

• CLINICAL RESEARCH •

Intra-tumor injection of H101, a recombinant adenovirus, in combination with chemotherapy in patients with advanced cancers: A pilot phase II clinical trial

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Abstract

AIM: H101, an E1B 55 kD gene deleted adenovirus, has been shown to possess oncolysis activity experimentally and proved to be safe in preliminary phase I study. The current study was designed to evaluate its anti-tumor activity and toxicity in combination with chemotherapy in patients with late stage cancers.

METHODS: H101 5.0×10^{11} virus particles were given by intra-tumor injection daily for five consecutive days at every three-week cycle, combined with routine chemotherapy, to one of the tumor lesions of 50 patients with different malignant tumors. Tumor lesions without H101 injection in the same individuals were used as controls. The efficacy and toxicity were recorded.

RESULTS: Forty-six patients were evaluable with a 30.4% response rate. H101 injection in combination with chemotherapy induced three complete response (CR) and 11 partial response (PR), giving an overall response rate of 28.0% (14/50) among intention-to-treat patients. The response rate for the control lesions was 13.0%, including one case with CR and five cases with PR, which was significantly lower than that for the injected lesions ($P < 0.05$). Main side effects were fever (30.2%) and pain at the injected sites (26.9%). Grade 1 hepatic dysfunction was found in four patients, grade 2 in one patient, and grade 4 in one patient. Hematological toxicity (grade 4) was found in four patients.

CONCLUSION: Intra-tumor injection of the genetically engineered adenovirus H101 exhibits potential anti-tumor activity to refractory malignant tumors in combination with chemotherapy. Low toxicity and good tolerance of patients to H101 were observed.

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INTRODUCTION

The fights against tumors are far from being finished. Biotherapy seems to be a potential anticancer weapon, but still needs strengthening. Engineered virus against cancer is one of the most hopeful therapeutic approaches. There are two different methods: (1) to use replication incompetent viruses as delivery agents for therapeutic genes to access to tumors, and (2) to destroy tumor by using replication-selective oncolytic viruses as therapeutic agents themselves^[1,2]. Multiple gene dysfunctions taking part in tumor formation have been known, single gene correction or modification can hardly reverse the malignancy. Viruses engineered for the purpose to replicate only in tumor cells and destroy the cells do not depend on the gene function they take on and have been shown to have great efficacy in both experimental and clinical studies^[3-5].

H101 is a recombinant human type-5 adenovirus (Ad5), in which E1B-55 kDs gene has been totally deleted. The H101 virus produced by Shanghai Sunway Biotech, also contains a deletion of 78.3-85.8 μ m gene segment in the E3 region. The E1B-55kD gene product is responsible for p53-binding and inactivation^[6]. If deleted, the virus would be unable to inactivate p53 for efficient replication in normal cells. However, cancer cells lacking functional p53 would hypothetically be sensitive to viral replication and subsequent cytopathic effects. p53 mutation is the most common genetic abnormality identified in human cancer^[7]. This characteristic can be utilized for H101 to identify the target. *In vitro* and *in vivo* studies have shown that H101 has anticancer activity, and has been proved to be safe through a five dosage of 5.0×10^7 - 1.5×10^{12} virus particles (VP)/d within 5 consecutive days in a clinical trial^[8]. We carried out this clinical trial to evaluate anti-tumor activity of H101 and its toxicity in combination with chemotherapy in patients with late stage cancers.

MATERIALS AND METHODS

Enrollment criteria

Histologically confirmed late stage cancer patients with more than two measurable lesions (at least one could be injected with H101), who had recurrent disease after surgery and/or radiotherapy for the primary tumor, or had progressed at or within 8 wk after completion of chemotherapy and/or radiotherapy, were recruited. Patients had to be ≥ 18 years old, with performance status above grade 2 according to The Eastern Cooperative Oncology Group (ECOG) standard, and life expectancy of ≥ 3 mo. Normal hematological and renal functions were also required. An informed consent was obtained from each patient or from the patient's legal guardian prior to enrollment. The p53 gene status was not critical for enrollment, because there were factors that inhibited p53 protein function including expression of the human papilloma virus E6 protein or *mdm-2* gene amplification^[9]. Institutional Review Board approval of the protocol and consent form were granted. This study was also approved by the State Food and Drug Administration of China.

