



Published in final edited form as:

Circ Res. 2015 October 9; 117(9): 744–746. doi:10.1161/CIRCRESAHA.115.307407.

Arteriosclerotic Calcification: A *Serpi(n)ginous Path To Cardiovascular Health?*

Dwight A. Towler, M.D., Ph.D.

Department of Internal Medicine | Endocrine Division, UT Southwestern Medical Center, Dallas, Texas 75390-8857

Keywords

Arteriosclerosis; Endothelial-Mesenchymal Transition; Matrix Gla Protein; Serine protease inhibitor; Serpin

Cardiovascular sclerosis increasingly afflicts our aging, dysmetabolic population – and this hardening of our hearts and arteries has significant physiological consequences¹. Myocardial stiffening reduces diastolic ventricular filling and function necessary for robust cardiac output during systole². Arterial stiffening impairs Windkessel physiology – the rubbery elasticity of conduit vessels that ensures smooth distal tissue perfusion throughout the cardiac cycle. Thus, in addition to the diastolic heart failure associated with cardiac sclerosis, a type of diastolic perfusion failure occurs with arteriosclerotic conduit vessel stiffening. The inability to store kinetic energy as potential energy in elastic conduit vessels during systole reduces the sustained pressure differential necessary to drive smooth distal perfusion throughout diastole -- and is manifested by increased arterial pulse wave velocity (PWV) during systole¹.

One clinical consequence of diastolic vascular perfusion failure can be well-appreciated in the central nervous system. In the Dallas Heart Study, increased aortic stiffness as quantified by PWV strongly portends increased brain MRI white matter hyperintensity volume³, a signature of ischemic (not hemorrhagic) histology⁴, independent of other cardiovascular risk factors including systolic blood pressure³. Cognitive decline is a clinical feature of conduit vessel stiffness^{5, 6} and white matter hyperintensity volume⁴. Large conduit artery biomechanics reflect the composite contributions of mural material properties, geometric properties, and dynamic endothelial and neuroendocrine inputs that control global and regional tissue perfusion⁷. Arterial calcification -- an active form of tissue biomineralization – has emerged as one important pathogenic feature of conduit vessel stiffening^{8, 9}. Over the past 2 decades, elegant work forthcoming from research teams at UCLA has identified that powerful signals provided by bone morphogenetic proteins (BMP) of the TGF-beta superfamily play critically important roles in arterial calcification¹⁰. Matrix Gla protein

Please address correspondence to: Dwight A. Towler MD, PhD, J.D. and Maggie E. Wilson Distinguished Chair in Biomedical Research, UT Southwestern Medical Center, Internal Medicine | Endocrine Division, 5323 Harry Hines Blvd, Dallas, TX 75390-8857, Dwight.Towler@utsouthwestern.edu, Fax: 214-648-8917.

Disclosures – D.A.T. previously consulted for Daiichi-Sankyo.

(MGP), a secreted calcium binding protein that inhibits BMP function in a Gla-(gamma-carboxyglutamate) dependent fashion, functions as a vascular co-morphogen and rate-limiting negative regulator of arterial mineral deposition in murine disease models^{11, 12}. *MGP*-null mice die precociously with pan-arterial calcification and aortic rupture¹³. MGP clearly impacts the BMP-directed osteochondrogenic programming; however, results from other groups have highlighted that the spectrum of MGP-regulated vascular actions relevant to arterial calcification may in fact extend beyond BMP modulation to encompass elastin matricrine signaling¹⁴ and osteogenic degradation products¹⁵. Since under-carboxylated MGP tracks arterial stiffness in humans¹⁶, a better understanding of MGP actions may yield novel therapeutic approaches to arteriosclerotic disease and its consequences.

In this issue of *Circulation Research*¹⁷, Yao, Boström, and colleagues once again advance our understanding of MGP actions in arteriosclerotic disease¹⁸. They identify that MGP serves to restrict expression and activities of vascular serine protease that promote the endothelial-mesenchymal transition (EndMT) – a key contributor to the cells and signals that drive arterial calcium deposition. In response to metabolic insult such as hyperglycemia, previous work from this group identified that MGP deficiency enabled endothelial cell (EC) phenotypic plasticity and subsequent osteogenic trans-differentiation, with concomitant induction of multiple Yamanaka factors including *Sox2*¹⁸. However, the regulatory circuits conveying this response were uncharacterized. Detailed vascular assessment by electron microscopy revealed early post-natal degradation of the remodeling arterial internal elastic lamina (IEL) in *MGP*^{-/-} mice with upregulation of Twist and *Slug/snai2* – key markers and mediators of the EndMT¹⁷. Implementing gene array analyses to interrogate for potential mediators, the team identified that mRNAs encoding a select cohort of five serine proteases -- *elastases* 1 and 2 and *kallikreins* 1, 5, and 6 – were markedly upregulated with MGP deficiency in aortic tissues. *In vitro*, treatment of human aortic ECs with this 5-protease protein cocktail upregulated the expression of key markers of multipotency – including *Sox2*¹⁷ -- along with the EndMT, thereby phenocopying the actions of MGP deficiency¹⁸. RNAi targeting either the protease pentad or *Sox2* abrogated osteogenic trans-differentiation of human aortic ECs induced by MGP knockdown. Importantly, BMP4- and glucose-induced EndMT was abrogated by administration of serpinA1 (alpha-1 antitrypsin) or diisopropylfluorophosphate – broad-specificity serine protease inhibitors – and this inhibitory action was fully reversed by transduction with a *Sox2*-expressing lentivirus. *In vivo*, EC-specific deletion of *Sox2* reduced arterial calcification in the global MGP-null background – and administration of serine protease inhibitors including serpinA1 reduced arterial calcification and delayed precocious cardiovascular death in *MGP*-null mice. Thus, the authors newly discover that a cadre of serine proteases participate in the EndMT and the *Sox2*-dependent phenotypic plasticity that drives arterial calcification in the absence of MGP – and demonstrate that serine protease inhibition limits arteriosclerotic disease and demise in this enlightening vasculopathy model¹⁷.

