UNI-L (2) 16S rDNA UNI-R (amplification primer) | 5'-AGAGTTTGATCATGGCTCAG-3' 5'-GTGTGACGGGCGGTGTGTAC-3' | 1 | 1,403 | [4] |
| (3) 16S rDNA-533F (4) 16S rDNA-907R (sequencing primer) | 5'-GTGCCAGCMGCCGCGGTAA-3' 5'-CCGTCAATTCMTTTGAGTTT-3' | _ | _ | [4] |
| (5) Gap-F (6) Gap-R (gap primer) | 5'-TCGAAGTTGTTGCAGTTAACGA-3' 5'-CCATTCGTTGTCGTACCAAG-3' | 2 | 830 | [4] |
| (7) ISR-23S-F (8) ISR-23S-R (intergenic spacer region primer) | 5'-CCTAGCCTGGTGGTTGGGTAG-3' 5'-GTGCGGGTAACCAGAAATAACTCTG- 3' | 3 | 345 | [6] |
| (9) C2700F | 5'-CGWATGAACATYGGBCAGGT-3' | | | |

Table 1: Oligonucleotide primer sequences and PCR conditions of the target genes used in the present study.

5'-TCCATYTCRCCRAARCGCTG-3'

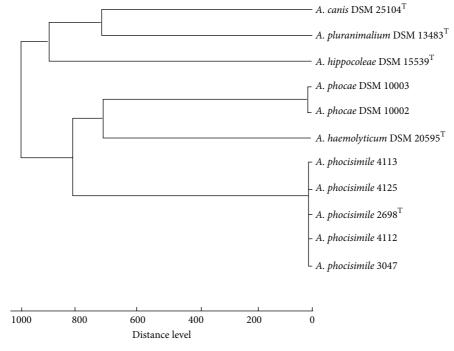


FIGURE 1: MALDI-TOF MS spectra from A. phocisimile 4113, A. phocisimile 4125, type strain A. phocisimile 2698^T, A. phocisimile 4112, A. phocisimile 3047, and all other species of genus Arcanobacterium.

of stomach. The harbour seal was found dead in Rantum, Sylt of the German North Sea. Also investigated *A. phocisimile* 4113 was recovered together with *Pasteurella* spp. and α -haemolytic streptococci from an anal swab and *A. phocisimile* 4125 together with *Pseudomonas* spp. and α -haemolytic streptococci also from an anal swab of two apparently healthy female harbour seals, respectively. *A. phocisimile* 4113 and *A. phocisimile* 4125 were isolated in 2007 during a monitoring program of free living harbour seals of the German North Sea.

2.2. Phenotypic and Genotypic Identification. All three newly investigated A. phocisimile strains were initially characterized phenotypically and by 16S rDNA sequencing [3, 4]. Both A. phocisimile strains previously mentioned in the species description [3] and the three A. phocisimile strains of the present study were further analysed by MALDI-TOF MS [5] and genotypically by amplification and sequencing of the previously described molecular target 16S-23S rDNA intergenic spacer region (ISR) and the genes rpoB and gap [4, 6, 7].

^{*}PCR program 1: x1 (95°C, 600 sec), x30 (95°C, 30 sec, 58°C, 60 sec, 72°C, 60 sec), and x1 (72°C, 420 sec). 2: x1 (94°C, 180 sec), x30 (94°C, 30 sec, 50°C, 40 sec, 72°C, 60 sec), and x1 (72°C, 420 sec). 3: x1 (95°C, 240 sec), x30 (95°C, 8 sec, 66°C, 10 sec, 72°C, 10 sec), x1 (72°C, 420 sec). 4: x1 (95°C, 600 sec), x35 (94°C, 30 sec, 50°C, 30 sec, 72°C, 120 sec), and x1 (72°C, 600 sec).