Baseline assessment

Baseline assessments were made prior to treatment, but these

results were not used as enrollment criteria. Baseline blood tests such as complete blood counts, neutralizing antibody titers, electrolytes, blood urea nitrogen, creatinine, and liver function tests were performed. In addition, plain chest radiography, electrocardiogram and type B ultrasonography of upper abdomen were performed.

H101

H101 was formulated as a sterile viral solution in PBS buffer and kept at -20°C . Each vial contained 0.5 mL of virus solution with 5×10^{11} VP and titered $<1:60$ TCID₅₀. Sterile purified lots of virus were produced for human clinical use by Shanghai Sunway Biotech (Shanghai, China), and tested for the titer, sterility, and general safety by National Institute For the Control of Pharmaceutical and Biological Products (Beijing, China).

Treatment regimen

In each patient, the most symptomatic and/or largest tumor mass was injected with H101, and the patient was treated together with routine systemic chemotherapy simultaneously. The tumor for injection was mapped into five equally spaced sections. Local anesthesia was applied to the skin as needed. The tumor was injected with 5×10^{11} virus particles into one section per day for 5 consecutive days, and these injections were repeated every 3 wk as one treatment cycle. The suspension volume of saline used for H101 administration was normalized to 30% of the estimated volume of the tumor mass to be injected. Tumor volume was estimated as: $1/2$ (maximal transverse diameter \times maximal vertical diameter \times depth).

Tumor assessments and toxicity evaluation

Tumor masses were measured serially by either physical examination or radiographic scanning (computed tomography or magnetic resonance imaging), whichever the principal investigator deemed most accurate for the measurement of the injected tumor mass. In general, superficial lesions were measured by physical examination, and deep tumors were measured most accurately by radiographic scanning. The tumor mass injected with H101 (injected lesion) and non-injected lesion were evaluated independently. Tumor measurements were performed either every 3 wk (physical examination) while patients were on active study treatment. After treatment completion, patients' tumor (s) were assessed every 4 wk or sooner if signs/symptoms of progression became evident. Radiographic scanning was assessed by independent radiologists, who were not investigators on the study. The degree of response within injected tumors was categorized as follows: complete regression (CR), complete disappearance of measurable tumor; partial regression (PR), $\geq 50\%$ but $<100\%$ decrease in cross-sectional tumor area; minor response (MR), $<50\%$ but $\geq 25\%$ decrease in tumor area; stable disease (SD), $<25\%$ decrease or 25% increase in tumor area; and progressive disease (PD), $\geq 25\%$ increase in tumor area *versus* the baseline area. Toxicity was assessed using the National Cancer Institute Toxicity Criteria.

Additional follow-up after treatment initiation

Neutralizing antibody titers were repeated at the end of each cycle, and viral dissemination in blood was tested immediately after injection on d 5 and d 22 for each cycle. The routine blood tests were repeated every week. Fine-needle aspirate biopsies at the injected sites on day 22 of the first treatment cycle were optional, based on patients' consent because of ethical considerations. These biopsies were analyzed for type Ad5 coat protein by immunohistochemistry.

PCR detection of H101 viral genomes in plasma

The blood taken before and one day after injection were collected

for PCR detection of H101 genomes (the amplicon overlaps the E1B region deletion and does not detect wild-type adenovirus sequences). The left primer was 5'ctggcgcagaagtattccat3', at Tm 60.24 $^{\circ}\text{C}$ and the right primer was 5'gtcacatccagcatcacagg3', at Tm 60.12 $^{\circ}\text{C}$. Viral DNA was extracted from samples, using the Sangon DNA mini kit (Shanghai, China). The amplification procedure was: at 94 $^{\circ}\text{C}$ for 10 min, then 94 $^{\circ}\text{C}$ for 60 s, 55 $^{\circ}\text{C}$ for 45 s, 72 $^{\circ}\text{C}$ for 60 s for 35 cycles; then at 72 $^{\circ}\text{C}$, for 10 min. The products were analyzed by 10 g/L agarose electrophoresis. The lower limit of detection was 100 particles of H101 per microlitre plasma.

