• BASIC RESEARCH •

Role of nitric oxide and peroxynitrite anion in lung injury induced by intestinal ischemia-reperfusion in rats

Jun-Lin Zhou, Guo-Hua Jin, Yi-Ling Yi, Jun-Lan Zhang, Xin-Li Huang

Jun-Lin Zhou, Department of Hand Surgery, Third Affiliated Hospital, Hebei Medical University, Shijiazhuang 050051, Hebei Province, China

Guo-Hua Jin, Department of Liver Medicine, Third Affiliated Hospital, Hebei Medical University, Shijiazhuang 050051, Hebei Province, China

Yi-Ling Yi, Jun-Lan Zhang, Xin-Li Huang, Department of Pathophysiology, Hebei Medical University, Shijiazhuang 050017, Hebei Province, China

Correspondence to: Dr. Jun-Lin Zhou, Department of Hand Surgery, Third Affiliated Hospital, Hebei Medical University, Shijiazhuang 050051, Hebei Province, China. zhjunlin@yahoo.com **Telephone:** +86+311-7027951 Ext 3117

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Abstract

AIM: To evaluate effects of nitric oxide (NO) and peroxynitrite anion (ONOO⁻) on lung injury following intestinal ischemia-reperfusion (IR) in rats.

METHODS: A rat model of intestinal ischemia was made by clamping superior mesenteric artery and lung injury was resulted from reperfusion. The animals were randomly divided into 3 groups: sham operation (Sham), 2 h ischemia followed by 2 h reperfusion (IR) and IR pretreated with aminoguanidine (AG) - an inhibitor of inducible NO synthase (iNOS) 15 minutes before reperfusion (IR+AG). The lung malondialdehyde (MDA) and nitrate/nitrite (NO₂⁻/NO₃⁻) contents and morphological changes were examined. Western blot was used to detect the iNOS protein expression. Immunohistochemical staining was used to determine the change of nitrotyrosine (NT)- a specific "footprint" of ONOO⁻.

RESULTS: The morphology revealed evidence for lung edema, hemorrhage and polymorphonuclear sequestration after intestinal IR. Compared with sham group, lung contents of MDA and NO₂/NO₃ in IR group were significantly increased (12.00±2.18 vs 23.44±1.25 and 76.39±6.08 vs 140.40±4.34, P<0.01) and the positive signals of iNOS and NT were also increased in the lung. Compared with IR group, the contents of MDA and NO₂⁻/NO₃ in IR+AG group were significantly decreased (23.44±1.25 vs 14.66±1.66 and 140.40±4.34 vs 80.00±8.56, P<0.01) and NT staining was also decreased.

CONCLUSION: Intestinal IR increases NO and ONOOproduction in the lung, which may be involved in intestinal IR-mediated lung injury.

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INTRODUCTION

A devastating consequence of tissue reperfusion is the damage

in organs uninvolved in the initial ischemic insult^[1]. Multiple organ dysfunction syndrome (MODS), as it is known, is the leading cause of death in critically ill patients and is a documented consequence of gut reperfusion^[2-5]. Although systemic inflammatory characteristic of MODS can result in damage to any organ, onset of the syndrome is usually heralded by the development of respiratory insufficiency^[2-9]. The pathophysiology of lung injury associated with intestinal ischemia-reperfusion (IIR) probably involves a variety of inflammatory and vasoactive mediators. Recently, the production of large amounts of nitric oxide (NO), a free radical produced by the inducible isoform of NO synthase (iNOS) has been implicated as a cytotoxic factor in a variety of pathophysiological processes, including various forms of inflammation and circulatory shock^[10-14]. The cytotoxic effects of NO are in part, mediated by peroxynitrite anion (ONOO), a reactive oxidant species formed from NO and superoxide at an almost diffusion-controlled rate^[15-18]. The production of ONOO has been demonstrated in various lung injury^[19-21]. However, no evidence exists about its change and role in the lung injury following IIR. The major aim of the present study, therefore, was to determine the change and role of iNOS and peroxynitrite in lung injury induced by IIR.

MATERIALS AND METHODS

Animal model^[22]

Male healthy Sprague-Dawley rats (250-300 g) were anesthetized with intraperitoneal administration of sodium pentobarbital (40 mg· kg⁻¹) and secured in a supine position on a heated restraining board at 37 °C after being shaved. Following midline laparotomy, a microvascular clip was placed across the superior mesenteric artery (SMA) for 120 minutes. Removal of the clip allowed reperfusion for120 minutes. This degree of IIR is consistently associated with intestinal, hepatic and pulmonary injury with a 12-hour mortality rate of 100 %.

