



Published in final edited form as:

*Transplantation*. 2015 September ; 99(9): 1753. doi:10.1097/TP.0000000000000810.

## Perfusion Decellularization of Discarded Human Kidneys: A Valuable Platform for Organ Regeneration

Harald C Ott, MD

Department of Surgery, Division of Thoracic Surgery, Massachusetts General Hospital, Harvard Medical School

In this issue of *Transplantation* Andrea Peloso and colleagues report on the successful application of perfusion decellularization in discarded human kidneys to generate scaffolds for organ regeneration. (1) The authors provide important insight into mechanical integrity and morphological features of the native vascular tree and examine matrix bound growth factors after perfusion decellularization of human kidneys. Both of these are fundamental scaffold properties that enable cell engraftment, tissue formation, and perfusion during culture and after transplantation.

The enthusiasm to build functional, transplantable kidney grafts from patient derived cells, is fueled by recent progress in the fields of kidney stem cell and developmental biology.[2] Through directed differentiation, human pluripotent stem cells have been successfully guided through key developmental milestones to generate cellular constituents of metanephric mesoderm, and ureteric bud.[3–5] Once generated in sufficient numbers and purity, these cells could be a nearly inexhaustible source of building blocks to regenerate autologous kidney tissue and possibly transplantable grafts.[6] As we begin to decipher pathways of both kidney development and adult kidney repair and regeneration, we will hopefully learn to control the process of tissue formation and eventually achieve graft homeostasis in regenerated or bioartificial constructs.[7–9]

One possible approach to organ engineering follows the principle of tissue engineering, in which an extracellular matrix scaffold provides the three dimensional foundation of structural boundaries, architecture, vasculature, and the cell specific niches for different endothelial, mesenchymal, and epithelial cell phenotypes.[10] Despite recent advances in additive manufacturing and other techniques, deriving scaffolds of comparable composition, scale, and complexity of mature human organs has been a challenge. Since the first report of detergent based perfusion decellularization of mammalian organs, and its use to engineer functional tissue in 2008,[11] this technology has been successfully applied to multiple organ systems including the heart,[12] lung,[13–19] kidney,[20–25] liver,[26] and pancreas. [27, 28] Detailed matrix analysis of some organ systems has been performed,[19, 29] but kidney scaffolds of human origin have yet to be fully examined. The present article provides valuable data points on composition and physical properties, and further underlines the translational value of this platform for kidney bioengineering. Future work comparing

composition, growth factor levels, and mechanical properties to native developing and adult kidneys similar will be helpful to even better understand which components are preserved, and which are lost during decellularization. This will guide us in developing strategies to optimize kidney scaffolds to accommodate the myriad of different cell types required for mature organ function.