Detection of Ad-specific neutralizing antibodies by ELISA

Triplicate plasma (5 μL) taken before and on d 22 after injection were collected, and tested for Ad-specific antibodies according to the procedures provided by Jingmei Biotech (Shenzhen, China). The absorbance at 450 nm was read on a Bio-Rad Model 550 microplate reader. The positive results were those above or equal to the average of A_{450nm} negative control plus 0.10. Otherwise, the samples were defined as negative.

Immunohistochemistry for Ad5

Injection site fine-needle aspiration biopsies were formalin-fixed, paraffin-embedded and cut into sections. Sections were then deparaffinized and hydrated. Slides were subjected to antigen retrieval at 120 $^{\circ}\text{C}$ for 10 min in citrate buffer and incubated with an Ad5 monoclonal antibody (NeoMarker, America) for 90 min at room temperature. This was followed by incubation with a biotinylated goat anti-mouse secondary antibody, and the streptavidin/horseradish peroxidase conjugate, then mounted in DPX mounting medium (BDH Chemicals, America). The percentage of brown-stained cells (positive for Ad5) was determined by counting the cells under high-power magnification ($\times 40$) of microscope. The average percentage of three high-power field assessments was then calculated. Tumors that had greater than 10% of positively stained cells were considered to be Ad5 positive.

Statistical analysis

All patients enrolled were calculated under the ITT principle. The rates were compared by χ^2 test.

RESULTS

Patient characteristics

Totally, 50 patients were enrolled, including 18 with head and neck cancer, eight esophageal cancer, five gastric cancer, five lung cancer, three colorectal cancer, three breast cancer, three soft tissue sarcoma, two malignant melanoma, one ovarian cancer, one lymphoma and one chordoma. Most cancers were at end-stage. The head and neck cancer and esophageal cancer enrolled were all squamous carcinoma. Seventy percent of patients were males. The median age was 52 years. All patients had ECOG Performance Status of grade 0-2. Thirty-nine (78%) patients had received pretreatment before, and 31 (62%) had received more than two kinds of treatment. The tumor mass had a median cross-sectional area of 12.5 cm² (range, 1.43-360 cm²) (Table 1).

Tumor response

Overall, 46 patients were evaluable. The response rate (CR+PR) among these patients was 30.4% (14/46), and the overall response rate according to ITT principle was 28.0%. For the control lesions, the response rate was 13.0%, which was significantly lower than the H101 treated lesions ($\chi^2 = 4.08$, $P < 0.05$) (Table 2). In the 14 cases with effective H101 injection, there were one CR, three PRs, two MRs, three SDs, and five PDs for

the control lesions, respectively. In these patients, combination of H101 injection with chemotherapy was more effective than chemotherapy alone ($\chi^2 = 15.6, P < 0.001$). The response rates to H101 injection combined with chemotherapy were different, no effect for gastric cancer was found in this study (Table 3). Figure 1 shows regression of the injected lesion in a patient with head and neck cancer.

Table 1 Patients' demographics

Characteristic	
Age (yr)	
Median	52
Range	18-76
Sex	
Male (%)	35 (70%)
Female (%)	15 (30%)
ECOG Performance Status	
Grade 0	15 (30%)
Grade 1	21 (42%)
Grade 2	14 (28%)
Pretreatment	
Total	39 (78%)
Surgical	24 (48%)
Chemotherapy	37 (74%)
Radiotherapy	20 (40%)
Biotherapy	8 (16%)
Two or more treatment	31 (62%)
Tumor size (cm ²)	
Median	12.5
Range	1.43-360

Table 2 Response of H101 injected lesion and control lesion

Lesion	n	Median area (cm ²)	Efficacy					Response rate (%)
			CR	PR	MR	SD	PD	
H101 injection	46	12.5	3	11	11	13	8	30.4
Control	46	11.3	1	5	7	21	12	13.0

CR, complete regression; PR, partial regression; MR minor response; SD, stable disease; and PD, progressive disease. Response rate was calculated from cases with CR and PR over cases in each group.

