

# Draft Genome Sequence of Enteropathogenic *Escherichia coli*, Isolated from the Bloody Stool Sample of a Common Marmoset (*Callithrix jacchus*)

Nobuhito Hayashimoto,<sup>a</sup> Hanako Morita,<sup>a</sup> Takashi Inoue,<sup>b</sup> Masahiko Yasuda,<sup>c</sup> Masafumi Yamamoto,<sup>a</sup> Toshio Itoh<sup>b</sup>

ICLAS Monitoring Center, Central Institute for Experimental Animals, Kanagawa, Japan<sup>a</sup>; Marmoset Research Department, Central Institute for Experimental Animals, Kanagawa, Japan<sup>b</sup>; Pathological Analysis Center, Central Institute for Experimental Animals, Kanagawa, Japan<sup>c</sup>

**Here, we report the draft genome sequence of *Escherichia coli* strain R811. This bacterium was isolated from the bloody stool sample of a common marmoset, and was categorized as enteropathogenic *E. coli* because it possessed *eae*.**

Received 20 August 2015 Accepted 25 August 2015 Published 8 October 2015

**Citation** Hayashimoto N, Morita H, Inoue T, Yasuda M, Yamamoto M, Itoh T. 2015. Draft genome sequence of enteropathogenic *Escherichia coli*, isolated from the bloody stool sample of a common marmoset (*Callithrix jacchus*). *Genome Announc* 3(5):e01161-15. doi:10.1128/genomeA.01161-15.

**Copyright** © 2015 Hayashimoto et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to Nobuhito Hayashimoto, nobuh@ciea.or.jp.

Enteropathogenic *Escherichia coli* (EPEC) strains are important causative pathogens of infantile diarrhea in developing countries (1). EPEC strains are also enteropathogens in common marmosets (*Callithrix jacchus*). However, very few studies have investigated the pathogenicity and characterization of EPEC strains in common marmosets (2).

The main virulence factor of EPEC is the ability to produce attaching and effacing (A/E) lesions (3). A/E lesions are characterized by tight attachment of the bacteria to the enterocyte surface, effacement of brush border microvilli, and formation of an actin-rich pedestal beneath the attached bacteria (2). An outer membrane protein called intimin, encoded by *eae*, mediates intestinal cell attachment. The *eae* is currently used for the molecular diagnosis of EPEC. All of the genetic elements required for the A/E lesions are encoded on a large genomic pathogenicity island called the locus of enterocyte effacement (LEE). The LEE pathogenicity island plays an important role in bacterial adherence to host intestinal epithelial cells (3). In order to study the genetic features of EPEC strains, especially those involved in pathogenicity, we sequenced a representative strain, R811, which was derived from the bloody stool sample of a common marmoset. Strain R811 was categorized as EPEC based on the positive PCR amplification of *eae* (4) and the absence of *VT1*, *VT2*, *ST*, and *LT*, which were determined with commercially available PCR kits (PowerCheck Diarrheal *E. coli* 4-plex Detection Kit; Kogenebiotech Co., Seoul, Korea).

The draft genome of *Escherichia coli* R811 was obtained by using an Illumina HiSeq 2500 sequencer (Illumina, San Diego, CA, USA), with sequencing runs for paired-end sequences. The bacterial DNA libraries were prepared with a TrueSeq DNA sample prep kit (Illumina). The genome was assembled into 386 contigs, ranging in size from 189 to 346,100 bp, by using *de novo* sequence assembler software (Velvet, EMBL-EBI) (5). Gene prediction was performed with the Rapid Annotation using Subsystem Technology server (<http://rast.nmpdr.org>) (6) and the Microbial Genome Annotation Pipeline (<http://www.migap.org>) (7). Sequencing and read assembly of the libraries were carried out by Hokkaido System Science (Sapporo, Japan).

The draft genome sequence of *E. coli* R811 was 5,444,756 bp (5.4 Mb) with a G+C content of 50.5%. The sequenced strain encoded, on average, 5,525 protein-coding genes, 7 rRNA genes, and 97 tRNA genes. Our analysis revealed several genes related to pathogenicity, including intimin (*eaeA*), RTX toxin (*rtxA1*), hemolysin E (*ClyA*), and *Shigella* enterotoxin 2 (*shET2*). Except for intimin, the other virulence factors listed here were identified for the first time in the strain in this study.

**Nucleotide sequence accession numbers.** The draft genome sequences for the *E. coli* R811 strain have been deposited in DDBJ/EMBL/GenBank under the accession numbers [BBYP01000001](https://www.ncbi.nlm.nih.gov/nuccore/BBYP01000001) to [BBYP01000539](https://www.ncbi.nlm.nih.gov/nuccore/BBYP01000539). The version described in this paper is the first version.

## ACKNOWLEDGMENT

This study was partially supported by JSPS KAKENHI (grant no. 24500499).

## REFERENCES

- Ochoa TJ, Contreras CA. 2011. Enteropathogenic *Escherichia coli* infection in children. *Curr Opin Infect Dis* 24:478–483. <http://dx.doi.org/10.1097/QCO.0b013e32834a8b8b>.
- Thomson JA, Scheffler JJ. 1996. Hemorrhagic typhlocolitis associated with attaching and effacing *Escherichia coli* in common marmosets. *Lab Anim Sci* 46:275–279.
- Hartland EL, Leong JM. 2013. Enteropathogenic and enterohemorrhagic *E. coli*: ecology, pathogenesis, and evolution. *Front Cell Infect Microbiol* 3:15. <http://dx.doi.org/10.3389/fcimb.2013.00015>.
- Nguyen TD, Vo TT, Vu-Khac H. 2011. Virulence factors in *Escherichia coli* isolated from calves with diarrhea in Vietnam. *J Vet Sci* 12:159–164. <http://dx.doi.org/10.4142/jvs.2011.12.2.159>.
- Zerbino DR, Birney E. 2008. Velvet: algorithms for *de novo* short read assembly using de Bruijn graphs. *Genome Res* 18:821–829. <http://dx.doi.org/10.1101/gr.074492.107>.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. *BMC Genomics* 9:75. <http://dx.doi.org/10.1186/1471-2164-9-75>.
- Sugawara H, Ohyama A, Mori H, Kurokawa K. 2009. Microbial genome annotation pipeline (MiGAP) for diverse users, abstr. 20<sup>th</sup> International Conference on Genome Informatics, Kanagawa, Japan.