Experimental protocols

To determine the effect of iNOS induction on the peroxynitrite formation and the lung injury, aminoguanidine (AG), an inhibitor of iNOS^[23], was used. The rats were randomly divided into 3 groups (n=8 in each): Sham, IR and IR+AG. The surgical sham group underwent full surgical preparation including isolation of the SMA without occlusion. After that they were followed for the same aggregate period of time. The IR group received 2 h of ischemia and 2 h of reperfusion. The IR + AG group was given AG (10 mg· kg⁻¹ intravenously, Sigma, USA) 15 min before reperfusion. The animals in IR group and sham group were treated with equal volume of the vehicle (normal saline solution, NS; 1 ml· kg⁻¹). All animals were killed by exsanguination at designated time. Both lungs were harvested immediately for the following detection.

Lung histology

The right middle lung lobe was harvested and fixed in 10 % formalin. After embedded in paraffin, sections of 8 μ m were stained with hematoxylin and eosin for light microscopy.

Assessments of lung malondialdehyde (MDA) content

The right base of lung was harvested and immediately homogenized on ice in 9 volumes saline. The homogenates were centrifuged at 4 000 r· min⁻¹ at 4 $^{\circ}$ C for 10 min. The MDA content in the supernatants was measured using MDA assay kit (Nanjing Jiancheng Corp. China).

Lung nitrite/nitrate (NO₂⁻/NO₃⁻) detection

 NO_2^{-}/NO_3^{-} production, an indicator of NO synthesis, was measured in the lung homogenate with a NO assay kit (Nanjing Jiancheng Corp. China) following the manufacturer's instruction.

Western blotting analysis for iNOS

The left lung was homogenized with PBS (pH 7.2) and centrifuged at 4 $^{\circ}$ C, 18 000 r· min⁻¹ for 10 min. After precipitated unsolubilized fraction was discarded, the protein concentration in the supernatant was determined by Coomassie blue dyebinding assay (Nanjing Jiancheng Corp. China). Aliquots (30 µg) of protein from each sample were electrophoresed on a 120 g· L⁻¹ SDS-polyacrylamide gel for 4 hour at 100V. The protein samples were transferred onto a nitrocellulose membrane (Amersham, USA). The membrane was then probed with polyclonal rabbit anti-rat iNOS antibody (1:50 dilution, Santa Cruz Co., USA) for 2 hours at 37 °C. After 3 washes with TPBS, blots were visualized with the use of an amplified HRP kit (Wuhan Boshide, China). The presence of iNOS was indicated by the development of brown color.

Immunohistochemical analysis of NT

Tyrosine nitration, a specific "footprint" of peroxynitrite formation^[16], was detected in the left lung by immunohistochemical technique. The left lung was harvested and fixed in 10 % formalin. Sections of 8 µm thickness of lung tissue were treated with 0.3 % H_2O_2 in methanol to block endogenous peroxide activity. Immunohistochemical staining was performed using rabbit polyclonal antibody against NT (1:50 dilution, Cayman Chemical, USA) by indirect streptavidin/peroxidase technique (SP kit, Zymed Co., USA). Experiments were performed following the manufacturer's instruction. Paraffin-embedded sections were incubated with polyclonal anti-rat NT antibody for 12 hours at 4 °C after antigen retrieval. Biotinylated IgG was added as second antibody. Horseradish peroxidase labeled streptomycin-avidin complex was used to detect second antibody. Slides were stained with diaminnobenzidine, and examined under light microscope. The brown or dark brown stained cytoplasm was considered as positive.

Statistical analysis

Values were expressed as mean \pm SD ($\bar{x}\pm s$). Statistical analyses were performed using paired Student' s *t* test. *P*<0.05 was considered significant.

RESULTS

Pathological alternations of lung tissue

The histological structure of alveolar and mesenchymal cells was normal in the lungs of sham group, while the lung tissues from the IR group were significantly damaged, with pulmonary edema, hemorrhage and inflammatory cell infiltration. Administrating AG before reperfusion could attenuate significantly the lung injury as showed by light microscope (Figure 1).

Change of MDA content

The lung MDA content was increased significantly in IR group

when compared with the sham control (P<0.05). Compared with the IR group, the MDA content was decreased markedly in IR +AG group (P<0.05) (Table 1).