Table 3 Efficacy of 46 evaluable patients treated with H101 and chemotherapy

Type of tumor	n	Response (CR+PR)
SCCHN ¹	15	4
Esophageal cancer	8	3
Gastric cancer	5	0/
Lung cancer	4	1
Colorectal cancer	3	1
Breast cancer	3	1
Soft tissue sarcoma	3	1
Malignant melanoma	2	1
Lymphoma	1	1
Chordoma	1	1
Ovarian cancer	1	0

¹SCCHN, squamous cell carcinoma of head and neck.

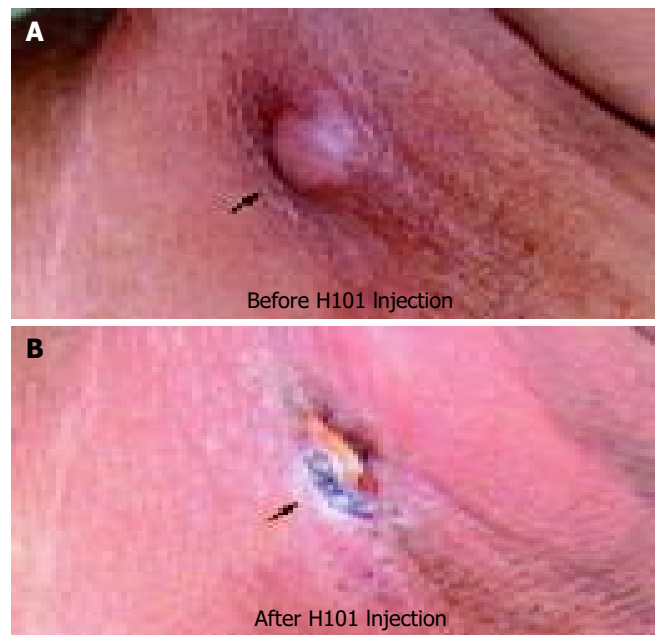


Figure 1 A 51 yr-old male patient with neck metastasis of soft palate squamous carcinoma. About 3 mo before enrollment, he had been treated with radiotherapy and Docetaxel plus DDP, but the metastatic tumor did not show any reaction to the treatment. Before the enrollment the tumor was about 2.2×1.5 cm² (A, arrow), then the tumor was injected with H101 5×10¹¹VP per day for 5 consecutive days with systemic administration of 5-Fu and DDP, after one cycle treatment the tumor regressed (B, arrow).

Adverse reaction

The most frequent adverse reaction was fever (30.2%), injection site pain (26.9%), flu-like symptoms (26.4%), nausea and vomiting (34.0%), leucopenia (49.1%), liver dysfunction (5.7%), alopecia (13.2%) (Table 4). Fever was moderate, which appeared at about 12 h post H101 injection, persisted for 2-4 h, and then returned to normal without treatment. There was a significant difference in the regression rate between patients with fever (69.2%, 9/13) and those without fever (21.2%, 7/33) ($\chi^2 = 9.48, P < 0.005$).

Table 4 Treatment-related toxicity

Adverse event	Grade				Total (%)
	I	II	III	IV	
Fever	10	5	1	0	16 (30.2)
Injection site pain	12	2	0	0	14 (26.4)
Nausea and vomiting	13	5	0	0	18 (34.0)
Leucopenia	12	7	3	4	26 (49.1)
Liver dysfunction	2	0	0	1	3 (5.7)
Flu-like symptom	13	2	0	0	15 (28.3)
Alopecia	3	3	1	0	7 (13.2)

Humoral immune response and plasma H101 viral genome

Fourteen patients were tested for the Ad-specific neutralizing antibody. Three (21.4%) of them were positive at baseline. Another six turned to be positive on day 22. Two patients positive at the baseline and two negative patients experienced tumor regression, and thus there was no correlation between baseline neutralizing antibody titers and induction of tumor response. Sixteen patients were tested for plasma H101 viral genome before injection and 30 min after. Only six cases were positive after injection (Table 5). All these patients were positive for blood Ad-specific neutralizing antibody on d 22.

