



Incidental Syphilis Diagnosed by Real-Time PCR Screening of Urine Samples

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dramatic increase in the incidence of sexually transmitted diseases (STDs), such as gonorrhea (1), syphilis (1, 2), and HIV seroconversion (3, 4), has been reported worldwide. The gold standard for syphilis diagnosis remains the serology test, particularly the Venereal Disease Research Laboratory (VDRL) test or rapid plasma reagin (RPR) test (5, 6). Since August 2013, following the reports of increases in STDs (7, 8), every sample addressed for detection of any one of the following pathogens has been automatically screened for all of them: Chlamydia trachomatis, Neisseria gonorrhoeae, and Treponema pallidum. DNA extraction as well as quality evaluation was performed as previously described (9). The possible presence of inhibitors in the samples was also tested using primers targeting a synthetic inhibition control (9). Treponema pallidum real-time PCR (RT-PCR) was performed using a CFX96 device according to the manufacturer's recommendations (Bio-Rad Clinical Diagnostics, Marnes-la-Coquette, France) targeting the polA gene (Tpa_polA_F, GAGTGTGCAGT CCGCTATGC; Tpa_polA_R, AGGCAAAAGCGGCATTTCTA; Tpa_polA_P, TCCGCTTGGAAACAGCAGGATTG). A synthetic positive control of 381 bp from the polA gene produced with a pUC57 plasmid was used, as well as a negative control for each run. Confirmation PCR targeting the *flaA* gene was performed as previously described (10). Over a period of 13 months, 1,706 samples from 1,409 patients were received, of which 887 (52%) were urine samples. Seven of these samples from five patients, including four urine samples and three lesion swabs, were positive for syphilis. The treponemal load in samples from patient 2 was 2 to 3 orders of magnitude higher than those from other subjects (Table 1). Treponema pallidum PCR was not requested for two of the five

patients, making these syphilis diagnoses incidental. These last findings were confirmed by follow-up serology performed at 1 and 4 months, respectively. Finally, all patients had both a positive VDRL test and a positive *T. pallidum* hemagglutination (TPHA) test, except patient 2, for whom serology was not performed.

During the same period, 337 sera from 147 patients were positive for syphilis by serology, defined by both a positive VDRL test and a positive TPHA test. Only 36 patients had further PCR screening. Forty-eight specimens were collected, including 25 urine samples, 16 swabs, and 7 others. Syphilis RT-PCR sensitivity was estimated at 16% (4/25) in urine and at 18.75% (3/16) in swabs.

Molecular amplification of *Treponema pallidum* DNA by RT-PCR has been proven useful in the diagnosis of early syphilis, particularly before the serological response has fully developed (11). Although genital ulcers or lesion swabs seem to be the more appropriate samples (12, 13), the sensitivity of urine specimens was previously estimated at 29% in cases of primary syphilis and at 44% in cases of secondary-stage disease (14) and offers the benefit of being noninvasive. Our experience also suggests the value of

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	TABLE 1 Characteristics	of the 7 samples	positive for Treponem	<i>a pallidum</i> detection ^{<i>a</i>}
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Patient no.	Age (yr)		Sex Specimen	Initial query	Syphilis RT-PCR result (C_T)		No. of days	Serology result by test:	
		Sex			polA gene	<i>flaA</i> gene	between serology and PCR	VDRL titer	TPHA
1	33	М	Urine	C. trachomatis	Positive (34)	Positive (36)	-3	1/32	R
			Endobuccal swab	T. pallidum	Positive (28)	Positive (31)			
2	24	М	Pharynx swab	N. gonorrhoeae, C. trachomatis, T. pallidum	Positive (26)	Positive (28)			
			Urine	N. gonorrhoeae, C. trachomatis, T. pallidum	Positive (24)	Positive (25)			
3	43	Μ	Urine	N. gonorrhoeae	Positive (31)	Positive (30)	124	1/512	R
4	53	Μ	Urine	N. gonorrhoeae, C. trachomatis	Positive (35)	Positive (34)	31	1/32	R
			Rectal swab	N. gonorrhoeae, C. trachomatis	Negative	Negative			
5	60	М	Genital swab	N. gonorrhoeae, C. trachomatis, T. pallidum	Positive (32)	Positive (33)	0	1/8	R
			Urine	N. gonorrhoeae, C. trachomatis, T. pallidum	Negative	Negative			

^a Patients whose demographic characteristics are in bold are the two incidental diagnoses. Abbreviations: M, male; C₇₂ cycle threshold; R, reactive.

screening multiple anatomical sites (15), as shown by discordant multisite RT-PCR results (Table 1).

Finally, this report highlights the usefulness of RT-PCR for the detection of *Treponema pallidum* from noninvasive samples, such as urine, in the context of PCR testing for STDs. The systematic screening reduces the risk of missing a diagnosis for a disease that tends to be overlooked.