Table 1 Changes of MDA and NO⁻²/NO⁻³ contents in lung tissue after IIR with pretreatment of AG in rats (\bar{x} ±*s*, *n*=8)

Groups	MDA (nmol.ml ⁻¹)	$\mathrm{NO}^{-2}/\mathrm{NO}^{-3}$ (umol.L ⁻¹)
Sham	12.00±2.18	76.39±6.08
IR	$23.44{\pm}1.25^{a}$	$140.40{\pm}4.34^{a}$
IR+AG	$14.66{\pm}1.66^{\rm c}$	$80.00 \pm 8.56^{\circ}$

^aP<0.05 vs Sham group. ^cP<0.05 vs IR group.



Figure 1 Light microscopic observation on the lung after IIR with pretreatment of AG in rats (HE×400). A. The normal lung tissue structure was found in sham group. B. Lung edema, hemorrhage and inflammatory cells sequestration were found in the IR group. C. Decreased morphological changes induced by the intestinal IR were found in the IR+AG group.

Change of nitrite/nitrate

Compared with the sham group, the lung content of nitrite/ nitrate in IR group was increased significantly (P<0.05). Compared with the IR group, the content of nitrite/nitrate in IR+AG group was decreased significantly (P<0.05) (Table 1).

Change of iNOS expression

Western blotting showed that weak positive signal was found in the lung in sham group. In contrast, marked increase of iNOS protein expression was found in the IR group. There was still notable positive signal in the IR+AG group (Figure 2).



Figure 2 Western blotting analysis of iNOS in rat lung after IIR with pretreatment of AG in rats. 1. Sham; 2. IR; 3. IR+AG.

Change of NT formation

The formation of peroxynitrite in the lung sections was demonstrated by immunohistochemical staining with monoclonal anti-NT antibodies. In the sham group, no brown deposits were present in lung sections. In contrast, very strong staining was observed in lung tissue sections from the IR group. Compared with the IR group, the immunostaining for NT in the IR+AG group decreased significantly, indicating that inhibition of iNOS activity and decrease of NO production could reduce the peroxynitrite formation (Figure 3).



Figure 3 Immunohistochemical analysis of NT in the lung after IIR with pretreatment of AG in rats. SP stain ×400. A. No

positive signal was found in the lung in sham group. B. Intense positive NT staining was found in the IR group. C. Positive NT staining decreased in the IR+AG group.

DISCUSSION

The lung is one of the very important target organs in multiple organ dysfunction syndrome (MODS) or multiple system organ failure (MOSF) caused by severe injury^[1-9]. It has been found that in addition to the direct trauma, the lung could also be damaged by indirect injury such as gut, liver and skeletal muscle reperfusion, as well as aortic occulation-reperfusion and circulatory shock^[1-9]. The present results showed that 2 h of intestinal ischemia followed by 2 h of reperfusion induced lung injury, manifested as the histological evidence of lung edema, hemorrhage and PMN infiltration. Moreover, the IIR mediated lung injury was oxygen-dependent, as indicated by the increase in the levels of MDA, an established marker of oxidative stress.

The mechanism of respiratory failure after intestinal I/R is complex and poorly understood. Under the condition of an inadequate mucosal blood flow, the gut barrier function can be progressively impaired and invaded by bacteria or endogenous endotoxin^[24-26]. This process is associated with activation of systemic inflammatory mediators including bacteriotoxin, inflammatory mediators, such as tumor necrosis factor (TNF) and interleukin (IL) and immunocytokines^[26-31]. In addition, several recent observations implicate that NO may be an important participant in the pulmonary response to IIR^[32]. It has been suggested that nitric oxide (NO), produced from endothelial constitutive nitric oxide synthase (ecNOS), may be an important protective molecule at the onset of the IR of the small bowel. In this regard, inhibitors of endogenous NO production greatly exacerbate the increase in epithelial permeability and cardiovascular dysfunction in the reperfused post-ischemia intestine^[32-34], while administration of NO donors prevents the early rise in epithelial permeability and tissue dysfunction^[35,36]. Excess NO production has been attributed to a second NOS (induced NOS, iNOS) that is not present under normal condition but can be induced in response to systemic inflammatory states, including IR. The induction of iNOS has been implicated in the pathogenesis of IIR and it was reported that inhibition of iNOS activity and nitric oxide production could attenuate the intestinal ischemia/reperfusion injury^[37-40]. In the present experiment, we studied the contribution of iNOS to the IIR-induced lung injury. The results demonstrated that 2 h of intestinal ischemia followed by 2 h of reperfusion upregulated significantly the lung iNOS expression, accompanied by the elevation of pulmonary nitrate/nitrite (stable metabolites of NO) level. This is consistent with the findings of Virlos IT, et al^[34] who demonstrated that pulmonary iNOS activity in rats subjected to IIR was significantly increased. The effect of increased iNOS expression on IIR-induced lung injury was indirectly assessed by administration of AG, a selective inhibitor of iNOS. The results that in vivo treatment with AG reduced markedly the nitrate/nitrite levels and the lung injury demonstrated that the induction of iNOS and the excessive NO production in the lung following intestinal IR involved in the IIR-induced lung injury.