## References

1. Peloso A, et al. Renal ECM scaffolds from discarded kidneys maintain glomerular morphometry and vascular resilience, and retains critical growth factors. *Transplantation*. In press – this issue.
2. Little MH, McMahon AP. Mammalian kidney development: principles, progress, and projections. *Cold Spring Harb Perspect Biol*. 2012; 4(5)
3. Takasato M, et al. Directing human embryonic stem cell differentiation towards a renal lineage generates a self-organizing kidney. *Nat Cell Biol*. 2014; 16(1):118–26. [PubMed: 24335651]
4. Xia Y, et al. Directed differentiation of human pluripotent cells to ureteric bud kidney progenitor-like cells. *Nat Cell Biol*. 2013; 15(12):1507–15. [PubMed: 24240476]
5. Taguchi A, et al. Redefining the in vivo origin of metanephric nephron progenitors enables generation of complex kidney structures from pluripotent stem cells. *Cell Stem Cell*. 2014; 14(1): 53–67. [PubMed: 24332837]
6. Lam AQ, Bonventre JV. Regenerating the nephron with human pluripotent stem cells. *Curr Opin Organ Transplant*. 2015
7. Kramann R, et al. Perivascular Gli1+ progenitors are key contributors to injury-induced organ fibrosis. *Cell Stem Cell*. 2015; 16(1):51–66. [PubMed: 25465115]
8. Diep CQ, et al. Identification of adult nephron progenitors capable of kidney regeneration in zebrafish. *Nature*. 2011; 470(7332):95–100. [PubMed: 21270795]
9. Uzarski JS, et al. New strategies in kidney regeneration and tissue engineering. *Curr Opin Nephrol Hypertens*. 2014; 23(4):399–405. [PubMed: 24848937]
10. Vacanti JP, Langer R. Tissue engineering: the design and fabrication of living replacement devices for surgical reconstruction and transplantation. *Lancet*. 1999; 354(Suppl 1):SI32–4. [PubMed: 10437854]
11. Ott HC, et al. Perfusion-decellularized matrix: using nature’s platform to engineer a bioartificial heart. *Nat Med*. 2008; 14(2):213–21. [PubMed: 18193059]
12. Guyette JP, et al. Perfusion decellularization of whole organs. *Nat Protoc*. 9(6):2014. 1451–68.
13. Ott HC, et al. Regeneration and orthotopic transplantation of a bioartificial lung. *Nat Med*. 2010; 16(8):927–33. [PubMed: 20628374]
14. Petersen TH, et al. Tissue-engineered lungs for in vivo implantation. *Science*. 2010; 329(5991): 538–41. [PubMed: 20576850]
15. Price AP, et al. Development of a Decellularized Lung Bioreactor System for Bioengineering the Lung: The Matrix Reloaded. *Tissue Eng Part A*. 2010
16. Wallis JM, et al. Comparative assessment of detergent-based protocols for mouse lung decellularization and re-cellularization. *Tissue Eng Part C Methods*. 2012; 18(6):420–32. [PubMed: 22165818]
17. Nichols JE, et al. Production and Assessment of Decellularized Pig and Human Lung Scaffolds. *Tissue Eng Part A*. 2013; 19(17–18):2045–62. [PubMed: 23638920]
18. Gilpin SE, et al. Perfusion decellularization of human and porcine lungs: Bringing the matrix to clinical scale. *J Heart Lung Transplant*. 2013
19. Gilpin SE, et al. Up-Scaling Decellularization and Whole Organ Culture for Human Lung Regeneration. *The Journal of heart and lung transplantation*. 2013; 32(4):S69–S70.
20. Orlando G, et al. Production and implantation of renal extracellular matrix scaffolds from porcine kidneys as a platform for renal bioengineering investigations. *Ann Surg*. 2012; 256(2):363–70. [PubMed: 22691371]

21. Song JJ, et al. Regeneration and experimental orthotopic transplantation of a bioengineered kidney. *Nat Med.* 2013; 19(5):646–651. [PubMed: 23584091]
22. Ross EA, et al. Embryonic Stem Cells Proliferate and Differentiate when Seeded into Kidney Scaffolds. *Journal of the American Society of Nephrology.* 2009; 20(11):2338–2347. [PubMed: 19729441]
23. Nakayama KH, et al. Decellularized rhesus monkey kidney as a three-dimensional scaffold for renal tissue engineering. *Tissue engineering Part A.* 2010; 16(7):2207–16. [PubMed: 20156112]
24. Bonandrini B, et al. Recellularization of well-preserved acellular kidney scaffold using embryonic stem cells. *Tissue Eng Part A.* 2014; 20(9–10):1486–98. [PubMed: 24320825]
25. Caralt M, et al. Optimization and critical evaluation of decellularization strategies to develop renal extracellular matrix scaffolds as biological templates for organ engineering and transplantation. *Am J Transplant.* 2015; 15(1):64–75. [PubMed: 25403742]
26. Uygun BE, et al. Organ reengineering through development of a transplantable recellularized liver graft using decellularized liver matrix. *Nat Med.* 2010; 16(7):814–820. [PubMed: 20543851]
27. Goh SK, et al. Perfusion-decellularized pancreas as a natural 3D scaffold for pancreatic tissue and whole organ engineering. *Biomaterials.* 2013; 34(28):6760–72. [PubMed: 23787110]
28. Claudius Conrad M PhD, Schuetz Christian MD, Clippinger Benjamin MS, Vacanti Joseph P MD, FACS, Markmann James F MD PhD, Ott Harald C MD. Bio-engineered endocrine pancreas based on decellularized pancreatic matrix and mesenchymal stem cell/islet cell coculture. 2010
29. Hill RC, et al. Quantification of Extracellular Matrix Proteins from a Rat Lung Scaffold to Provide a Molecular Readout for Tissue Engineering. *Mol Cell Proteomics.* 2015