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REFERENCES

- 1. Savage E, Marsh K, Duffell S, Ison CA, Zaman A, Hughes G. 2011. Rapid increase in gonorrhoea and syphilis diagnoses in England in 2011. Euro Surveill 25:35–44. http://www.eurosurveillance.org /ViewArticle.aspx?ArticleId=20224.
- Simms I, Wallace L, Thomas DR, Emmett L, Shankar AG, Vinson M, Padfield S, Andrady U, Whiteside C, Williams CJ, Midgley C, Johnman C, McLellan A, Currie A, Logan J, Leslie G, Licence K, Hughes G. 2014. Recent outbreaks of infectious syphilis, United Kingdom, January 2012 to April 2014. Euro Surveill 19:24. http://www .eurosurveillance.org/ViewArticle.aspx?ArticleId=20833.
- Chow EPF, Wilson DP, Zhang J, Jing J, Zhang L. 2011. Human immunodeficiency virus prevalence is increasing among men who have sex with men in China: findings from a review and meta-analysis. Sex Transm Dis 38:845–857. http://dx.doi.org/10.1097/OLQ.0b013e31821a4f43.
- 4. Dukers NH, Spaargaren J, Geskus RB, Beijnen J, Coutinho RA, Fennema HS. 2002. HIV incidence on the increase among homosexual men attending an Amsterdam sexually transmitted disease clinic: using a novel approach for detecting recent infections. AIDS 16:F19–F24. http://dx.doi .org/10.1097/00002030-200207050-00001.
- 5. Larsen SA, Steiner BM, Rudolph AH. 1995. Laboratory diagnosis and interpretation of tests for syphilis. Clin Microbiol Rev 8:1–21.
- Janier M, Hegyi V, Dupin N, Unemo M, Tiplica GS, Potočnik M, French P, Patel R. 2014. 2014 European guideline on the management of syphilis. J Eur Acad Dermatol Venereol 28:1581–1593. http://dx.doi.org /10.1111/jdv.12734.
- 7. Colson P, Gouriet F, Badiaga S, Tamalet C, Stein A, Raoult D. 2013.

Real-time laboratory surveillance of sexually-transmissible infections in Marseille University hospitals reveals rise of gonorrhoea, syphilis and human immunodeficiency virus seroconversions in 2012. Euro Surveill 18:4. http://www.eurosurveillance.org/ViewArticle.aspx?ArticleId=20402.

- Dubourg G, Colson P, Tamalet C, Fournier PE, Raoult D. 2014. Increase in sexually transmitted infections during Europride 2013 in Marseille, France. Lancet Infect Dis 14:677–678. http://dx.doi.org/10.1016 /S1473-3099(14)70839-4.
- Morel A, Dubourg G, Edouard S, Prudent E, Fenollar F, Gouriet F, Casalta J, Fournier P, Drancourt M, Raoult D. 2015. Complementarity between targeted real-time specific PCR and conventional broad-range 16S rDNA PCR in the syndrome-driven diagnosis of infectious diseases. Eur J Clin Microbiol Infect Dis 34:561–570. http://dx.doi.org/10.1007 /s10096-014-2263-z.
- Salazar JC, Rathi A, Michael NL, Radolf JD, Jagodzinski LL. 2007. Assessment of the kinetics of *Treponema pallidum* dissemination into blood and tissues in experimental syphilis by real-time quantitative PCR. Infect Immun 75:2954–2958. http://dx.doi.org/10.1128/IAI.00090-07.
- Shields M, Guy RJ, Jeoffreys NJ, Finlayson RJ, Donovan B. 2012. A longitudinal evaluation of *Treponema pallidum* PCR testing in early syphilis. BMC Infect Dis 12:353. http://dx.doi.org/10.1186/1471-2334-12-353.
- Heymans R, Van der Helm JJ, De Vries HJC, Fennema HSA, Coutinho RA, Bruisten SM. 2010. Clinical value of *Treponema pallidum* real-time PCR for diagnosis of syphilis. J Clin Microbiol 48:497–502. http://dx.doi .org/10.1128/JCM.00720-09.
- Gayet-Ageron A, Ninet B, Toutous-Trellu L, Lautenschlager S, Furrer H, Piguet V, Schrenzel J, Hirschel B. 2009. Assessment of a real-time PCR test to diagnose syphilis from diverse biological samples. Sex Transm Infect 85:264–269. http://dx.doi.org/10.1136/sti.2008.034314.
- 14. Glatz M, Juricevic N, Altwegg M, Bruisten S, Komericki P, Lautenschlager S, Weber R, Bosshard PP. 2014. A multicenter prospective trial to asses a new real-time polymerase chain reaction for detection of *Treponema pallidum*, herpes simplex-1/2 and *Haemophilus ducreyi* in genital, anal and oropharyngeal ulcers. Clin Microbiol Infect **20**:O1020–O1027. http://dx.doi.org/10.1111/1469-0691.12710.
- Peters RP, Nijsten N, Mutsaers J, Jansen CL, Morré SA, van Leeuwen AP. 2011. Screening of oropharynx and anorectum increases prevalence of *Chlamydia trachomatis* and *Neisseria gonorrhoeae* infection in female STD clinic visitors. Sex Transm Dis 38:783–787. http://dx.doi .org/10.1097/OLQ.0b013e31821890e9.