Though the cytotoxic actions of the excessive NO production may be involved in several potential mechanisms, recent studies suggested that peroxynitrite, may be a key mediator for cytotoxicity of excess NO generation and is involved in many pathological processes^[15-18]. The cytotoxic processes triggered by peroxynitrite include initiation of lipid peroxidation, inhibition of mitochondrial respiration, inhibition of membrane pumps, depletion of glutathione, and damage to DNA with subsequent activation of poly (ADP-ribose) synthetase and

concomitant cellular energy depletion^[19-21, 41, 42]. Peroxynitrite is now generally considered an oxidant more toxic than either NO or superoxide anion alone. Furthermore, it has been shown that a major product of the reaction of peroxynitrite with protein is the addition of a nitro group in the ortho position of tyrosine to form NT which could be detected by antibodies specific to protein NT^[16]. The present results of immunohistochemical staining with monoclonal anti-NT antibodies demonstrated that peroxynitrite formation occurred in the reperfusion phase of ischemic intestine. The results also suggested that the formation of peroxynitrite was correlated with iNOS expression and involved in the IIR-induced lung injury, because inhibiting iNOS activity with AG reduced significantly the NT immunoreactivity and attenuated the IIR-induced lung injury. The cytotoxic actions of iNOS-NO and peroxynitrite may be associated with their damages to the endothelial barrier, their oxidant to the membrane lipid and the triggering of the PMN sequestration in the lung, manifested as the detection of lung MDA content and the observation of the lung pathological changes.

In summary, the present results demonstrate that the systemic inflammatory response and lung injury occur following IIR with an increase of NO, iNOS expression and the formation of peroxynitrite in the lung. Inhibition of iNOS prevented the lung injury and the formation of NT, suggesting that excessive NO production and peroxynitrite formation are cytotoxic to the cell and tissue and are involved in the secondary lung injury induced by the IIR. These results may partially explain the mechanism of MOF and suggest that inhibiting peroxynitrite may be a novel pharmacological approach to prevent cell injury and MOF.

REFERENCES

- Carden DL, Granger DN. Pathophysiology of ischemiareperfusion injury. J Pathol 2000; 190: 255-266
- 2 **Rotstein OD**. Pathogenesis of multiple organ dysfunction syndrome: gut origin, protection, and decontamination. *Surg Infect (Larchmt)* 2000; **1**: 217-225
- 3 Fu XB, Yang YH, Sun TZ, Gu XM, Jiang LX, Sun X Q, Sheng ZY. Effect of intestinal ischemia-reperfusion on expression of endogenous basic fibroblast growth factor and transforming growth factor betain lung and its relation with lung repair. World J Gastroenterol 2000; 6: 353-355
- 4 **Wu XT**, Li JS, Zhao XF, Zhuang W, Feng XL. Modified techniques of heterotopic total small intestinal transplantation in rats. *World J Gastroenterol* 2002; **8**: 758-762
- 5 Mitsuoka H, Schmid-Schonbein GW. Mechanisms for blockade of in vivo activator production in the ischemic intestine and multiorgan failure. *Shock* 2000; 14: 522-527
- 6 Yassin MM, Harkin DW, Barros D' Sa AA, Halliday MI, Rowlands BJ. Lower limb ischemia-reperfusion injury triggers a systemic inflammatory response and multiple organ dysfunction. *World J Surg* 2002; 26: 115-121
- 7 Zhou JL, Zhu XG, Ling T, Zhang JQ, Chang JY. Effect of endogenous carbon monoxide on oxidant-mediated multiple organ injury following limb ischemia-reperfusion in rats. *Zhongguo Xiufu Chongjian Waie Zazhi* 2002; 16: 273-276
- 8 Rezende-Neto JB, Moore EE, Melo de Andrade MV, Teixeira MM, Lisboa FA, Arantes RM, de Souza DG, da Cunha-Melo JR. Systemic inflammatory response secondary to abdominal compartment syndrome: stage for multiple organ failure. *J Trauma* 2002; 53: 1121-1128
- 9 Weinbroum AA, Shapira I, Abraham RB, Szold A. Mannitol dosedependently attenuates lung reperfusion injury following liver ischemia reperfusion: a dose-response study in an isolated perfused double-organ model. Lung 2002; 180: 327-338
- 10 Tavaf-Motamen H, Miner TJ, Starnes BW, Shea-Donohue T. Nitric oxide mediates acute lung injury by modulation of inflammation. J Surg Res 1998; 78: 137-142
- 11 Wangle F, Patel M, Razavi HM, Weicker S, Joseph MG,