Table 2: Phenotypical properties of three *A. phocisimile* strains investigated in the present study and *A. phocisimile* 4112 and *A. phocisimile* 2698^T described previously.

| Phenotypic properties | 3047 | 4113 | 4125 | 4112** | 2698 ^{T**} |
|--|-----------|------------------|-----------|--------------|---------------------|
| Hemolysis on sheep blood agar | + | + | + | + | + |
| Hemolysis on rabbit blood agar | + | + | + | + | + |
| CAMP-like reaction with:* | | | | | |
| <i>Staphylococcus aureus</i> β -hemolysin | _ | _ | _ | _ | _ |
| Streptococcus agalactiae | + | + | + | + | + |
| Rhodococcus equi | + | + | + | + | + |
| Arcanobacterium haemolyticum | _ | _ | _ | _ | _ |
| Reverse CAMP reaction | + | + | + | + | + |
| Nitrate reduction | _1 | $-^{1}$ | $-^{1}$ | _1 | - ¹ |
| Pyrazinamidase | $(+)^{1}$ | $(+)^1$ | $+^1$ | $+^1$ | $+^1$ |
| Pyrrolidonyl arylamidase | _1 | _1 | $(+)^{1}$ | _1 | _1 |
| Alkaline phosphatase | _1 | _1 | $-^{1}$ | $(+)^{1}$ | $(+)^{1}$ |
| β -Glucuronidase (β -GUR) | _1,3 | - ^{1,3} | $-^{1,3}$ | $-^{1,3}$ | - ^{1,3} |
| β -Galactosidase (β -GAL) | +1,3 | +1,3 | +1,3 | +1,3 | +1,3 |
| α -Glucosidase (α -GLU) | +1,2,3 | +1,2,3 | +1,2,3 | +1,2,3 | +1,2,3 |
| β -Glucosidase (β -GLU) | _2 | _2 | _2 | _2 | _2 |
| N-Acetyl- β -glucosaminidase (β -NAG) | -1, +3 | -1, +3 | -1, +3 | $-1^{1}, +3$ | -1, +3 |
| Esculin (β -glucosidase) | _1 | _1 | _1 | _1 | _1 |
| Urease | _1 | _1 | +1 | _1 | _1 |
| Gelatine | _1 | _1 | _1 | _1 | _1 |
| Fermentation of: | | | | | |
| D-Glucose | +1 | $+^1$ | $+^1$ | +1 | +1 |
| D-Ribose | _1 | $+^1$ | $+^1$ | +1 | +1 |
| D-Xylose | _1 | _1 | _1 | _1 | _1 |
| D-Mannitol | _1 | _1 | _1 | _1 | _1 |
| D-Maltose | +1 | $+^1$ | $+^1$ | +1 | $+^1$ |
| D-Lactose | +1 | $+^1$ | +1 | +1 | +1 |
| D-Saccharose | +1 | $+^1$ | $+^1$ | +1 | +1 |
| Glycogen | +1 | $+^1$ | $+^1$ | +1 | +1 |
| α-Mannosidase | +2 | +2 | +2 | +2 | +2 |
| Catalase | + | + | + | + | + |
| Serolysis on Loeffler agar | _ | _ | _ | _ | _ |
| Caseinase | _ | _ | _ | _ | _ |
| Starch hydrolysis | + | + | + | + | + |

The reactions are shown as follows: *synergistic CAMP-like reaction with indicator strains; **results mostly obtained from Hijazin et al., 2013 [3]; +: positive reaction; (+): weak positive reaction; -: negative reaction; 1: Api Coryne test system (Biomerieux, Nürtingen, Germany); 2: tablets containing substrates (Rosco Diagnostica A/S, Taastrup, Denmark); 3: 4-methylumbelliferyl conjugated substrates (Sigma, Steinheim, Germany).

The primer sequences and the thermocycler programs are given in Table 1.

3. Results and Discussion

All three strains newly characterized in the present study could reliably be identified as *A. phocisimile* by phenotypic properties and by 16S rDNA sequencing. The phenotypic properties appeared to be almost identical to both previously characterized *A. phocisimile* strains (Table 2). However, a positive pyrazinamidase reaction of *A. phocisimile* seems to be the only reliable biochemical property for differentiation of *A. phocisimile* from pyrazinamidase negative *A. phocae*.