Table 5 Humoral immune response and plasma H101 viral genome test

	Before injection		After injection	
	Negative	Positive	Negative	Positive
Ad neutralizing titer	11	3	5	9
Plasma H101 PCR	11	0	7	4

H101 immunohistochemistry detection

Totally, three fine-needle aspiration biopsies of tumor were obtained at the end of treatment on d 22 or d 44, and detected for Ad5 coat protein by immunohistochemistry for adenovirus presence. Two of them were positive (Figure 2).

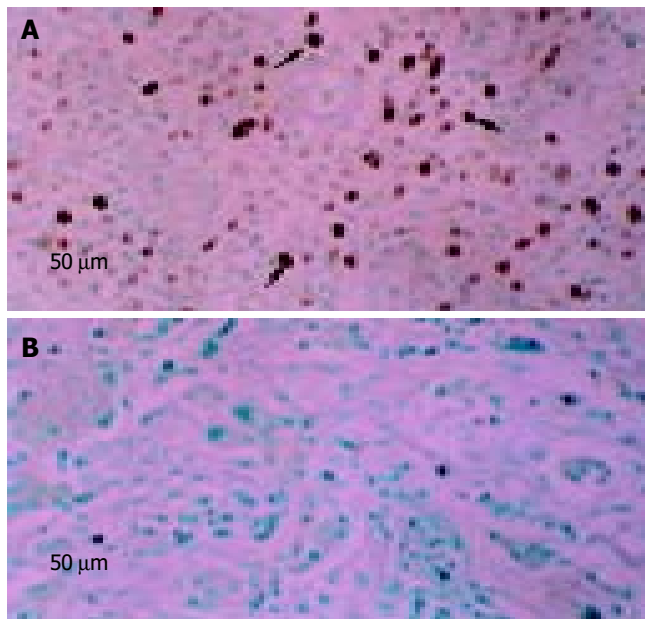


Figure 2 Immunohistochemical staining of Ad5 capsid from the fine-needle aspiration of the tumor tissue injected with H101 at the end of treatment cycle. The dark-brown stained granules (arrows in Figure A) represent a positive staining indicating adenovirus replication and package in the tumor cells. However negative staining was also obtained in one of the three samples (B).

DISCUSSION

Selective replication of E1B deleted adenovirus in the p53 dysfunctional human cancer for cancer therapy is one of promising treatment approaches. Its safety has been shown in a number of clinical trials^[5,10-12]. Although anticancer activity of the virus has been proved, the clinical efficacy is still not predominant. Therefore, the oncolytic ability needs to be enhanced. Current studies are focusing on arming these viruses with therapeutic genes to increase its potency.^[13-15]

But before that, the virus itself can be reinforced by augmentation or elimination of specific viral functions to enhance the anticancer efficacy. To enhance the virus-induced host anti-tumor immune response is one of the key points. However, the roles of the immune response to virotherapy are profound. Cutting down the functions of the virus to escape from immune surveillance can impede the spread of viral infection on the one hand, but augment tumor cell destruction through the recruitment of T cells “vaccinated” against tumor antigens on the other^[16]. The E3 region is related to the inhibition of host immunity, which enhances the virus replication and spread in tumor^[17]. But this is not necessary for intra-tumor injection of oncolytic viral. The virus replication and spread effect can be enhanced by

repeated injection. By sacrificing the spread ability, the virus may activate the host immune response to virus infected tumor cells and help the host immune system to recognize tumor cells themselves, and thus may benefit patients under such therapy. Metastasis is prevalent in malignant tumor patients, which is the main cause of treatment failure or even death. Moreover, patients may have more than one tumor lesion, and the lesion that cannot be injected could exist. Therefore, the ability of activating the host immune response seems crucial. So treatment with the E3 region deleted adenovirus, H101, may have additional benefit to patients.