The precise proteases driving arterial calcification as responsive to serine protease inhibition *in vivo* have yet to be unambiguously identified, and future studies will undoubtedly focus upon this important aspect. However, it is intriguing to reflect upon the responses to recombinant serpinA1 in *MGP*^{-/-} mice and their significant implications. Firstly, serpin-

6. Zeki Al Hazzouri A, Newman AB, Simonsick E, Sink KM, Sutton Tyrrell K, Watson N, Satterfield S, Harris T, Yaffe K, Health ABCS. Pulse wave velocity and cognitive decline in elders: The health, aging, and body composition study. *Stroke*. 2013; 44:388–393. [PubMed: 23321445]
7. Greenwald SE. Ageing of the conduit arteries. *J Pathol*. 2007; 211:157–172. [PubMed: 17200940]
8. Cheng SL, Ramachandran B, Behrmann A, Shao JS, Mead M, Smith C, Krcchma K, Bello Arredondo Y, Kovacs A, Kapoor K, Brill LM, Perera R, Williams BO, Towler DA. Vascular smooth muscle Irf6 limits arteriosclerotic calcification in diabetic ldlr^{-/-} mice by restraining noncanonical wnt signals. *Circ Res*. 2015; 117:142–156. [PubMed: 26034040]
9. Cheng SL, Behrmann A, Shao JS, Ramachandran B, Krcchma K, Bello Arredondo Y, Kovacs A, Mead M, Maxson R, Towler DA. Targeted reduction of vascular msx1 and msx2 mitigates arteriosclerotic calcification and aortic stiffness in ldlr-deficient mice fed diabetogenic diets. *Diabetes*. 2014; 63:4326–4337. [PubMed: 25056439]
10. Demer LL, Tintut Y. Vascular calcification: Pathobiology of a multifaceted disease. *Circulation*. 2008; 117:2938–2948. [PubMed: 18519861]
11. Yao Y, Jumabay M, Wang A, Bostrom KI. Matrix gla protein deficiency causes arteriovenous malformations in mice. *J Clin Invest*. 2011; 121:2993–3004. [PubMed: 21765215]
12. Yao Y, Bennett BJ, Wang X, Rosenfeld ME, Giachelli C, Luscis AJ, Bostrom KI. Inhibition of bone morphogenetic proteins protects against atherosclerosis and vascular calcification. *Circ Res*. 2010; 107:485–494. [PubMed: 20576934]
13. Luo G, Ducy P, McKee MD, Pinero GJ, Loyer E, Behringer RR, Karsenty G. Spontaneous calcification of arteries and cartilage in mice lacking matrix gla protein. *Nature*. 1997; 386:78–81. [PubMed: 9052783]
14. Khavandgar Z, Roman H, Li J, Lee S, Vali H, Brinckmann J, Davis EC, Murshed M. Elastin haploinsufficiency impedes the progression of arterial calcification in mgp-deficient mice. *J Bone Miner Res*. 2014; 29:327–337. [PubMed: 23857752]
15. Sinha A, Vyavahare NR. High-glucose levels and elastin degradation products accelerate osteogenesis in vascular smooth muscle cells. *Diab Vasc Dis Res*. 2013; 10:410–419. [PubMed: 23754846]
16. Pivin E, Ponte B, Pruijm M, Ackermann D, Guessous I, Ehret G, Liu YP, Drummen NE, Knapen MH, Pechere-Bertschi A, Paccaud F, Mohaupt M, Vermeer C, Staessen JA, Vogt B, Martin PY, Burnier M, Bochud M. Inactive matrix gla-protein is associated with arterial stiffness in an adult population-based study. *Hypertension*. 2015; 66:85–92. [PubMed: 25987667]
17. Yao J, Guihard P, Blazquez-Medela AM, Guo Y, Moon JH, Jumabay M, Bostrom KI, Yao Y. Serine protease activation essential for endothelial-mesenchymal transition in vascular calcification. *Circ Res*. 2015; 117:xxx–xxx. [in this issue].
18. Yao Y, Jumabay M, Ly A, Radparvar M, Cubberly MR, Bostrom KI. A role for the endothelium in vascular calcification. *Circ Res*. 2013; 113:495–504. [PubMed: 23852538]
19. Stockley RA. The multiple facets of alpha-1-antitrypsin. *Ann Transl Med*. 2015; 3:130. [PubMed: 26207223]
20. Luo LY, Jiang W. Inhibition profiles of human tissue kallikreins by serine protease inhibitors. *Biol Chem*. 2006; 387:813–816. [PubMed: 16800745]
21. Lose F, Srinivasan S, O'Mara T, Marquart L, Chambers S, Gardiner RA, Aitken JF, Spurdle AB, Batra J, Clements JA. Australian Prostate Cancer B. Genetic association of the klk4 locus with risk of prostate cancer. *PLoS One*. 2012; 7:e44520. [PubMed: 22970239]
22. Inouye M, Ripatti S, Kettunen J, Lyytikainen LP, Oksala N, Laurila PP, Kangas AJ, Soinen P, Savolainen MJ, Viikari J, Kahonen M, Perola M, Salomaa V, Raitakari O, Lehtimäki T, Taskinen MR, Jarvelin MR, Ala-Korpela M, Palotie A, de Bakker PI. Novel loci for metabolic networks and multi-tissue expression studies reveal genes for atherosclerosis. *PLoS Genet*. 2012; 8:e1002907. [PubMed: 22916037]