As shown by numerous authors MALDI-TOF MS is a powerful tool for species characterization of a broad spectrum of gram-positive and gram-negative bacteria [8–10]. This technique had previously been successfully used for rapid and reliable identification of bacteria of genera *Arcanobacterium* and *Trueperella* [5, 11]. The MALDI-TOF MS analysis of the present study revealed that by using the current Bruker data base, all five strains of this hitherto unknown species could not be identified to species level. However, using the MALDI Biotyper 3.1 software package the log (score) values of *A. phocisimile* 4112, *A. phocisimile* 3047, *A. phocisimile* 4113, and *A. phocisimile* 4125 matched against *A. phocisimile* 2698^T with log (score) values between 2.69

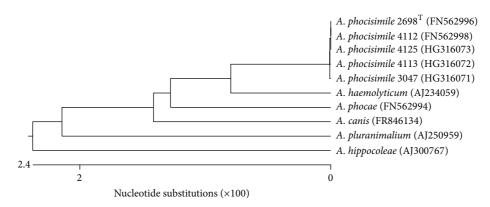


FIGURE 2: Dendrogram analysis of 16S rDNA sequences of the *A. phocisimile* strains of the present study and reference strains of genus *Arcanobacterium* obtained from NCBI GenBank.

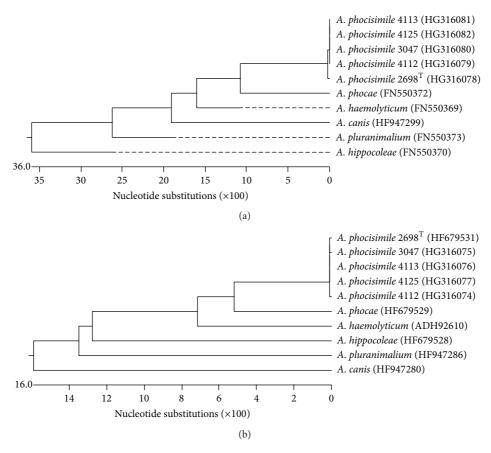


FIGURE 3: Dendrogram analysis of sequences of the genes *rpoB* (a) and *gap* (b) of the *A. phocisimile* strains of the present study and all other species of genus *Arcanobacterium* obtained from NCBI GenBank.

and 2.74 indicating that all five strains belong to this newly described species. Inclusion of *A. phocisimile* in the Bruker reference database will allow for the identification of this new species in future. A dendrogram analysis of the MALDI-TOF MS results is presented in Figure 1.

The genotypic classification by 16S rDNA sequencing revealed that the three novel *A. phocisimile* strains of the present study yielded 100% identity to both *A. phocisimile* strains described previously [3], also including the type strain *A. phocisimile* 2698^T (Figure 2).

Comparable to previously described *A. canis* [11] all five *A. phocisimile* from the present study could additionally be classified by amplification and sequencing of ISR (FN563000, FN563002, HG316083, HG316084, and HG316085), gene *rpoB* (HG316078, HG316079, HG316080, HG316081, and HG316082), and gene *gap* (HF679531, HG316074, HG316075, HG316076, and HG316077) yielding for all three molecular targets an identity of \geq 99.4%, \geq 99.8%, and \geq 99.8%, respectively, for all five strains among each other. A typical dendrogram using the sequencing results of the target genes *rpoB* and *gap* is shown in Figure 3.

The results of the present study revealed that phenotypic properties, the determination of peptidic spectra by MALDITOF MS, and the various genotypic targets allow for a reliable identification of *A. phocisimile* and a further differentiation of *A. phocisimile* from closely related *A. phocae* which could also be isolated from marine mammals [12]. However, all *A. phocisimile* strains of the present study were isolated together with various other bacteria, partly from obviously healthy animals, indicating that the pathogenic importance of this species for marine mammals remains unclear.

4. Interpretive Summary

Arcanobacterium phocisimile type strain and four additional A. phocisimile strains isolated from harbour seals were identified phenotypically, by matrix assisted laser desorption ionization time of flight mass spectrometry (MALDI-TOF MS), by sequencing 16S rDNA, and, as novel molecular targets, by sequencing 16S-23S rDNA intergenic spacer region and the genes *rpoB* and *gap* indicating that MALDI TOF MS and the molecular targets might help to identify this novel species.

Conflict of Interests

The authors declare that they have no conflict of interests. The authors certify that they have no affiliation with or financial involvement in any organization or entity with a direct financial interest in the subject matter or materials discussed in the paper.

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