The main purpose of this pilot study was to test the effect of H101 on a wide type of advanced cancers. Results showed that the total response rate was only 28.0% under the ITT principle, which was significantly higher than the lesions that received chemotherapy alone ($P < 0.05$). This indicates that H101 may have potential anticancer activity. The total regression rate observed is not salient for the treatment. This may be due to the late stage of the diseases, and most of the patients had been vigorously treated previously but failed at last. The other reason is the wide enrollment of the tumor types, some of which might not be sensitive to H101. For instance, gastric cancer showed no response.

However, some patients presented notable therapeutic efficacy without grievous adverse reactions. Moreover, in those who had fever during H101 injection, the efficacy was significantly higher than those who did not have fever ($P < 0.005$). Although there is not enough evidence to estimate the effect of H101 on host immunity to tumors, our results suggest that there is a relationship between the immune reaction to H101 and the efficacy, which was not well recognized in previous studies. In the beginning of last century, it was noticed that patients with various malignancies experienced spontaneous tumor regression after rabies vaccination, a viral illness or even bacterial infection^[18,19]. In these cases, virus infection may activate the host immune system, and elevated cell-mediated immunity may play a role in the tumor regression. But the mechanism is still unclear. On the basis of those results, immune modulation strategies should be further studied and developed.

Our study also shows that H101 intra-tumor injection is well tolerated. No severe toxicity was observed, and the main adverse reactions that related to H101 were injection site pain, nausea, fever and flu-like symptoms. Fever and flu-like symptoms were obviously caused by the virus injection and consequently transitory viraemia. H101 presence did not cause severe inflammation in peritumoral normal tissue, despite multiple directive injection. Thus, H101 may benefit the patient without adding severe affliction in clinical application.

Treatment for cancers with the recombinant oncolytic adenovirus is hopeful, but still immature. Experiences should be accumulated before it is applied in cancer therapy. Since patients enrolled in our clinical trial were in their end-stage of diseases, there were difficulties in patient selection and unifying the chemotherapy drugs due to ethical consideration, and immunosuppression was prevalent in those patients. The clinical benefit of intra-tumor injection with H101 should be further determined in randomized trials and, possibly, in earlier stage patients. The dosage, medication methods, treatment cycle and combined chemotherapy or immunotherapy should be explored in further studies as well. Genetically engineered and reinforced viruses may become a novel therapeutic platform for the treatment of cancers.

REFERENCES

- 1 Roth J, Cristiano RJ. Gene therapy for cancer: what have we done and where are we going? *J Natl Cancer Inst* 1997; **89**: 21-39