McCormack DG, Mehta S. Role of inducible nitric oxide synthase in pulmonary microvascular protein leak in murine sepsis. *Am J Respir Crit Care Med* 2002; **165**: 1634-1639

- 12 Li H, Forstermann U. Nitric oxide in the pathogenesis of vascular disease. J Pathol 2000; 190: 244-254
- 13 Hofseth LJ, Saito S, Hussain SP, Espey MG, Miranda KM, Araki Y, Jhappan C, Higashimoto Y, He P, Linke SP, Quezado MM, Zurer I, Rotter V, Wink DA, Appella E, Harris CC. Nitric oxide-induced cellular stress and p53 activation in chronic inflammation. *Proc Natl Acad Sci U S A* 2003; 100: 143-148
- 14 Qin JM, Zhang YD. Intestinal expressions of eNOSmRNA and iNOSmRNA in rats with acute liver failure. *World J Gastroenterol* 2001; 7: 652-656
- 15 Cuzzocrea S, Zingarelli B, Caputi AP. Role of constitutive nitric oxide synthase and peroxynitrite production in a rat model of splanchnic artery occlusion shock. *Life Sci* 1998; 63: 789-799
- 16 Pryor WA, Squadrito GL. The chemistry of peroxynitrite: a product from the reaction of nitric oxide with superoxide. *Am J Physiol* 1995; **268**: L699-L722
- 17 Szabo C. The pathophysiological role of peroxynitrite in shock, inflammation, and ischemia-reperfusion injury. *Shock* 1996; 6: 79-88
- 18 Zhao KS, Liu J, Yang GY, Jin C, Huang Q, Huang X. Peroxynitrite leads to arteriolar smooth muscle cell membrane hyperpolarization and low vasoreactivity in severe shock. *Clin Hemorheol Microcirc* 2000; 23: 259-267
- 19 Gu ZY, Ling Y, Cong B. Peroxynitrite mediated acute lung injury induced by lipopolysaccharides in rats. *Zhonghua Yixue Zazhi* 2000; 80: 58-61
- 20 Laskin DL, Fakhrzadeh L, Laskin JD. Nitric oxide and peroxynitrite in ozone-induced lung injury. *Adv Exp Med Biol* 2001; 500: 183-190
- 21 Gu ZY. Peroxynitrite-mediated pulmonary vascular injury induced by endotoxin and the protective role of cholecystokinin. *Shengli Kexue Jinzhan* 2001; 32: 135-137
- 22 **Turnage RH**, Kadesky KM, Bartula L, Myers SI. Intestinal reperfusion up regulates inducible nitric oxide synthase activity within the lung. *Surgery* 1995; **118**: 288-293
- 23 **Zhang GL**, Wang YH, Teng HL, Lin ZB. Effects of aminoguanidine on nitric oxide production induced by inflammatory cytokines and endotoxin in cultured rat hepatocytes. *World J Gastroenterol* 2001; **7**: 331-334
- 24 Turnage RH, Guice KS, Oldham KT. Endotoxemia and remote organ injury following intestinal reperfusion. J Surg Res 1994; 56: 571-578
- 25 Olanders K, Sun Z, Borjesson A, Dib M, Andersson E, Lasson A, Ohlsson T, Andersson R. The effect of intestinal ischemia and reperfusion injury on ICAM-1 expression, endothelial barrier function, neutrophil tissue influx, and protease inhibitor levels in rats. *Shock* 2002; **18**: 86-92
- 26 Yao Y, Yu Y, Chen J. The effect of intestinal ischemia/reperfusion on increased sensitivity to endotoxin and its potential mechanism. *Zhonghua Zhengxing Shaoshang Waike Zazhi* 1999; 15: 301-304
- 27 Ishii H, Ishibashi M, Takayama M, Nishida T, Yoshida M. The role of cytokine-induced neutrophil chemoattractant-1 in neutrophil-mediated remote lung injury after intestinal ischaemia/ reperfusion in rats. *Respirology* 2000; 5: 325-331
- 28 Yan GT, Hao XH, Xue H, Wang LH, Li YL, Shi LP. Interleukin-1beta expression and phospholipase A(2) activation after intestinal ischemia/reperfusion injury. *Shengli Xuebao* 2002; 54: 28-32
- 29 Souza DG, Cassali GD, Poole S, Teixeira MM. Effects of inhibition of PDE4 and TNF-alpha on local and remote injuries following ischaemia and reperfusion injury. *Br J Pharmacol* 2001; 134: 985-994
- 30 Wada K, Montalto MC, Stahl GL. Inhibition of complement C5 reduces local and remote organ injury after intestinal ischemia/ reperfusion in the rat. *Gastroenterology* 2001; 120: 126-133
- 31 Zhao H, Montalto MC, Pfeiffer KJ, Hao L, Stahl GL. Murine model of gastrointestinal ischemia associated with complement-dependent injury. J Appl Physiol 2002; 93: 338-345
- 32 Khanna A, Rossman JE, Fung HL, Caty MG. Attenuated nitric oxide synthase activity and protein expression accompany intestinal ischemia/reperfusion injury in rats. *Biochem Biophys Res Commun* 2000; 269: 160-164

33 Tavaf-Motamen H, Miner TJ, Starnes BW, Shea-Donohue T. Nitric oxide mediates acute lung injury by modulation of inflammation. J Surg Res 1998; 78: 137-142

34 Virlos IT, Inglott FS, Williamson RC, Mathie RT. Differential expression of pulmonary nitric oxide synthase isoforms after intestinal ischemia-reperfusion. *Hepatogastroenterol* 2003; 50: 31-36

- 35 Kalia N, Brown NJ, Hopkinson K, Stephenson TJ, Wood RF, Pockley AG. FK409 inhibits both local and remote organ damage after intestinal ischaemia. J Pathol 2002; 197: 595-602
- 36 Ward DT, Lawson SA, Gallagher CM, Conner WC, Shea-Donohue T. Sustained nitric oxide production via l-arginine administration ameliorates effects of intestinal ischemia-reperfusion. *J Surg Res* 2000; 89: 13-19
- 37 Turnage RH, Wright JK, Iglesias J, LaNoue JL, Nguyen H, Kim L, Myers S. Intestinal reperfusion-induced pulmonary edema is related to increased pulmonary inducible nitric oxide synthase activity. *Surgery* 1998; **124**: 457-462
- 38 Xia G, Lara-Marquez M, Luquette MH, Glenn S, Haque A, Besner GE. Heparin-binding EGF-like growth factor decreases inducible nitric oxide synthase and nitric oxide production after intestinal is-

chemia/reperfusion injury. Antioxid Redox Signal 2001; 3: 919-930

- 39 Takada K, Yamashita K, Sakurai-Yamashita Y, Shigematsu K, Hamada Y, Hioki K, Taniyama K. Participation of nitric oxide in the mucosal injury of rat intestine induced by ischemiareperfusion. J Pharmacol Exp Ther 1998; 287: 403-407
- 40 Suzuki Y, Deitch EA, Mishima S, Lu Q, Xu D. Inducible nitric oxide synthase gene knockout mice have increased resistance to gut injury and bacterial translocation after an intestinal ischemiareperfusion injury. Crit Care Med 2000; 28: 3692-3696
- 41 Jagtap P, Soriano FG, Virag L, Liaudet L, Mabley J, Szabo E, Hasko G, Marton A, Lorigados CB, Gallyas F Jr, Sumegi B, Hoyt DG, Baloglu E, VanDuzer J, Salzman AL, Southan GJ, Szabo C. Novel phenanthridinone inhibitors of poly (adenosine 5'-diphosphate-ribose) synthetase: potent cytoprotective and antishock agents. *Crit Care Med* 2002; **30**: 1071-1082
- 42 Cuzzocrea S, McDonald MC, Mazzon E, Dugo L, Serraino I, Threadgill M, Caputi AP, Thiemermann C. Effects of 5aminoisoquinolinone, a water-soluble, potent inhibitor of the activity of poly (ADP-ribose) polymerase, in a rodent model of lung injury. *Biochem Pharmacol* 2002; 63: 293-304

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