- 2 **Kirn D**, Martuza RL, Zwiebel J. Replication-selective virotherapy for cancer: biological principles, risk management and future directions. *Nat Med* 2001; **7**: 781-787
- 3 **Makower D**, Rozenblit A, Kaufman H, Edelman M, Lane ME, Zwiebel J, Haynes H, Wadler S. Phase II clinical trial of intralesional administration of the oncolytic adenovirus ONYX-015 in patients with hepatobiliary tumors with correlative p53 studies. *Clin Cancer Res* 2003; **9**: 693-702
- 4 **Habib NA**, Mitry RR, Sarraf CE, Jiao LR, Havlik R, Nicholls J, Jensen SL. Assessment of growth inhibition and morphological changes in *in vitro* and *in vivo* hepatocellular carcinoma models post treatment with dl1520 adenovirus. *Cancer Gene Ther* 2002; **9**: 414-420
- 5 **Hamid O**, Varterasian ML, Wadler S, Hecht JR, Benson A 3rd, Galanis E, Uprichard M, Omer C, Bycott P, Hackman RC, Shields AF. Phase II trial of intravenous CI-1042 in patients with metastatic colorectal cancer. *J Clin Oncol* 2003; **21**: 1498-1504
- 6 **Dobner T**, Horikoshi N, Rubenwolf S, Shenk T. Blockage by adenovirus E4orf6 of transcriptional activation by the p53 tumor suppressor. *Science* 1996; **272**: 1470-1473
- 7 **Hollstein M**, Sidransky D, Vogelstein B, Harris CC. p53 mutations in human cancer. *Science* 1991; **253**: 49-53
- 8 **Yuan ZY**, Zhang L, Li S, Qian XZ, Guan ZZ. Safety of an E1B deleted adenovirus administered intratumorally to patients with cancer. *Aizheng* 2003; **22**: 310-313
- 9 **Leach FS**, Tokino T, Meltzer P, Burrell M, Oliner JD, Smith S, Hill DE, Sidransky D, Kinzler KW, Vogelstein B. p53 mutation and MDM2 amplification in human soft tissue sarcomas. *Cancer Res* 1993; **53**: 2231-2234
- 10 **Kirn D**. Oncolytic virotherapy for cancer with the adenovirus dl1520 (Onyx-015): results of phase I and II trials. *Expert Opin Biol Ther* 2001; **1**: 525-538
- 11 **Nemunaitis J**, Khuri F, Ganly I, Arseneau J, Posner M, Vokes E, Kuhn J, McCarty T, Landers S, Blackburn A, Romel L, Randlev B, Kaye S, Kirn D. Phase II trial of intratumoral administration of ONYX-015, a replication-selective adenovirus, in patients with refractory head and neck cancer. *J Clin Oncol* 2001; **19**: 289-298
- 12 **Khuri FR**, Nemunaitis J, Ganly I, Arseneau J, Tannock IF, Romel L, Gore M, Ironside J, MacDougall RH, Heise C, Randlev B, Gillenwater AM, Bruso P, Kaye SB, Hong WK, Kirn DH. A controlled trial of intratumoral ONYX-015, a selectively-replicating adenovirus, in combination with cisplatin and 5-fluorouracil in patients with recurrent head and neck cancer. *Nat Med* 2000; **6**: 879-885
- 13 **Hermiston TW**, Kuhn I. Armed therapeutic viruses: strategies and challenges to arming oncolytic viruses with therapeutic genes. *Cancer Gene Ther* 2002; **9**: 1022-1035
- 14 **Stubdal H**, Perin N, Lemmon M, Holman P, Bauzon M, Potter PM, Danks MK, Fattaey A, Dubensky T, Johnson L. A prodrug strategy using ONYX-015-based replicating adenoviruses to deliver rabbit carboxylesterase to tumor cells for conversion of CPT-11 to SN-38. *Cancer Res* 2003; **63**: 6900-6908
- 15 **Bauzon M**, Castro D, Karr M, Hawkins LK, Hermiston TW. Multigene expression from a replicating adenovirus using native viral promoters. *Mol Ther* 2003; **7**: 526-534
- 16 **John TM**, Kenneth KT. Viral Oncolysis. *The Oncologist* 2002; **7**: 106-119
- 17 **Benedict CA**, Norris PS, Prigozy TI, Bodmer JL, Mahr JA, Garnett CT, Martinon F, Tschopp J, Gooding LR, Ware CF. Three adenovirus E3 proteins cooperate to evade apoptosis by tumor necrosis factor-related apoptosis-inducing ligand receptor-1 and -2. *J Biol Chem* 2001; **276**: 3270-3278
- 18 **Chabalgoity JA**, Dougan G, Mastroeni P. Live bacteria as the basis for immunotherapies against cancer. *Expert Rev Vaccines* 2002; **1**: 495-505
- 19 **Lamon EW**, Hale P, Whitten HD. Antibody-dependent, cell-mediated cytotoxicity with autochthonous lymphocytes and sera after infection with moloney sarcoma virus. *J Natl Cancer Inst* 1976; **56**: 349-355

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